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Lewis Acid-Catalyzed Nucleophilic Addition of Indoles to in situ Generated 2-Amidoallyl Cations

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ABSTRACT: We report herein the first Lewis acid-catalyzed generation of 2-amidoallyl cations through ring-opening of 4-benzylidene-2-oxazolines with $Sc(OTf)_3$. Upon nucleophilic addition of indoles indolylenamides were obtained with yields of 60 to 99% and excellent (*Z*)-selectivity. In addition, the novel strategy was also successfully applied to pyrroles and naphtholes as π -nucleophiles. A Brønsted acid-catalyzed process using TfOH formed in situ was ruled out by control experiments.

Oxyallyl cations have emerged as highly reactive intermediates in C-C-bond forming processes due to their unique electrophilic character. These species are known to undergo $(4+3)^1$ - and (3+2)-cycloadditions² easily and therefore many synthetic applications have been developed for the construction of complex molecules. More recently, the nucleophilic capture of oxyallyl cations has become a versatile method in synthesizing highly functionalized ketones which are otherwise not accessible by classical strategies.^{3–10} For example, Kartika et al. demonstrated the first Brønsted-acid catalyzed, regioselective addition of π - and σ -nucleophiles to introduce α -quaternary centers in ketone-derived compounds.³ Moreover, the group of MacMillan has very recently developed the first enantioselective, nucleophilic addition of *N*-protected indoles to oxyallyl cations generated in situ using a Jørgensen-Hayashi type catalyst.¹⁰

Despite of their enormous potential as templates in organic synthesis, 2-amino- and 2-amidoallyl cations are less well-studied because methods for their generation are limited. Typical procedures comprise ionization of rather unstable α -chloroenamines or α -chloroimines with stoichiometric amounts of silver(I) salts as Lewis acids followed by (4+3)-cycloaddition with furan or dienes. Shipman and co-workers reported a BF₃·OEt₂-promoted or Sc(OTf)₃-catalyzed intramolecular (4+3) cycloaddition of 2-aminoallyl cations which are formed in situ by the release of ring strain from 2-methyleneaziridines. Blakey and Stoll developed a Rh-catalyzed allene amination which affords cyclopropaneimines via 2-amidoallyl cation species. These intermediates were also described in (4+3)-, Shipman and (3+2)-cycloadditions as well as addition reactions with nucleophiles.

We envisioned 4-alkylidene substituted 2-oxazolines as suitable precursors in the generation of 2-amidoallyl cations. Although 2-oxazolines are particularly known to undergo cationic polymerization reactions by Lewis acid-catalyzed ring opening,¹⁸ the addition of π -nucleophiles to trap the resulting cations has never been achieved in an effective and catalytic manner.¹⁹ Hence, we decided to install the 4-alkylidene group to stabilize the cation

more efficiently by resonance effects which would allow the attack of a nucleophile under mild conditions. We now report our results on the first Lewis-acid catalyzed nucleophilic capture of in situ generated 2-amidoallyl cations with indole.

Commercially available α -acetamidocinnamic acid **1** was converted into 4-benzylidene-2-oxazoline **2** in a 5-step prodecure. Esterification of the carboxylic acid and reduction with LiAlH₄ afforded the allyl alcohol as a single double bond isomer. Selective oxidation was achieved with high yields by treating with MnO₂. The addition of phenyl magnesium bromide delivered the secondary alcohol which was then dehydrated through methanesulfonyl chloride and NEt₃ to give rise to oxazoline **2**. The resulting (*Z*)- and (*E*)-isomers of **2** were separated by column chromatography and their relative configuration was determined by NOE-experiments (Scheme 1).

Scheme 1. Synthesis of Oxazoline 2.

We initially started our investigations with the Brønsted-acid catalyzed activation of (*Z*)-2 with diphenylphosphate (DPP) followed by the reaction with indole. It turned out that conventional solvents (CH₂Cl₂, CH₃CN, THF, toluene) failed to yield the desired addition product, even under refluxing conditions. Based upon procedures in which oxyallyl cations were generally formed in protic solvents,⁵⁻⁹ we decided to employ trifluoroethanol (TFE), which successfully gave rise to the formation of indolylenamide 3a with 91% yield and high (*Z*)-selectivity (Scheme 2, eq. 1). The (*Z*)-configuration of the enamide double bond, which

was rigorously determined by X-ray crystal structure analysis (see Supporting Information, Figure S1), is favored to minimize 1,3-allylic strain. No reaction was observed without DPP in TFE.

In addition to Brønsted acid-catalysis, we decided to study the ring-opening of oxazoline 2 with Lewis acids in non-protic organic solvents mainly for two reasons: 1) During our investigations we occasionally observed the addition of TFE to the intermediate 2-amidoallyl cations in competition to other nucleophiles, a side reaction that is also described in the nucleophilic capture of oxyallyl cations.^{5,9} 2) We wanted to exclude the potential hydrolysis of the enamide moiety in the presence of the Brønsted acid and the protic solvent. After extensive optimization studies (see Supporting Information, Table S1) we eventually settled on 5 mol% of Sc(OTf)₃ providing 3a in 95% yield for the subsequent investigations (Scheme 2, eq. 2).

Scheme 2. Synthesis of 3a with DPP and Sc(OTf)₃ under optimized conditions.

Metal triflates and other Lewis acids are known to release traces of protons that might act as the active catalyst. ^{20,21} We carried out some control experiments to exclude this type of "hidden Brønsted acid" catalysis in the Sc(OTf)₃-catalyzed process (see Supporting Information, Table S2). In the presence of 2,6-di-*tert*-butylpyridine (DTBP), which is known to be a non-coordinating and selective proton scavenger, ^{20,22} no difference in reaction time and yield was obtained in comparison to the result without the base. Moreover, we performed the

same reaction with 10 mol% TfOH, which gave rise to **3a** in 87% yield within one day. Addition of DTBP to the TfOH-catalyzed reaction significantly decreased the conversion of starting material and furnished the product with less than 10% yield after 3 days. These experiments strongly indicated that Sc(OTf)₃ itself was the active catalyst but traces of TfOH generated in situ can also assist in this transformation.

