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Synthesis of the fungus metabolite cladosin C⁺

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Cladosin C is one of the few known enaminotetramic acids, isolated from extracts of the deep sea fungus Cladosporium sphaerospermum. It was synthesised in ten steps and 14% overall yield by a late-stage amination of the corresponding 3-acyltetramic acid. This was obtained by a Dieckmann condensation of an N-B-ketoacylaminoester derived from dehydrovalinate and the thioester-terminated side chain containing the stereogenic centre which stemmed from poly-(R)-3-hydroxybutyrate.

Introduction

The cladosins A-D (1-4) (Fig. 1) were isolated as mixtures of two geometric isomers from the deep sea derived fungus Cladosporium sphaerospermum 2005-01-E3 and structurally characterised by D. Li *et al.* in 2013.¹ They are distinguished by their 3-enamino side chains and their biosynthetic derivation from valine, two features quite rare for naturally occurring tetramic acids.² Synthetic 3-enaminotetramic acids were recently shown by Moloney et al. to have a different mode of antibacterial action to the more common analogous 3-acyltetramic



Fig. 1 Structures of cladosins A-D (1-4).

Department of Chemistry, University Bayreuth, Universitaetsstr. 30, D-95440 Bayreuth, Germany. E-mail: Rainer.Schobert@uni-bayreuth.de †Electronic supplementary information (ESI) available: 1H and 13C NMR spectra acids.³ Also, their bioactivity appears to be less pronounced than that of the far better investigated⁴ 3-acyl congeners. Of the cladosins A-D, only cladosin C (3) showed moderate bioactivity, namely an anti-influenza A H1N1 virus effect.¹ Here, we report a short first synthesis of cladosin C.

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Results and discussion

Fig. 2 shows our retrosynthetic approach. Cladosin C was to be prepared by a late-stage amination of the corresponding 3-acyltetramic acid 5 which in turn should be obtainable by a Dieckmann condensation of the N-(β-ketoacyl)amino ester 6. The latter was thought to be accessible by aminolysis of the thioester-terminated side chain 7 with amino ester 8 according to Lev et al.5

The β -ketothioester 7 was prepared in six steps. The cheap, commercially available poly-(R)-3-hydroxybutyrate (9) was depolymerised to give ethyl ester R-10 in 81% yield (Scheme 1).



Retrosynthetic approach to cladosin C (3). Fia. 2

of new compounds. See DOI: 10.1039/c7ob01795b



Scheme 1 Synthesis of side chain thioester 7.

Its stereogenic centre was inverted by quantitatively mesylating the OH group to leave **11** which was treated with CaCO₃ in water to undergo an S_N2-type hydrolysis furnishing ethyl ester *S*-**10** in 82% yield.⁶ TBS protection of its hydroxy group with TBSCl and imidazole afforded ester **12** in 90% yield. The latter was reduced with DIBAL–H to the corresponding aldehyde **13** in 95% yield according to literature.⁷ An *E*-selective Horner– Wadsworth–Emmons olefination of aldehyde **13** with Ley's *t*-butyl 4-diethylphosphono-3-oxobutanthioate (**14**),⁵ prepared by a modified protocol,⁸ afforded β -ketothioester **7** in excellent 92% yield when *n*-BuLi was used as the base. Other bases, such as NaH or KHMDS gave poor yields below 50%.

The required dehydrovaline methyl ester **8** was synthesised in five steps starting from L-valine (**15**), which was converted to the methyl ester hydrochloride with SOCl₂ and then Bocprotected with Boc₂O under basic conditions to give methyl *N*-Boc-valinate (**16**) in 98% yield over two steps. In order to oxidise the amino group of **16** to an imine which can tautomerize to the desired dehydrovaline, we chlorinated the nitrogen with Ca(OCl)₂ in the presence of Al₂O₃ in 86% yield.⁹ Other methods were either cumbersome (*t*-BuOCl) or did not work at all such as the chlorination with NaOCl and Oxon® in the presence of NaCl or trichloroisocyanuric acid. This chlorination only works for N-atoms being part of amides or carbamates.

N-Chloroamino ester **17** was dehydrohalogenated with DBU to afford the protected dehydrovaline methyl ester **18** in 97% yield.¹⁰ Since Ley's aminolysis of thioesters does not work for electron-poor amino groups we had to remove the electron withdrawing Boc-group first. Treatment of **18** with acetyl chloride and MeOH furnished methyl dehydrovalinate hydrochloride **8** × HCl in 99% yield and thus in 81% over all five steps (Scheme 2).

The *N*-acylation of **8** with thioester **7** in the presence of AgO_2CCF_3 according to Ley *et al.*⁵ afforded the *N*-(β -ketoacyl) amino ester **6** in 89% yield when carried out under strict exclusion of moisture (Scheme 3). The Dieckmann cyclisation of **6** was carried out with five equivalents of NaOMe in methanol under reflux for one hour to give the corresponding pure



Scheme 2 Synthesis of methyl dehydrovalinate hydrochloride.



Scheme 3 Ley N-acylation, Dieckmann cyclisation and amination to give 3.

3-acyltetramic acid 5 in 99% yield without further purification. Its conversion into the desired enamine turned out to be far from straightforward. Treatment of 5 with NH₄OH according to Beyer *et al.*¹¹ gave very little product, while its reaction with HMDS failed completely even after three days refluxing in dichloroethane. However, reaction of 5 with 2,4-dimethoxybenzylamine (DMB-NH₂) afforded the DMB-enamine tetramic acid **19** in 40% yield. Moloney *et al.*¹² also showed that the yield of reactions of 3-acyltetramic acids with alkyl amines may vary from 25% to 95%. A simultaneous deprotection of hydroxy and amino groups of **19** with CH₂Cl₂/TFA 9:1 eventually afforded cladosin C (**3**) in 77% yield as a 2:1 mixture of *3E*- and *3Z*-isomers.

