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1-Toluene-sulfonyl-3-[(3'-hydroxy-5'-substituted)-γ-butyrolactone]indoles: Synthesis, COX-2 inhibition and anti-cancer activities

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Abstract—Indoles carrying a cyclic ester (γ -butyrolactone) at C-3 position have been synthesized by the allylation of 3-indoleglyoxylate followed by iodocyclisation and the nucleophilic replacement of the iodo-group. Screening of these molecules for COX-2 inhibition and anti-cancer activities has identified compounds **10** and **11** as highly potent and selective for COX-2 as well as showing remarkable anti-cancer activities (better than that of indomethacin). © 2007 Elsevier Ltd. All rights reserved.

The critical role of indole moiety in indole acetic acid¹ (a plant growth regulator hormone), in tryptophan¹ (an amino acid) and as a part of a number of alkaloids² has subjected it to several derivatisations for procuring medicinally important molecules. The development of indomethacin³ (1) as a non-steroidal anti-inflammatory drug (NSAID) (non-selective COX-2 inhibitor) has been followed by the modifications in the different parts of the molecule, especially in the C-3 substituent, for increasing its efficacy⁴ as well as for transforming it into selective COX-2 inhibitors (1, n = 2, 3, branched alkyl groups, R = NH₂, alkyl).⁵

In our ongoing research programme for developing COX-2 inhibitors, it has been planned to introduce a γ -butyrolactone at C-3 and a tolylsulfonyl group at N-1 of indole (2). The γ -butyrolactone moiety (cyclic ester) mimics the furanone moiety present in rofecoxib⁶ (a selective COX-2 inhibitor), while tolylsulfonyl is a key pharmacophore in most of the COX-2 inhibitors. The rationally designed molecules (2) have been synthesized and evaluated for their COX-1, COX-2 inhibitory activities and also screened for anti-cancer properties over 59 human tumour cell lines (Fig. 1).

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The synthesis of the target molecules has been achieved by the indium mediated diastereoselective allylation of 3-indoleglyoxylate, followed by diastereoselective iodocyclisation⁷ and nucleophilic replacement of the iodogroup. Commercially available 3-indoleglyoxylate (3) was *N*-alkylated by reaction with *p*-toluene-sulfonyl chloride to get indole substituted α -keto ester 4 (Scheme 1). A solution of 4, allyl bromide and indium metal (suspension) (1:1.5:1) in THF–H₂O (2:1) on stirring at 0 °C for 6–8 h, after usual workup and chromatography, provided compound 5 (87%) as a thick liquid [M⁺ *m*/*z* 399] (Scheme 1). The participation of Cram's chelation model in this reaction explains the diastereoselective formation of compound 5.⁷

Stirring of solution of 5 in C_2H_5OH -THF (2:1) with 2% NaOH for 2 h, after acidification and extraction with



Figure 1.

Keywords: Indole; Butyrolactone; Synthesis; COX-2 inhibition; Dockings; Anti-cancer activities.

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Scheme 1. Reagents and conditions: (a) *p*-tolunenesulfonyl chloride, NaH, CH₃CN; (b) allyl bromide, In, THF/H₂O (2:1), 0 °C, stir; (c) ethanol–THF, 2% NaOH, stir.

ethyl acetate, gave the corresponding acid 6 (70%) $[M^+-OH m/z 368]$ (Scheme 1). Compound 6 on treatment with I₂ (3 equiv) and NaHCO₃ (3 equiv) in dry CH₃CN for 3-4 h after usual workup gave a mixture of two diastereomers in the ratio 3:1 (¹H NMR integration) (84%) (Scheme 2).⁸ In higher R_f component 7; 61%, mp 130 °C; the observation of NOE between C-5'H and aromatic protons and no NOE between CH₂I unit and aromatic protons indicates that C-5'H and indole moiety are placed on the same face of furanone and helps in assigning stereochemistry 3'S*, 5'R* at the two chiral centers. In case of lower R_f component 8, 18%, NOE has been observed at ArHs when CH₂I protons were irradiated and accordingly stereochemistry 3'S*, 5'S* has been assigned at the two chiral centres. Compound **6** also underwent iodocyclisation with I_2 in the presence of silica gel (for TLC) in ethyl acetate-methanol to give a mixture of 7 and 8 (85%) in the diastereomeric ratio 3:1.

