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Highly Regioselective α -Formylation and α -Acylation of BODIPY Dyes via Tandem Cross-Dehydrogenative Coupling with *in situ* Deprotection

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A metal-free C-H formylation and acylation of BODIPY dyes using a variety of dioxolane derivatives as aldehyde equivalent is reported, providing a postfunctionalization method for controllable synthesis of BODIPYs with carbonyl groups at 3,5positions via a radical process. The photophysical properties of resultant dyes from this efficient one-pot, chemo- and siteselective transformation have been studied.

Introduction

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Small-molecule fluorophores are critically important tools to investigate biological processes and are prevalent in many modern applications, including bioimaging, sensing and therapy.¹ Recently, boron dipyrromethene (BODIPY)² dyes have been widely used in highly diverse research fields,³⁻⁶ due to their excellent spectroscopic and photophysical properties. Rapid and reliable access to functionalized BODIPY chromophores,7,8 therefore, greatly expedites their applications in chemical biology and materials chemistry. To this end, aldehydes and ketones are versatile and high-value functional handles that can be rapidly elaborated or tethered to compounds of biological interest.⁹ The synthesis of α acylated BODIPYs from ready-made BODIPY frameworks has not yet been described. However, acylation with acid chloride through traditional Friedel-Crafts electrophilic aromatic substitution using BF3 OEt2 as Lewis acid catalyst gave regioselectively 2-acylated BODIPYs due to the least positive charge on the 2(,6)-position(s) of the BODIPy core (Scheme 1a).^{10a} A potential, as yet not reported, *de novo* synthesis route toward α -acylated BODIPYs might comprise direct (di)acylation of dipyrromethane, followed by classic oxidation, basepromoted deprotonation and complexation with BF3.OEt2 (Scheme 1b). However, mixtures of products were formed

during the direct acylation of dipyrromethane using acyl chloride.^{10b} In addition, limited stability of dipyrromethanes and their acylated derivatives further complicated the synthesis. Formylation of dipyrromethanes which is highly α -regioselective, followed by standard oxidation, base-promoted deprotonation and complexation with BF₃.OEt₂ yielded 3(,5)-(di)formylated BODIPYs.^{9d,e} A fully alkylated BODIPY having a formyl group at the 3-position was created by 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ) oxidation of the corresponding 3-methyl group.^{9g}



Scheme 1. Reported β -acylation of BODIPY (a), potential α -acylated BODIPYs through *de novo* synthesis (b) and our tandem cross-dehydrogenative coupling between BODIPY and 1,3-dioxolane derivative with *in situ* deprotection (c). HAT: hydrogen atom transfer.

Due to the most positive charge of α -position of BODIPY core, various α -regioselective postfunctionalizations, such as S_NAr reaction¹¹ and radical reaction,¹² have been developed. Particularly, radical reactions on BODIPYs have received increasing interest due to their advantages of high reactivity, high regioselectivity and atom-economy. The highly regioselective cross-dehydrogenative coupling (CDC) has played a vital role in the construction of C–C^{12a-e} and C–X (N, S)^{12f-h} bonds on BODIPY core. Acetals and hemiacetals are very common protective groups of carbonyls. The weak C–H bonds

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Email: haoehong@ahnu.edu.cn; jiao421@ahnu.edu.cn; noel.boens@kuleuven.be Electronic Supplementary Information (ESI) available: Experimental details, spectroscopic/photophysical properties of selected BODIPYs, NMR and HRMS spectra for all new compounds. CCDC 1811936-1811938 and 1815587. For ESI and crystallographic data in CIF or other electronic format, see DOI: 10.1039/x00xx00000x

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 $\boldsymbol{\alpha}$ to both O atoms in acetals and hemiacetals can undergo activation through hydrogen atom transfer in the presence of reactive radical species.¹³ We envisioned a complementary approach that harnesses the direct activation of weak C-H bonds in 1,3-dioxolane derivatives and couples these intermediates with a CDC mechanism to achieve α -formylation and α -acylation of BODIPYs after in situ deprotection (Scheme 1c).