Conclusions

The marine fungus metabolite cladosin C (3) was synthesised in ten steps (longest linear sequence) starting from cheap, commercially available compounds. The synthetic approach should be applicable also to other 3-enaminotetramic acids.

Three details of this synthesis are remarkable and probably of a general nature. The *N*-chlorination of methyl valinate was

possible only after a preceding Boc protection and only with $Ca(OCl)_2$ in the presence of Al_2O_3 , however in high yield and under mild conditions. The Dieckmann cyclisation proceeded quantitatively since an excess of base and heating could be applied because of the absence of an acidic H-atom at C5 of the 3-acyltetramic acid 5. Finally, its amination with DMB-NH₂, although of moderate yield in this particular case, took place under mild and water-free conditions and offers various debenzylation options.

Experimental section

General information

¹H and ¹³C NMR spectra were recorded on a Bruker Avance 300 (300 MHz) or on a Bruker Avance DRX-500 spectrometer (500 MHz, with cryoprobe). Chemical shifts are given in parts per million using the residual solvent peak as an internal standard. Coupling constants (1) are quoted in Hz. Abbreviations for the multiplicity used: s singlet, d doublet, t triplet, q quartet, sx sextet, and m multiplet. Mass spectra were measured with a Varian MAT 8500 (EI, 70 eV). High resolution mass spectra were obtained with a UPLC/Orbitrap MS system in ESI mode. IR spectra were recorded with an FT-IR spectrophotometer equipped with an ATR unit. Optical rotations were measured at 589 nm (Na-D line) on a PerkinElmer 241 Polarimeter; $\left[\alpha\right]_{\rm D}$ values are given in 10^{-1} deg cm² g⁻¹. For chromatography silica gel 60 (230–400 mesh) and RP-silica gel C 18 endcapped polygorep 100-50 were used. All reagents were purchased from commercial sources and were used without further purification. Reactions were routinely carried out under an atmosphere of dry argon unless stated otherwise. All glassware was flame-dried before use. Analytical TLC was carried out using Merck silica gel 60GF₂₅₄ pre-coated aluminium-backed plates or Merck 60RP-18 F_{254s} foil plates.

Ethyl (R)-3-hydroxybutyrate (R-10). A suspension of poly-(R)-3-hydroxybutyrate (9) (6.0 g, 79.7 mmol, 1 equiv.) in 1,2dichlorethane (60 mL) was treated with H₂SO₄ (1.3 mL, 23.9 mmol, 0.3 equiv.) and ethanol p.a. (20 mL) and stirred for 75 h at reflux. After adding brine (70 mL) the mixture was stirred for a further 10 min and then extracted with CH₂Cl₂. The combined organic layers were washed with brine, aqueous NaHCO₃ solution, and a second time with brine. After drying over MgSO₄ the solvent was removed under reduced pressure to give R-10 (8.52 g, 81%) as a light orange oil. $R_{\rm f} = 0.30$ (*n*-hexane/ethyl acetate 2:1); $[\alpha]_{D}^{25}$ -36.9 (*c* 1.0 in CHCl₃) (lit⁶ $[\alpha]_{\rm D}$ -44.5 (c 1.0 in CHCl₃)); IR ($\nu_{\rm max}$ /cm⁻¹) 3340, 2978, 1732, 1373, 1296, 1178, 1116, 1068, 1027, 947, 845, 593; ¹H NMR (300 MHz, $CDCl_3$) δ 1.14 (3H, d, J = 6.3 Hz, $CHCH_3$), 1.19 (3H, t, J = 7.2 Hz, CH_2CH_3), 2.35 (1H, dd, J = 16.2, 8.3 Hz, OCOCH), 2.40 (1H, dd, J = 16.2, 4.3 Hz, OCOCH), 3.23 (1H, br. s, OH), 4.08 (2H, q, J = 7.2 Hz, CH_2CH_3), 4.02–4.16 (1H, m, CH); ¹³C NMR (75 MHz, CDCl₃) δ 14.0 (CH₂CH₃), 22.4 (CHCH₃), 42.8 (CH₂CH), 60.4 (CH₂CH₃), 64.0 (CH), 172.5 (CO); HRMS (ESI) m/z $[M + H]^+$ calcd for C₆H₁₃O₃ 133.08592, found 133.08596.

Ethyl (R)-3-mesyloxybutyrate (11). Ester R-10 (2.0 g, 15.13 mmol, 1 equiv.) was dissolved in CH₂Cl₂ (50 mL), cooled to 0 °C and MsCl (1.41 mL, 18.16 mmol, 1.2 equiv.) and NEt₃ (2.52 mL, 18.16 mmol, 1.2 equiv.) were added dropwise via a syringe. The reaction was stirred for 2 h and slowly warmed up to room temperature. The suspension was filtered over Celite and the filtrate was evaporated in vacuo. The residue was desolved in Et₂O (50 mL) and filtered a second time over Celite. After removing the solvent under reduced pressure, 11 (3.14 g, 99%) was obtained as a colourless oil. $R_{\rm f} = 0.39$ (*n*-hexane/ EtOAc 2:1); $[\alpha]_{D}^{25}$ -26.0 (c 1.0 in CHCl₃) (lit¹³ $[\alpha]_{D}$ -32.1 (c 1.0 in CHCl₃)); IR ($\nu_{\text{max}}/\text{cm}^{-1}$) 2984, 1731, 1346, 1300, 1194, 1170, 1105, 1028, 974, 914, 895, 795, 730; ¹H NMR (300 MHz, CDCl₃) δ 1.23 (3H, t, J = 7.2 Hz, CH₂CH₃), 1.46 (3H, d, J = 6.1 Hz, CHCH₃), 2.53 (1H, dd, J = 16.4, 4.6 Hz, OCOCH), 2.73 (1H, dd, J = 16.4, 8.7 Hz, OCOCH), 2.99 (3H, s, SCH₃), 4.12 (2H, q, J = 7.2 Hz, CH_2CH_3), 5.15–5.03 (1H, m, CH); ¹³C NMR (75 MHz, CDCl₃) δ 14.0 (CH₂CH₃), 21.2 (CHCH₃), 38.0 (SCH₃), 41.0 (CH₂CH), 60.7 (CH₂CH₃), 75.8 (CH), 169.5 (CO); HRMS (ESI) $m/z [M + Na]^+$ calcd for C₇H₁₄O₅SNa 233.04542, found 233.04500.

