



Cite this: *Org. Biomol. Chem.*, 2014, **12**, 9439

Received 18th August 2014,
Accepted 25th September 2014

DOI: 10.1039/c4ob01763c

www.rsc.org/obc

Conformationally locked bicyclo[4.3.0]nonane carbanucleosides: synthesis and bio-evaluation†

Tony K. M. Shing,^{*a} Anthony W. H. Wong,^a Huiyan Li,^b Z. F. Liu^b and Paul K. S. Chan^c

D-Ribose was converted into 3 novel carbobicyclic nucleosides bearing a bicyclo[4.3.0]nonane framework in 16–19 steps with 5–12% overall yields involving a Wittig olefination and an intramolecular Diels–Alder reaction as the key steps. The present synthesis also provides an efficient entry for chiral hydrindenones. The conformational studies of these carbanucleosides and their bio-evaluation as potential antiviral agents are reported.

Introduction

In recent years, carbocyclic nucleosides or carbanucleosides have warranted tremendous attention from both synthetic and medicinal chemists.^{1,2} The replacement of the oxygen atom in the furanose ring by a methylene unit provides both enzymatic and chemical stability, attributable to the loss of a true glycosidic linkage.³ However, there is a significant change in the conformation due to the loss of the *gauche* and anomeric effects which exist in conventional nucleosides. These effects maintain the sugar moiety of the nucleoside in either a C3'-*endo* (North) or a C2'-*endo* (South) conformation.⁴ In the absence of these effects, the conformation of carbocyclic nucleosides is mainly governed by the steric bulk of the nucleobase, which prefers to occupy the equatorial position and adopts an atypical C1'-*exo* conformation. The conformation of nucleosides is intriguing since any structure–activity relationship (SAR) study of antiretroviral nucleosides is complicated by the complexity of the anabolic process of activation. It involves three enzymatic steps to transform the nucleoside into its 5'-triphosphate (NTP), plus the final interaction of the NTP with the target enzyme, the reverse transcriptase (RT). Therefore, the preferences of conformation exhibited by the nucleoside, or its nucleotides, must be identified at each intervening enzymatic step. The only step which is invariant and common to all nucleoside RT inhibitors is the final interaction of NTP with

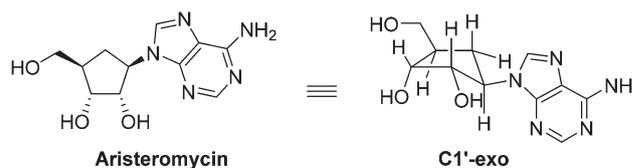


Fig. 1 Naturally occurring carbocyclic nucleoside aristeromycin.

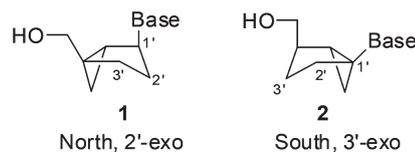


Fig. 2 Conformationally locked nucleosides with a bicyclo[3.1.0]hexane system.

RT. The route to NTP anabolite, however, involves different kinases, all of which are highly dependent on the nature of the heterocyclic base.⁵

Aristeromycin is a representative and its conformation is illustrated in Fig. 1.⁶ It has been reported to be a potent inhibitor of *S*-adenosyl-*L*-homocysteine (AdoHcy) hydrolase, which plays an important regulatory role in the *S*-adenosyl-*L*-methionine (AdoMet)-dependent methylation process.⁷ The methylation process is important as most plant and animal viruses require a methylated cap structure at the 5'-terminus of their mRNA for viral replication. However, their cytotoxicity precludes clinical use.^{8–10}

There are not many accounts on the syntheses of bicyclic carbanucleosides. In 1993, Rodriguez *et al.* pioneered the construction of dideoxyribonucleoside analogues with a bicyclo[3.1.0]hexane backbone (Fig. 2) which constrains the conformation of the analogue back to the C2'-*exo* (North).¹¹ From

^aDepartment of Chemistry and Center of Novel Functional Molecules, The Chinese University of Hong Kong, Shatin, Hong Kong, China

^bDepartment of Chemistry and Centre for Scientific Modeling and Computation, The Chinese University of Hong Kong, Shatin, Hong Kong, China

^cDepartment of Microbiology, The Chinese University of Hong Kong, China

†Electronic supplementary information (ESI) available: Experimental procedures, characterisation data, copies of NMR spectra for all compounds, detailed DFT modelling and CIF files. CCDC 727043 and 1000117. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c4ob01763c