Results and discussion

We explored the viability of the above process with mesomesityl BODIPY 2a and 1,3-dioxolane as substrates in the presence of a radical initiator. To our delight, when the reaction was conducted using Bu₄NI (20 mol%) as catalyst and tert-butyl hydroperoxide (TBHP, 4 equiv) as oxidant at 90 °C for 8 h, the major product 3a was isolated in 56% yield (Table 1, entry 1). Other catalysts, including Bu₄NBr, KI, Pd(OAc)₂ and Cu(OAc)₂, gave lower yields (entries 2-5). In the absence of catalyst, the yield of corresponding product 3a was drastically decreased to 15% yield (entry 6). Reducing the amount of catalyst to 10 mol% also decreased the yield and, conversely, no significant improvement in product yield was observed with 30 mol% of the catalyst (entries 7-8). These results demonstrate that catalysts play an important role in this transformation. Subsequently, other oxidants were studied including DDQ, K₂S₂O₈ and even its analogue di-tert-butyl peroxide (DTBP) either failed in promoting this reaction or

Table 1. Optimization of the reaction conditions^a

1

	$(\mathbf{x}_{\mathbf{N}},\mathbf{y}_{\mathbf{N}})$	+ 000 - 2a	catalyst oxidant temp. F F 3a	-0
entry	catalysts	oxidants	temp. (°C)	yield [%)] ^b
1	Bu₄NI	твнр	90	56
2	Bu₄NBr	TBHP	90	36
3	KI	TBHP	90	< 5
4	Pd(OAc) ₂	TBHP	90	25
5	Cu(OAc) ₂	TBHP	90	32
6		TBHP	90	15
7	Bu₄NI	TBHP	90	33 ^c
8	Bu ₄ NI	TBHP	90	53 ^d
9	Bu ₄ NI	$K_2S_2O_8$	90	NR
10	Bu ₄ NI	DDQ	90	NR
11	Bu ₄ NI	DTBP	90	18
12	Bu ₄ NI	TBHP	90	33 ^e
13	Bu ₄ NI	TBHP	90	52 ^f
14	Bu ₄ NI	TBHP	80	45
15	Bu ₄ NI	TBHP	100	53

 \downarrow

^a Reaction conditions: 1a (0.1 mmol), 3 mL of solvent (1,3-dioxolane serves both as reagent and solvent), catalyst (0.02 mmol), oxidant (4 equiv), 8 h. ^b Isolated yield. ^c 10 mol% catalyst. ^d 30 mol% catalyst. ^e 3 equiv TBHP. ^f 5 equiv TBHP. TBHP tert-butyl hydroperoxide (70% in aqueous solution), DTBP = di-tert-butyl peroxide. NR = no reaction.

showed inferior results (entries 9-11).

We further optimized the ratio of TBHP (6Atres/92213) 278 found the optimal ratio of TBHP was 4 equiv. After a brief survey of reaction temperature, we found that the optimal reaction temperature was 90 °C (entries 14-15). Finally, we obtained the optimized reaction conditions to be 0.2 equiv of Bu₄NI, 4 equiv of TBHP, at 90 °C in 8 h.









Scheme 4. One-pot synthesis of BODIPYs 5 from BODIPY 1a and 1,3-dioxolane 2a and its derivatives 2b, 2d and 2e.

Next, we evaluated a number of Lewis acids for hydrolysis of the dioxolane functional group on BODIPY 3a. Trifluoroacetic acid (TFA) was found to be able to convert **3a** to α formylBODIPY 4a in 90% yield at room temperature (Scheme 2). Other reagents including HCl aqueous solution and BF₃.Et₂O were also effective in this deprotection reaction (details in ESI). A one-pot reaction for synthesis of 4a from BODIPY 1a was then developed without the isolation of 3a (Scheme 2), from which 4a was obtained with 6 equiv of TFA in 48% yield. The regiochemistry of 4a was confirmed by XRD structure determination (Fig. 1).