Ethyl (S)-3-hydroxybutyrate (S-10). Mesylate 11 (12.61 g, 60.0 mmol, 1 equiv.) was dissolved in H₂O (120 mL) and CaCO₃ (3.30 g, 33.0 mmol, 0.55 equiv.) was added. After stirring at 80 °C for 3 h the mixture was extracted with ethyl acetate and the combined organic layers were washed with brine and dried over MgSO₄. After removal of the volatiles under reduced pressure S-10 (6.51 g, 82%) was obtained as a colourless oil. $R_{\rm f} = 0.34$ (*n*-hexane/EtOAc 2 : 1); $[\alpha]_{\rm D}^{25} + 33.1$ (*c* 3.0 in CHCl₃) (lit¹⁴ $[\alpha]_{D}^{25}$ +32.8 (c 3.0 in CHCl₃)); IR (ν_{max}/cm^{-1}) 3428, 2979, 1731, 1373, 1296, 1176, 1116, 1067, 1028, 948, 845, 594, 563; ¹H NMR (500 MHz, CDCl₃) δ 1.14 (3H, d, J = 6.2 Hz, CHCH₃), 1.18 (3H, t, J = 7.2 Hz, CH₂CH₃), 2.35 (1H, d, J =4.1 Hz, CH₂CH), 2.37 (1H, d, J = 1.3, CH₂CH), 3.22 (1H, br. s, OH), 4.08 (2H, q, J = 7.2 Hz, CH_2CH_3), 4.01–4.17 (1H, m, CH); ¹³C NMR (75 MHz, CDCl₃) δ 14.1 (CH₂CH₃), 22.4 (CHCH₃), 42.8 (CH₂CH), 60.5 (CH₂CH₃), 64.1 (CH), 172.5 (CO); HRMS (ESI) $m/z [M + H]^+$ calcd for C₆H₁₃O₃ 133.08592, found 133.08589.

Ethyl (S)-3-(t-butyldimethylsilyloxy)butyrate (12). A solution of S-10 (6.13 g, 46.4 mmol, 1 equiv.) in CH₂Cl₂ (120 mL) was cooled to 0 °C and treated with imidazole (3.79 g, 55.7 mmol, 1.2 equiv.) and TBSCl (7.69 g, 51.0 mmol, 1.1 equiv.). The reaction mixture was stirred at room temperature for 15 h and then quenched with NaHCO3 and washed with H2O and brine. The aqueous layer was reextracted with CH₂Cl₂. The combined organic layers were dried over MgSO4 and concentrated under reduced pressure. The crude product was purified by column chromatography, eluting with *n*-hexane/ethyl acetate 8:1, to give 12 (10.28 g, 90%) as a colourless oil. $R_f = 0.49$ (n-hexane/ ethyl acetate 19:1); $[\alpha]_{D}^{25}$ +24.6 (c 1.0 in CHCl₃) (lit¹⁵ $[\alpha]_{D}^{25}$ +24.1 (c 1.0 in CHCl₃)); IR ($\nu_{\rm max}$ /cm⁻¹) 2931, 2858, 1737, 1473, 1376, 1254, 1182, 1081, 1001, 831, 809, 774, 659; ¹H NMR (300 MHz, CDCl₃) δ 0.04 (3H, s, SiCH₃), 0.06 (3H, s, SiCH₃), 0.86 (9H, s, SiC(CH₃)₃), 1.19 (3H, d, J = 6.2 Hz, CHCH₃), 1.26 (3H, t, J = 7.2 Hz, CH_2CH_3), 2.36 (1H, dd, J = 14.4, 5.3 Hz, OCOH), 2.46 (1H, dd, J = 14.4, 7.2 Hz, OCOH), 4.17-4.06 (2H, m, CH₂CH₃),

4.31–4.24 (1H, m, CH); ¹³C NMR (75 MHz, CDCl₃) δ –5.0 (SiCH₃), –4.4 (SiCH₃), 14.3 (CH₂CH₃), 18.1 (SiC(CH₃)₃), 24.0 (CHCH₃), 25.8 (SiC(CH₃)₃), 45.1 (CH₂CH), 60.4 (CH₂CH₃), 65.9 (CH), 171.8 (CO); HRMS (ESI) *m*/*z* [M + H]⁺ calcd for C₁₂H₂₇O₃Si 247.17240, found 247.17202.

