RSC Advances

PAPER

Cite this: RSC Adv., 2014, 4, 14513

Received 29th January 2014 Accepted 10th March 2014 DOI: 10.1039/c4ra00849a

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Introduction

The double helix is ubiquitous in biological molecules and its sophisticated structure is often closely related to its biological function.¹ Inspired by nature, chemists have great interest to prepare artificial molecules with double helical structure.²⁻⁸ Self-assembly is the most commonly used approach in which non-covalent interactions, such as electrostatic,^{9,10} metal-ligand,^{7,8,11-16} π - π stacking¹⁷⁻¹⁹ and hydrogen bonding,²⁰⁻²⁴ drive the two strands to intertwine in the formation process. These interactions of the two strands can come from motifs that are incorporated into the strands during synthesis²⁰ or from a third component, like a metal ion, which induces the interactions.^{7,8}

Important to both biological and chemical process,^{25,26} anions can form hydrogen-bond and this hydrogen-bonding interaction has been shown to play curial role in the formation of many supramolecular system^{27,28} which include double helices.²⁹⁻³² In the double helix examples, chloride, fluoride or sulfate, are locate at the helical axis, and form multiple strong N-H…Cl, N-H…F and N-H…O hydrogen

Acid-induced formation of hydrogen-bonded double helix based on chiral polyphenyl-bridged bis(2,2'-bipyridine) ligands[†]

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A series of chiral polyphenyl-bridged bis(2,2'-bipyridine) ligands comprising one to four phenyl units were synthesized. The ligands give a weak signal in the CD spectra, but upon addition of tetrachloroferric acid or perchloric acid, a more intense CD signal is observed for ligands having two or more phenyl units. Titration experiments show that the CD signal comes from a monoprotonated species which gives broadened and upfield shifted ¹H NMR signals. Variable temperature NMR experiments split the broadened signals into two sets of signals when the temperature is decreased. One of the sets is remarkably upfield while the other has chemical shifts similar to those of the free ligand. The X-ray crystal structures of a free ligand (mono phenyl), a monoprotonated ligand (biphenyl) and a biprotonated ligand (tetraphenyl) were obtained and the structure of the monoprotonated ligand shows that it is a double-stranded helix, which is stabilized by interior hydrogen bonding between the pyridinium N–H and the pyridine N of another ligand strand, and exterior CH…Cl hydrogen bonding between FeCl₄⁻ and the two ligand strands. Theoretical DFT calculations show that there is such stabilization in solution as well. With the perchlorate anion, the helix formation process is reversible with Et₃N which accompanies an on/off CD signal change.

bonds, respectively, to both ligand strands to stabilize the double helices.

Proton, the smallest cation, can interact with the lone pair electron of a ligand strand and lead to hydrogen bonds. Huc et al.33 and Aida et al.34 have both reported the use of proton to induced formation of single-stranded helices, however, to the best of our knowledge, double helix formation induced by proton is not known. We have previously reported the synthesis of Mn double-stranded helicates with mono- to triphenyl-bridged bis(2,2'-bipyridine) L1-3 ligands.³⁵ Herein, together with a tetraphenyl-bridged bis(2,2'-bipyridine) L4, we report the synthesis of this series of ligands. Upon protonation, ligands L2-4 give intense CD signal in the presence of FeCl₄⁻ or ClO₄⁻ anion. CD and NMR titration experiments suggest that a monoprontonated ligand species is responsible for the intense CD signals and X-ray crystal structures of monoprotonated L2 shows that a doublestranded helix is responsible for the signal.

Experimental

Chemicals and starting materials

Solvents used for synthesis were of analytical grade. All starting chemicals were of reagent-grade quality and were obtained commercially and used as received without further purification. Synthesis of chiral bromobipyridine **1** was reported previously.³⁶

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[†] Electronic supplementary information (ESI) available. CCDC 983706–983708. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c4ra00849a

Physical measurements and instrumentation

¹H, COSY and NOESY NMR spectra were recorded on Bruker 400 MHz instrument. The ¹H and chemical shift was referred to TMS as reference. Electrospray (ESI) mass spectra were measured by a PE SCIEX API 150 EX system. CD spectra were recorded on a Biokin MOS-450 instrument with a 1 mm cell.

Crystal structure determination

For crystal structure of L1, data was collected at 293 K with an Oxford Diffraction Gemini S Ultra X-ray single crystal diffractometer using graphite monochromatized Mo-Ka radiation $(\lambda = 0.71073 \text{ Å})$. For structure of $[(L2)_2H_2](ClO_4)_2$ and [(L4)H₂(Cl)](FeCl₄), data were collected at 133 K with an Oxford Diffraction Gemini S Ultra X-ray single crystal diffractometer using graphite monochromatized Cu-K α radiation ($\lambda = 1.54178$ Å). All collected frames were processed with the software SAINT, and absorption correction was applied (SAD-ABS) to the collected reflections. The structure of the complex was solved by direct methods (SHELXTL) in conjunction with standard difference Fourier syntheses. All non-hydrogen atoms were assigned with anisotropic displacement parameters. The hydrogen atoms were generated in their idealized positions and allowed to ride on the respective carbon atoms. Crystal data of L1. $C_{80}H_{76}N_8O$, M = 1165.49, orthorhombic, a = 10.0224(4), b = 13.5194(6), c = 24.219(1) Å, U = 3281.6(2) Å³, space group $P2_12_12_1$, Z = 4, 10071 reflections measured, 5493 unique $(R_{\rm int} = 0.0228)$ which were used in all calculation. The final $wR(F_2)$ was 0.0804 (all data). Crystal data of $[(L2)_2H_2](FeCl_4)_2$. $C_{96}H_{94}Cl_8Fe_2N_8O_{1.75}, M = 1783.09, monoclinic, a = 30.7135(9),$ $b = 21.8661(6), c = 15.6061(5) \text{ Å}, U = 10290.2(5) \text{ Å}^3$, space group C2, Z = 4, 24178 reflections measured, 14245 unique $(R_{\rm int} = 0.0346)$ which were used in all calculation. The final $wR(F_2)$ was 0.1416 (all data). Crystal data of $[(L2)_2H_2](FeCl_4)_2$. $C_{96}H_{94}Cl_8Fe_2N_8O_{1.75}, M = 1783.09$, monoclinic, a = 30.7135(9), b = 21.8661(6), c = 15.6061(5) Å, U = 10290.2(5) Å³, space group C2, Z = 4, 24178 reflections measured, 14245 unique ($R_{int} =$ 0.0346) which were used in all calculation. The final $wR(F_2)$ was 0.1416 (all data). Crystal data of $[(L4)H_2](Cl)(FeCl_4)$. $C_{59}H_{56}Cl_5FeN_4O$, M = 1070.18, monoclinic, a = 11.4756(3), $b = 17.6487(5), c = 26.0159(7) \text{ Å}, U = 5182.0(2) \text{ Å}^3$, space group $C12_11$, Z = 4, 12966 reflections measured, 9565 unique $(R_{\rm int} = 0.0259)$ which were used in all calculation. The final $wR(F_2)$ was 0.099 (all data).