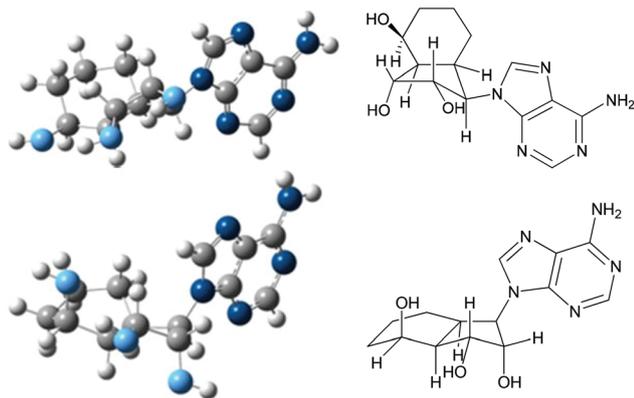


Fig. 3 Conformation of bicyclic carbanucleosides calculated using the DFT model: top: C1'-exo conformation; bottom: C3'-endo conformation.

then on, different kinds of conformationally restricted carbocyclic nucleosides that lock the conformation as either North or South were reported.¹²

Coxsackie B viruses¹³ are single strand positive-sense RNA viruses and contain six serotypes of pathogenic enteroviruses that trigger illness ranging from gastrointestinal distress to full-fledged pericarditis and myocarditis. They resist a wide variety of chemical treatment and to the best of our knowledge, there is still no well accepted treatment for the Coxsackie B viruses and it is interesting to see whether our compounds could be potential drugs to treat the viruses.¹⁴

However, to date, only carbobicyclic nucleosides with a bicyclo[3.1.0]hexane and a bicyclo[3.3.0]octane system have been described. There is still no publication on the syntheses of carbanucleosides with a bicyclo[4.3.0]nonane system. Therefore, it is fascinating to see if they exhibit any antiviral activity. Furthermore, according to our theoretical calculations, carbanucleosides with a cyclohexane ring *cis*-fused to a cyclopentane ring should exist in either a C1'-exo or a C3'-endo conformation, which is in close agreement with the North conformer of conventional nucleosides (Fig. 3) and may show RNA antiviral activity.

To synthesize this class of carbobicyclic nucleosides for biological studies, we first have to synthesize the bicyclo[4.3.0]nonane core. Functionalized hydrindenones (bicyclo[4.3.0]nonenones) are versatile intermediates in complex natural product synthesis, *e.g.*, the CD ring in steroids. It is noteworthy that syntheses of chiral functionalized hydrindenones were not an easy task *via* an intramolecular cycloaddition.¹⁵ To date, only a few publications have reported the intramolecular Diels–Alder (IMDA) reaction on sugars and most of these papers documented the syntheses of *trans*-decaline systems.¹⁶ Only one report described the synthesis of a bicyclo[4.3.0]nonenone, involving a lengthy synthetic route.¹⁷ Jarosz *et al.* reported the syntheses of chiral bicyclo[4.3.0]non-2-enes from a sugar allyltin. However, the choice for protecting groups was limited as Lewis acid was employed in the preparation of the IMDA precursors.¹⁸ In this paper, we report a shorter route

with milder conditions for the construction of chiral hydrindenone intermediates.

Results and discussion

Theoretical calculation

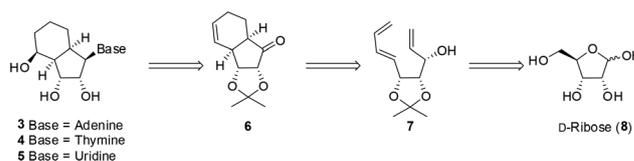
Gaussian 09 program package¹⁹ was used to calculate the structures of the carbanucleosides with the density functional theory (DFT) quantum chemical method. Equilibrium geometries of all molecules were fully optimized at the B3LYP/6-31G(d) level of theory.²⁰ Stationary points were characterized as either minima (no imaginary frequency) or transition structures (only one imaginary frequency) by calculation of the vibrational frequencies using analytical gradient procedures at the same level. Minimum energy pathways connecting the reactants and products were confirmed by intrinsic reaction coordinate (IRC) calculations.²¹

Synthesis

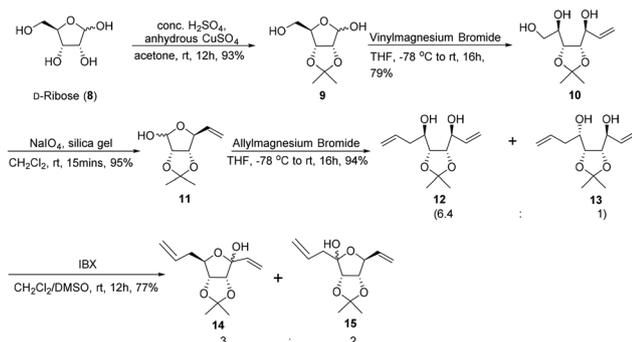
Retrosynthesis of the target carbanucleosides (3–5) shows that it could be synthesized from cycloadduct **6**, which would be prepared from D-(–)-ribose (**8**) *via* triene **7**, using an IMDA reaction as the key step (Scheme 1).