(S)-3-(t-Butyldimethylsilyloxy)butanal (13). A solution of 12 (4.0 g, 16.23 mmol, 1 equiv.) in CH_2Cl_2 was cooled to -78 °C and slowly treated with DIBAL-H (17.86 mL, 1 M in Hexan, 1.1 equiv.) by means of a syringe pump over a 90 min period. After this time the reaction was stirred for a further 30 min, treated with methanol (5 mL) and stirred another 15 min. After warming up to room temperature Rochelle salt (40 mg) was added and the mixture was allowed to stir another 20 min. The turbid reaction mixture was extracted with CH₂Cl₂ and the combined organic layers were washed with H₂O and brine and dried over MgSO₄. After the solvent had been removed under reduced pressure product 13 (3.12 g, 95%) was obtained as a colourless oil. $R_{\rm f} = 0.70$ (*n*-hexane/ethyl acetate 9:1); $[\alpha]_{\rm D}^{25}$ +16.7 (c 1.0 in CHCl₃) (lit¹⁶ $[\alpha]_{D}^{25}$ +14 (c 1.0 in CHCl₃)); IR (ν_{max} / cm⁻¹) 2931, 2858, 1728, 1264, 1098, 1026, 833, 774, 680, 560; ¹H NMR (300 MHz, CDCl₃) δ 0.06 (3H, s, SiCH₃), 0.07 (3H, s, SiCH₃), 0.87 (9H, s, C(CH₃)₃), 1.24 (3H, d, J = 6.0 Hz, CHCH₃), 2.46 (1H, ddd, J = 15.8, 5.1, 2.2 Hz, OHCCH), 2.55 (1H, ddd, J = 15.8, 7.1, 2.8 Hz, OHCCH), 4.39-4.32 (1H, m, CH), 9.80 (1H, dd, J = 2.8, 2.2 Hz, CHO); ¹³C NMR (75 MHz, CDCl₃) δ -5.1 (SiCH₃), -4.5 (SiCH₃), 17.9 (C(CH₃)₃), 24.1 (CHCH₃), 25.7 $(C(CH_3)_3)$, 52.9 $(OHCCH_2)$, 64.6 (CH), 202.3 (C=O); MS: m/z $(\%) = 202 (2) [M^+], 185 (9), 159 (100) [C_8H_{19}OSi], 145 (79)$ $[C_6H_{13}O_2Si]$, 119 (51), 101 (47), 75 (94) $[C_2H_7OSi]$, 73 (55) $[C_3H_5O_2]$, 57 (40), 41 (20).

Methyl L-N-(t-butyloxocarbonyl)valinate (16). To a suspension of L-valine (15.0 g, 0.128 mol, 1 equiv.) in methanol SOCl₂ (9.29 mL, 0.13 mmol, 1 equiv.) was slowly added at 0 °C. Stirring was continued for 1 h at room temperature and then for another 22 h at boiling temperature. The solvent was reduced under vacuum to afford the methyl valinate hydrochloride as a white solid which was taken up in CH₂Cl₂ (120 mL) and treated at 0 °C with NEt₃ (34.9 mL, 0.25 mmol, 2 equiv.) and Boc₂O (30.3 g, 0.14 mmol, 1.1 equiv.) and stirred for 17 h at room temperature. Water (100 mL) was added and the resulting mixture was extracted with CH₂Cl₂. To remove residual NEt₃ the organic layers were washed with 1 M citric acid and dried over MgSO₄. All volatiles were removed under vacuum to obtain product 16 (29.0 g, 98%) as a colourless oil. $R_{\rm f} = 0.57$ (*n*-hexane/ethyl acetate 4:1); $\left[\alpha\right]_{\rm D}^{25}$ +10.6 (c 1.0 in CHCl₃) (lit¹⁷ $[\alpha]_{D}^{25}$ +15.9 (c 1.0 in CHCl₃)); IR $(\nu_{\rm max}/{\rm cm}^{-1})$ 3377, 2967, 1744, 1710, 1499, 1366, 1156, 1015, 779; ¹H NMR (300 MHz, CDCl₃) δ 0.80 (3H, d, J = 6.6 Hz, $CHCH_3$), 0.86 (3H, d, J = 6.6 Hz, $CHCH_3$), 1.35 (9H, s, $C(CH_3)_3$), 2.12-1.92 (1H, m, (CH₃)₂CH), 3.64 (3H, s, OCH₃), 4.17-4.08 (1H, m, NCH), 5.03 (1H, br. d, J = 7.7 Hz, NH); ¹³C NMR (75 MHz, $CDCl_3$) δ 17.7 (CHCH₃), 18.9 (CHCH₃), 28.3 (C(CH₃)₃), 31.2 ((CH₃)₂CH), 51.8 (OCH₃), 58.4 (NHCH), 79.7 (C(CH₃)₃), 155.7 (NCO), 172.9 (CO₂CH₃); HRMS (ESI) m/z [M + Na]⁺ calcd for C₁₁H₂₁NO₄Na 254.13594, found 254.13628.