We next studied the influence of different substituted indoles on the nucleophilic attack to oxazolines (Z)- and (E)-2. It turned out that electron-donating as well as electron-withdrawing groups were well tolerated, leading to the corresponding products with good to excellent yields and high (Z)-selectivities (Table 1). Reactions that started from the (E)-isomer of oxazoline 2 afforded the products 3 within generally longer reactions times but with similar yields as indicated by comparing entries 1 and 2. The lower reactivity of (E)-2 likely resulted from the enhanced 1,3-allylic strain between the two phenyl groups. This led to double bond isomerization into the more reactive (Z)-configured oxazoline 2, which was observed by TLC monitoring of the reaction mixtures. Indoles with electron-donating substituents typically furnished the products with very high yields and in most cases exclusively as (Z)-isomers (entries 3-10). N-Methylated as well as 2-methyl indole were suitable nucleophiles but required longer reaction times (entries 3-4). However, placing a phenyl group at the 2-position of indole provided the corresponding product in quantitative yield within one day (entry 10). C-4 Substituted indoles showed slower conversion of starting material and a slight decrease of product yield because of steric hindrance to the nucleophilic center (entries 9 and 11). The 4bromoindole substituted product 3j was the only case in which a significantly lower Z/E ratio was observed (entry 11). Other functional groups like halides (entries 12-13) or esters (entries 14-15) also delivered the corresponding products **3k-3n** with excellent yields. Indoles carrying strongly electron-withdrawing cyano or nitro groups showed a significant decrease in reactivity with oxazoline (Z)-2. After column chromatography, 30 and 3p were isolated in 62% and 60% yield, respectively, and high (Z)-selectivities (entries 16-17).

Table 1. Substrate Scope with Indoles toward Indolylenamides 3. [a]

entry	product	2	R	time [h]	yield [%] ^[b]	$Z/E^{[\mathfrak{c}]}$
1	3a	Z	Н	15	95	>95:5
2	3a	E	Н	39	91	>95:5
3	3 b	E	1-CH ₃	96	82	>95:5
4	3c	E	2-CH ₃	$96^{[d,e]}$	72	>95:5
5	3d	Z	5-CH ₃	21	92	>95:5
6	3e	Z	7-CH ₃	20	97	>95:5
7	3f	E	5-OCH ₃	22	89	94:6
8	3g	E	7 -OCH $_3$	69 ^[d]	80	>95:5
9	3h	Z	$4,5-(OCH_2)_2$	44	72	94:6
10	3i	Z	2-Ph	15	99	>95:5
11	3j	Z	4-Br	$96^{[d,e]}$	82	85:15
12	3k	E	6-C1	64	93	94:6
13	31	Z	5-F	18	90	92:8
14	3m	Z	$5-CO_2Me$	48 ^[d]	95	>95:5
15	3n	Z	$6-CO_2Me$	69 ^[d]	92	>95:5
16	30	Z	5-CN	$96^{[d,f]}$	62	>95:5
17	3 p	Z	$5-NO_2$	$96^{[d,f]}$	60	>95:5

[a] Reaction conditions: oxazoline **2** (0.20 mmol), indole (0.24 mmol), Sc(OTf)₃ (5 mol%) in 0.5 mL CHCl₃ at 60 °C. [b] Isolated yield after column chromatography. [c] *Z/E* ratio was determined by ¹H NMR spectroscopy after column chromatography. [d] Catalyst loading was increased to 10 mol% Sc(OTf)₃ after 40 h. [e] 0.8 additional equivalents of the corresponding indole were added after 40 h. [f] Incomplete conversion of starting material.

We also investigated other π - as well as σ -nucleophiles to extend the substrate scope of our established protocol. When oxazolines (*Z*)- and (*E*)-2 were treated with pyrrole and 10 mol% $Sc(OTf)_3$ in CHCl₃ at 60 °C, *E/Z*-mixtures of pyrrolylenamide 4 were obtained with good overall yields and high *Z/E* ratios after 17 h and 64 h, respectively. Both isomers could be

separated by column chromatography. Under identical conditions, the nucleophilic addition of 1-naphthol to (Z)-2 resulted in a separable mixture of *ortho*-product 5 and *para*-product 6 in a ratio of 62:38 after 17 h. Both isomers were isolated in moderate yields and excellent (Z)-selectivities (Scheme 3). Other potential π -nucleophiles such as β -diketones, enol ethers, and enamines did not furnish any product, but exclusively gave rise to decomposition of starting material.

Scheme 3. Addition Products of 2 with Pyrrole and 1-Naphthol.

Mechanistically, we propose that the Lewis acid will first coordinate to the basic nitrogen atom of oxazolines (Z)- and (E)-2 resulting in ring-opening of the heterocycle and formation of 2-amidoallyl cation 7. This symmetrical and highly reactive intermediate can exist in 3 different conformations. The W-form 7a is generally preferred over the sickle form 7b and the U-form 7c due to minimized 1,3-allylic strain between the two Ph-groups. However, in analogy to reported Lewis-acid catalyzed formations of 2-aminoallyl cations, the W-form 7a might be less favored compared to the sickle form 7b because of additional 1,3-allylic strain between one Ph-group and the bulky, coordinating Lewis acid. 12,13 Intramolecular attack of the acetyl group to 7 regenerates the oxazoline 2. The addition of indole finally forms the product 3a in an irreversible step (Scheme 4). It is unclear whether the (Z)-configuration of 3a is directly related to the 2-amidoallyl cation intermediate 7 or whether double bond isomerization of a potential (E)-3a-isomer had taken place in the reaction medium. Although a possible S_N2 or S_N2 mechanism cannot be fully ruled out in this reaction, we rather assume

a S_N1 -type mechanism due to a high resonance stabilization of intermediate 7. Moreover, our successful experiments with DPP in TFE underline a unimolecular mechanism based on stabilization of 7 by the protic solvent (see Scheme 2, eq. 1).

Scheme 4: Proposed Mechanism in the Formation of Indolylenamide **3a**.

(Z)-2
$$\stackrel{\triangle}{\longrightarrow}$$
 $\stackrel{\triangle}{\longrightarrow}$ $\stackrel{\triangle}{\longrightarrow}$

CONCLUSION

In summary, we have reported a novel strategy to form 2-amidoallyl cations by $Sc(OTf)_3$ -catalyzed ring-opening of 4-alkylidene substituted oxazolines. A broad range of indoles as well as pyrrole and 1-naphthol proved to be suitable π -nucleophiles to give rise to highly α -substituted enamides with good to excellent yields and (Z)-selectivities. We could prove that $Sc(OTf)_3$ itself acts as the active catalytic species. Furthermore, we were able to run the same reaction under mild Brønsted acidic conditions using catalytic amounts of diphenylphosphate in trifluoroethanol providing addition product 3a with high yield and (Z)-selectivity.