Starting from D-(–)-ribose (**8**), the 2,3-diol in **8** was isopropylidened to afford acetonide **9**,²² which was then subjected to vinylation to give alkene **10** in good yield.²³ Glycol cleavage oxidation of the vicinal diol in **10** gave lactol **11**, which with allylmagnesium bromide furnished dienes **12** and **13** in a ratio of 6:1, respectively, and in excellent overall yields. Aqueous indium allylation only led to decomposition of the starting material. Oxidation of **12** with IBX provided a mixture of lactols **14** and **15** in a 3:2 ratio, which could not be separated by column chromatography (Scheme 2). We hoped to utilize the equilibrated open-chain form of **14** and to eliminate the homoallylic alcohol to form IMDA precursor **17** (Scheme 3). Different elimination conditions were examined, but no positive results were observed after exhaustive experimentation.

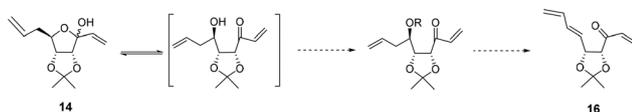
Since the ring opening–elimination approach failed, we changed our synthetic avenue to a protection and deprotection approach (Scheme 4). The allylic hydroxyl group in the diol **12** was protected as silyl ether **17** in good yield. The homoallylic hydroxyl group was then attempted to be eliminated. However, after trying different methods, silyl ether **17** only reacted with trifluoroacetic anhydride (TFAA) to give trifluoroacetate **18** which was then subjected to elimination in the presence of DBU. However, no triene **19** was detected, beyond recovery of



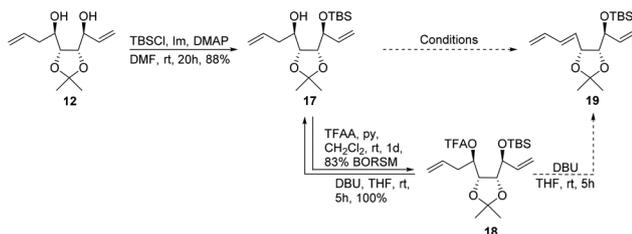
Scheme 1 Retrosynthesis of the target nucleosides.



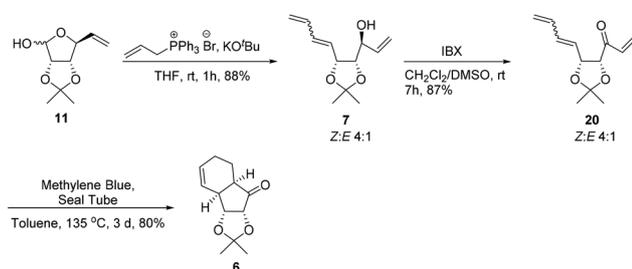
Scheme 2 Synthesis of lactol 14 and 15.



Scheme 3 Equilibrium between the ring and chain form of 14.



Scheme 4 Attempted synthesis of triene 19.



Scheme 5 Synthesis of 6.

alcohol 17 quantitatively. The use of other bases did not yield any desired 19.

We therefore revised our synthetic strategy and discovered that lactol 11 was a good candidate for the Wittig reaction²⁴ with allyltriphenylphosphonium bromide²⁵ to introduce the diene functionality (Scheme 5). After screening with different bases, potassium *tert*-butoxide (KO^tBu) in THF gave the best yield of triene 7 of 88% with a *Z* to *E* ratio of 4:1. The geometric isomers were not separable by column chromatography. In order to enhance the selectivity, Horner–Wadsworth–

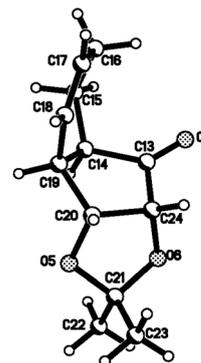


Fig. 4 X-ray structure of 6.

Emmons (HWE) olefination²⁶ was tried but no reaction was observed with different kinds of bases such as dimethyl sodium and *n*-butyllithium. The mixture of trienes 7 was subjected to oxidation with IBX as an oxidant and furnished trienone 20 in 87% yield. Trienone 20 was highly unstable and polymerized quickly after standing in vacuum. Trienone 20 underwent the IMDA reaction in toluene in the presence of methylene blue in a sealed tube to give hydrindenone 6 in good yield. The structure and absolute configuration of 6 was confirmed by X-ray crystallography (Fig. 4).