Methyl L-N-chloro-N-(t-butyloxocarbonyl)valinate (17).Ca(OCl)₂ (12.56 g, 87.8 mmol, 3 equiv.) and Al₂O₃ (87.9 g, Brockmann V) were suspended in CH₂Cl₂ and 16 (6.77 g, 29.3 mmol, 1 equiv.) was added. The suspension was stirred for 16 h at 40 °C. The reaction mixture was filtered over Celite and the filtrate was evaporated under reduced pressure to afford product 17 (66.4 g, 86%) as a colourless oil. $R_{\rm f}$ = 0.66 (*n*-hexane/ethyl acetate 4:1); $[\alpha]_{D}^{25}$ -71.5 (*c* 1.0 in CHCl₃); IR $(\nu_{\text{max}}/\text{cm}^{-1})$ 2978, 1746, 1705, 1369, 1280, 1254, 1203, 1157, 1131, 1008, 848, 750; ¹H NMR (300 MHz, CDCl₃) δ 0.93 (3H, d, J = 6.8 Hz, CHCH₃), 0.98 (3H, d, J = 6.8 Hz, CHCH₃), 1.44 (9H, s, C(CH₃)₃), 2.38-2.24 (1H, m, (CH₃)₂CH), 3.69 (3H, s, OCH₃), 4.40 (1H, d, J = 10.0 Hz, NHCH); ¹³C NMR (75 MHz, CDCl₃) δ 19.0 (CHCH₃), 19.6 (CHCH₃), 27.5 (CH(CH₃)₂), 27.9 (C(CH₃)₃), 52.0 (OCH₃), 68.0 (NCH), 83.4 (C(CH₃)₃), 154.7 (NCO), 169.9 (CO_2CH_3); HRMS (ESI) $m/z [M + Na]^+$ calcd for C₁₁H₂₀ClNO₄Na 288.09731, found 288.09672.

Methyl N-(t-butyloxocarbonyl)dehydrovalinate (18). A solution of 17 (885 mg, 3.33 mmol, 1 equiv.) in Et₂O (30 mL) was treated with DBU (547 µL, 3.66 mmoL, 1.1 equiv.) and stirred at room temperature for 3 h. The mixture was filtered and the filtrate was concentrated in vacuo. The crude product was purified by column chromatography, eluting with n-hexane/ethyl acetate 2:1, to give 18 (742 mg, 97%) as a white solid of m.p. 81 °C. $R_{\rm f}$ = 0.37 (*n*-hexane/ethyl acetate 5:1); IR ($\nu_{\rm max}$ / cm⁻¹) 3351, 1705, 1497, 1315, 1258, 1226, 1160, 1092, 1054, 756, 574; ¹H NMR (300 MHz, CDCl₃) δ 1.43 (9H, s, C(CH₃)₃), 1.85 (3H, s, =CCH₃), 2.10 (3H, s, =CCH₃), 3.73 (3H, s, OCH₃), 5.77 (1H, br. s, NH); 13 C NMR (75 MHz, CDCl₃) δ 21.2 $(=CCH_3)$, 22.4 $(=CCH_3)$, 28.2 $(C(CH_3)_3)$, 51.8 (OCH_3) , 80.2 (C(CH₃)₃), 121.5 (NC=C), 145.0 (NC=C), 154.1 (NCO), 165.6 (CO_2CH_3) ; HRMS (ESI) m/z [M + Na]⁺ calcd for C₁₁H₁₉NO₄Na 252.12063, found 252.12028.

Dehydrovaline methyl ester hydrochloride (8 × HCl). Acetyl chloride (9.3 mL, 0.13 mol, 30 equiv.) and dry methanol (5.6 mL, 0.13 mol, 30 equiv.) were carefully added to dioxane at 0 °C and stirred for 5 min. Then carbamate **18** (1.00 g, 4.36 mmol, 1 equiv.) was added in one portion and the resulting mixture was stirred for a further 90 min. The solvent was evaporated under reduced pressure to afford **8** × HCl (715 mg, 99%) as colourless crystals of m.p. 115–120 °C. IR (ν_{max}/cm^{-1}) 2980, 2785, 2594, 1986, 1729, 1622, 1436, 1304, 1228, 1146, 1126, 1043, 877, 770, 617; ¹H NMR (300 MHz, D₂O) δ 2.02 (3H, s, CCH₃), 2.25 (3H, s, CCH₃), 3.84 (3H, s, OCH₃); ¹³C NMR (75 MHz, D₂O) δ 21.6 (CCH₃), 22.0 (CCH₃), 53.0 (OCH₃), 114.8 (NC=C), 152.8 (NC=C), 164.1 (CO₂CH₃); HRMS (ESI) *m/z* [M + H]⁺ calcd for C₆H₁₂NO₂ 130.08626, found 130.08632.

S-t-Butyl (*S,E*)-7-(*t*-butyldimethylsilyloxy)-3-oxo-oct-4-enthioate (7). A solution of *S-t*-butyl 4-diethylphosphonio-3-oxobutanthioate 14^5 (2.87 g, 9.3 mmol, 1.4 equiv.) in THF (90 mL) was cooled to -78 °C and slowly treated with *n*-butyl lithium (7.41 mL, 18.5 mmol, 2.5 M in hexane, 2.8 equiv.). The reaction mixture was stirred for 30 min and then treated with aldehyde 13 (1.34 g, 6.6 mmol, 1 equiv.). After 3 h the reaction was quenched with NH₄Cl, extracted with Et₂O and dried over MgSO₄. The solvent was removed under reduced pressure to