Based upon the previous results,⁹ docking studies and the literature reports,¹⁰ it was planned to introduce the



Scheme 3. Reagents and conditions: (a) ethanthiol, CH₃CN, K₂CO₃, stir, rt (for 9); (b) NaCN, CH₃CN, stir, rt (for 10); (c) KSCN, CH₃CN, stir, rt (for 11).

groups like $CH_2SC_2H_5$, CH_2SCN , and CH_2CN at C-5' of butyrolactone ring. Nucleophilic substitutions of 7 with ethanthiol, NaCN and KSCN in CH_3CN gave compounds 9, 10 and 11 possessing, respectively, SC_2H_5 , CN and SCN groups in place of iodine in 7 (Scheme 3).

Compounds 7, 9, 10 and 11 have been evaluated for in-vitro COX-1, COX-2 inhibitory activities using COX (ovine) inhibitor screening assay kit (Cayman Chemicals, cat. No. 560101). The results of these investigations are given in Table 1. Out of these four compounds, 10 and 11 with, respectively, CH₂CN and CH₂SCN groups at C-5' show the best results with 90% and 85% inhibition of COX-2 at 10^{-8} M concentration and IC_{50} values <0.01 μ M. 60 and 65% COX-1 inhibitions at 10^{-5} M concentrations have been observed for compounds 10 and 11, respectively. Significantly, the results of these two compounds are better than that of indomethacin and rofecoxib. The COX-2 inhibition by compound 9 is also considerable (65% at 10^{-8} M) while compound 7 exhibits poor inhibition. The results of these investigations indicate that along with other components, the C-5' substituent significantly controls the COX-2 inhibitory activities of these compounds.

In order to investigate the interactions of these compounds in the active site of COX-2 and to rationalize the role of C-5' substituent towards COX-2 inhibition, we carried out the dockings of these molecules in the



Scheme 2. Reagents and conditions: (a) I₂, NaHCO₃, CH₃CN, 0 °C, stir; (b) I₂, silica, ethyl acetate-methanol, stir, rt.

Compound			% Inhibition	IC ₅₀ (µM)		COX-2 selectivity*		
	COX-2				COX-1	COX-2	COX-1	
	$10^{-5} {\rm M}$	10^{-6} M	$10^{-7} \mathrm{M}$	$10^{-8} {\rm M}$	10^{-5} M			
7	30	32	35	44	40	>10	>10	
9	65	62	67	75	35	< 0.001	>10	<1000
10	90	87	88	95	60	< 0.001	<10	>1000
11	85	90	89	92	65	< 0.001	<10	>1000
Indomethacin5d				97	100	0.75	0.05	0.067
Rofecoxib			75	100	75	0.3	40	~133

Table 1. In-vitro COX-2 inhibitory activities of indole derivatives

* COX-2 selectivity = IC_{50} (COX-1)/ IC_{50} (COX-2).

active site of COX-2.¹¹ The crystal structure of COX-2 with indomethacin shows the interactions of the carboxyl group of indomethacin with the guanidine moiety of R120, amino acid active during the turnover phase of the enzyme. Figure 2 validates our programme of docking, where indomethacin docked in the active site of COX-2 nearly overlaps with the one present in the crystal structure of the enzyme (rms deviation 0.88).

Docking of compound 10 in the active site of COX-2 indicates that this molecule fits in the COX-2 active site in the same fashion as indomethacin. In compound 10, the oxygen of C-3' OH group is present at a distance of 2.10 Å from the H of guanidine moiety of R120 and interacts through H-bond formation while the CN group present at C-5' position approaches the guanidine moiety at a distance of 2.43 Å (Fig. 3). The aromatic part of the tosyl group present at N-1 of compound 10 fits in the hydrophobic pocket formed by F518, W387 and Y385 residues. While similar types of interactions are observed during the docking of compound 11 in the active site of COX-2, compounds 7 and 9 are devoid of such interactions.