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To test the versatility of this one-pot reaction, mesophenylBODIPY 1b and meso-2,6-dichlorophenylBODIPY 1c reacted with 1,3-dioxolane 2a, which, after in situ deprotection, provided the α -formylated products **4b** and **4c** in 43 and 38% yields, respectively (Scheme 3a). Next, 2substitued 1,3-dioxolane derivatives 2b-f (Fig. S1, ESI) were applied to obtain α -acylated BODIPYs, which showed higher yields than that of 1,3-dioxolane 2a: BODIPYs 4d-h with alkylor aryl-substituted carbonyl groups at the $\alpha\text{-position}$ were obtained in 40-55% yields (Scheme 3a). Unsymmetric BODIPY 1d was also suitable for this reaction, giving 4i in 34% yield using 1,3-dioxolane 2a (Scheme 3b). By simply extending the reaction time to 16 h followed by hydrolysis in dichloromethane in the presence of TFA, the corresponding 3,5-diacylation products were major products and 5a-d were regioselectively produced from 1a in 23-45% yields, respectively (Scheme 4).

Finally, using **4a** as an example, further transformation of the resultant dyes was demonstrated (Scheme 5). BODIPY **4a** smoothly reacted with diethylamine in DMF at room temperature for 2 h to give BODIPY **6** in 64% yield through oxidative nucleophilic hydrogen substitution at the 5-position of the BODIPY core (Scheme 5a).^{12b} A novel α -meso directedly linked BODIPY dimer **7** was also synthesized in 78% overall yield from the acid-catalyzed condensation of BODIPY **4a** with pyrrole (to give intermediate **8**),¹⁵ followed by DDQ oxidation and subsequent difluoroboration (Scheme 5b, Fig. 1d).



Figure 1. Top and front views of XRD structures of BODIPY 4a (a), 4b (b), 5c (c) and 7 (d). C, light gray; N, blue; B, dark yellow; F, bright green, O, red. Hydrogen atoms have been removed for clarity.

Crystals of 3-formyIBODIPYs **4a** and **4b**, 3,5-diacylated BODIPY **5c** and BODIPY dimer **7** (Figure 1) suitable for XRD analysis were obtained. As usual, B atoms all have slightly distorted tetrahedron geometry with the B–N distances within 1.53–1.57 Å (Table S1, ESI). The dihedral angle of two pyrrole

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rings in the crystal structure of **4b** is only 2.7°, indicating an almost planar BODIPY core. Slight distortions were found of 44a and **5c** with those dihedral angles of 9.8° and 19.3°, respectively. The dihedral angles between the *meso*-aryl group and the BODIPY core are around 76°, 55°, 80° and 83° for **4a**, **4b**, **5c** and **7**, respectively. The dihedral angle of the two BODIPY core planes in dimer **7** is 61.9°, whereas each BODIPY core also has an almost planar plane with the dihedral angle of the two pyrrole rings of 6.1° and 12.1°, respectively.

Fundamental spectroscopic properties of these newly synthesized BODIPYs were investigated in dichloromethane and are summarized in Table 2. In comparison with BODIPYs 1, BODIPYs **4** generally led to red shifts of the absorption (λ_{abs}^{max}) and fluorescence emission (λ_{em}^{max}) maxima of approximately 15-20 nm, wheras disubstituted BODIPYs 5 provided additional red shifts of about 10-30 nm. Most BODIPYs 4 and 5 showed intense fluorescence emission. However, electron-rich 4methoxybenzoyl substituted BODIPYs 4g and 5d were weakly fluorescent. Diethylamine substituted BODIPY 6 displayed broad absorption bands with λ_{abs}^{max} around 470 nm (Fig. S2, ESI), which is about 50 nm blue-shifted compared with λ_{abs}^{max} of starting BODIPY 4a. With the increase of the polarity of the solvent from cyclohexane to methanol, the λ_{em}^{max} values of **6** were significantly blue-shifted from 560 nm (in cyclohexane) to 519 nm (in methanol) and the fluorescence quantum yields were reduced from 0.18 (in cyclohexane) to 0.002 (in methanol) (Table S2, ESI). The spectroscopic features of 6 are similar to our previously reported 3-aminoBODIPY derivatives.16