EXPERIMENTAL SECTION

General methods. Unless otherwise noted, all reactions were carried out in oven dried glassware and in dry solvents under an argon atmosphere. ¹H- and ¹³C-NMR spectra were recorded in DMSO-d6 or CDCl₃ at 26 °C using a Mercury plus 300 MHz and a Bruker

Avance DRX 400 MHz spectrometer. The spectra were referenced to residual DMSO (2.50 ppm, ¹H; 39.52 ppm, ¹³C) or CHCl₃ (7.26 ppm, ¹H; 77.16 ppm, ¹³C), respectively. Chemical shifts are reported in ppm, multiplicities are indicated by s (singlet), br s (broad singlet), d (doublet), t (triplet), q (quartet), p (pentet), m (multiplet), and related permutations. Coupling constants, J, are reported in Hertz. All high-resolution mass spectra (HRMS) were recorded on a Bruker Daltonics Apex II FT-ICR. Melting points were determined uncorrected on a Boetius measurement device. The used solvents dichloromethane and tetrahydrofuran (THF) were dried using a MBraun Solvent Purification System (SPS) 800. Dry chloroform and methanol were purchased from Acros Organics and stored over molecular sieve. Acetonitrile was technical grade. Solvents for column chromatography were of technical grade and distilled from the indicated drying reagents: dichloromethane (CaH₂), diethyl ether (KOH), methyl-tert-butyl ether (KOH), ethyl acetate (CaCl₂) and n-hexane (KOH). Methanol was technical grade. Triethylamine was distilled freshly prior to use. Flash column chromatography was performed by using silica gel (Fluka, 60 Å, 230 – 400 mesh size). Analytical thin-layer chromatography (TLC) was performed on Macherey-Nagel pre-coated TLC-sheets AlugramXtra SIL G/UV₂₅₄. Visualization of the spots was achieved by UV-light and treatment with a vanillin staining solution.

Synthesis of Oxazoline 2

Methyl α -acetamidocinnamate was synthesized from α -acetamidocinnamic acid 1 according to known literature.²³

A solution of methyl α-acetamidocinnamate (17.5 g, 80.0 mmol, 1.00 equiv) in 150 mL dry THF was added dropwise to a suspension of LiAlH₄ (10.6 g, 280 mmol, 3.50 equiv) in 400 mL dry THF over 30 min at 0 °C. The reaction mixture was stirred at 0 °C for 1 h and then transferred into 300 mL ice-cold EtOAc. It was filtered over Celite and the residue was washed with EtOAc and MeOH. The filtrate was treated with sat. NH₄Cl solution. The

organic phase was separated and the aqueous phase was extracted twice with EtOAc. The combined organic phases were dried over Na₂SO₄, filtered and the solvent was removed under reduced pressure. After purification by flash column chromatography (30% EtOAc/n-hexane \rightarrow EtOAc) N-(3-hydroxy-1-phenylprop-1-en-2-yl)acetamide was isolated as a white solid (9.65 g, 63%). R_f (EtOAc) 0.42; mp 80 °C; ¹H NMR (400 MHz, DMSO-d6) δ 9.14 (bs, 1H), 7.40 (m, 2H), 7.32 (m, 2H), 7.21 (m, 1H), 6.27 (s, 1H), 5.12 (t, 1H, J= 6.0 Hz), 4.13 (d, 2H, J= 6.0 Hz), 1.95 (s, 3H); ¹³C{¹H} NMR (100 MHz, DMSO-d6) δ 169.0, 137.6, 135.6, 128.3, 126.7, 118.3, 62.3, 22.9; IR (KBr) \tilde{v} 3239, 3172, 3062, 3049, 3021, 2940, 2919, 1646, 1556, 1489, 1437, 1373, 1348, 1310, 1183, 1037, 850, 755, 692, 526 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₁₁H₁₃NO₂Na 214.0838; Found 214.0835.

To a solution of N-(3-hydroxy-1-phenylprop-1-en-2-yl)acetamide (9.65 g, 50.5 mmol, 1.00 equiv) in 200 mL CH₃CN was added activated MnO₂ (88% w/w, 40.0 g, 404 mmol, 8.00 equiv) and the mixture was stirred at rt for 15 h. It was filtered over Celite and the residue was washed with EtOAc. The filtrate was concentrated in vacuo and the crude product was purified by flash column chromatography (30% EtOAc/n-hexane \rightarrow EtOAc). N-(3-Oxo-1-phenylprop-1-en-2-yl)acetamide was isolated as a white solid (8.36 g, 88%). R_f (80% EtOAc/n-hexane) 0.50; mp 104-105 °C; ¹H NMR (400 MHz, DMSO-d₆) δ 9.55 (bs, 1H), 9.45 (s, 1H), 7.69 (m, 2H), 7.48-7.43 (m, 3H), 7.32 (s, 1H), 2.05 (s, 3H); ¹³C{¹H} NMR (100 MHz, DMSO-d₆) δ 190.8, 168.9, 139.7, 134.7, 133.4, 130.2, 128.7, 22.6; IR (KBr) \tilde{v} 3464, 3250, 3016, 2847, 1698, 1687, 1659, 1634, 1490, 1419, 1382, 1372, 1321, 1277, 1189, 980, 764, 746, 684, 536, 438 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₁₁H₁₁NO₂Na 212.0682; Found 212.0681.

Bromobenzene (13.4 mL, 128 mmol, 3.50 equiv) was added dropwise to a mixture of activated magnesium turnings (3.20 g, 132 mmol, 3.60 equiv) in 80 mL dry THF over 20 min. After complete addition, the resulting black suspension was heated under reflux in an oil bath for 90 min and then cooled down to 0 °C. *N*-(3-Oxo-1-phenylprop-1-en-2-yl)acetamide (6.79

g, 36.6 mmol, 1.0 equiv) was dissolved in 80 mL dry THF and added dropwise to the Grignard reagent at 0 °C. The reaction mixture was stirred at 0 °C for 1 h and then quenched with 200 mL sat. NH₄Cl solution. The organic phase was separated and the aqueous phase was extracted twice with EtOAc. The combined organic phases were dried over Na₂SO₄, filtered and the solvent was removed under reduced pressure. After purification by flash column chromatography (10% EtOAc/n-hexane \rightarrow EtOAc) N-(3-hydroxy-1,3-diphenylprop-1-en-2-yl)acetamide was isolated as a yellowish solid (9.31 g, 95%). R_f (70% Et₂O/n-hexane) 0.22; mp 115-117 °C; ¹H NMR (300 MHz, DMSO-d6) δ 9.01 (bs, 1H), 7.43-7.40 (m, 2H), 7.35-7.28 (m, 6H), 7.27-7.17 (m, 2H), 6.58 (s, 1H), 5.92 (d, 1H, J = 5.0 Hz), 5.48 (d, 1H, J = 5.0 Hz), 1.85 (s, 3H); ¹³C{¹H} NMR (75 MHz, DMSO-d6) δ 169.0, 143.3, 139.2, 135.6, 128.4, 128.2, 127.9, 127.0, 126.8, 126.7, 119.3, 72.8, 22.8; IR (KBr) \tilde{v} 3396, 3246, 3166, 3054, 3021, 1660, 1645, 1540, 1489, 1447, 1410, 1375, 1319, 1305, 1276, 1179, 1041, 1014, 756, 712, 696 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₁₇H₁₇NO₂Na 290.1151; Found 290.1148.