DFT calculations

All calculations were done at M06-2X (hybrid meta exchangecorrelation functional with double the amount of nonlocal exchange) functional³⁷ using LanL2DZ basis set for Fe,³⁸⁻⁴⁰ 6-31G(d) basis sets for H, C, N, and 6-31 + G(d) basis sets for O, Cl atoms. The solvent effect is taken account by the Polarizable Continuum Model.^{41,42} Atom-in-molecule (AIM) analysis is performed with AIM2000 program.⁴³ The wavefunction was taken from the optimized structure at the M06-2X level with using LanL2DZ basis set for Fe and 6-31G(d) basis set for all nonmetal elements.

Procedure for synthesis of 2 and 3

A solution of 1 (1.45 mmol) and tetrakis(triphenylphospine) palladium(0) (0.06 mmol) in degased toluene (6 ml) was treated with a solution of sodium carbonate (2.0 mmol) in H₂O (3 ml). A methanolic solution (3 ml) of 3-chlorophenylboronic acid (1.45 mmol) or 3-bromophenylboronic acid (1.19 mmol) was added. The mixture was stirred at 80 °C for 3 h under nitrogen. After cooling to room temperature, aqueous NH₃ (30 ml) was added and the mixture was extracted by CH₂Cl₂. The combined organic layers were dried by MgSO4. Solvent was removed under reduced pressure. The crude product was purified by column chromatography (petroleum ether : ethyl acetate = 10:1). Products were isolated as white solid in 95% and 83% yield for 2 and 3, respectively. ¹H NMR of 2 (300 MHz, $CDCl_3$) δ 8.94 (d, J =7.8 Hz, 1H), 8.60 (s, 1H), 8.49 (s, 1H), 8.05 (m, 2H), 7.95 (m, 1H), 7.85 (d, J = 8 Hz, 1H), 7.42 (m, 2H), 3.29 (d, J = 2.4 Hz, 2H), 3.02 (t, J = 5.4 Hz, 1H), 2.8 (m, 1H), 2.41 (s, 1H), 1.43 (s, 3H), 1.23 (d, J)= 10.3 Hz, 1H), 0.64 (s, 3H). ¹H NMR of 3 (300 MHz, CD_2Cl_2): δ 8.37 (s, 1H), 8.35 (d, 1H, J = 8.1 Hz), 8.31 (t, 1H, J = 1.8 Hz), 8.22 (s, 1H), 8.02 (d, 1H, J = 7.8 Hz), 7.85 (t, 1H, J = 7.8 Hz), 7.67 (d, 1H, 6.9 Hz), 7.55 (d, 1H, J = 7.8 Hz), 7.36 (t, 1H, J = 7.8 Hz), 3.10 (d, 1H, J = 2.7 Hz), 2.88 (t, 1H, J = 5.4 Hz), 2.70 (m, 1H), 2.33 (m, 1H)1H), 1.42 (s, 3H), 1.21 (d, 1H, J = 6.9 Hz), 0.67 (s, 3H).

Procedure for synthesis of 4

A solution of 3 (11.6 mmol) and tetrakis(triphenylphospine) palladium(0) (5 mol%) in degased toluene (36 ml) was treated with a solution of sodium carbonate (20 mmol) in H₂O (16.5 ml). A methanolic solution (16.5 ml) of 3-chlorophenylboronic acid (11.6 mmol) was added. The mixture was stirred at 95 °C for 24 h under nitrogen. After cooling to room temperature, aqueous NH3 (50 ml) was added and the mixture was extracted by CH₂Cl₂. The combined organic layers were dried by MgSO₄. Solvent was removed under reduced pressure. The crude product was purified by column chromatography (petroleum ether : ethyl acetate = 10:1). Products were isolated as white solid with 93% yield. ¹H NMR (400 MHz, $CDCl_3$) δ 8.56 (d, J = 7.8 Hz, 1H), 8.49 (s, 1H), 8.33 (s, 1H), 8.29 (t, J = 1.5 Hz, 1H), 8.16 (d, J = 7.3 Hz, 1H), 7.98 (t, J = 7.8 Hz, 1H), 7.85 (d, J = 7.2 Hz, 1H), 7.70 (t, J = 1.7 Hz, 1H), 7.66 (d, J = 7.7 Hz, 1H), 7.63 (d, J = 7.6 Hz, 1H), 7.59 (d, J = 7.6 Hz, 1H), 7.43 (t, J = 7.7 Hz, 1H), 7.38 (d, J = 8.0 Hz, 1H), 3.18 (d, J = 2.5 Hz, 2H), 2.96 (t, J = 5.4 Hz, 1H), 2.8 (m, 1H), 2.4 (m, 1H), 1.46 (s, 3H), 1.27 (t, J = 8.2 Hz, 1H), 0.69 (s, 3H).