Table 2. Photophysical properties of BODIPYs in $\mathsf{CH}_2\mathsf{Cl}_2$ at room temperature^a

dyes	$\lambda_{abs}{}^{max}$	$\lambda_{\text{em}}^{\text{max}}$	ϵ_{abs}^{max}	$\Phi^{\rm b}$	Stokes shift ^e
	[nm]	[nm]	[M ⁻¹ cm ⁻¹] ^a		[cm ⁻¹]
1a	500	522	54100	0.74	840
3a	505	522	68700	0.81	610
4a	519	536	52700	0.68	610
4b	518	538	51700	0.25	820
4c	530	551	58500	0.44	720
4d	514	533	49600	0.48	650
4e	514	532	56500	0.36	660
4f	515	534	49600	0.51	690
4g	514	535	49200	0.03	960
4h	514	532	50500	0.45	690
4i	501	549	39000	0.09	1750
5a	548	563	77800	0.40	490
5b	537	555	47700	0.37	600
5c	526	559	42200	0.14	1120
5d	525	554	50300	0.02	1000
6	470	535	42100	0.004	2580
7	514	606	75400	0.005	2950

^a Molar absorption coefficient values rounded to the nearest 100 M⁻¹ cm⁻¹. ^b Fluorescence quantum yields determined using fluorescein (Φ = 0.90 in 0.1 N NaOH aqueous solution) as reference, excited at 480 nm. Standard errors are less than 10%. ^c Stokes shift values rounded to nearest 10 cm⁻¹.

Orthogonal BODIPY dimers have been demonstrated to be good photosensitizers because of their intrinsic intramolecular

charge transfer character through the spin-orbit chargetransfer intersystem crossing (SOCT-ISC) mechanism.^{17,17-19} With dimer **7** in hand, we first measured its photophysical properties. Dimer **7** has a typical λ_{abs}^{max} at 514 nm, and a dramatically red-shifted, but very weak emission with λ_{em}^{max} at 606 nm (Fig. S18, ESI). The extremely low fluorescent quantum yield of 0.005 in dichloromethane inspired us to further study its singlet oxygen generation properties. As shown in Fig. 2, the photooxidation of 1,3-diphenylisobenzofuran (DPBF, a singlet oxygen trap molecule) was monitored at 60 s intervals in the presence of dimer **7**. The calculated singlet oxygen quantum yield for dimer **7** in toluene is 0.15 using Rose Bengal as reference.



Figure 2. (a) Changes in the absorption spectrum of DPBF (5 × 10⁻⁵ M) upon irradiation in the presence of BODIPY dimer 7 (1 × 10⁻⁵ M) in toluene (recorded at 60 s intervals). (b) Plot of change in absorbance of DPBF at 416 nm vs irradiation time (λ_{irr} = 532 nm) in the presence of BODIPY dimer 7 in toluene.

Conclusions

In conclusion, we have developed an efficient protocol for the C–H formylation and acylation of BODIPY dyes using dioxolane derivatives as aldehyde equivalent. Importantly, this reaction is devoid of precious metals and regioselectively occurs at the α -positions of the BODIPY core *via* a radical pathway.

Experimental

General experimental method

Reagents and solvents were used as received from Energy Chemicals (Shanghai, China). All reactions were performed in oven-dried or flame-dried glassware and were monitored by TLC using 0.25 mm silica gel plates with UV indicator (60F-254). ¹H and ¹³C NMR spectra were recorded on a 300, 400 or 500 MHz NMR spectrometer at room temperature. Chemical shifts (δ) are given in ppm relative to CDCl₃ (7.26 ppm for ¹H and 77 ppm for ¹³C) or d6-DMSO (2.54 ppm for ¹H and 39.9 ppm for ¹³C) or to internal TMS (δ = 0 ppm) as internal standard. Data are reported as follows: chemical shift, multiplicity, coupling constants and integration. High-resolution mass spectra (HRMS) were obtained using APCI-TOF in positive mode.