Initially we believed that *cis* to *trans* isomerization of the diene in 20 should precede the cycloaddition. However, theoretical calculations disagreed with this idea since the relative energy of the isomerization is surprisingly high (99 kcal mol⁻¹), which was 50 kcal mol⁻¹ higher than the published value.²⁷ Therefore, the stereoselectivity of the cycloadduct could be explained by the proposed transition state models as shown in Fig. 5. For *E*-20, two possible transition states, TS-1 and TS-2, in the *endo* mode of cycloaddition, would lead to the formation of two thermodynamically favorable *cis*-fused cycloadducts 6 and 21. Theoretical calculations also agreed with these results. However, the steric repulsion between the isopropylidene and the newly formed 6-membered ring in the transition state might inhibit the formation of 21. TS-3 might give 22 but unfortunately 22 was unable to detect. For *Z*-20, TS-5 was not favorable due to the steric repulsion between the isopropylidene and the newly formed 6-membered ring and in TS-6, the highly twisted conformation might inhibit the formation of the *trans*-fused cycloadduct 22.

Since there was no *trans*-fused cycloadduct 22 isolated in the previous step, epimerization was tried but failed to occur after many attempts and under a wide variety of conditions (Scheme 6).²⁸

Since epimerization was not successful, syntheses of bicyclic carbanucleosides were carried on with the *cis*-fused bicyclic carbocycle 6. The hydrindenone 6 was epoxidized to give epoxide 23 in good yield (Scheme 7). The β-epoxide was the only isolated product and no α-epoxide was detected. The extremely high diastereoselectivity might be explained by the ketone directing effect proposed by Armstrong.²⁹

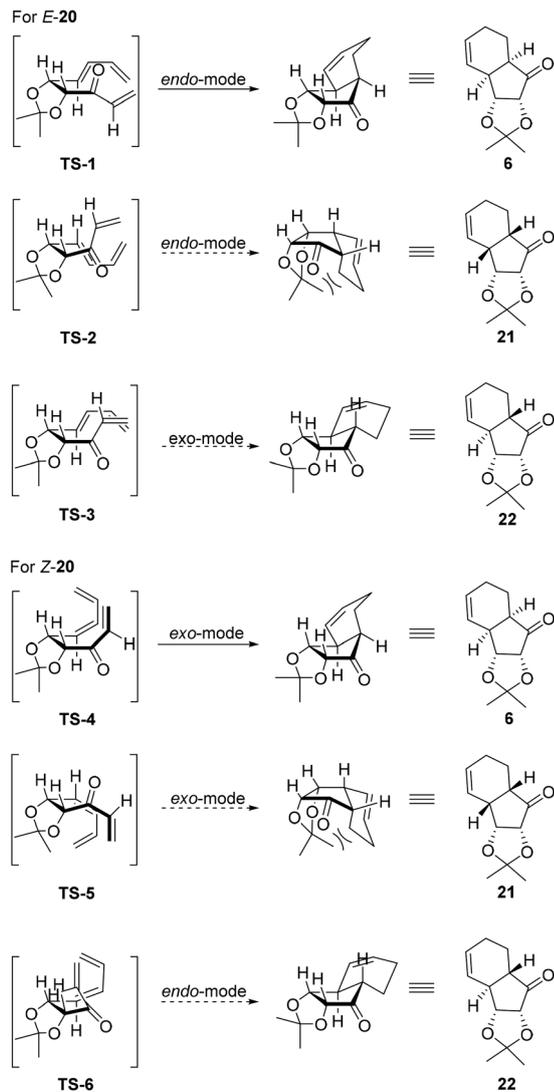
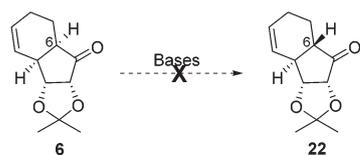
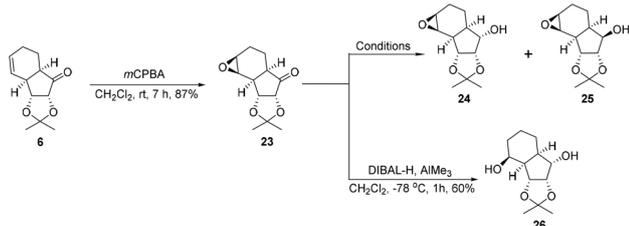


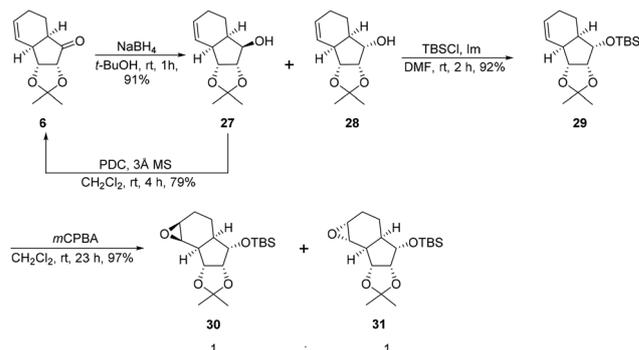
Fig. 5 Proposed transition state models of the cyclization of trienone 20.