The role of COX-2 in the propagation of cancer¹² and the evaluations of several COX-2 inhibitors for reducing cancer propagation led us to investigate the present



Figure 2. Indomethacin docked in the active site of COX-2 exactly overlaps with the one present in the crystal structure of the enzyme.



Figure 3. Compound 10 docked in the active site of COX-2. OH and CN groups present on the lactone ring approach R120 at a distance of 2.1 and 2.43 Å, respectively. Hs are omitted for clarity.

compounds for their anti-cancer activities. The screening for anti-cancer properties was performed on 59 human tumour cell lines at NIH, Bethesda, USA, using the standard procedure.¹³ Out of the four compounds (7, 9-11), compounds 10 and 11 with CH₂CN and CH₂SCN group at C-5' exhibit remarkable anti-cancer activities with average GI₅₀ over all the cell lines as 1.9 and 9.1 µM, respectively, which is much better than the average GI₅₀ value of indomethacin $(64.3 \,\mu\text{M})^{14}$ (Table 2). Compound 10 is highly specific for all the cell lines of leukaemia, MALME-3M and M14 of melanoma, RXF393 of renal cancer, PC-3 of prostate cancer and MCF7, MDA-MB-435 of breast cancer where it exhibits GI₅₀ in sub-micromolar concentration (0.1- $0.8 \,\mu\text{M}$). Moreover, the relatively high LC₅₀ values of 10 and 11 show their non-toxicities. However, compounds 7 and 9, investigated at 10^{-5} M concentration only, do not show much anti-cancer activity. Therefore, in parallel with the COX-2 inhibitory activities, compounds 10 and 11 are also highly effective towards various cancer cell lines of human tumour cell panels.

Table 2. Anti-cancer activities of compounds 10 and 11 (average GI_{50})

Compound	GI ₅₀ (µM)	LC ₅₀ (µM)		
10	1.9	95		
11	9.1	74		
Indomethacin	64.3	100		

 Table 3. Lipinski values for compounds 7, 9–11

Compound	$\log P$	TPSA (Å ²)	nON	nOHNH
7	2.49	85.6	6	1
9	2.38	85.6	6	1
10	0.94	109.4	7	1
11	1.78	109.4	7	1

Calculations of Lipinski values of these compounds $(Table 3)^{15}$ indicate a difference in log *P* and Total Polar Surface Area (TPSA) of compounds **10** and **11** from **7** and **9** which might be contributing towards the difference in the bioactivities of these compounds.

In conclusion, we have constructed a γ -lactone moiety at C-3 of indole by the allylation of 3-indoleglyoxylate followed by iodocyclisation. Replacement of iodo-group with nucleophiles like CN, SCN, SC₂H₅ and their investigations for COX-2 inhibition have identified compounds **10** and **11** as highly potent and selective for COX-2. Along with COX-2 inhibition, the remarkable anti-cancer activities of **10** and **11** in comparison to indomethacin enable them to be used as leads for further investigations and also support the design of these molecules.

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References and notes

- Parry, R. J. In *Heterocyclic Compounds, Indole, Part Two*; Houlihan, W. J., Ed.; Wiley Interscience: New York, 1972; pp 2–9.
- (a) Gupta, L.; Talwar, A.; Chauhan, M. S. Curr. Med. Chem. 2007, 14, 1789; (b) Jiricek, J.; Blechert, S. J. Am. Chem. Soc. 2004, 126, 3534; (c) Yu, J.; Wearing, X. Z.; Cook, J. M. J. Am. Chem. Soc. 2004, 126, 1358; (d) Pindur, U.; Lemster, T. Curr. Med. Chem. 2001, 8, 1681.
- Shen, T. Y.; Ellis, R. L.; Windholz, T. B.; Matzuk, A. R.; Rosegay, A.; Lucas, S.; Witzel, B. E.; Stammer, C. H.; Wilson, A. N.; Holly, F. W.; Willett, J. D.; Sarett, L. H.; Holtz, W. J.; Risley, E. A.; Nuss, G. W.; Winter, C. A. J. Am. Chem. Soc. 1963, 85, 488.
- (a) Juby, P. F.; Hudyma, T. W. J. Med. Chem. 1969, 12, 396; (b) Glamkowski, E. J.; Gal, G.; Sletzinger, M. J. Med. Chem. 1973, 16, 176; (c) Paris, G. Y.; Garmaise, D. L.; Cimon, D. G. J. Med. Chem. 1980, 23, 9; (d) Kramer, J. B.; Boschelli, D. H.; Connor, D. T.; Kostlan, C. R.; Flynn, D. L.; Dyer, R. D.; Bornemeier, D. A.; Kennedy, J. A.; Wright, C. D.; Kuipers, P. J. Bioorg. Med. Chem. Lett. 1992, 12, 1655; (e) Tammara, V. K.; Narurkar, M. M.; Crider, A. M.; Khan, M. A. Pharm. Res. 1993, 10, 1191.