leave 7 (2.18 g, 92%) as a red oil and as a 4:1 mixture of enol and keto tautomers. $R_{\rm f}$ = 0.26 (ketone), 0.41 (enol) (n-hexane/ Et₂O 98:2); $[\alpha]_{D}^{25}$ +7.4 (c 1.0 in CHCl₃); IR (ν_{max}/cm^{-1}) 2957, 2928, 2858, 1656, 1583, 1364, 1255, 1075, 1002, 968, 832, 773; ¹H NMR (300 MHz, CDCl₃) δ enol: 0.03 (3H, s, SiCH₃), 0.04 $(3H, s, SiCH_3)$, 0.88 $(9H, s, SiC(CH_3)_3)$, 1.15 (3H, d, J = 6.1 Hz)CHCH₃), 1.52 (9H, s, SC(CH₃)₃), 2.24–2.35 (2H, m, C=CHCH₂), 3.89 (1H, sx, J = 6.1 Hz, CHOSi), 5.35 (1H, s, COCHCOH), 5.70 (1H, dd, J = 15.4, 1.5 Hz, CH=CH), 6.68 (1H, dt, J = 15.4, 7.6 Hz, CH=CH), 12.56 (1H, d, J = 1.5 Hz, OH); ketone: 0.04 (3H, s, SiCH₃), 0.05 (3H, s, SiCH₃), 0.88 (9H, s, SiC(CH₃)₃), 1.16 (3H, d, J = 6.1 Hz, CHCH₃), 1.55 (9H, s, SC(CH₃)₃), 2.31–2.38 (2H, m, C=CHCH₂), 3.92-3.98 (1H, m, CHOSi), 5.31 (2H, s, COCH₂CO), 5.62 (1H, dd, *J* = 12.1, 1.8 Hz, CH=CH), 6.91 (1H, dt, J = 15.9, 7.5 Hz, CH=CH); ¹³C NMR (75 MHz, CDCl₃) δ enol: -4.8 (SiCH₃), -4.6 (SiCH₃), 24.0 (CHCH₃), 26.0 (SiC $(CH_3)_3$, 29.9 (SiC(CH₃)₃), 30.3 (SC(CH₃)₃), 43.2 (CH=CHCH₂), 48.4 (SC(CH₃)₃), 68.1 (CHOSi), 100.6 (COCHCOH), 126.3 (CH=CHCH₂), 139.4 (CH=CHCH₂), 166.6 (SCO), 196.6 (CHCOH); ketone: -4.8 (SiCH₃), -4.6 (SiCH₃), 24.0 (CHCH₃), 26.0 (SiC(CH₃)₃), 29.8 (SC(CH₃)₃), 29.9 (SiC(CH₃)₃), 42.9 (CH=CHCH₂), 48.9, (SC(CH₃)₃), 56.0 (COCHCOH), 67.6 (CHOSi), 131.6 (CH=CHCH₂), 147.3 (CH=CHCH₂), 191.6 (CHCOH), 192.6 (SCO); HRMS (ESI) $m/z [M + Na]^+$ calcd for C18H34O3NaSSi 381.18901, found 381.18832.

Methyl (S,E)-N-[7-(t-butyldimethylsilyloxy)-3-oxo-oct-4-enoyl] dehydrovalinate (6). A suspension of dehydrovaline methyl ester hydrochloride ($8 \times HCl$) (2.0 g, 12.1 mmol, 2.5 equiv.) and powdered 4 Å molecular sieve in THF (130 mL) was cooled to 0 °C and treated first with NEt₃ (1.74 mL, 12.6 mmol, 2.6 equiv.) and after stirring for 5 min with β -ketothioester 7 (1.73 g, 4.84 mmol, 1 equiv.). The reaction flask was wrapped in light-tight foil and then silver trifluoroacetate (2.14 g, 9.7 mmol, 2.0 equiv.) was added in one portion. After 3 h the reaction mixture was filtered over Celite and the residue was washed with CH₂Cl₂. The solvent was evaporated in vacuo and the remainder was purified by column chromatography, eluting with *n*-hexane/ethyl acetate 9:1, to leave 6 (1.71 g, 89%) as a white solid of m.p. 81 $^{\circ}$ C with a ratio ketone/enol of 2:1. $R_{\rm f} = 0.46$ (*n*-hexane/ethyl acetate 3:1); $[\alpha]_{\rm D}^{25}$ +3.2 (*c* 1.0 in CHCl₃); IR ($\nu_{\text{max}}/\text{cm}^{-1}$) 3243, 2928, 2856, 1721, 1608, 1542, 1307, 1220, 1088, 831, 774; ¹H NMR (300 MHz, CDCl₃) δ ketone: 0.04 (3H, s, SiCH₃), 0.05 (3H, s, SiCH₃), 0.88 (9H, s, SiC(CH₃)₃), 1.17 (3H, d, J = 6.2 Hz, CHCH₃), 1.85 (3H, s, C=CCH₃), 2.14 (3H, s, C=CCH₃), 2.36-2.40 (2H, m, C=CHCH₂), 3.63 (2H, s, COCH₂CO), 3.73 (3H, s, OCH₃), 3.97 (1H, q, J = 5.9 Hz, CHOSi), 6.18 (1H, d, J = 15.9 Hz, CH=CHCH₂), 7.01 (1H, dt, *J* = 15.9, 7.6 Hz, CH=CHCH₂), 8.46 (1H, s, NH); enol: 0.03 (3H, s, SiCH₃), 0.05 (3H, s, SiCH₃), 0.88 (9H, s, SiC(CH₃)₃), 1.15 (3H, d, *J* = 6.1 Hz, CHCH₃), 1.87 (3H, s, C=CCH₃), 2.18 (3H, s, C=CCH₃), 2.23-2.31 (2H, m, C=CHC H_2), 3.75 (3H, s, OCH₃), 3.90 (1H, q, J = 6.4 Hz, CHOSi), 4.94 (1H, s, CHCOH), 5.78 (1H, d, J = 15.4 Hz, CH=CHCH₂), 6.31 (1H, s, NH), 6.60 (1H, dt, J = 15.4, 7.7 Hz, CH=CHCH₂); ¹³C NMR (75 MHz, CDCl₃) δ ketone: -4.7 $(SiCH_3)$, -4.3 $(SiCH_3)$, 18.2 $(SiC(CH_3)_3)$, 21.3 $(C=CH_3)$, 22.4