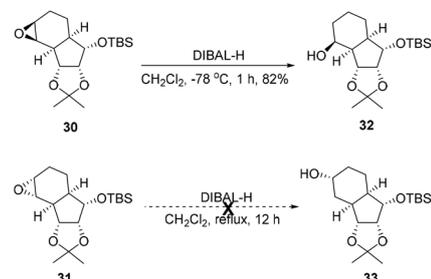
Scheme 6 Attempted epimerization of 6 to 22.



Scheme 7 Synthesis of diol 26.



Scheme 8 Synthesis of epoxides 30 and 31.



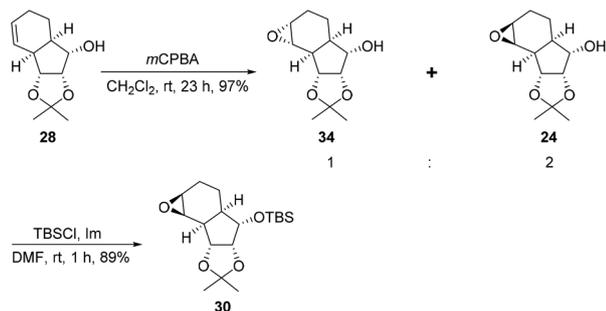
Scheme 9 Reduction of epoxides 32 and 33.

Reduction of epoxide was attempted with LiBH_4 , NaBH_4 , super hydride or LiAlH_4 under various conditions, but unfortunately only alcohols 24 and 25 were isolated. Interestingly, reduction with DIBAL-H in the presence of AlMe_3 provided the desired diol 26 in moderate yield.

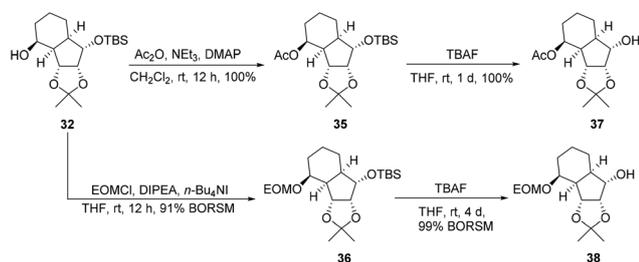
As the reduction was not as easy as expected and the yield was not satisfactory, an alternative approach was performed (Scheme 8). Hydrindenone 6 was firstly reduced with NaBH_4 to the corresponding alcohols 27 and 28 in a 1 : 4 ratio. Alcohol 27 could be oxidized back to 6 by PDC in good yield. Protection of 28 as silyl ether 29 proceeded smoothly and this was followed by epoxidation to give epoxides 30 and 31 in a 1 : 1 ratio. When both epoxides were subjected to reduction, only the β -epoxide 30 was susceptible to reduction, and alcohol 32 was obtained in good yield. The α -epoxide 31 was reluctant to be opened with DIBAL-H even under refluxing CH_2Cl_2 (Scheme 9).

The difficulty of ring-opening in α -epoxide 33 might be attributable to the steric hindrance of the adjacent cyclopentane ring so that the hydride was unable to approach the reacting site. To minimize the formation of α -epoxide, we chose to epoxidize alkene-alcohol 28, and the ratio of epoxide 34 to 24 was slightly increased to 1 : 2 (Scheme 10). The hydroxyl group in 24 was protected as TBS ether to furnish silyl ether 30 again.

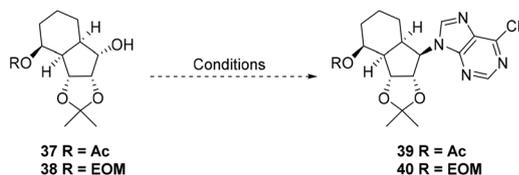
After the reduction of the epoxide, the newly formed hydroxyl group in 32 was protected as acetate 35 and as ethoxy methyl (EOM) ether 36, respectively, in excellent yields (Scheme 11). Removal of the silyl protecting group in both



Scheme 10 Epoxidation of 28.



Scheme 11 Synthesis of 37 and 38.

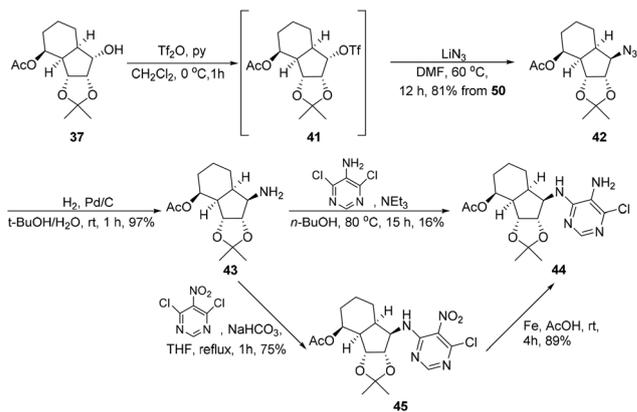


Scheme 12 Attempted to synthesize pyrimidine 39 and 40.