To a solution of N-(3-hydroxy-1,3-diphenylprop-1-en-2-yl)acetamide (3.00 g, 11.2 mmol, 1.00 equiv) in 50 mL dry CH₂Cl₂, triethylamine (6.20 mL, 44.8 mmol, 4.00 equiv) was added dropwise at 0 °C followed by methanesulfonyl chloride (1.30 mL, 16.8 mmol, 1.50 equiv). After 10 min stirring the reaction mixture was quenched with sat. NaHCO₃ solution. The organic phase was separated and the aqueous phase was extracted twice with CH₂Cl₂. The combined organic phases were dried over Na₂SO₄, filtered and the solvent was removed under reduced pressure. After purification by flash column chromatography (10% \rightarrow 50% Et₂O/n-hexane), oxazoline (Z)-2 was isolated as a white solid (1.23 g, 44%). (Z)-Isomer: R_f (70% Et₂O/n-hexane) 0.85; mp 85 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.81 (m, 2H), 7.45-7.37 (m, 5H), 7.33 (m, 2H), 7.18 (m, 1H), 6.01 (d, 1H, J = 2.5 Hz), 5.58 (d, 1H, J = 2.5 Hz), 2.31 (s, 3H); n-13C (n-14) NMR (100 MHz, CDCl₃) δ 172.2, 152.9, 138.8, 136.4, 129.2, 129.0, 128.6, 128.4,

127.7, 126.4, 112.1, 85.8, 15.2; IR (KBr) \tilde{v} 3083, 3063, 3039, 3029, 3019, 1613, 1593, 1492, 1387, 1307, 1295, 1273, 1248, 1205, 977, 926, 755, 695 cm⁻¹; HRMS (ESI+) m/z: [M+H]⁺ Calcd for C₁₇H₁₆NO 250.1226; Found 250.1223. (*E*)-Isomer: R_f (70% Et₂O/*n*-hexane) 0.41; mp 80 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.39-7.37 (m, 2H), 7.34-7.31 (m, 3H), 7.15-7.11 (m, 2H), 7.07-7.01 (m, 3H), 6.74 (d, 1H, J = 2.5 Hz), 6.34 (d, 1H, J = 2.5 Hz), 2.17 (s, 3H); 13 C{ 1 H} NMR (100 MHz, CDCl₃) δ 169.4, 152.3, 136.1, 135.6, 129.3, 129.1, 128.4, 128.3, 127.7, 126.4, 116.3, 84.5, 14.7; IR (KBr) \tilde{v} 3433, 3086, 3063, 3046, 3026, 1665, 1607, 1589, 1572, 1492, 1446, 1422, 1387, 1318, 1308, 1387, 1230, 1187, 1141, 960, 917, 909, 870, 765, 759, 712, 692, 621, 605, 513 cm⁻¹; HRMS (ESI+) m/z: [M+H]⁺ Calcd for C₁₇H₁₆NO 250.1226; Found 250.1228.

General Procedure for the Synthesis of Indolylenamides 3

Oxazoline (*Z*)-2 or (*E*)-2 (50 mg, 0.20 mmol, 1.00 equiv), Sc(OTf)₃ (4.9 mg, 10 μ mol, 0.05 equiv) and indole (1.20 equiv) were dissolved in dry CHCl₃ (0.5 mL) and stirred at 60 °C in an oil bath for the indicated time. The reaction mixture was quenched with sat. NaHCO₃ solution. The organic phase was separated and the aqueous phase was extracted twice with CH₂Cl₂. The combined organic phases were dried over Na₂SO₄, filtered and the solvent was removed under reduced pressure. The crude products were purified by flash column chromatography (CH₂Cl₂ \rightarrow 1% MeOH/CH₂Cl₂) and dried under high vacuum (0.1 mbar) at 60 °C overnight.

Indolylenamide 3a was obtained from oxazoline (*Z*)-2 and indole (28 mg, 0.24 mmol) after 16 h as a white solid (69 mg, 95%, Z/E >95:5). R_f (1% MeOH/CH₂Cl₂) 0.28; mp 175-177 °C; ¹H NMR (400 MHz, DMSO-d6) δ 10.90 (bs, 1H), 9.22 (bs, 1H), 7.62 (d, 1H, J = 8.0 Hz), 7.34-7.24 (m, 10H), 7.19-7.14 (m, 1H), 7.05 (t, 1H, J = 7.5 Hz), 6.92 (t, 1H, J = 7.5 Hz) 6.73 (s, 1H), 5.84 (s, 1H), 5.62 (s, 1H), 1.86 (s, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 168.9,

142.1, 139.4, 136.6, 135.8, 128.8, 128.2, 128.1, 126.8, 126.6, 126.4, 124.2, 122.5, 121.1, 119.7, 118.3, 116.1, 111.4, 47.3, 23.0; IR (KBr) \tilde{v} 3413, 3056, 3024, 2924, 2853, 1655, 1491, 1455, 1267, 745, 698 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₂₅H₂₂N₂ONa 389.1624; Found 389.1626.

Indolylenamide 3b was obtained from oxazoline (*E*)-2 and 1-methylindole (31 μL, 0.24 mmol) after 96 h as a white solid (62 mg, 82%, Z/E > 95:5). R_f (1% MeOH/CH₂Cl₂) 0.44; mp 154-157 °C; ¹H NMR (400 MHz, DMSO-d6) δ 9.22 (bs, 1H), 7.69 (d, 1H, J = 8.0 Hz), 7.39-7.35 (m, 5H), 7.30-7.24 (m, 5H), 7.19-7.14 (m, 1H), 7.13 (t, 1H, J = 7.5 Hz), 6.97 (t, 1H, J = 7.5 Hz), 6.71 (s, 1H), 5.88 (s, 1H), 5.64 (s, 1H), 3.71 (s, 3H), 1.86 (s, 3H); ¹³C { ¹H } NMR (100 MHz, DMSO-d6) δ 169.0, 142.0, 139.4, 137.1, 135.7, 128.7, 128.5, 128.3, 128.3, 128.1, 127.1, 126.7, 126.4, 122.6, 121.2, 120.0, 118.5, 115.5, 109.5, 47.2, 32.3, 23.0; IR (KBr) \tilde{v} 3406, 3055, 3023, 2930, 1655, 1614, 1490, 1449, 1371, 1330, 1273, 743, 700 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₂₆H₂₄N₂ONa 403.1781; Found 403.1782.

