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COMMUNICATION

Selective aqueous acetylation controls the photoanomerization of α -cytidine-5'-phosphate

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Nucleic acids are central to information transfer and replication in living systems, providing the molecular foundations of Darwinian evolution. Here we report that prebiotic acetylation of the non-natural, but prebiotically plausible, ribonucleotide α -cytidine-5'-phosphate, selectively protects the vicinal diol moiety. Vicinal diol acetylation blocks oxazolidinone formation and prevents C2'-epimerization upon irradiation with UV-light. Consequently, acetylation enhances (4-fold) the photoanomerization of α -cytidine-5'-phosphate to produce the natural β -pyrimidine ribonucleotide-5'-phosphates required for RNA synthesis.

Developing a chemical rationale for the origins of the canonical nucleotides is one of the key challenges at the heart of understanding the origins of life on Earth.¹ In particular, RNA is thought to have played an essential role in the early evolution of life. Despite the remarkable progress made in elucidating prebiotic pathways towards ribonucleotide-2',3'-cyclic phosphates,²⁻⁴ much less attention has been paid to understanding the origins of the canonical 5'-phosphorylation pattern observed in biological nucleotides.⁵ We have recently reported an oxidative phosphorylation of nucleotide precursors that delivers regiospecific 5'-phosphorylation in water.⁶ Our reaction pathway begins with prebiotically plausible acrolein **1** and 2-aminooxazole **2**, and furnishes *ribo*-3-5P with excellent *ribo*-selectivity through a combination of kinetic and thermodynamic control (Fig. 1). Direct nucleophilic phosphorylation with inorganic phosphate in water sidesteps the requirement for electrophilic activation of phosphate, and as a direct consequence of exploiting the intrinsic nucleophilicity of phosphate, overcomes the widely cited "phosphorylation in water problem" during nucleotide synthesis.⁷ However, access to RNA through this route still requires an important stereochemical issue to be resolved. The reaction strategy relies upon (a tethered) simultaneous assembly of the nucleobase and sugar moieties of the cytidine nucleotide. The constrained architecture of intermediate *ribo*-3-5P provides the

stereochemical control required to direct the stereoselective assembly of the *ribo*-diastereomer, but delivers the non-natural anomeric stereochemistry. Accordingly, pyrimidine ribonucleotide α -*ribo*-4 is readily synthesized in high yield upon reaction of *ribo*-3-5P with cyanoacetylene (**5**) in water (Fig. 1), and though α -*ribo*-4 has the natural 5'-phosphorylation pattern, it has non-natural C1'-stereochemistry which requires stereochemical inversion to access the canonical nucleotide.

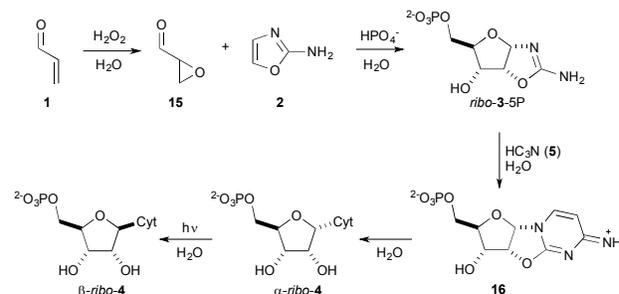


Figure 1. Previous work: Synthesis of cytidine-5'-phosphates α -*ribo*-4 and β -*ribo*-4 by oxidative phosphorylation and nucleobase photoanomerization in water.^{2,6} Cyt=cytosine.

The absence of (biogenic) O₂ and O₃ UV-absorbance in a primitive atmosphere and the greater UV output from the young Sun,^{8,9} suggest that UV-light would be an important component of prebiotic environments and may have been the most abundant energy source available to drive prebiotic chemistry.⁸ Attempts to photoanomerize α -*ribo*-4 with UV irradiation to furnish β -*ribo*-4 is low yielding (4-6%),¹⁰ and also results in concomitant C2'-epimerization to give a complex mixture of *ribo*- and *arabino*-nucleotides (Fig. 2). Furthermore, photoanomerization of α -*ribo*-4 is severely prohibited by the formation of oxazolidinone **6**, which gradually and irrevocably destroys the cytosine nucleobase through intramolecular reaction with the C2'-hydroxyl moiety of α -*ribo*-4.¹¹ To overcome the poor photoanomerization of α -*ribo*-4, Sutherland and co-workers recently reported a remarkably efficient photoanomerization of non-natural nucleoside, α -2-thiocytidine, however, although certainly interesting, this strategy requires a non-aqueous reaction step (to yield a 2-thiopyrimidine