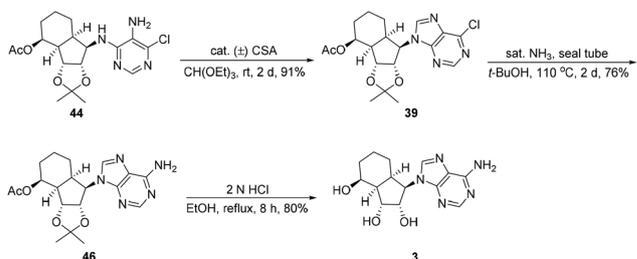
substrates proceeded smoothly to furnish alcohols 37 and 38 in excellent yields, which were the precursors for coupling with the nucleobases.

With alcohol 37 and 38 in hand, direct coupling of nucleobases with the free alcohols was examined (Scheme 12). Firstly, the Mitsunobu reaction³⁰ was used but no desired product was isolated using either PPh₃ or PBu₃ with DIAD in refluxing toluene. Alcohols 37 and 38 were then activated by conversion into sulfonate esters (R = Ms, Tf) which were displaced with purine bases. Unfortunately, both sulfonate esters failed to form purine 39 and 40, respectively.

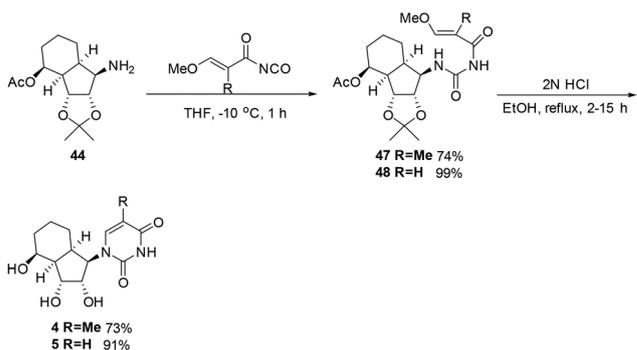
As the convergent route failed to give any desired nucleoside, the linear approach was adopted to synthesize the target molecules. Triflate 41 was successfully converted into azide 42 in good overall yield from alcohol 37 (Scheme 13). Azide 42 was reduced to the corresponding amine 43 by catalytic hydrogenation in excellent yield. Reaction of amine 43 with 5-amino-4,6-dichloropyrimidine furnished pyrimidine 44 in miserable yield and could be explained by the low electrophilicity of 5-amino-4,6-dichloropyrimidine. In order to improve



Scheme 13 Synthesis of pyrimidine 44.



Scheme 14 Synthesis of the target adenosine analogue 3.



Scheme 15 Syntheses of thymidine analogue 4 and uridine analogue 5.

the reactivity, a stronger electrophile, 5-nitro-4,6-dichloropyrimidine, was used instead. Pyrimidine 45 was then obtained in good yield, and reduction of the nitro group to the amino group was achieved using iron powder as a reductant in AcOH (Scheme 13).³¹ Using a standard procedure, pyrimidine 44 was transformed into the target carbanucleoside 3 (Scheme 14).

The syntheses of thymidine analogue 4 and uridine analogue 5 were similar and the results are shown in Scheme 15. Amine 44 was reacted with 3-methoxymethacryloyl isocyanate³² and 3-methoxy-2-propenoyl isocyanate³³ to produce urea 47 and 48 in 74% and 99%, respectively. Both ureas 47 and 48 were then subjected to acid hydrolysis to remove all the protecting groups as well as to facilitate the cyclization. Thymi-

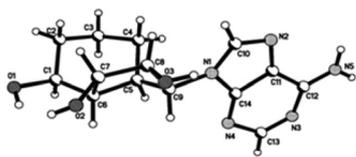


Fig. 6 X-ray structure of adenosine 3.

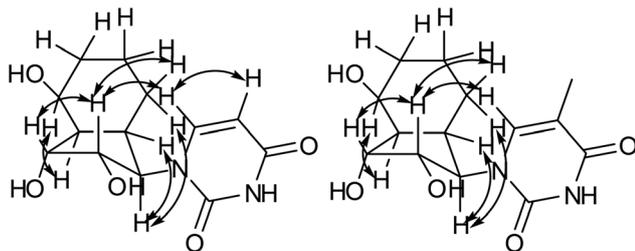


Fig. 7 NOE results of nucleosides 4 and 5.

dine carbanucleoside 4 was obtained in moderate yield (73%) and uridine carbanucleoside 5 was obtained in excellent yield (91%).

Determination of conformation

With all the carbanucleosides in hand, their conformations were determined by both X-ray crystallography and NOE experiments. Disappointing results were obtained as all the carbanucleosides displayed the undesired C1'-*exo* conformation. The crystal structure of adenine 3 is shown in Fig. 6. The NOE results of both nucleosides 4 and 5 also agreed with the C1'-*exo* conformation (Fig. 7).