Indolylenamide 3c was obtained from oxazoline (*E*)-2, 2-methylindole (53 mg, 0.40 mmol, 2.00 equiv) and Sc(OTf)₃ (9.8 mg, 20 μmol, 0.10 equiv) after 96 h as a brown solid (55 mg, 72%, Z/E > 95:5). R_f (1% MeOH/CH₂Cl₂) 0.21; mp 103-108 °C; ¹H NMR (400 MHz, DMSOd6) δ 10.87 (bs, 1H), 9.18 (bs, 1H), 7.33-7.23 (m, 10H) 7.20-7.15 (m, 2H), 6.94 (t, 1H, J = 7.5 Hz) 6.82 (t, 1H, J = 7.5 Hz), 5.91 (s, 1H), 5.81 (s, 1H), 2.26 (s, 3H), 1.81 (s, 3H); 13 C{ 1 H} NMR (100 MHz, DMSO-d6) δ 168.7, 142.3, 139.3, 135.8, 135.3, 133.2, 128.9, 128.2, 128.1, 128.0, 126.6, 125.9, 122.5, 119.7, 118.7, 118.2, 110.5, 110.0, 46.0, 23.0, 11.8; IR (KBr) \tilde{v} 3399, 3056, 3023, 1660, 1621, 1490, 1458, 746, 698 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₂₆H₂₄N₂ONa 403.1781; Found 403.1780.

Indolylenamide 3d was obtained from oxazoline (*Z*)-2 and 5-methylindole (32 mg, 0.24 mmol) after 21 h as a white solid (70 mg, 92%, Z/E > 95:5). R_f (1% MeOH/CH₂Cl₂) 0.45; mp 88-95 °C; ¹H NMR (400 MHz, DMSO-d6) δ 10.79 (bs, 1H), 9.23 (bs, 1H), 7.40 (s, 1H), 7.34-7.33 (m, 4H), 7.31-7.24 (m, 6H), 7.18-7.14 (m, 1H), 6.89 (dd, 1H, J = 8.5, 1.5 Hz, 1H), 6.72 (d, 1H, J = 2.0 Hz) 5.84 (s, 1H), 5.64 (s, 1H), 2.31 (s, 3H), 1.87 (s, 3H); ¹³C{¹H} NMR (100 MHz, DMSO-d6) δ 168.8, 142.3, 139.6, 135.9, 135.0, 128.8, 128.2, 128.1, 127.1, 126.7, 126.6, 126.3, 124.3, 122.7, 122.4, 119.1, 115.5, 111.1, 47.1, 23.0, 21.4; IR (KBr) \tilde{v} 3407, 3296, 3055, 3023, 1661, 1599, 1491, 1448, 1424, 1369, 796, 753, 699 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₂₆H₂₄N₂ONa 403.1781; Found 403.1777.

Indolylenamide 3e was obtained from oxazoline (*Z*)-2 and 7-methylindole (32 mg, 0.24 mmol) after 20 h as a white solid (74 mg, 97%, Z/E > 95:5). R_f (1% MeOH/CH₂Cl₂) 0.36; mp 95-98 °C; ¹H NMR (400 MHz, DMSO-d6) δ 10.87 (bs, 1H), 9.24 (bs, 1H), 7.46 (dd, 1H, J = 7.0, 2.0 Hz), 7.35-7.23 (m, 9H), 7.17-7.14 (m, 1H), 6.87-6.82 (m, 2H), 6.75 (d, 1H, J = 2.0 Hz), 5.86 (s, 1H), 5.65 (s, 1H), 2.46 (s, 3H), 1.88 (s, 3H); ¹³C{¹H} NMR (100 MHz, DMSO-d6) δ 168.9, 142.2, 139.5, 136.2, 135.9, 128.8, 128.2, 128.1, 126.6, 126.5, 126.4, 123.9, 122.6, 121.6, 120.4, 118.6, 117.3, 116.6, 47.4, 23.0, 16.8; IR (KBr) \tilde{v} 3420, 3054, 3024, 1655, 1492, 1448, 749, 699 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₂₆H₂₄N₂ONa 403.1781; Found 403.1777.

Indolylenamide 3f was obtained from oxazoline (*E*)-2 and 5-methoxyindole (35 mg, 0.24 mmol) after 15 h as a white solid (70 mg, 89%, Z/E 94:6). R_f (1% MeOH/CH₂Cl₂) 0.22; mp 86-90 °C; ¹H NMR (400 MHz, DMSO-d6) δ 10.70 (bs, 1H), 9.20 (bs, 1H), 7.36-7.35 (m, 4H), 7.28-7.22 (m, 7H), 7.18-7.14 (m, 1H), 6.71 (dd, 1H, J = 8.5, 2.5 Hz), 6.63 (d, 1H, J = 2.0 Hz), 5.69 (s, 1H), 5.51 (s, 1H), 3.66 (s, 3H), 1.87 (s, 3H); ¹³C{¹H} NMR (100 MHz, DMSO-d6) δ 169.1, 152.8, 142.0, 139.3, 135.9, 131.8, 128.8, 128.2, 128.0, 127.2, 126.7, 126.4, 124.8,

122.9, 116.1, 111.9, 111.0, 102.0, 55.2, 47.6, 22.9; IR (KBr) \tilde{v} 3411, 3056, 3024, 2936, 2829, 1656, 1626, 1485, 1451, 1209, 1171, 699 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for $C_{26}H_{24}N_2O_2Na$ 419.1730; Found 419.1729.

Indolylenamide 3g was obtained from oxazoline (*E*)-2, 7-methoxyindole (35 mg, 0.24 mmol) and Sc(OTf)₃ (9.8 mg, 20 μmol, 0.10 equiv) after 69 h as a white solid (63 mg, 80%, Z/E >95:5). R_f (1% MeOH/CH₂Cl₂) 0.29; mp 94-98 °C; ¹H NMR (400 MHz, DMSO-d6) δ 10.97 (bs, 1H), 9.21 (bs, 1H), 7.34-7.33 (m, 4H), 7.27-7.20 (m, 6H), 7.17-7.14 (m, 1H), 6.85 (t, 1H, J= 8.0 Hz), 6.63-6.61 (m, 2H), 5.83 (s, 1H), 5.60 (s, 1H), 3.89 (s, 3H), 1.86 (s, 3H); ¹³C{¹H} NMR (100 MHz, DMSO-d6) δ 168.9, 146.0, 142.2, 139.4, 135.8, 128.8, 128.3, 128.2, 128.1, 126.7, 126.6, 126.4, 123.8, 122.5, 118.9, 116.7, 112.6, 101.6, 55.0, 47.3, 23.0; IR (KBr) \tilde{v} 3422, 3055, 3024, 2934, 1655, 1578, 1494, 1465, 1448, 1370, 1261, 1232, 1091, 1046, 782, 752, 731, 716, 700 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₂₆H₂₄N₂O₂Na 419.1730; Found 419.1729.