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intermediate) and ultimately requires synthesis of the non-natural 2',3'-cyclic phosphates, rather than nucleotides with the canonical 5'-phosphorylation pattern observed in biology.⁴

With the specific goal of only undertaking all reactions in water, the most prebiotically plausible and biological relevant solvent, and retaining the 5'-phosphorylation pattern installed by oxidative phosphorylation,⁶ we consider the effect of nucleotide sugar moiety modification on photoanomerization *en route* to RNA. We have previously found that blocking the C2'-hydroxyl moiety of α -ribo-4 by non-natural 2'-phosphorylation, led to improved photoanomerization.¹² However, to synthesize the targeted canonical nucleotide (β -ribo-4) the *cis*-2',3'-diol moiety must be preserved, therefore 2'-phosphorylation diverts nucleotide synthesis away from the canonical structures observed in biology. Ideally, a temporary blockade of the C2'-hydroxyl moiety of α -ribo-4 would increase the efficiency of the photoanomerization, but then be removed (tracelessly) through a simple chemical transformation giving access to the target nucleotide structure (β -ribo-4). These considerations prompted us to further investigate the photochemistry of α -ribo-4 and its prebiotic derivatives.

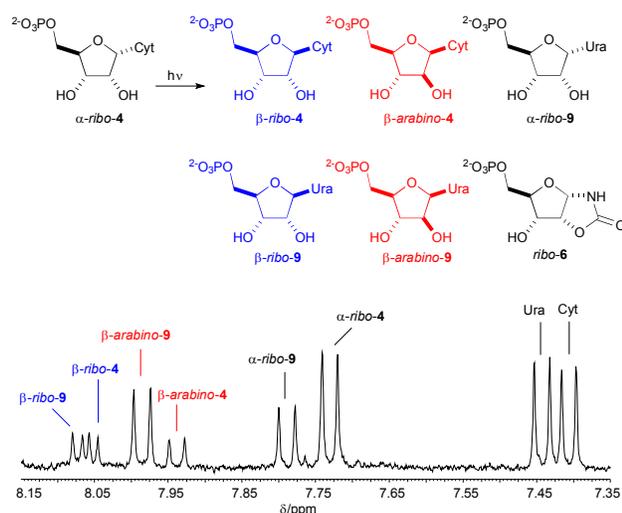


Figure 2. ¹H NMR (600 MHz, D₂O, 7.35–8.20 ppm) spectrum showing concomitant anomerization (blue) and C2'-epimerization (red) observed after irradiation of α -ribo-4 for 16 h at pH 6.5 with λ =254 nm light. Cyt=cytosine, Ura=uracil.

We have previously demonstrated that C2'-acetylation can play an important role controlling the 3'-5'-specific ligation of RNA under prebiotic constraint,¹³ and C2'-acetylation has recently been implicated in (prebiotic) proofing of RNAs during energy-dissipative recycling.¹⁴ Therefore, we considered how C2'-acetylation would effect the photoanomerization of prebiotic α -ribo-4 to biological β -ribo-4. The simplicity and prebiotic plausibility of acyl-transfer reactions suggested that, if direct acetylation of the 2',3'-diol moiety of non-natural α -ribo-4 could be readily and selectively achieved in water, then acetylation would be an excellent prebiotic strategy to facilitate photoanomerization of *ribo*-4 by prohibiting the formation of oxazolidinone *ribo*-6 (Fig. 2). Subsequent slow ester hydrolysis (or rapid ester ammonolysis) would liberate the

free hydroxyl moieties from these simple esters, rendering acetylation ideally suited to act as a traceless directing group in an aqueous environment.¹³ We have previously reported the selective acetylation of β -ribo-4, and, pleasingly, we observed that incubation of α -ribo-4 (100mM) with *N*-acetylimidazole¹⁵ (4 – 10 eq.) in water at near-neutral pH furnished 2',3'-(di-*O*-acetyl)- α -cytidine-5'-phosphate (α -ribo-7) in 60 – 91% yield (Fig. 3; Supplementary Information, Fig. S3), even with only 2 eq. of *N*-acetylimidazole significant 2',3'-di-*O*-acetylation (29%) is observed. Imidazoles are widely exploited in prebiotic activation chemistry and thought to have potentially played a key role in the origins of life,¹⁶ however we observed comparable acetylation of α -ribo-4 with thioacetic acid and ferricyanide¹⁷ (Supplementary Fig. S1).