Photophysical Measurements

UV-visible absorption and fluorescence emission spectra were recorded on commercial spectrophotometers (Shimadzu UV-2450 and Edinburgh FS5 spectrometers). All measurements were made at 25 oC, using 5×10 mm cuvettes. Relative fluorescence quantum efficiencies of BODIPY derivatives were obtained by comparing the areas under the corrected emission

fluorescein ($\Phi r = 0.90$ in 0.1 N NaOH aquéous souvente emine fluorescein ($\Phi r = 0.90$ in 0.1 N NaOH aquéous souvents and 10 mm optical path length quartz cuvettes were used. Dilute solutions ($0.01 < A(\lambda ex) < 0.05$) were used to minimize the inner-filter effects. Quantum yields Φx were determined according to equation (1):²⁰

$$\Phi_{x} = \Phi_{r} \times \frac{F_{x}}{F_{r}} \times \frac{1 - 10^{-A_{r}(\lambda_{ex})}}{1 - 10^{-A_{x}(\lambda_{ex})}} \times \frac{n_{x}^{2}}{n_{r}^{2}}$$
(1)

where the subscripts x and r refer respectively to the BODIPY sample x and reference (standard) fluorophore r with known quantum yield Φr in a specific solvent; F stands for the spectrally corrected, integrated fluorescence spectra; A(λex) denotes the absorbance at the used excitation wavelength λex and n represents the refractive index of the solvent (in principle at the average emission wavelength).

Crystallography

Crystals of BODIPYs 4a, 4b, 5c and 7 suitable for X-ray analysis were obtained via the slow diffusion of petroleum ether into their dichloromethane solutions. The vial containing this solution was placed, loosely capped, to promote the crystallization. A suitable crystal was chosen and mounted on a glass fiber using grease. Data were collected using a diffractometer with equipped а graphite crystal monochromator situated in the incident beam for data collection at room temperature. Cell parameters were retrieved using SMART²¹ software and refined using SAINT on all observed reflections. The determination of unit cell parameters and data collections were performed with Mo Ka radiation (λ) at 0.71073 Å. Data reduction was performed using the SAINT software,²² which corrects for Lp and decay. The structure was solved by the direct method using the SHELXS-974 program and refined by least squares method on F², SHELXL-97,²³ incorporated in SHELXTL V5.10.²⁴ CCDC-1811936 (4a), CCDC-1811937 (4b), CCDC-1811938 (5c) and CCDC-1815587 (7) contain the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data request/cif.

Synthesis and Characterization

BODIPYs **1a-d** were synthesized according to literature procedures.^{9d,11d} Compounds **2-f** were prepared from the aldol condensation reaction of aldehyde and ethylene glycol by following literature procedures.²⁵

Synthesis of 3a. BODIPY 1a (78 mg, 0.25 mmol), Bu_4NI (18 mg, 0.05 mmol) and the oxidant tert-butyl hydroperoxide (TBHP, 0.12 mL, 1.25 mmol) were dissolved in 2 mL 1,3-dioxolane 2a, which serves both as reagent and solvent. The reaction mixture was stirred at 90 °C for 8 h. Upon completion, the reaction mixture was cooled to room temperature and was poured into dichloromethane (100 mL), washed three times with water (100 mL), dried over Na_2SO_4 , filtered, and evaporated to dryness. The crude product was purified by

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column chromatographically (silica; petroleum ether/ethyl acetate; 10:1-5:1, v/v) to provide **3a** in 55% yield (52 mg). mp 68.1-70.5 °C. ¹H NMR (300 MHz, CDCl₃) δ 7.92 (s, 1H), 6.94 (s, 2H), 6.67 (s, 1H), 6.63 (d, *J* = 3.5 Hz, 1H), 6.57 (d, *J* = 3.6 Hz, 1H), 6.48 (s, 1H), 6.38 (s, 1H), 4.20 - 4.07 (m, 4H), 2.35 (s, 3H), 2.09 (s, 6H); ¹³C NMR (126 MHz, *d*₆-DMSO) δ 157.02, 147.89, 146.81, 139.44, 136.35, 135.54, 135.37, 131.38, 130.81, 129.94, 129.07, 120.93, 118.13, 97.11, 66.08, 21.58, 20.33. HRMS calcd. for C₂₁H₂₁BF₂N₂O₂Na⁺ [M+Na]⁺ 405.1556, found 405.1557.