Indolylenamide 3h was obtained from oxazoline (*Z*)-2 and 4,5-ethylenedioxyindole (42 mg, 0.24 mmol) after 44 h as a white solid (61 mg, 72%, *Z/E* 94:6). R_f (1% MeOH/CH₂Cl₂) 0.18; mp 104-109 °C; ¹H NMR (400 MHz, DMSO-d6) δ 10.77 (bs, 1H), 9.13 (bs, 1H), 7.31-6.57 (m, 10H), 6.81-6.77 (m, 2H), 6.58 (d, 1H, J= 8.5 Hz), 5.77 (s, 1H), 5.64 (s, 1H), 4.12-4.08 (m, 3H) 4.02-3.98 (m, 1H), 1.80 (s, 3H); ¹³C{¹H} NMR (100 MHz, DMSO-d6) δ 167.8, 143.6, 140.0, 136.5, 136.3, 134.9, 134.9, 128.9, 128.0, 127.8, 127.7, 126.3, 125.8, 123.7, 122.2, 117.1, 114.8, 112.3, 104.0, 64.0, 63.7, 48.4, 23.1; IR (KBr) \tilde{v} 3417,3055, 3023, 2926, 2870, 1668, 1503, 1446, 1368, 1338, 1274, 1257, 1222, 1085, 962, 714, 699 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₂₇H₂₄N₂O₃Na 447.1679; Found 447.1680.

Indolylenamide 3i was obtained from oxazoline (*Z*)-2 and 2-phenylindole (46 mg, 0.24 mmol) after 15 h as a white solid (87 mg, 99%, Z/E > 95:5). R_f (1% MeOH/CH₂Cl₂) 0.24; mp 107-112 °C; ¹H NMR (400 MHz, DMSO-d6) δ 11.34 (bs, 1H), 9.14 (bs, 1H), 7.59 (m, 2H), 7.50-7.46 (m, 3H), 7.42-7.37 (m, 2H), 7.28-7.22 (m, 8H), 7.19-7.13 (m, 2H), 7.06 (t, 1H, J = 7.5 Hz), 6.90 (t, 1H, J = 7.5 Hz), 6.10 (s, 1H), 5.84 (s, 1H), 1.70 (s, 3H); ¹³C{¹H} NMR (100 MHz, DMSO-d6) δ 168.5, 142.4, 139.0, 136.5, 135.8, 132.6, 128.9, 128.6, 128.5, 128.2, 128.1, 128.0, 127.7, 126.7, 125.9, 123.4, 121.1, 120.5, 118.8, 111.5, 110.6, 46.8, 22.8; IR (KBr) \tilde{v} 3397, 3281, 3056, 3023, 1668, 1600, 1490, 1449, 1426, 764, 746, 699 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₃₁H₂₆N₂ONa 465.1937; Found 465.1941.

Indolylenamide 3j was obtained from oxazoline (*Z*)-2, 4-bromoindole (50 μL, 0.40 mmol, 2.00 equiv) and Sc(OTf)₃ (9.8 mg, 20 μmol, 0.10 equiv) after 96 h as a white solid (73 mg, 82%, *Z/E* 85:15). (*Z*)-isomer: R_f (1% MeOH/CH₂Cl₂) 0.26; mp 93-95 °C; ¹H NMR (400 MHz, DMSO-d6) δ 11.41 (bs, 1H), 9.22 (bs, 1H), 7.40 (d, 1H, J = 8.0 Hz), 7.08-7.31 (m, 11H) 7.01 (d, 1H, J = 2.0 Hz), 6.95 (t, 1H, J = 8.0 Hz), 6.16 (s, 1H), 5.52 (s, 1H), 1.79 (s, 3H); ¹³C{¹H} NMR (100 MHz, DMSO-d6) δ 167.8, 143.1, 139.9, 138.2, 136.4, 129.2, 128.0, 127.9, 126.4, 126.0, 124.5, 122.9, 122.7, 122.2, 115.5, 113.1, 111.3, 47.3, 23.2; IR (KBr) \tilde{v} 3419, 3056, 3024, 1669, 1652, 1491, 1335, 1184, 746, 699 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₂₅H₂₁⁷⁹BrN₂ONa 467.0730; Found 467.0726.

Indolylenamide 3k was obtained from oxazoline (*E*)-2 and 6-chloroindole (36 mg, 0.24 mmol) after 64 h as a white solid (74 mg, 93%, *Z/E* 94:6). R_f (1% MeOH/CH₂Cl₂) 0.33; mp 205-206 °C; ¹H NMR (400 MHz, DMSO-d6) δ 11.05 (bs, 1H), 9.24 (bs, 1H), 7.66 (d, 1H, J = 8.5 Hz), 7.40 (d, 1H, J = 1.5 Hz), 7.36-7.33 (m, 4H), 7.29-7.24 (m, 5H), 7.18-7.15 (m, 1H), 6.95 (dd, 1H, J = 8.5, 1.5 Hz), 6.74 (d, 1H, J = 1.5 Hz), 5.83 (s, 1H), 5.59 (s, 1H), 1.87 (s, 3H); ¹³C{¹H} NMR (100 MHz, DMSO-d6) δ 169.1, 141.7, 139.3, 137.0, 135.7, 128.8,

128.3, 128.2, 128.1, 126.7, 126.5, 125.9, 125.6, 125.4, 122.8, 121.1, 118.7, 116.6, 111.0, 47.2, 22.9; IR (KBr) \tilde{v} 3420, 3223, 3057, 3022, 1649, 1518, 1491, 1450, 907, 812, 749, 716, 700 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₂₅H₂₁³⁵ClN₂ONa 423.1235; Found 423.1231.

Indolylenamide 31 was obtained from oxazoline (*Z*)-2 and 5-fluoroindole (32 mg, 0.24 mmol) after 18 h as a white solid (69 mg, 90%, *Z/E* 92:8). R_f (1% MeOH/CH₂Cl₂) 0.29; mp 84-89 °C; ¹H NMR (400 MHz, DMSO-d6) δ 10.97 (bs, 1H), 9.25 (bs, 1H), 7.46 (dd, 1H, J = 10.0, 2.5 Hz), 7.38-7.24 (m, 10H), 7.20-7.16 (m, 1H), 6.90 (td, 1H, J = 9.0, 2.5 Hz), 6.76 (d, 1H, J = 2.5 Hz), 5.88 (s, 1H), 5.54 (s, 1H), 1.89 (s, 3H); ¹³C{¹H} NMR (100 MHz, DMSO-d6) δ 169.2, 156.5 (d, J = 231.0 Hz), 141.7, 139.2, 135.7, 133.3, 128.8, 128.3, 128.2, 128.1, 127.0 (d, J = 10.0 Hz), 126.8, 126.6, 126.3, 122.8, 116.7 (d, J = 4.5 Hz), 112.4 (d, J = 10.0 Hz), 109.3 (d, J = 26.0 Hz), 104.5 (d, J = 23.5 Hz), 47.4, 22.9; IR (KBr) \tilde{v} 3417, 3383, 3283, 3056, 3024, 2922, 1661, 1582, 1486, 1451, 1369, 1263, 1166, 940, 798, 751, 699 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₂₅H₂₁FN₂ONa 407.1530; Found 407.1533.