- 5. After the identification of COX-2 as causing inflammation during arachidonic acid metabolism (a) Black, W. C.; Bayly, C.; Belley, M.; Chan, C.-C.; Charleson, S.; Denis, D.; Gauthier, J. Y.; Gordon, R.; Guay, D.; Kargman, S.; Lau, C. K.; Leblanc, Y.; Mancini, J.; Ouellet, M.; Percival, D.; Roy, P.; Skorey, K.; Tagari, P.; Vickers, P.; Wong, E.; Xu, L.; Prasit, P. Bioorg. Med. Chem. Lett. 1996, 6, 725; (b) Leblanc, Y.; Black, W. C.; Chan, C. C.; Charleson, S.; Delorme, D.; Denis, D.; Gauthier, J. Y.; Grimm, E. L.; Gordon, R.; Guay, D.; Hamel, P.; Kargman, S.; Lau, C. K.; Mancini, J.; Ouellet, M.; Percival, D.; Roy, P.; Skorey, K.; Tagari, P.; Vickers, P.; Wong, E.; Xu, L.; Prasit, P. Bioorg. Med. Chem. Lett. 1996, 6, 731; (c) Kalgutkar, A. S.; Crews, B. C.; Rowlinson, S. W.; Marnett, A. B.; Kozak, K. R.; Remmel, R. P.; Marnett, L. J. Proc. Natl. Acad. Sci. 2000, 97, 925; (d) Kalgutkar, A. S.; Marnett, A. B.; Crews, B. C.; Remmel, R. P.; Marnett, L. J. J. Med. *Chem.* **2000**, *43*, 2860; (e) Palomer, A.; Cabre, F.; Pascual, J.; Campos, J.; Trujillo, M. A.; Entrena, A.; Gallo, M. A.; Garcia, L.; David, M.; Espinosa, A. J. Med. Chem. 2002, 45, 1402; (f) Olgen, S.; Nebioglu, D. Il Farmaco 2002, 57, 677; (g) Kalgutkar, A. S.; Crews, B. C.; Saleh, S.; Prudhomme, D.; Marnett, L. J. Bioorg. Med. Chem. 2005, 13, 6810; (h) Khanna, S.; Madan, M.; Vangoori, A.; Banerjee, R.; Thaimattam, R.; Basha, S. K. J. S.; Ramesh, M.; Casturi, S. R.; Pal, M. Bioorg. Med. Chem. 2006, 14, 4820.
- Prasit, P.; Wang, Z.; Brideau, C.; Chan, C.-C.; Charleson, S.; Cromlish, W.; Ethier, D.; Evans, J. F.; Ford-Hutchinson, A. W.; Gauthier, J. Y.; Gordon, R.; Guay, J.; Gresser, M.; Kargman, S.; Kennedy, B.; Leblanc, Y.; Leger, S.; Mancini, J.; O'Neill, G. P.; Ouellet, M.; Percival, M. D.; Perrier, H.; Riendeau, D.; Rodger, I.; Tagari, P.; Therien, M.; Vickers, P.; Wong, E.; Xu, L.-J.; Young, R. N.; Zamboni, R.; Boyce, S.; Rupniak, N.; Forrest, M.; Visco, D.; Patrick, D. *Bioorg. Med. Chem. Lett.* 1999, *9*, 1773.
- 7. (a) Kaur, P.; Singh, P.; Kumar, S. *Tetrahedron* 2005, *61*, 8231; (b) Singh, P.; Mittal, A.; Kaur, P.; Kumar, S. *Tetrahedron* 2006, *62*, 1063.