Procedure for synthesis of L1 and L3

Degas toluene (16 ml), MeOH (6 ml) and water (6 ml) were added to a flask containing Pd(PPh₃)₄ (0.17 g, 3 mol%), **1** or **3** (4.4 mmol), **1**,3-phenyldiboronic acid (0.34 g, 2 mmol) and sodium carbonate (0.92 g). The mixture was heated at 80 °C for 24 h. It was cooled to room temperature. Ammonia solution in saturated Na₂CO₃ solution was added and the solution was stirred for 5 min. It was extracted by CH₂Cl₂. Solvent was dried under vacuo, and the compound was purified by column chromatograph with solvent (dichloromethane : diethyl ether = 10 : 1). The crude yellow solution can be further purified by washing with MeCN. Products were isolated as pale yellow solid with 57% and 30% yield for L1 and L2, respectively. ¹H NMR of L1 (400 MHz, $CDCl_3$): δ 8.81 (t, 1H, J = 1.6 Hz), 8.85 (s, 2H), 8.38 (d, 2H, J = 6.7Hz), 8.27 (d, 2H, J = 7.8 Hz), 8.25 (s, 2H), 7.93 (t, 2H, J = 7.7 Hz), 7.87 (d, 2H, J = 6.8 Hz), 7.69 (t, 1H, J = 7.7 Hz) 3.11 (d, 4H, J = 2.6 Hz), 2.90 (t, 2H, J = 5.4 Hz), 2.73 (m, 2H), 2.34 (m, 2H), 1.44 (s, 6H), 1.27 (d, 2H, J = 9.6 Hz), 0.69 (s, 6H). ¹³C NMR (CDCl₃): δ 156.65, 156.61, 154.86, 145.68, 145.57, 143.30, 140.30, 140.36, 137.86, 129.38, 127.93, 125.80, 120.90, 120.42, 119.50, 44.82, 40.38, 39.54, 33.30, 32.08, 26.28, 21.66. ¹H NMR of L3 (300 MHz, $CDCl_3$): δ 8.43 (m, 4H), 8.36 (d, 2H, J = 6 Hz), 8.22 (s, 2H), 8.16 (d, 2H, J = 6 Hz), 8.05 (t, 1H, J = 1.8 Hz), 7.90 (t, 2H, J = 7.8 Hz),7.81 (d, 2H, J = 7.8 Hz), 7.75 (t, 4H, J = 7.5 Hz), 7.63 (t, 3H, J = 7.5 Hz), 3.05 (d, 4H, I = 2.7 Hz), 2.88 (t, 2H, I = 5.4 Hz), 2.70 (m, 2H), 2.30 (m, 2H), 1.41 (s, 6H), 1.21 (d, 2H, J = 6.9 Hz), 0.65 (s, 6H). ¹³C NMR (CDCl₃): δ 156.62, 156.54, 154.75, 145.65, 145.58, 143.31, 142.15, 141.91, 140.53, 137.87, 129.52, 129.43, 128.09, 126.67, 126.44, 126.26, 120.88, 120.35, 119.48, 44.80, 40.37, 39.49, 33.23, 32.06, 26.25, 21.63.

Procedure for synthesis of L2

To a solution of NiCl₂·6H₂O (1.2 mmol) in degassed DMF (15 ml) at 70 °C under nitrogen, triphenylphosphine (4.8 mmol) was added to give a blue solution. Zinc powder (2.6 mmol) was then added and the resulting mixture was stirred for an hour, in which dark-brown mixture was formed. **2** (1 mmol) in degassed DMF (5 ml) was added slowly and the mixture stirred at 70 °C for 72 hours. The mixture was then allowed to cool to room temperature and 25% aqueous NH₃ (25 ml) was added. The layers were separated, and the aqueous layers were extracted with CH₂Cl₂. The solution was dried with MgSO₄ and solvent was removed under reduced pressure. Crude product was purified by column chromatography. Product was isolated as a pale yellow solid. Yield: 37%. ¹H NMR of L2 (300 MHz, CDCl₃): δ 8.43 (m, 2H), 8.37 (d, 2H, *J* = 7.5 Hz), 8.22 (s, 1H), 8.18 (d, 1H, *J* = 8.1 Hz), 7.90 (t, 1H, *J* = 7.8 Hz), 7.83 (d, 1H, *J* = 7.2 Hz), 7.78 (d,

1H, J = 7.8 Hz), 7.64 (t, 1H, J = 7.5 Hz), 3.05 (d, 2H, J = 2.7 Hz), 2.88 (t, 1H, J = 5.4 Hz), 2.70 (m, 1H), 2.31 (m, 1H), 1.42 (s, 3H), 1.21 (d, 1H, J = 6.9 Hz), 0.66 (s, 3H). ¹³C NMR (CDCl₃): δ 156.43, 156.38, 154.63, 145.46, 145.39, 143.11, 140.13, 137.68, 129.18, 127.73, 125.57, 120.70, 120.23, 119.28, 44.56, 40.12, 39.30, 33.07, 31.85, 26.05, 21.44.

Procedure for synthesis of L4

To a solution of NiCl₂·6H₂O (3.8 mmol) in degassed DMF (17 ml) at 70 °C under nitrogen, triphenylphosphine (16 mmol) was added to give a blue solution. Zinc powder (6.4 mmol) was then added and the resulting mixture was stirred for an hour, in which dark-brown mixture was formed. Compound 4 (3.2 mmol) in degassed DMF (9 ml) was added slowly and the mixture stirred at 70 °C for 72 hours. The mixture was then allowed to cool to room temperature and 25% aqueous NH₃ (100 ml) was added. The layers were separated, and the organic layers were washed with CH₂Cl₂. The solution was dried with MgSO₄ and solvent was removed under reduced pressure. Crude product was purified by column chromatography (dichloromethane : *n*-hexane : diethyl ether = 30:6:1). Product was isolated as a pale brown solid in 26% yield. ¹H NMR (400 MHz, CDCl₃) δ 8.42 (d, J = 2.8 Hz, 1H), 8.37 (d, J = 7.6Hz, 1H), 8.22 (s, 1H), 8.16 (d, J = 7.3 Hz, 1H), 8.02 (s, 1H), 7.90 (t, J = 7.8 Hz, 1H), 7.80 (d, J = 7.7 Hz, 1H), 7.74 (t, J = 7.2 Hz, 1)2H), 7.62 (d, J = 13.8, 7.6 Hz), 3.06 (d, J = 2.3 Hz, 1H), 2.89 (t, J = 5.4 Hz, 1H), 2.7 (m, 1H), 2.32 (s, 1H), 1.43 (s, 2H), 1.25 (d, J = 9.8 Hz, 1H), 0.67 (s, 2H). ¹³C NMR (CDCl₃): δ 156.40, 156.33, 254.57, 145.46, 125.40, 143.12, 141.98, 141.84, 141.65, 140.31, 137.71, 129.28, 127.90, 126.54, 126.43, 126.30, 126.05, 120.72, 120.19, 119.33, 44.57, 40.14, 39.28, 33.02, 31.87, 23.06, 21.45.

Procedure for reversibility experiment

To a solution of L (L2–4, 3×10^{-4} M in CH₂Cl₂) in a 3 mm cell, 1 equiv. of HClO₄ was added, and the CD spectrum of the solution was obtained. After that, 1.2 equiv. of NEt₃ was added to the



Scheme 1 Preparation of polyphenyl-bridged bis(2,2'-bipyridine) L1-4.

above solution to fully restore the original spectrum. The experiments were repeated by following the sequence, and the absorption at 334 nm which is the maximum of the induced CD signals was plotted.