Indolylenamide 3m was obtained from oxazoline (*Z*)-2, methyl indole-5-carboxylate (42 mg, 0.24 mmol) and Sc(OTf)₃ (9.8 mg, 20 μmol, 0.10 equiv) after 48 h as a white solid (81 mg, 95%, Z/E > 95:5). R_f (1% MeOH/CH₂Cl₂) 0.11; mp 199-201 °C; ¹H NMR (400 MHz, DMSOd6) δ 11.37 (bs, 1H), 9.26 (bs, 1H), 8.32 (s, 1H), 7.71 (d, 1H, J = 8.5 Hz), 7.44 (d, 1H, J = 8.5 Hz), 7.35-7.24 (m, 9H), 7.18-7.14 (m, 1H), 6.94 (d, 1H, J = 2.5 Hz), 5.81 (s, 1H), 5.74 (s, 1H), 3.79 (s, 3H), 1.86 (s, 3H); ¹³C{¹H} NMR (100 MHz, DMSOd6) δ 168.8, 167.2, 141.9, 139.3, 139.2, 135.7, 128.7, 128.3, 128.2, 128.1, 126.7, 126.5, 126.4, 126.2, 122.5, 122.2, 122.1, 120.1, 117.4, 111.5, 51.6, 46.8, 23.0; IR (KBr) \tilde{v} 3420, 3023, 2949, 1691, 1654, 1617, 1491, 1436, 1279, 1244, 1109, 754, 699 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₂₇H₂₄N₂O₃Na 447.1679; Found 447.1680.

Indolylenamide 3n was obtained from oxazoline (*Z*)-2, methyl indole-6-carboxylate (42 mg, 0.24 mmol) and Sc(OTf)₃ (9.8 mg, 20 μmol, 0.10 equiv) after 69 h as a white solid (78 mg, 92%, Z/E > 95:5). R_f (1% MeOH/CH₂Cl₂) 0.19; mp 99-104 °C; ¹H NMR (400 MHz, DMSOd6) δ 11.32 (bs, 1H), 9.25 (bs, 1H), 8.05 (s, 1H), 7.73 (d, 1H, J = 8.5 Hz), 7.44 (d, 1H, J = 8.5 Hz), 7.36-7.35 (m, 4H), 7.29-7.25 (m, 5H), 7.18-7.14 (m, 1H), 7.01 (d, 1H, J = 2.0 Hz), 5.84 (s, 1H), 5.67 (s, 1H), 3.83 (s, 3H), 1.87 (s, 3H); ¹³C{¹H} NMR (100 MHz, DMSO-d6) δ 169.0, 167.2, 141.1, 139.2, 135.9, 135.7, 130.3, 128.8, 128.3, 128.2, 128.1, 126.8, 126.6, 122.8, 122.2, 119.5, 119.1, 116.8, 113.5, 51.7, 47.1, 23.0; IR (KBr) \tilde{v} 3411, 3323, 3057, 3024, 2949, 1694, 1665, 1623, 1492, 1455, 1435, 1320, 1276, 1218, 1086, 776, 749, 699 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₂₇H₂₄N₂O₃Na 447.1679; Found 447.1677.

Indolylenamide 30 was obtained from oxazoline (*Z*)-2, 5-cyanoindole (34 mg, 0.24 mmol) and Sc(OTf)₃ (9.8 mg, 20 μmol, 0.10 equiv) after 96 h as a pink solid (48 mg, 62%, *Z/E* >95:5). R_f (1% MeOH/CH₂Cl₂) 0.18; mp 107-110 °C; ¹H NMR (400 MHz, DMSO-d6) δ 11.47 (bs, 1H), 9.30 (bs, 1H), 8.26 (s, 1H), 7.53 (d, 1H, J = 8.5 Hz), 7.42 (d, 1H, J = 8.5 Hz), 7.39-7.35 (m, 4H), 7.32-7.25 (m, 5H), 7.20-7.16 (m, 1H), 6.87 (d, 1H, J = 2.5 Hz), 5.87 (s, 1H), 5.60 (s, 1H), 1.90 (s, 3H); ¹³C{¹H} NMR (100 MHz, DMSO-d6) δ 169.4, 141.2, 139.2, 138.4, 135.5, 128.8, 128.4, 128.3, 128.1, 126.9, 126.8, 126.6, 125.5, 123.9, 123.2, 120.9, 117.8, 112.8, 100.5, 47.2, 22.9; IR (KBr) \tilde{v} 3387, 3296, 3057, 3025, 2220, 1656, 1617, 1492, 1473, 1448, 1368, 807, 753, 699 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₂₆H₂₁N₃ONa 414.1577; Found 414.1578.

Indolylenamide 3p was obtained from oxazoline (*Z*)-**2**, 5-nitroindole (39 mg, 0.24 mmol) and Sc(OTf)₃ (9.8 mg, 20 μ mol, 0.10 equiv) after 96 h as a yellow solid (49 mg, 60%, *Z/E* >95:5). R_f (1% MeOH/CH₂Cl₂) 0.20; mp 109-114 °C; ¹H NMR (400 MHz, DMSO-d6) δ

11.32 (bs, 1H), 9.30 (bs, 1H), 8.76 (d, 1H, J = 2.0 Hz), 7.98 (dd, 1H, J = 9.0, 2.0 Hz), 7.53 (d, 1H, J = 9.0 Hz), 7.38-7.36 (m, 4H), 7.31-7.25 (m, 5H), 7.19-7.15 (m, 1H), 6.97 (s, 1H), 5.87 (s, 1H), 5.71 (s, 1H), 1.90 (s, 3H); 13 C{ 1 H} NMR (100 MHz, DMSO-d6) δ 169.3, 141.3, 140.4, 139.8, 139.1, 135.5, 128.8, 128.5, 128.2, 128.1, 126.9, 126.8, 126.1, 123.0, 119.2, 117.0, 116.7, 112.0, 47.0, 22.9; IR (KBr) \tilde{v} 3408, 3058, 3025, 1654, 1624, 1518, 1492, 1472, 1332, 1100, 700 cm $^{-1}$; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₂₅H₂₁N₃O₃Na 434.1475; Found 434.1472.