Synthesis of 4a from 3a. To a solution of 3a (38 mg, 0.1 mmol) in dichloromethane (3 mL) was added trifluoroacetic acid (TFA, 68 mg, 0.6 mmol). The reaction mixture was stirred at room temperature for 1 h. Upon completion, the reaction mixture was poured into dichloromethane (30 mL), washed three times with water (50 mL), dried over Na₂SO₄, filtered, and evaporated to dryness. The crude product was purified by column chromatographically (silica; petroleum ether/ethyl acetate; 15:1-9:1 v/v) to provide 4a in 90% yield (30 mg). mp 165.7-167.6 °C. ¹H NMR (500 MHz, CDCl₃) δ 10.37 (s, 1H), 8.21 (s, 1H), 7.02 (d, J = 4.2 Hz, 1H), 6.98 (s, 2H), 6.92 (d, J = 4.4 Hz, 1H), 6.67 (d, J = 4.4 Hz, 1H), 6.57 (d, J = 4.2 Hz, 1H), 2.37 (s, 3H), 2.10 (s, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 184.55, 150.86, 150.29, 148.26, 139.94, 138.20, 137.42, 136.57, 134.19, 129.43, 128.84, 127.65, 122.38, 118.76, 21.54, 20.46. HRMS calcd. for C₁₉H₁₈BF₂N₂O [M+H]⁺: 339.1475, found 339.1477.

General Procedure for the One-pot Synthesis of 4a-h. We are using 4a (R = H, Ar = mesityl) as an example to show the general procedure for the preparation of 3-formylBODIPYs 4ac and 3-acylBODIPYs 4d-h: BODIPY 1a (78 mg, 0.25 mmol), Bu₄NI (18 mg, 0.05 mmol) and the oxidant TBHP (0.12 mL, 1.25 mmol) were dissolved in 2a (2 mL). The reaction mixture was stirred at 90 °C for 8 h and then was cooled to room temperature. After addition of 6 equiv of TFA, the reaction mixture was stirred at room temperature for 1 h. Upon completion, the reaction mixture was poured into dichloromethane (100 mL), washed three times with water (100 mL), dried over Na_2SO_4 , filtered, and evaporated to dryness. The crude product was purified by column chromatographically (silica; petroleum ether/ethyl acetate; 15:1-9:1 v/v) to provide 4a in 48% yield (40 mg). mp 165.7-167.6 °C. ¹H NMR (500 MHz, CDCl₃) δ10.37 (s, 1H), 8.21 (s, 1H), 7.02 (d, J = 4.2 Hz, 1H), 6.98 (s, 2H), 6.92 (d, J = 4.4 Hz, 1H), 6.67 (d, J = 4.4 Hz, 1H), 6.57 (d, J = 4.2 Hz, 1H), 2.37 (s, 3H), 2.10 (s, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 184.55, 150.86, 150.29, 148.26, 139.94, 138.20, 137.42, 136.57, 134.19, 129.43, 128.84, 127.65, 122.38, 118.76, 21.54, 20.46. HRMS calcd. for C₁₉H₁₈BF₂N₂O [M+H]⁺: 339.1475, found 339.1477.

4b (R = H, Ar = Ph) was prepared in 43% yield (31 mg) from **1b** and **2a**. mp 153.4-155.2 °C. ¹H NMR (300 MHz, CDCl3) δ 10.39 (s, 1H), 8.23 (s, 1H), 7.67 – 7.57 (m, 5H), 7.16 (d, *J* = 4.3 Hz, 1H), 7.10 (d, *J* = 4.2 Hz, 1H), 6.86 (d, *J* = 4.1 Hz, 1H), 6.75 (d, *J* = 4.2 Hz, 1H); ¹³C NMR (126 MHz, CDCl₃) δ 184.29, 150.03, 149.11, 147.62, 136.97, 136.62, 135.21, 133.12, 131.44, 130.57, 128.87, 128.69, 121.82, 118.21. HRMS calcd. for C₁₆H₁₂BF₂N₂O [M+H]⁺: 297.1005, found 297.1008.