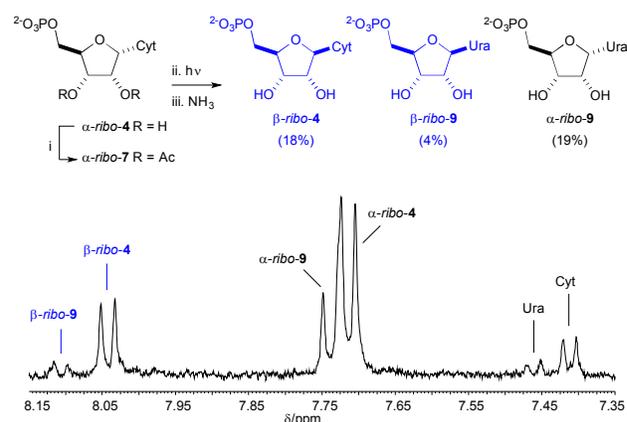


Figure 3. Selective anomerization observed after irradiation of α -ribo-7 for 16 h at pH 6.5 with λ =254 nm light, followed by acetyl cleavage yielding 22% β -ribo-4 and β -ribo-9. i) *N*-acetyl imidazole, H₂O, pH 8, 91%; or AcSH, K₃[Fe(CN)₆], H₂O pH 8.0, 81% Cyt=cytosine, Ura=uracil.

Importantly, acetylation of the cytosine moiety of α -ribo-4 was not observed in water, even with a large excess of acetylating agent (20 eq.). By comparison, under traditional acetylation conditions (pyridine, Ac₂O) high yielding conversion to triacetylated nucleotide α -ribo-8 was found to occur with concomitant nucleobase modification (Fig. 4; Supplementary Information, Fig. S4). The difference in product distribution further suggests that selective aqueous acetylation, which does not prevent the Watson-Crick base pairing of canonical nucleotides, may have played a role in the first steps towards establishing biochemistry.

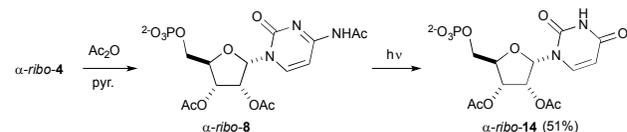


Figure 4. Acetylation of α -cytidine-5'-phosphate α -ribo-4, in pyridine (pyr.) and subsequent rapid deamination upon UV irradiation (λ =254 nm) in water.

The selective di-*O*-acetylation of α -ribo-4 in water spurred us to investigate the UV irradiation of α -ribo-7. Accordingly, α -ribo-7 (2.5–5mM) was irradiated with UV light (principle emission at λ =254



nm) for 16 h in aqueous solution at pH 6.5 or 8.0. The products of irradiation were then subjected to ammoniacal deacetylation, and we were pleased to observe 22% (pH 6.5) and 17% (pH 8.0) anomerization to yield the natural cytidine and uridine ribonucleotides (β -*ribo*-4/ β -*ribo*-9, Fig. 2, blue). Acetylation inhibited the formation of oxazolidinone *ribo*-6 by blocking the C2'-hydroxyl group, but remarkably, acetylation also completely suppressed C2'-epimerization. It is of note suppressing C2'-epimerization preserves the *ribo*-stereochemistry of α -*ribo*-7 (Fig. 3), and prevents formation of *arabino*-isomers during photoanomerization.