Results and discussion

Synthesis of polyphenyl-bridged bis(2,2'-bipyridine) ligands

Chiral polyphenyl-bridged bis(2,2'-bipyridine) ligands L1-4 with pinene-based chiral substituents at the 4,5-position of terminal pyridine rings were prepared from bipyridine intermediate 1, which was obtained from reported Kröhnke condensation between of pyridinium iodide and a, \beta-unsaturated ketone.36 The Pd-catalyzed Suzuki coupling between 1 and *m*-phenylenediboronic acid with Na₂CO₃ in a mixture of H₂O, MeOH and toluene yielded the monophenyl-bridged L1 in 57% yield. For the synthesis of biphenyl-bridged L2, Pd-catalyzed Suzuki coupling between 1 and 3-chlorophenylboronic acid resulted in the chlorophenylbipyridine intermediate 2. Ni(0)mediated homocoupling of 2 in DMF gave L2 in 37% yield. For the synthesis of triphenyl-bridged L3, the reaction between 2 and *m*-phenylenediboronic acid with $Pd(PPh_3)_4$ as catalyst was first tried, but no reaction was observed. Then, bromophenylbipyridine intermediate 3, prepared by Pd-catalyzed Suzuki coupling between 1 and 3-bromophenylboronic acid, was employed. A sub-stoichiometric amount of 3-bromophenylboronic acid was used to reduce the formation of the bromobiphenylbipyridine and Suzuki coupling between 3 and mphenylenediboronic acid yielded L3 in 30% yield. For the synthesis of tetraphenyl-bridged L4, intermediate 4, synthesized by the reaction between 3 and 3-chlorophenylboronic acid with Pd(PPh₃)₄ as catalyst, was used. Ni(0)-mediated homocoupling of 4 yielded L4 in 26% yield (Scheme 1).

Effect of acid on the CD spectra of polyphenyl-bridged bis-(2,2'-bipyridine) ligands

Although L1-4 contains chiral substitution at the 4,5-position of the terminal pyridine rings, they gave only a weak CD absorption. Fig. 1a-d show the effect of different acids on the CD spectrum of the ligands. The phenyl-bridged L1 does not show much effect with addition of acid. Only very small change in the CD spectrum of L1 is observed (Fig. 1a). However, L2-4 show much greater change with addition of some acid. Fig. 1b shows the results obtained with the biphenyl-bridged L2. Addition of HFeCl₄ gives an intense induced CD signal absorption at 334 nm with $\Delta \epsilon = 6.0 \text{ M}^{-1} \text{ cm}^{-1}.^{44} \text{ HClO}_4$ and HOTf give similar change but with weaker intensity, $\Delta \varepsilon = 4.0$ and 2.0 M⁻¹ cm⁻¹, respectively. Fig. 1c shows the results obtained with the triphenyl-bridged L3. Similar to the results of L2, addition of HFeCl₄, HClO₄ and HOTf to L3 leads to induced CD signals at 333 nm with $\Delta \varepsilon = 10.2$, 4.3, and 3.0 M⁻¹ cm⁻¹, respectively. These signals have larger intensity when compared to L2. Fig. 1d shows the results obtained with the tetraphenyl-bridged L4. Intense induced CD signals with $\Delta \varepsilon = 9.0, 5.0, \text{ and } 1.7 \text{ M}^{-1}$ cm⁻¹ are observed at 330 nm with addition of HFeCl₄, HClO₄ and HOTf, respectively, which are comparable to L3. The trend



Fig. 1 CD spectra of L1–4 (3 \times 10 $^{-4}$ M) with addition of different acids (1 equiv.).

of HFeCl₄ giving the strongest absorption, followed by $HClO_4$ and then HOTf, is very similar to L2 and L3. With the additions of acids other than HFeCl₄, $HClO_4$ and HOTf, the CD spectra of L2–4 give very minimal change.



Fig. 2 ESI-MS spectrum of a CH_2Cl_2 solution of L1–4 with addition of HFeCl4.

When comparing the polyphenyl-bridged ligands, much stronger acid-induced CD spectral change are observed with L2– 4 than L1 and it seems to suggest that L2–4 give similar species











Fig. 5 CD spectrum of L4 in CH₂Cl₂ (3 \times 10⁻⁴ M) with addition of HFeCl₄ (0 to 4 equiv.). The inset shows the change in absorption at 332 nm.

upon protonation. For L2–4, the CD signal change does not seem to follow the trend of the pK_a value; for example, HClO₄ ($pK_a = -10.0$) give a more intense signal than HOTf ($pK_a =$ -14.0). Other acids like HOAc ($pK_a = -4.8$), HCl ($pK_a = -8.0$), HNO₃ ($pK_a = -1.3$), H₂SO₄ ($pK_a = -3.0$) give only very small or no CD change. These results suggest that the CD signal change is not triggered by protonation alone. Anion seems to have a role in the change of the CD signal as well.

ESI-MS study

With HFeCl₄ giving the largest CD signals, the ESI-MS spectra of the ligands with 1 equiv. of HFeCl₄ were obtained for L1–4 $(3 \times 10^{-4} \text{ M})$. The spectrum with L1 is shown in Fig. 2a. The spectrum show peaks at *m*/*z* 575.8 and 1151.6, which can be



Fig. 6 1 H NMR spectra (400 MHz) of (a) L2 (3 × 10⁻⁴ M) in CD₂Cl₂ with addition of (b) 1 equiv. (c) 2 equiv. (d) 3 equiv. (e) 4 equiv. of HClO₄.



Fig. 7 1 H NMR spectra for (a) L2 (2 × 10⁻² M) in CD₂Cl₂ with addition of (b) 0.5 equiv. (c) 1.0 equiv. (d) 1.5 equiv. (e) 2.0 equiv. of HClO₄.



Fig. 8 1 H NMR spectra for (a) L3 (3 \times 10⁻⁴ M) in CD₂Cl₂ with addition of (b) 0.5 equiv. (c) 1 equiv. (d) 2 equiv. (e) 3 equiv. of HClO₄.

assigned to $[(L1)H]^+$ and $[(L1)_2H]^+$, respectively, indicating protonation of L1 by HFeCl₄. Fig. 2b shows the spectrum with L2. Similar to L1, peaks corresponding to $[(L2)H]^+$ and $[(L2)_2H]^+$ are observed, however, in addition to these peaks, a peak at 1500.8 which can be assigned to $[(L2)_2H_2](FeCl_4)^+$ is also observed. This formula indicates that a dimeric form of monoprotonated L2 may have been formed. The spectrum with L3, which is shown in Fig. 2c, has peaks at m/z 729.1, 1452.4 which corresponds to $[(L3)H]^+$ and $[(L3)_2H]^+$, respectively. Again, the spectrum of L3 shows peak corresponded to a dimeric species, $[(L3)_2H_2](FeCl_4)^+$, at 1654.3. For L4, the spectrum show peaks corresponded to $[(L4)H]^+$, $[(L4)_2H]^+$, and $[(L4)_2H_2](FeCl_4)^+$ at m/z 805.7, 1606.4, and 1806.5 respectively (Fig. 2d). The signals of $[(L)_2H_2](FeCl_4)^+$ (L = L2-4) observed in the spectra suggest the presences of dimeric species $[(L)_2H_2](FeCl_4)_2$, but this species is not observed with L1.