Biological studies

Antiviral activity against enterovirus, Coxsackie B3 virus, was assessed using the plaque reduction assay. Briefly, African green monkey cells (Vero) were used to cultivate virus-compound mixtures at various dilutions. After 72 hours of incubation at 37 °C, infected cells were fixed and the number of plaques was counted. The percentage of reduction in the number of plaques in wells with compounds was calculated. Compounds suspected to have antiviral effect were further examined for cytotoxicity using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay.

As a result, 3 nucleosides 3, 4 and 5 showed screening results suggestive of having weak *in vitro* antiviral activity against Coxsackie B3 virus. Adenosine 3 (concentration = 1 µg mL⁻¹) showed 4.9% of inhibition and thymidine 4 (concentration = 1 µg mL⁻¹) showed 5.5% of inhibition and higher concentration (concentration = 100 µg mL⁻¹) led to highly toxic uridine 5 (concentration = 1 µg mL⁻¹) which gave the best result, showing more than 12.3% of inhibition and further structural modification is recommended.

Conclusion

To conclude, bicyclo[4.3.0]nonane carbanucleosides 3, 4, 5 were synthesized from D-(-)-ribose in 16 steps or 19 steps with 5%, 7% and 12% overall yield, respectively, involving a Wittig alkenation and an IMDA reaction as the key steps. The cycloadduct 6, a chiral functionalized hydrindenone (bicyclo[4.3.0]nonenone), should open an avenue as a versatile intermediate in complex natural product synthesis. X-ray crystallography and NOE studies of these carbanucleosides show that they display the C1'-*exo* conformation. These compounds demonstrated weak *in vitro* antiviral activity against Coxsackie B3 virus. It is worth modifying the compounds and reassess for a wider range of viruses. Research in this direction is in progress.

Acknowledgements

This work was supported by a Hong Kong RGC General Research Fund (Ref. CUHK 4030/09P).

Notes and references

- 1 D. K. Tosh, H. O. Kim, S. Pal, J. A. Lee and L. S. Jeong, in *Modified Nucleosides*, Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim, Germany, 2008, pp. 525–560.
- 2 Review: (a) E. De Clercq, *Nucleosides, Nucleotides Nucleic Acids*, 2009, **28**, 586–600; (b) D. M. Huryn and M. Okabe, *Chem. Rev.*, 1992, **92**, 1745–1768; (c) A. D. Borthwick and K. Biggadike, *Tetrahedron*, 1992, **48**, 571–623; (d) L. Agrofoglio, E. Suhas, A. Farese, R. Condom, S. R. Challand, R. A. Earl and R. Guedj, *Tetrahedron*, 1994, **50**, 10611–10670; (e) V. E. Marquez and M. I. Lim, *Med. Res. Rev.*, 1986, **6**, 1–40.
- 3 H. Bricaud, P. Herdewijn and E. De Clercq, *Biochem. Pharmacol.*, 1983, **32**, 3583–3590.
- 4 W. Saenger, in *Principle of Nucleic Acid Structure*, Springer-Verlag, New York, 1984, pp. 9–28.
- 5 H. Mitsuya, in *Anti-HIV Nucleosides: Past, Present and Future*, R. G. Landes Co., Texas, 1997.
- 6 A. Kalman, T. Koritsanszky, J. Beres and G. Sagi, *Nucleosides Nucleotides*, 1990, **9**, 235–243.
- 7 R. M. Ransohoff, P. Narayan, D. F. Ayers, F. M. Rottman and T. W. Nilsen, *Antiviral Res.*, 1987, **1**, 317–327.
- 8 D. L. Hill, S. Straight, P. W. Allan and L. L. Bennett Jr., *Mol. Pharmacol.*, 1971, **7**, 375–380.
- 9 S. Yaginuma, N. Muto, M. Tsujino, Y. Sudate, M. Hayashi and M. Otani, *J. Antibiot.*, 1981, **34**, 359–366.
- 10 E. De Clercq, *Antimicrob. Agents Chemother.*, 1985, **28**, 84–89.
- 11 J. B. Rodriguez, V. E. Marquez, M. C. Micklaus, H. Mitsuya and J. J. Barchi Jr., *Tetrahedron Lett.*, 1993, **34**, 6233–6236.
- 12 (a) Q. Chao and V. Nair, *Tetrahedron*, 1997, **53**, 1957–1970; (b) M. Šála, H. Hřebabecký, M. Dračinský, M. Masojídková,