(C=CH₃), 24.0 (CHCH₃); 25.9 (SiC(CH₃)₃), 42.9 (CH=CHCH₂), 45.7 (COCH₂CO), 52.0 (OCH₃), 67.5 (CHOSi), 121.2 (NC=C), 131.9 (CH=CHCH₂), 144.5 (NC=C), 148.1 (CH=CHCH₂), 164.5 (COOCH₃), 165.2 (NCO), 195.7 (COCH₂CO); enol: -4.7 (SiCH₃), -4.3 (SiCH₃), 21.3 (C=CH₃), 22.4 (C=CH₃), 24.0 (CHCH₃); 25.9 (SiC(CH₃)₃), 39.7 (SiC(CH₃)₃), 49.4 (CH=CHCH₂), 52.0 (OCH₃), 67.5 (CHOSi), 91.2 (COCHCOH), 121.2 (NC=C), 126.9 (CH=CHCH₂), 137.0 (CH=CHCH₂), 144.5 (NC=C), 164.5 (COOCH₃), 165.2 (NCO), 195.7 (COCH₂CO); HRMS (ESI) m/z [M + H]⁺ calcd for C₂₀H₃₅NO₅Si 398.23573, found 398.23503.

3-[(S,E)-5-(t-Butyldimethylsilyloxy)hex-2-enoyl]-5-(propan-2yliden)-pyrrolidin-2,4-dione (5). A solution of β -ketoamide 6 (100 mg, 0.25 mmol, 1 equiv.) in methanol (10 mL) was treated with NaOMe (68 mg, 1.26 mmol, 5 equiv.) and refluxed for 1 h. The reaction was quenched with H₂O and 10% HCl, extracted with Et₂O and washed with brine. After drying with MgSO₄ the solvent was removed under reduced pressure to afford 5 (92 mg, 99%) as a yellow oil. $R_f = 0.54$ (tailing) (CH₂Cl₂/methanol 95:5); $[\alpha]_{D}^{25}$ +9.4 (c 1.0 in methanol); IR (ν_{max} /cm⁻¹) 3195, 2955, 2930, 2858, 1694, 1646, 1583, 1374, 1252, 1128, 1088, 991, 833, 806, 773, 662; ¹H NMR (500 MHz, MeOD) δ 0.07 (3H, s, SiCH₃), 0.08 (3H, s, SiCH₃), 0.89 (9H, s, SiC(CH₃)₃), 1.20 (3H, d, J = 6.1 Hz, CHCH₃), 1.89 (3H, s, C=CCH₃), 2.25 (3H, s, C=CH₃), 2.40–2.52 (2H, m, CH=CHCH₂), 4.05 (1H, q, J = 6.1Hz, CHOSi), 7.23 (2H, br. s, CH=CH); ¹³C NMR (75 MHz, DMSO-d₆) δ *E-isomer*: -4.8 (SiCH₃), -4.5 (SiCH₃), 17.9 $(SiC(CH_3)_3)$, 18.2 (C=CCH₃), 21.2 (C=CCH₃), 24.0 (CHCH₃), 25.8 (SiC(CH₃)₃), 42.8 (CH=CHCH₂), 67.5 (CHOSi), 100.8 (NCOC), 122.7 (CH=CHCH₂), 125.0 (NC=C), 129.8 (NC=C), 147.0 (CH=CHCH₂), 164.5 (NCO), 172.3 (COC=CCH₃), 175.8 (C=COH); Z-isomer: -3.1 (SiCH₃), 17.9 (SiC(CH₃)₃), 18.9 (C=CCH₃), 21.2 (C=CCH₃), 24.0 (CHCH₃), 25.8 (SiC(CH₃)₃), 42.8 (CH=CHCH₂), 65.3 (CHOSi), 103.1 (NCOC), 123.3 (CH=CHCH₂), 126.2 (NC=C), 129.8 (NC=C), 147.6 $(CH = CHCH_2)$, 165.7 (NCO), 171.9 ($COC = CCH_3$), 182.1 (C=COH); HRMS (ESI) m/z [M + H]⁺ calcd for C₁₉H₃₂NO₄Si 366.20951, found 366.20935.

Cladosin C (3). Tetramic acid 5 (90 mg, 0.25 mmol, 1 equiv.) was dissolved in toluene (4 mL). 3 Å molecular sieve pellets and 2,4-dimethoxybenyzlamine (37 μ L, 0.25 mmol, 1 equiv.) were added and the mixture was refluxed for 80 min. The resulting solution was filtrated and washed with CH₂Cl₂. Upon removal of the solvent, tetramic acid **19** (50 mg, 40%) was obtained as a yellow oil. The crude product was used for the next step without further purification.

Crude compound **19** (40 mg, 0.08 mmol, 1 equiv.) was added to a solution of TFA (0.2 mL) in CH₂Cl₂ (1.8 mL) and the mixture was stirred at room temperature for 1 h. The solvent was removed as an azeotrope with toluene under reduced pressure and the resulting crude product was purified by RP–column chromatography, eluting with methanol/H₂O 40 : 60, to leave cladosin C (3) (15 mg, 77%) as a pale yellow oil as a 2 : 1 mixture of 3*E*- and 3*Z*- isomers. $R_{\rm f} = 0.25$ (CH₂Cl₂/ methanol 95 : 5); $[\alpha]_{\rm D}^{25}$ +7.9 (*c* 0.1 in methanol) (lit¹ $[\alpha]_{\rm D}^{25}$ +10.5 (*c* 0.1 in methanol)); IR ($\nu_{\rm max}/{\rm cm}^{-1}$) 3204, 2973, 1659, 1615,