Acid titration monitored by CD

In order to have a better understanding of the change in CD signals, titration experiments were carried out. As shown in Fig. 3, addition of various concentration of HFeCl_4 into a 3×10^{-4} M solution of L2 first give positive absorption, then decrease in absorption. The inset shows the change in intensity of the absorption at 334 nm. The intensity of the signal reaches maximum when there is 1 equiv. of HFeCl_4 . Further addition decreases the signal intensity, and finally the positive CD absorption completely disappeared when 3 equiv. of HFeCl_4 is added. A similar trend in the



Fig. 10 NOESY spectrum (400 MHz) of L2 in CD₂Cl₂.



Fig. 11 NOESY spectrum (400 MHz) of L2 with addition of 2 equiv. of $HClO_4$ in CD_2Cl_2 .

change of CD signals was also observed in the titration experiments of L3 and L4. Fig. 4 and 5 show the change of the intensity of the CD signals with concentration of HFeCl₄. The signal reach maximum when there is 1 equiv. of HFeCl₄, and the intensity of signal decreases with further increase of HFeCl₄. These titration experiments suggest that the CD signals are originated from a monoprotonated state, and the acid-induced signal is lost upon further protonation.

Acid titration monitored by NMR

With the paramagnetic nature of HFeCl₄, NMR informations were obtained by using HClO₄. With L2-4, titration experiments were carried out under the same concentration as the CD experiments. Fig. 6 shows the results with L2. With addition of HClO₄, there is an immediate upfield shift and broadening of the ¹H NMR signals. The most upfield and broadened signals appear when 2 equiv. of HClO₄ is added. Further addition of HClO₄ leads to a downfield shift and sharpening of the signals. Sharpened signals with chemical shift similar to the unprotonated L2 is obtained when more acid is added. However, unlike CD titration where the concentration is limited, the NMR titration can be carried out at a much higher concentration. Fig. 7 shows the spectra when the experiment is carried out at 2 $\times 10^{-2}$ M of L2, only 1 equiv. of HClO₄ is needed to lead to the upfield signals. Fig. 8 and 9 are the spectra of titrations with L3 and L4 which also show similar change, but in both of this case only one equiv. of HClO₄ is needed for reaching the most upfield signal at the CD experiments concentration. These results suggest that the upfield signals may come from the monoprotonation of the polyphenyl-bridged bis(2,2'-bipyridine) ligands. By considering that the induced CD signals and the upfield ¹H signals occurred at the same time, we believe that they come from the same protonated species.

¹H NMR assignment

To have a better understanding of the species, 2D NMR was carried out. The NOESY spectrum of L2 is shown in Fig. 10. The correlation signals between the pyridine proton H^5 and both protons at the 2- and 6-position of the bridging phenyl ring, H^6 and H^9 , suggests that L2 interconverts between two conformations with H^5 syn either to H^6 or H^9 . Although broadened signals observed with addition of 1 equiv. of HClO₄ cannot be assigned, the sharpened signals observed at 2 equiv. of HClO₄ were fully assigned. The *syn*-conformation of the mono-protonated 2,2′- bipyridine is well established by both X-ray crystal structure and



Fig. 12 ¹H NMR spectra for L2 (3×10^{-4} M) in CD₂Cl₂ with HClO₄ (2 equiv.) at variable temperature. The species giving upfield and downfield signals are labelled \Box and \bigcirc , respectively.



Fig. 13 ¹H NMR spectra for L3 (3 × 10⁻⁴ M) in CD₂Cl₂ with HClO₄ (1 equiv.) at variable temperature. The species giving upfield and downfield signals are labelled \square and \bigcirc , respectively.



Fig. 14 ¹H NMR spectra for L4 (3 \times 10⁻⁴ M) in CD₂Cl₂ with HClO₄ (1 equiv.) at variable temperature. The species giving upfield and downfield signals are labelled \square and \bigcirc , respectively.

theoretical calculation.^{45,46} The correlation signal between H² and H³ observed in the NOESY spectrum (Fig. 11) suggests that the pyridyl rings of both bipyridine units are in *syn*-conformation which is consistence with the mono-protonation of the bipyridine units. By considering that proton at the 2-position of the phenyl ring, H⁹, gives correlation signals to both protons at the 4-position of the phenyl ring, H⁸, and the 5'-position of the bipyridine, H⁵, a biprotonated species with a linear conformation is proposed. This biprotonated species comes from the further protonation of the species giving the upfield signals, which is consistent with the suggestion that the upfield signal is most likely a monoprotonated species of **L2**.

Low temperature ¹H NMR study

Apart from the upfield shift, broadening of signals of were also observed in NMR spectra of L2-4 when HClO₄ was added.

Variable temperature NMR experiments were then carried out to study the broadened signals. Fig. 12-14 shows the effect of the temperature on the ¹H NMR signals. Fig. 12 shows that the broadened signals of L2 with 2 equiv. of HClO₄ become even broader when the sample is cooled to 0 °C. Further decrease in temperature leads to sharpening and splitting of the signals into two sets with one set significantly upfield shifted while the other set shifted downfield to chemical shift similar to that of the free ligand. Fig. 13 shows the spectra obtained with L3 and 1 equiv. of HClO₄. Similar signals splitting is observed, but it starts at higher temperature of 0 °C. The most sharpened signals are observed at -40 °C. Fig. 14 shows that signals of L4 with 1 equiv. HClO₄ also split into in two sets at 0 °C. The lineshape of the signals does not change much when the temperature is below -30 °C. These results suggest that the broadening of signals observed with L2-4 and HClO₄ come from the exchange between an upfielded and a downfielded species. The



Fig. 15 The ORTEP plot of crystal structure of L1. Hydrogen atoms are removed for clarity.

differences in temperature in giving the splitting and change in line-shape of the signals suggest that the exchange rates are different. Analysis of the linewidth of the signals at temperature below coalescence give the free energy of activation for the exchange process^{47,48} of 13.0, 13.4, and 14.0 kcal mol⁻¹ for L2, L3 and L4, respectively. By considering that the signals broadening and the upfield shift of signals occur at the same time, we believe that the exchanges involve monoprotonated species of L2–4.

After the variable temperature experiments, the NMR solutions were then characterized by ESI-MS at room temperature (Fig. S1[†]). The spectra show similar results to that of HFeCl₄ as both $[(L)_2H_2](ClO_4)^+$ and $[(L)H]^+$ were observed. These monoprotonated species can be considered monomer and dimer. We



Fig. 16 Capped stick drawing of X-ray crystal structure of (a) P- and (b) M- [(L4)H₂](Cl)(FeCl₄). Dotted lines show hydrogen bonding with the chloride. The FeCl₄⁻ ion is not shown for clearance. (c) and (d) are the spacefilling model of P- and M-[(L4)H₂](Cl)(FeCl₄) respectively.