We next turned our attention to the role of the 5'-phosphate in the irradiation of α -*ribo*-7. Selective tri-*O*-acetylation of α -cytidine (α -*ribo*-10) was readily observed upon reaction *N*-acetylimidazole (10 eq.) in water at pH 8 to furnish 2',3',5'-(triacetyl)- α -cytidine (α -*ribo*-11) in 90% yield (65% isolated yield, Fig. 5). However, interestingly, subsequent irradiation – at initial pH 6.5 – now lead to significant levels of ester hydrolysis and concomitant formation of oxazolidinone by-product *ribo*-12 (33%; Supplementary information Fig. S9). This hydrolysis had not been observed upon irradiation of α -*ribo*-7 at pH 6.5, and was significantly suppressed at pH 8.0, which is in line with the buffering capacity of the 5'-phosphate moiety. We recognized that Dewar pyrimidine intermediates are observed during the photochemical irradiation of pyrimidine nucleotides,^{2,11,18} and therefore suspected that hydrolysis was incurred upon pH fluctuation during irradiation. Photochemical irradiation was suspected to yield transient concentrations of hydroxide capable of hydrolyzing the ester moieties, for example, upon Dewar pyrimidine **13** C5-protonation (Fig. 5). This suggested that efficient acetyl protection of the 2'-hydroxyl moiety of cytidine required a sufficient buffering capacity during irradiation to prevent photochemical induced pH fluctuations. To test this hypothesis we next introduced phosphate buffer into the irradiation of α -*ribo*-11, and we were pleased to observe that near complete ester stability was now observed during a comparable irradiation (Supplementary information Fig. S9).

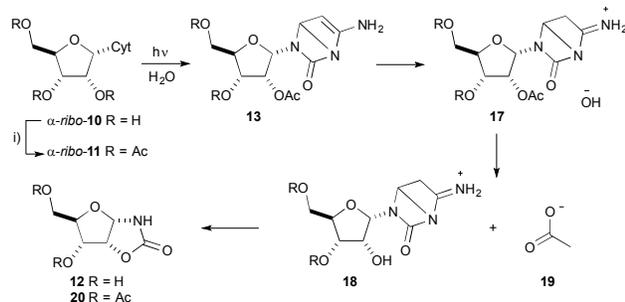


Figure 5. Photochemical induced ester hydrolysis and oxazolidinone formation upon irradiation of cytidine nucleotides.¹⁵ i) *N*-acetyl imidazole, H₂O, pH 8, 90%. Cyt=cytosine.

Finally, having observed the suppression of C2'-epimerization upon diol acetylation, we investigated the effect of nucleobase acetylation. Nucleobase acetylation had not been observed upon prebiotic (aqueous) acetylation of α -*ribo*-4, but was facile under non-aqueous acetylation protocols. Numerous investigations into

prebiotic chemistry have proposed non-aqueous reaction steps, often to overcome perceived problematic effects of reaction modulation by water and the so-called “water problem”.¹⁹ Therefore we were intrigued to investigate the effect of the non-aqueous acetylation on photoanomerization. Interestingly, irradiation of α -*ribo*-8 lead to remarkably low photoanomerization yield with only trace yield observed after 16 h (Fig. 4, Supplementary information S10). However, we observed unprecedentedly high nucleobase photohydrolysis to uridine nucleotide α -*ribo*-14 (51%), alongside significant nucleobase loss (19%) and oxazolidinone *ribo*-6 (6%) after 16 h.

We recognize the contingency of changing nucleotide concentration during acetylation and irradiation. High concentrations are not realistic in the context of the earth Earth's oceans, however are reasonable to consider in fresh water environments, where the process of dilution with water is not difficult to imagine in a prebiotic context. Moreover the UV screening effect of the water column further suggests fresh water, rather than a deep ocean, environment for prebiotic nucleotide photochemistry. Therefore, finally we carried out the one-pot acetylation-photoanomerization-deacetylation (Supplementary Figure S8), which was observed to yield comparable anomerization results. Again no *arabino*-nucleotides were observed to form during irradiation. The influx of high-pH eluent, to liberate or introduce simple nucleophiles (such as ammonia),¹³ are also not thought to be unreasonable and provide a simple, effective and potentially prebiotic method to liberate the vicinal diol of β -*ribo*-4. Given the observed propensity of nucleobase acetylation to inhibit anomerization, it is of note that aqueous (prebiotically plausible) acetylation yielded only α -*ribo*-7 and not α -*ribo*-8. Moreover, as life is a chemical system that has evolved to exploit reactions, products and processes that occur in water, it is should not be seen as a surprise that aqueous reactions (more so than reactions in organic solvents) reflect the structure and pattern of biological chemistry. It is necessary that prebiotic chemistry embraces water as a solvent, and changes the mindset that invokes water as a “problem”. Prebiotic chemistry must come to terms with the principle that water is a part of the solution, and not the problem.

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Conflicts of interest

There are no conflicts to declare.

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