1512, 1457, 1381, 1289, 1203, 1137, 1034, 837, 801, 722, 618, 585, 557; ¹H NMR (500 MHz, DMSO-d₆) δ *E-isomer*: 1.10 (3H, d, J = 6.2 Hz, CHCH₃), 1.74 (3H, s, C=CCH₃), 2.15 (3H, s, C=CCH₃), 2.29-2.34 (2H, m, CH=CHCH₂), 3.72-3.79 (1H, m, CHOH), 4.71 (1H, d, J = 5.0 Hz, OH), 6.87-6.98 (1H, m, CH=CHCH₂), 7.42 (1H, d, J = 16.2 Hz, CH=CHCH₂), 8.70 (1H, br. s, NH2), 9.21 (1H, s, NH), 9.88 (1H, br. s, NH2); Z-isomer: 1.10 (3H, d, J = 6.2 Hz, CHCH₃), 1.73 (3H, s, C=CCH₃), 2.12 (3H, s, C=CCH₃), 2.29-2.34 (2H, m, CH=CHCH₂), 3.72-3.79 (1H, m, CHOH), 4.71 (1H, d, J = 5.0 Hz, OH), 6.87–6.98 (1H, m, CH=CHCH₂), 7.42 (1H, d, J = 16.2 Hz, CH=CHCH₂), 8.60 (1H, br. s, NH₂), 9.25 (1H, s, NH₂), 9.39 (1H, br. s, NH); $^{13}\mathrm{C}$ NMR (75 MHz, DMSO-d₆) δ *E-isomer*: 18.3 (C=CCH₃), 21.3 (C=CCH₃), 23.8 (CHCH₃), 43.3 (CH=CHCH₂), 66.0 (CHOH), 96.8 (C=CNH₂), 118.3 (C=CCH₃), 123.2 (CH=CHCH₂), 130.2 (C=CCH₃), 142.1 (CH=CHCH₂), 161.2 (C=CNH₂), 168.3 (NHCO), 187.2 (COC=CCH₃); Z-isomer: 18.4 (C=CCH₃), 21.2 (C=CCH₃), 23.8 (CHCH₃), 43.4 (CH=CHCH₂), 66.0 (CHOH), 96.5 (C=CNH₂), 118.6 (C=CCH₃), 123.5 (CH=CHCH₂), 130.8 (C=CCH₃), 142.1 (CH=CHCH₂), 161.4 (C=CNH₂), 171.6 (NHCO), 183.7 (COC=CCH₃); HRMS (ESI) m/z [M + H]⁺ calcd for C₁₃H₁₉O₃N₂ 251.13902, found 251.13831.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

1 G. Wu, X. Sun, G. Yu, W. Wang, T. Zhu, Q. Gu and D. Li, *J. Nat. Prod.*, 2014, 77, 270–275.

- 2 For examples of 3-enaminotetramic acids see:
 (a) L. Hagmann and F. Jüttner, *Tetrahedron Lett.*, 1996, 37, 6539–6542;
 (b) C. W. Holzapfel, R. D. Hutchison and D. C. Wilkins, *Tetrahedron*, 1970, 26, 5239–5246.
- 3 (a) Y.-C. Jeong, M. Anwar and M. G. Moloney, *Bioorg. Med. Chem. Lett.*, 2014, 24, 1901–1906; (b) S. W. B. Tan, C. L. L. Chai and M. G. Moloney, *Org. Biomol. Chem.*, 2014, 12, 1711–1716.
- 4 (a) B. J. L. Royles, *Chem. Rev.*, 1995, 95, 1981–2001;
 (b) R. Schobert, *Naturwissenschaften*, 2007, 94, 1–11;
 (c) R. Schobert and A. Schlenk, *Bioorg. Med. Chem.*, 2008, 16, 4203–4221; (d) M. Petermichl and R. Schobert, *Synlett*, 2017, 654–663.
- 5 S. V. Ley, S. C. Smith and P. R. Woodward, *Tetrahedron*, 1992, **48**, 1145–1174.
- 6 A. J. Carnell, R. Head, D. Bassett and M. Schneider, *Tetrahedron: Asymmetry*, 2004, **15**, 821–825.
- 7 R. A. Fernandes and S. V. Mulay, *J. Org. Chem.*, 2010, 20, 7029–7032.
- 8 M. Petermichl, S. Loscher and R. Schobert, *Angew. Chem.*, *Int. Ed.*, 2016, **34**, 10122–10125.
- 9 O. V. Larionov, S. I. Kozhushkov and A. de Meijere, *Synthesis*, 2003, 1916–1919.
- 10 S.-P. Lu and A. H. Lewin, *Tetrahedron*, 1998, 54, 15097–15104.
- 11 C. Beyer, K. Woithe, B. Lüke, M. Schindler, H. Antonicek and J. Scherkenbeck, *Tetrahedron*, 2011, **67**, 3062–3070.
- 12 Y.-C. Jeong, M. Anwar, Z. Bikadi, E. Hazai and M. G. Moloney, *Chem. Sci.*, 2013, 4, 1008–1015.
- 13 H. Urata, D. Goto and T. Fuchikami, *Tetrahedron Lett.*, 1991, 26, 3091–3094.
- 14 J. S. Yadav, S. Nanda, P. T. Reddy and A. B. Rao, *J. Org. Chem.*, 2002, **67**, 3900–3903.
- 15 R. A. Fernandes and S. V. Mulay, *J. Org. Chem.*, 2010, 75, 7029–7032.
- 16 S. Wattanasereekul and M. E. Maier, *Adv. Synth. Catal.*, 2004, **346**, 855–861.
- 17 J. Zheng, B. Yin, W. Huang, X. Li, H. Yao, Z. Liu, J. Zhang and S. Jiang, *Tetrahedron Lett.*, 2009, **50**, 5094–5097.