4c (R = H, Ar = 2,6-dichlorophenyl) was prepared in 38% yield (34 mg) from 1c and 2a. mp 146.7-148.9 °C. 1 H NMR (400

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MHz, CDCl₃) δ 10.36 (s, 1H), 8.27 (s, 1H), 7.53-7,45,4(m, 3H), 7.06 (d, *J* = 4.3 Hz, 1H), 6.94 (d, *J* = 4.2 Hz, 1H), 6.93 (0, 99) (27) Hz, 1H), 6.60 (d, *J* = 4.2 Hz, 1H); ¹³C NMR (126 MHz, CDCl₃) δ 183.85, 151.87, 148.35, 142.42, 140.72, 137.52, 134.94, 133.56, 131.73, 130.54, 128.37, 126.97, 122.59, 118.63. HRMS calcd. for C₁₆H₁₀BCl₂F₂N₂O [M+H]⁺: 365.0226, found 365.0229.

4d (R = Me, Ar = mesityl) was prepared in 40% yield (35 mg) from **1a** and **2b**. mp 91.6-93.4 °C. ¹H NMR (300 MHz, CDCl₃) δ 8.16 (s, 1H), 6.97 (s, 2H), 6.92 (s, 1H), 6.85 (s, 1H), 6.63 (s, 1H), 6.55 (s, 1H), 2.70 (s, 3H), 2.37 (s, 3H), 2.09 (s, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 190.78, 150.17, 149.58, 149.23, 149.13, 139.26, 137.15, 136.18, 132.75, 129.38, 128.28, 127.26, 121.84, 121.31, 28.94, 21.07, 19.95. HRMS calcd. for C₂₀H₂₀BF₂N₂O [M+H]⁺: 353.1631, found 353.1635.

4e (R = Bu, Ar = mesityl) was prepared in 48% yield (47 mg) from **1a** and **2c**. mp 148.4-150.2 °C. ¹H NMR (300 MHz, CDCl₃) δ 8.15 (s, 1H), 6.97 (s, 2H), 6.88 (d, *J* = 4.1 Hz, 1H), 6.82 (d, *J* = 4.3 Hz, 1H), 6.61 (d, *J* = 3.6 Hz, 1H), 6.55 (d, *J* = 4.1 Hz, 1H), 3.00 (t, *J* = 7.3 Hz, 2H), 2.37 (s, 3H), 2.09 (s, 6H), 1.78-1.73 (m, 2H), 1.44-1.39 (m, 2H), 0.95 (t, *J* = 7.3 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 193.76, 150.56, 149.84, 149.63, 139.65, 137.73, 137.44, 136.65, 133.08, 129.88, 128.71, 127.81, 122.22, 121.04, 41.12, 26.57, 22.75, 21.55, 20.44, 14.37. HRMS calcd. for C₂₃H₂₆BF₂N₂O [M+H]⁺: 395.2101, found 395.2107.

4f (R = Ph, Ar = mesityl) was prepared in 50% yield (53 mg) from **1a** and **2d**. mp 154.8-156.5 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.14 (s, 1H), 7.95–7.93 (m, 2H), 7.62–7.58 (m, 1H), 7.50–7.46 (m, 2H), 6.99 (s, 2H), 6.85 (d, *J* = 4.3 Hz, 1H), 6.64 (d, *J* = 4.2 Hz, 1H), 6.62–6.59 (m, 2H), 2.38 (s, 3H), 2.14 (s, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 187.45, 171.30, 149.91, 139.21, 137.78, 136.24, 133.69, 132.979, 132.76, 130.13, 129.92, 129.22, 128.43, 128.27, 128.22, 126.97, 121.94, 121.53, 21.08, 20.03. HRMS calcd. for C₂₅H₂₁BFN₂O [M-F]⁺: 395.1729, found 395.1725.