- A. M. De Palma, J. Neyts and A. Holý, *Collect. Czech. Chem. Commun.*, 2010, **75**, 1–20.
- 13 H. A. Rotbart, *Antiviral Res.*, 2002, **53**, 83–98.
- 14 B. N. Fields, D. M. Knipe, R. M. Chanock, J. L. Melnick, B. Roizman and R. E. Shope, in *Fields Virology*, Raven Press, New York, 1985, pp. 739–794.
- 15 (a) C. C. Browder, F. P. Marmsaeter and F. G. West, *Org. Lett.*, 2001, **3**, 3033–3035; (b) B. B. Snider and T. C. A. Kirk, *J. Am. Chem. Soc.*, 1983, **105**, 2364–2368; (c) E. J. Corey and T. A. Engler, *Tetrahedron Lett.*, 1984, **25**, 149–152; (d) D. A. Smith, K. Sakan and K. N. Houk, *Tetrahedron Lett.*, 1986, **27**, 4877–4880.
- 16 (a) S. Mondal, R. N. Yadav and S. Ghosh, *Org. Lett.*, 2011, **13**, 6078–6081; (b) R. S. Nandurdikar, A. V. Subrahmanyam and K. P. Kaliappan, *Eur. J. Org. Chem.*, 2010, 2788–2799; (c) C. Taillefumier and Y. Chapleur, *Can. J. Chem.*, 2000, **78**, 708–722.
- 17 M. D. Clay, D. Riber and A. G. Fallis, *Can. J. Chem.*, 2005, **83**, 559–568.
- 18 S. Jarosz, E. Kozłowska and A. Jezewski, *Tetrahedron*, 1997, **53**, 10775–10782.
- 19 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, A. F. Izmaylov, J. Bloino, G. Zheng, J. L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, Jr. J. A. Montgomery, J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, N. Rega, N. J. Millam, M. Klene, J. E. Knox, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, R. L. Martin, K. Morokuma, V. G. Zakrzewski, G. A. Voth, P. Salvador, J. J. Dannenberg, S. Dapprich, A. D. Daniels, O. Farkas, J. B. Foresman, J. V. Ortiz, J. Cioslowski and D. J. Fox, *GAUSSIAN 09 (Revision B.01)*, Gaussian Inc., Pittsburgh, PA, 2010.
- 20 A. D. Becke, *J. Chem. Phys.*, 1993, **98**, 5648–5652.
- 21 C. Gonzalez and H. B. Schlegel, *J. Phys. Chem.*, 1991, **94**, 5523–5527.
- 22 P. A. Levene and R. S. Tipson, *J. Biol. Chem.*, 1936, **115**, 731–747.
- 23 T. K. M. Shing, D. A. Elsley and J. G. Gillhouley, *J. Chem. Soc., Chem. Commun.*, 1989, 1280–1282.
- 24 (a) G. Wittig and G. Geissler, *Ann.*, 1953, **580**, 44–57; (b) G. Wittig and U. Schollkopf, *Chem. Ber.*, 1954, **87**, 1318–1330.
- 25 (a) S. D. Tilley and M. B. Francis, *J. Am. Chem. Soc.*, 2006, **128**, 1080–1081; (b) J. G. Atkinson, M. H. Fisher, D. Horley, A. T. Morse, R. S. Stuart and E. Synnes, *Can. J. Chem.*, 1965, **43**, 1614–1624.
- 26 (a) L. Horner, H. Hoffmann and H. G. Wippel, *Chem. Ber.*, 1958, **91**, 61–63; (b) L. Horner, H. Hoffmann, H. G. Wippel and G. Klahre, *Chem. Ber.*, 1959, **92**, 2499–2505.
- 27 W. M. Marley and P. M. Jeffers, *J. Phys. Chem.*, 1975, **79**, 2085–2087.
- 28 Several bases were tried such as NaH, NaOMe, LDA, KO^tBu and DBU. We also tried to deprotonate the α -H with NaH and quenched by D₂O. The proton signal disappeared which proved that deprotonation did exist.
- 29 A. Armstrong, P. A. Barsanti, P. A. Clarke and A. Wood, *J. Chem. Soc., Perkin Trans. 1*, 1996, 1373–1380.
- 30 W.-B. Lu, S. Sengupta, J. L. Petersen, N. Akhmedov and X. Shi, *J. Org. Chem.*, 2007, **72**, 5012–5015.
- 31 (a) Y. Liu, Y. Lu, M. Prashad, O. Repic and T. A. Blacklock, *Adv. Synth. Catal.*, 2005, **347**, 217–219; (b) S. E. Hazlet and C. A. Dornfeld, *J. Am. Chem. Soc.*, 1944, **66**, 1781–1782.
- 32 G. Shaw and N. R. Warrener, *J. Chem. Soc.*, 1958, 157–161.
- 33 (a) C. Cesario and M. J. Miller, *J. Org. Chem.*, 2009, **74**, 5730–5733; (b) L. Santana, M. Tejeria and E. A. Uriarte, *J. Heterocycl. Chem.*, 1999, **36**, 293–295.