Fig. 17 (a) Capped stick drawing of X-ray crystal structure of $[(L2)_2H_2](FeCl_4)_2$. Dotted lines show hydrogen bonding within the double helix. (b) Spacefilling model of X-ray crystal structure of $[(L2)_2H_2](FeCl_4)_2$.

believe that the broadening of signals may be due to the exchange between the dimer and its monomeric form, and the stacking between the strands of the dimeric species is the reason that leads to anisotropic effects of the aromatic rings and upfield signals.

X-ray characterization

Single crystals suitable for X-ray analysis were obtained for the nonprotonated state of L1, the biprotonated state of L4, and the dimeric form of the monoprotonated L2. Crystals of L1 were obtained by slowly evaporation of a diethyl ether solution. It is crystalized in a $P2_12_12_1$ space group. Fig. 15 shows the ORTEP diagram. The pyridine rings adopt a transoid geometry and L1 is not coplanar. Twisting is observed between the aromatic rings with the torsional angles between the pyridyl rings being 2.8° and 19.1°, and between the phenyl and pyridyl rings being 22.7° and 24.8°. The ligand is not long enough to have a complete helical turn, and there is no stacking interaction.

Single crystals of the biprotonated L4 were obtained by ether diffusion into a methanol and chloroform solution of L4 and HFeCl₄. The compound crystallized in a P12₁1 space group. Although the mixture was prepared using a one to one molar ratio of L4 and HFeCl₄, the biprotonated species of L4 is a structure with a chloride and a tetrachloroferrate anions. The biprotonated L4 coils up around the chloride ion to give two helical structures, the P- and M-helices, which are coexist in the crystal lattice. The structures of P- and M-helices are shown in Fig. 16a-d. Beside the methyl groups of the P-form is pointing towards the chloride and the methyl groups of the M-form is pointing away from the chloride, the structures of the two helices are very similar. In both structures, the pyridyl rings adopt a cisoid geometry and point toward the chloride ion. The coiling of L4 comes from the twisting between the aromatic rings with most of it is contributed by the bridging phenyl rings. The dihedral angles between the phenyl rings are in the range 33.53–48.49°. The chloride anion has a close proximity with the terminal pyridine rings with N-H…Cl distances and angles in range 2.28–2.41 Å and 139.37–147.24°, respectively, which suggests the presence of hydrogen bonding. Although these



Fig. 18 Crystal structure of $[(L2)_2H_2](\text{FeCl}_4)_2$ showing crystal packing and location of FeCl_4^- anions surrounding the $[(L2)_2H_2]^{2+}$ core.

Table 1Hydrogen bonding parameters of $[(L2)_2H_2]^{2+}$ and surrounding $FeCl_4^-$ observed in X-ray crystal structure of $[(L2)_2H_2](FeCl_4)_2$

D−H···A	H···A/Å	$\mathbf{D}-\mathbf{H}\cdots\mathbf{A}/^{\circ}$
N1-H1…N5	1.987	156.14
N8-H8…N4	1.989	154.98
C-H77…Cl1	3.036	165.66
C-H11····Cl2	2.713	162.04
C-H14…Cl3	3.119	124.83
C-H78…Cl5	2.906	139.35
C-H79…Cl6	3.097	136.80
C-H16…Cl7	3.057	132.81
C-H71…Cl6	3.042	128.50
C-H72…Cl6	3.090	126.66
C-H67…Cl8	3.031	137.48
C-H32…Cl1	2.880	162.13
C-H62····Cl3	2.996	136.51
C-H65…Cl4	2.747	128.67
C-H27…Cl8	2.994	171.65
C-H20…Cl4	3.125	152.81
C-H21····Cl4	2.768	166.47

species have helical structures, by considering that the anions are different, and the coexistence of the P- and M-form, these species are not related to the CD signals observed in the CD experiments. In addition, these biprotonated species shows that the tetraphenyl-bridged L4 is long enough to give a complete helical turn, and there are stacking interactions between the terminal pyridine rings, however, no upfield signals shift is observed while the biprotonated species is formed in the ¹H NMR studies with HClO₄. It is not likely that this helical form of biprotonated L4 is presented when only ClO_4^- anion is presented.

Single crystals of the monoprotonated L2 was obtained by ether diffusion into a solution of L2 in the presence of 1 equiv. of HFeCl₄. It crystallizes in a monoclinic *C*2 space group and it is a dimeric form of formula, $[(L2)_2H_2](FeCl_4)_2$. The structure, shown in Fig. 17, has *syn* pyridine rings and the two protonated strands twist around each other resulting in the doublestranded helical structure with P-helical chirality (Flack



Fig. 19 Crystal structure of $[(L2)_2H_2](FeCl_4)_2$ showing crystal packing and location of $FeCl_4^-$ anions surrounding $[(L2)_2H_2]^{2+}$ core. Dotted lines shows some of the hydrogen bonding between $FeCl_4^-$ and the helix.



Fig. 20 Energy profile of the formation of $[(L2)_2H_2](X)_2$ ($X = ClO_4^-$ or FeCl₄⁻) obtained by calculation at M06-2X/6-31G(d)/LANL2DZ with solvent effect. The red line indicates the formation of $[(L2)_2H_2](ClO_4)_2$ and black line indicates the formation of $[(L2)_2H_2](FeCl_4)_2$. The relative energy is given in kcal mol⁻¹.

parameter = 0.026(7)). The two molecules of L2 are held tightly together by intermolecular hydrogen bonding between the pyridinium proton of one strand and pyridine ring of the other strand. The N–H…N distances are 1.987 and 1.989 Å, and the angles N-H…N are 154.14 and 156.98°. The helix is also stabilized by π - π stacking interactions. There are extensive stacking interactions between the aromatic rings of the two molecules which start from the stacking of the pyridinium of one strand with the forth aromatic ring of the other, and extend along the whole ligand strand. Fig. 18 shows the crystal lattice of $[(L2)_2H_2](FeCl_4)_2$. There are $FeCl_4^-$ anions surrounding the helix core. Some of the chloride atoms of the ${\rm FeCl_4}^-$ are close to the aromatic hydrogens of the helix, and their distances and angles are summarized in Table 1. The C-H…Cl distances and angles are in the range 2.713-3.125 Å and 124.83-171.65° respectively. These results suggest the presence of the C-H…Cl hydrogen bonding interactions between the helix and FeCl₄⁻ anions.⁴⁹⁻⁵¹ Fig. 19 shows some of these interactions.