Pyrrolylenamides 4. Oxazoline (*Z*)-2 (50 mg, 0.20 mmol, 1.00 equiv), Sc(OTf)₃ (9.8 mg, 20 μmol, 0.10 equiv) and pyrrole (28 μL, 0.40 mmol, 2.00 equiv) were dissolved in dry CHCl₃ (0.5 mL) and stirred at 60 °C in an oil bath for 17 h. The reaction mixture was guenched with sat. NaHCO₃ solution. The organic phase was separated and the aqueous phase was extracted twice with CH₂Cl₂. The combined organic phases were dried over Na₂SO₄, filtered and the solvent was removed under reduced pressure. The crude products were purified by flash column chromatography ($20\% \rightarrow 50\%$ MTBE/n-hexane) and dried under high vacuum (0.1 mbar) at 60 °C overnight. Compound (Z)-4 was obtained as an orange solid (38 mg, 59%). R_f (50% MTBE/n-hexane) 0.25; mp 49-52 °C; ¹H NMR (300 MHz, DMSO-d6) δ 10.61 (bs, 1H), 9.21 (bs, 1H), 7.33-7.16 (m, 10H), 6.67-6.64 (m, 1H), 5.97-5.94 (m, 1H), 5.78-5.75 (m, 2H), 5.46 (s, 1H), 1.83 (s, 3H); ¹³C{¹H} NMR (75 MHz, DMSO-d6) δ 168.8, 142.2, 139.5, 135.7, 131.2, 128.7, 128.21, 128.15, 128.07, 126.7, 126.3, 122.6, 117.3, 107.1, 106.9, 48.8, 23.0; IR (KBr) v 3419, 3372, 3269, 3057, 3024, 2924, 2853, 1656, 1516, 1492, 1369, 1276, 1075, 1028, 751, 711, 698 cm⁻¹; HRMS (ESI+) m/z; [M+Na]⁺ Calcd for C₂₁H₂₀N₂ONa 339.1468; Found 339.1466; Compound (E)-4 was obtained as an orange solid (5 mg, 7%). R_f (50%) MTBE/n-hexane) 0.40; mp 102-105 °C; ¹H NMR (400 MHz, DMSO-d6) δ 10.74 (bs, 1H), 7.81 (bs, 1H), 7.50 (s, 1H), 7.35-7.18 (m, 8H), 7.10 (m, 2H), 6.75-6.73 (m, 1H), 6.03-6.01 (m, 1H), 5.91-5.89 (m, 1H), 5.45 (s, 1H), 1.92 (s, 3H); ${}^{13}C\{{}^{1}H\}$ NMR (100 MHz, DMSO-d6) δ

169.1, 140.6, 136.7, 135.4, 129.4, 128.5, 128.4, 127.8, 126.7, 126.3, 118.0, 117.3, 107.6, 107.4, 44.0, 24.2; IR (KBr) \tilde{v} 3410, 3344, 3024, 2924, 1675, 1647, 1517, 1494, 1267, 734, 700 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₂₁H₂₀N₂ONa 339.1468; Found 339.1466.

1-Hydroxynaphthalenylenamides 5 and 6. Oxazoline (Z)-2 (50 mg, 0.20 mmol, 1.00 equiv), Sc(OTf)₃ (9.8 mg, 20 µmol, 0.10 equiv), and 1-naphthol (35 mg, 0.24 mmol, 1.20 equiv) were dissolved in dry CHCl₃ (0.5 mL) and stirred at 60 °C in an oil bath for 17 h. The reaction mixture was quenched with sat. NaHCO₃ solution. The organic phase was separated and the aqueous phase was extracted twice with CH₂Cl₂. The combined organic phases were dried over Na₂SO₄, filtered and the solvent was removed under reduced pressure. The crude products were purified by flash column chromatography ($10\% \rightarrow 70\%$ MTBE/n-hexane) and dried under high vacuum (0.1 mbar) at 60 °C overnight. Compound 5 was obtained as a white solid (28 mg, 36%, Z/E >95:5). R_f (30% MTBE/n-hexane) 0.26; mp 210-212 °C; ¹H NMR (400 MHz, DMSO-d6) δ 10.30 (bs, 1H), 10.00 (bs, 1H), 8.23-8.21 (m, 1H), 7.78-7.76 (m, 1H), 7.46-7.40 (m, 4H), 7.37-7.26 (m, 8H), 7.23-7.20 (m, 1H), 6.80 (d, 1H, J = 8.5 Hz), 5.94 (s, 1H), 5.65 (s, 1H), 2.03 (s, 3H); ${}^{13}C\{{}^{1}H\}$ NMR (100 MHz, DMSO-d6) δ 171.9, 150.1, 139.9, 138.2, 134.7, 133.2, 129.4, 128.6, 128.4, 128.2, 127.4, 127.2, 127.0, 126.8, 125.8, 125.6, 124.83, 124.77, 122.2, 121.3, 118.2, 49.9, 22.3; IR (KBr) \tilde{v} 3407, 3250, 3056, 3025, 2925, 1654, 1569, 1508, 1492, 1383, 1279, 752, 698 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₂₇H₂₃NO₂Na 416.1621; Found 416.1611; Compound 6 was obtained as a white solid (17 mg, 22%, Z/E >95:5). R_f (70% MTBE/n-hexane) 0.56; mp 127-133 °C; ¹H NMR $(400 \text{ MHz}, DMSO-d6) \delta 10.05 \text{ (bs, 1H)}, 9.30 \text{ (bs, 1H)}, 8.36 \text{ (d, 1H } J = 8.5 \text{ Hz)}, 8.16 \text{ (dd, 1H, } J = 8.5 \text{ Hz)}$ J = 8.0, 1.5 Hz), 7.48-7.39 (m, 2H), 7.36-7.32 (m, 2H), 7.25-7.23 (m, 7H), 7.18-7.13 (m, 1H), 6.89 (d, 1H, J = 8.0 Hz), 6.80 (d, 1H, J = 8.0 Hz), 6.26 (s, 1H), 5.53 (s, 1H), 1.82 (s, 3H); ¹³C{¹H} NMR (100 MHz, DMSO-d6) δ 169.0, 152.2, 141.9, 139.6, 135.6, 132.9, 129.4, 128.4, 128.2, 128.1, 127.9, 127.1, 126.8, 126.5, 126.2, 125.1, 124.6, 124.2, 123.5, 122.3,

107.3, 50.6, 23.0; IR (KBr) \tilde{v} 3384, 3058, 3025, 2925, 1656, 1625, 1598, 1586, 1516, 1491, 1378, 1271, 765, 754, 700 cm⁻¹; HRMS (ESI+) m/z: [M+Na]⁺ Calcd for C₂₇H₂₃NO₂Na 416.1621; Found 416.1626.

ASSOCIATED CONTENT

¹H and ¹³C NMR spectra for all new compounds and crystallographic data for compound **3a** (deposition numbers: CCDC 1541861) (PDF).

Crystal data and structure refinement for indolylenamide 3a (CIF).

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