- 8. Selected data for compound 7. ¹H NMR (300 MHz, CDCl₃) δ 2.36 (s, 3H, CH₃), 2.53 (dd, 1H, ²J = 13.2 Hz, ³J = 9 Hz, 1H of 4'-H), 3.04 (dd, 1H, ²J = 13.2 Hz, ³J = 5.4 Hz, 1H of 4'-H), 3.14 (br s 1H, OH, exchanges with D₂O), 3.37 (dd, 1H, ²J = 10.2 Hz, ³J = 7.5 Hz, 1H of CH₂I), 4.29–4.38 (m, 1H, ⁵/-H), 7.25–7.32 (m, 3H, ArH), 7.39 (t, 1H, J = 7.5 Hz, ArH), 7.53 (s, 1H, ArH), 7.78 (t, 3H, J = 8.1 Hz, ArH), 8.01 (d, 1H, J = 8.7 Hz, ArH). The multiple signals at δ 4.29–4.38 on decoupling converted all the four double doublets into doublets. MS (FAB) 511 [M⁺]. Compound 8. ¹H NMR (300 MHz, CDCl₃) δ 1.60 (br s, 1H, OH, exchanges with D₂O), 2.32 (dd, 1H, ²J = 14.1 Hz, ³J = 8.1 Hz, 1H of 4'-H), 3.26 (dd, 1H, ²J = 10.5 Hz, ³J = 7.5 Hz, 1H of CH₂I), 3.41 (dd, 1H, ²J = 10.5 Hz, ³J = 4.5 Hz, 1H of CH₂I), 4.77–4.86 (m, 1H, 5'-H), 7.24–7.30 (m, 3H, ArH), 7.38 (d, 1H, J = 8.4 Hz), 7.65 (d, 1H,
- J = 7.8 Hz, ArH), 7.76 (s, 1H, ArH), 7.81 (d, 2H, J = 8.4 Hz, ArH), 8.02 (d, 1H, J = 8.1 Hz, ArH). MS (FAB) 511 (M⁺).
- Singh, P.; Mittal, A.; Kaur, S.; Kumar, S. Bioorg. Med. Chem. 2006, 14, 7910.
- Navidpour, L.; Shafaroodi, H.; Abdi, K.; Amini, M.; Ghahremani, M. H.; Dehpour, A. R.; Shafiee, A. *Bioorg. Med. Chem.* 2006, 14, 2507.
- 11. Crystal structure of COX-2 with indomethacin in the active site was downloaded from protein data bank (pdb

ID 4COX) and refined as per the usual procedure. The dockings were carried out using 'Dock in active site' module of BioMed CaChe 7.0.5.85.

- (a) Meric, J.-B.; Rottey, S.; Olaussen, K.; Soria, J.-C.; Khayat, D.; Rixe, O.; Spano, J.-P. *Crit. Rev. Oncol.*/ *Hepat.* 2006, 59, 51; (b) Thun, M. J.; Henley, S. J.; Patrono, C. J. Natl. Cancer Inst. 2002, 94, 252; (c) Taketo, M. M. J. Natl. Cancer Inst. 1998, 90, 1609.
- (a) Ally, M. C.; Scudiero, D. A.; Monks, P. A.; Hursy, M. L.; Czerwinski, M. J.; Fine, D. A.; Abbott, B. J.; Mayo, J. G.; Showmaker, R. H.; Boyd, M. R. *Cancer Res.* **1988**, 48, 589; (b) Grever, M. R.; Schepartz, S. A.; Chabner, B. A. *Semin. Oncol.* **1992**, 19, 622; (c) Boyd, M. R.; Paull, K. D. *Drug Dev. Res.* **1995**, 34, 91.
- 14. NIH data base, NSC 77541.
- 15. Lipinski values were calculated using molinspiration.