Solution state structure by DFT calculation

From the crystal structure of $[(L2)_2H_2]$ (FeCl₄)₂, we believe that a double-stranded helix is formed in solution when L2 is



Fig. 21 Energy profile of the formation of $[(L3)_2H_2](X)_2$ (X = ClO_4^- or $FeCl_4^-$) obtained by calculation at M06-2X/6-31G(d)/LANL2DZ with solvent effect. The red line indicates the formation of $[(L3)_2H_2](ClO_4)_2$ and black line indicates the formation of $[(L3)_2H_2](FeCl_4)_2$. The relative energy is given in kcal mol⁻¹.



Fig. 22 Energy profile of the formation of $[(L4)_2H_2](X)_2$ (X = ClO_4^- or $FeCl_4^-$) obtained by calculation at M06-2X/6-31G(d)/LANL2DZ with solvent effect. The red line indicates the formation of $[(L4)_2H_2](ClO_4)_2$ and black line indicates the formation of $[(L4)_2H_2](FeCl_4)_2$. The relative energy is given in kcal mol⁻¹.

monoprotonated in the presence of $FeCl_4^-$ or ClO_4^- , and the formation of the double helix is the origin the spectro-change observed in CD and NMR. The P-helical chirality of the $[(L2)_{2}H_{2}]$ (FeCl₄)₂ is consistent with the first positive CD signals observed in the CD spectra⁵² which is similar to the spectra of the rigid Mn double-stranded helicates of L2 and L3.35 The extensive π - π stacking of the aromatic rings of the strands of the double-stranded helical structure which lead to anisotropic effect also agree well with the upfield signals observed in the NMR spectra. Considering the observation and results obtained with L2, L3 and L4 in the experiments, we believe that the monoprotonated form of triphenyl-bridged L3 and tetraphenylbridged L4 also form double-stranded helical structure. To show that this is the case and to have a better understanding to the helices in solution, DFT calculations were carried out on the monoprotonated species of L2, L3 and L4 using both the FeCl₄ and ClO_4^- anions.

Models of $[(L)_2H_2](FeCl_4)_2$ and $[(L)_2H_2](ClO_4)_2$ (L = L2-4) were first constructed, then their formation were investigated using theoretical DFT calculation at M06-2X level, with solvent effect taken account by the Polarizable Continuum Model. Fig. 20 shows the calculation with the biphenylbridged L2. The calculation starts from protonation of L2 and formation of the double-stranded helical core $[(L2)_2H_2]^{2+}$. The formation of the helical core stabilizes the system by 164.4 and 107.8 kcal mol⁻¹, respectively, with HFeCl₄ and



Fig. 23 Minimized energy model of (a) $[(L2)_2H_2](FeCl_4)_2$, and (b) $[(L2)_2H_2](ClO_4)_2$ obtained from theoretical calculation at M06-2X/6-31G(d)/LANL2DZ. Hydrogen bonding interactions are shown with dotted lines on only one of the anions for clearance.



Fig. 24 Minimized energy model of (a) $[(L3)_2H_2](FeCl_4)_2$, and (b) $[(L3)_2H_2](ClO_4)_2$ obtained from theoretical calculation at M06-2X/6-31G(d)/LANL2DZ. Hydrogen bonding interactions are shown with dotted lines on only one of the anions for clearance.



Fig. 25 Minimized energy model of (a) $[(L4)_2H_2](FeCl_4)_2$, and (b) $[(L4)_2H_2](ClO_4)_2$ obtained from theoretical calculation at M06-2X/6-31G(d)/LANL2DZ. Hydrogen bonding interactions are shown with dotted lines on only one of the anions for clearance.

 $HClO_4$. The energy different is the sum of dissociation of the $HFeCl_4$, protonation of L2, the stacking between the ligands and the pyridinium pyridine hydrogen bondings. Interactions between $[(L2)_2H_2]^{2+}$ with $FeCl_4^-$ and ClO_4^- anion further stabilize the system by 30.1 and 26.1 kcal mol⁻¹, respectively, which give $[(L2)_2H_2](FeCl_4)_2$ and $[(L2)_2H_2](ClO_4)_2$ with an overall stabilization 194.1 and 133.9 kcal mol⁻¹, respectively.

Table 2 A summary of the selected bond lengths and angles of X-ray and calculated data of [(L2)₂H₂](FeCl₄)₂

X-ray structure of $[(L2)_2H_2](FeCl_4)_2$		Model of $[(\mathbf{L2})_2\mathbf{H}_2](\operatorname{FeCl}_4)_2$	
Atoms	Bond lengths or angles	Atoms	Bond lengths or angles
N1-H1…N5	1.987 Å	N89-H195…N194	1.890 Å
N1-H1···N5	156.14°	N89-H195…N194	157.00°
N8-H8····N4	1.989 Å	N181-H196…N92	1.894 Å
N8-H8…N4	154.98°	N181-H196…N92	156.48°
N1-C12-C13-N2	17.58°	N89-C26-C27-N90	23.83°
N2-C17-C18-C23	20.49°	N90-C34-C35-C43	12.87°
C23-C22-C24-C29	32.74°	C43-C42-C45-C53	29.67°
C29-C28-C30-N3	24.58°	C53-C52-C55-N91	20.82°
N3-C34-C35-N4	21.82°	N91-C62-C63-N92	21.30°
N5-C58-C59-N6	24.04°	N181-C116-C119-N182	23.85°
N6-C63-C64-C69	22.81°	N182-C126-C127-C135	11.47°
C69-C68-C70-C75	34.24°	C135-C134-C137-C145	30.08°
C75-C74-C76-N7	13.29°	C145-C144-C147-N183	21.48°
N7-C80-C81-N8	20.35°	N183-C154-C155-N184	20.64°



Scheme 2 Formation of hydrogen-bonded double-stranded helix $[(L2)_2H_2](CIO_4)_2$ with CIO_4^- anions represented by

Similar calculations were carried out using L3 and L4 and the results are shown in Fig. 21 and 22. The results suggest that the formation of the double-stranded helices with L3 and L4 are also feasible, and also give additional information to the difference between these systems. In the presence of HFeCl₄, the formation of the double-stranded helical core $[(L)_2H_2]^{2+}$ with L4 being the most stable $(-173.4 \text{ kcal mol}^{-1})$, which is followed by L3 (-172.1 kcal mol⁻¹), and then followed by L2 $(-164.4 \text{ kcal mol}^{-1})$. The same trend also occurs with HClO₄. These trends seem most likely due to the increase in the number of the bridging phenyl-rings of the ligands which increase the stacking interactions. Interactions between $[(L)_2H_2]^{2+}$ and FeCl₄⁻ are in range -30.1 to -31.7 kcal mol⁻¹. With ClO_4^{-} , the interactions are in the range -25.3 to -28.3kcal mol⁻¹. The FeCl₄⁻ anion seems to have better stabilization than ClO_4^{-} . With the same anion, the differences in energy among L2-4 suggest that the helices interact differently with different anions. The energy of -133.9 kcal mol⁻¹ for $[(L2)_2H_2](ClO_4)_2$ is less stable than other helices which agrees quite well with the observations in the CD experiments.

The models of $[(L)_2H_2](FeCl_4)_2$ and $[(L)_2H_2](ClO_4)_2$ (L = L2-4) are shown in Fig. 23–25. Fig. 23a shows the model of $[(L2)_2H_2](FeCl_4)_2$. The optimized geometry is in satisfactory agreement with its crystal structure, and the X-ray and calculated data are summarized in Table 2. Beside the pyridinium N⁺H and pyridine hydrogen bonding, there are C-H…Cl type hydrogen bonding interactions between the $[(L2)_2H_2]^{2+}$ core and the two FeCl₄⁻ anions very much similar to that observed in the X-ray structure. The bond distances and angles are summarized in Table S1.† The presence of C-H…Cl hydrogen bondings are also confirmed by Atom in Molecules (AIM) analysis, and the results are shown in Table S2.† The FeCl₄⁻ anions use three out of the four chloride atoms to hold the double-stranded helical structure by forming multi-centered hydrogen bonding to the protons on the first and the second aromatic rings of one strands, and the protons on the forth and firth aromatic rings of the other. The model of $[(L2)_2H_2](ClO_4)_2$ is shown in Fig. 21b. There are C-H…O type hydrogen bonding interactions. The smaller $ClO_4^$ anions use three out of the four its oxygen atoms to form



Fig. 26 CD absorption at 334 nm of (a) L2, (b) L3, and (c) L4, with alternate addition of $HClO_4$ and $NEt_3.$

hydrogen bondings to the same aromatic protons on the backbone of both strands as $FeCl_4^-$ but with slightly different orientation. The data are summarized in Tables S3 and S4.[†]

Models with L3 are shown in Fig. 22a and b. The stacking between the aromatic rings observed in $[(L3)_2H_2]^{2+}$ are different from $[(L2)_2H_2]^{2+}$. In the case of $[(L2)_2H_2]^{2+}$, the pyridinium of one strand stacks with the forth aromatic ring of the other, but the pyridinium stacks with the third aromatic ring in case of $[(L3)_2H_2]^{2+}$. Exterior hydrogen bindings between the helices and the anions are also observed. Data are summarized in Table S5-8.† Some differences are observed when comparing to L2. For L3, the FeCl₄⁻ forms hydrogen bonds to the protons on the first and the second aromatic rings of one strands, but to the protons on the third and the forth aromatic rings of the other. In the presence of ClO_4^- , it forms hydrogen bonds to the same aromatic rings as $FeCl_4^-$ but with slightly different protons.

Models with L4 are shown in Fig. 23a and b. The stacking observed in the helix of L4 is different from L2 and L3. The pyridinium of one strand stacks with the second aromatic ring of the other. The close proximity of the anions with the aromatic protons again suggest the presents of hydrogen bondings. The data are summarized in Table S9-11.[†] However, due to the size of the model of $[(L4)_2H_2](FeCl_4)_2$ excess the limit of the software, AIM analysis was only carried out on $[(L4)_2H_2](ClO_4)_2$. The FeCl₄⁻ forms hydrogen bonds to the protons on the first and the second aromatic rings of one strand, but to the protons on the third aromatic rings of the other, which is different from helix of L2 and L3. The ClO_4^{-} also form hydrogen bonds to the same aromatic protons. Although the C-H···O and C-H···Cl hydrogen bonding are generally considered as weak hydrogen bonding,49 the calculation results show that the sum of the interactions from the two anions are not weak (in range -30.1 to -31.7 kcal mol⁻¹ for FeCl₄⁻, and -25.3 to -28.3 kcal mol⁻¹ for ClO₄⁻), and is comparable to a strong hydrogen bond.53 As shown in Scheme 2, we believe that the formation of the double-stranded helix is a delicate combination of the two types of hydrogen bonding, the interior hydrogen bonding between pyridinium and pyridine, and the exterior hydrogen bonding between the core and anions.

Reversibility of the formation of the hydrogen-bonded double helix

Since reversible interconversion between multistable states of a molecule in response to protonation is of interest,⁵⁴⁻⁵⁶ we tried to see if **L2–4** can be developed into reversible interconverting systems with CD signal change by sequential addition of acid and base. With HFeCl₄ and NEt₃ as base, excess NEt₃ (5 equiv.) is needed to turn off the CD signal, but addition of the HFeCl₄ cannot fully restore the original CD signal suggesting that the process is not reversible with HFeCl₄. However, with HClO₄, the results were quite different. Fig. 26a shows the results of reaction of L2 with HClO₄ and NEt₃. When HClO₄ is used, addition of NEt₃ turns the signal "off". Addition of another equiv. of HClO₄ turns the signal "on" again restoring the CD signal. This cycle can be repeated without the decrease in signal's intensity for at least four to five cycles. This on and off CD signal switching properties can be demonstrated by using L3 and L4

(Fig. 26b and c). The results suggest that these systems have potential to be developed as chiroptical switches.

Conclusion

We have presented the synthesis of a series of chiral polyphenylbridged bis(2,2'-bipyridine) ligands. Protonation of the ligands give intense CD signals. The protonation processes were studied in detail by both CD and ¹H NMR. The results suggest that the CD signals come from monoprotonated species. X-ray crystal structures of free ligand and protonated ligands were obtained. The structure of the monoprotonated ligand shows that it is a double-stranded helix that is stabilized by a delicate combination of interior hydrogen bonds between the ligand strands and exterior hydrogen bonds between the helical core and anions. Theoretical DFT calculations on the formation of hydrogenbonded double helices with the series of ligands were carried out. We believe that hydrogen-bonded double helices are formed upon monoprotonation of the ligands and they are stabile in solution as well. With perchlorate anion, the system can be interconverted between non-helical and helical states by addition and removal of proton, making it an on/off chiroptical switch. Further investigation of these properties for other applications line is now in progress.

Acknowledgements

The work described in this paper was supported by grants from the University Grants Committee of Hong Kong Special Adminstrative Region, China (Project no. [AoE/P-03/08] and CityU 7004018).

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Published on 11 March 2014. Downloaded by Florida State University on 21/10/2014 10:31:27.

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