4g (R = 4-methoxyphenyl, Ar = mesityl) was prepared in 55% yield (54 mg) from **1a** and **2e**. mp 143.2-145.8 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.07 (s, 1H), 7.96 (d, *J* = 8.9 Hz, 2H), 6.98 (s, 2H), 6.96 (d, *J* = 9.0 Hz, 2H), 6.82 (d, *J* = 4.3 Hz, 1H), 6.63–6.60 (m, 2H), 6.57 (d, *J* = 4.1 Hz, 1H), 3.89 (s, 3H), 2.38 (s, 3H), 2.14 (s, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 187.03, 164.24, 149.23, 139.58, 137.41, 136.72, 132.88, 132.71, 130.81, 129.93, 128.70, 127.97, 121.32, 114.15, 114.01, 55.92, 21.54, 20.49. HRMS calcd. for C₂₆H₂₄BF₂N₂O₂ [M+H]⁺: 445.1893, found 445.1896.

4h (R = 4-chlorophenyl, Ar = mesityl) was prepared in 47% yield (52 mg) from **1a** and **2f**. mp 163.1-165.4 °C. ¹H NMR (300 MHz, CDCl₃) δ 8.15 (s, 1H), 7.88 (d, J = 8.4 Hz, 2H), 7.46 (d, J = 8.3 Hz, 2H), 6.99 (s, 2H), 6.87 (s, 1H), 6.63-6.60 (m, 3H), 2.38 (s, 3H), 2.13 (s, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 186.66, 150.73, 149.65, 148.59, 139.90, 139.75, 138.09, 137.37, 136.67, 136.62, 133.47, 131.71, 129.75, 129.25, 129.04, 128.76, 127.29, 122.24, 21.53, 20.47. HRMS calcd. for C₂₅H₂₁BClF₂N₂O [M+H]⁺: 449.1398, found 449.1397.

Synthesis of 4i. BODIPY 1d (62 mg, 0.25 mmol), Bu_4NI (18 mg, 0.05 mmol) and the oxidant TBHP (0.12 mL, 1.25 mmol) were dissolved in 2a (2 mL). The reaction mixture was stirred at 90 °C for 8 h and then was cooled to room temperature. After addition of 6 equiv of TFA, the reaction mixture was stirred at room temperature for 1 h. Upon completion, the

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reaction mixture was poured into dichloromethane (100 mL), washed three times with water (100 mL), dried over Na₂SO₄, filtered, and evaporated to dryness. The crude product was purified by column chromatographically (silica; petroleum ether/ethyl acetate; 15:1-9:1 v/v) to provide 4i in 34% yield (25 mg). mp 163.7-165.6 °C. ¹H NMR (300 MHz, CDCl₃) δ 10.03 (s, 1H), 7.81 (s, 1H), 7.44 (s, 1H), 6.80 (s, 1H), 2.64 (s, 3H), 2.44 (q, J = 7.6 Hz, 2H), 2.26 (s, 3H), 1.11 (t, J = 7.6 Hz, 3H); ¹³C NMR (126 MHz, CDCl_3) δ 187.23, 170.19, 143.97, 140.26, 137.57, 133.72, 131.21, 130.96, 122.43, 117.83, 17.34, 13.97, 13.88, 9.72. HRMS calcd. for C14H15BFN2O [M-F]+: 277.1318, found 277.1319.

General Procedure for the One-pot Synthesis of 5a-d. We are using 5a (R = H, Ar = mesityl) as an example to show the general procedure for the preparation of 3,5-diformyIBODIPY 5a and 3,5-diacylBODIPYs 5b-d: BODIPY 1a (78 mg, 0.25 mmol), Bu₄NI (18 mg, 0.05 mmol) and the oxidant TBHP (0.12 mL, 1.25 mmol) were dissolved in 2a (2 mL). The reaction mixture was stirred at 90 °C for 16 h and was cooled to room temperature. After the addition of 6 equiv of TFA, the reaction mixture was stirred at room temperature for 1 h. Upon completion, the reaction mixture was then poured into dichloromethane (100 mL), washed three times with water (100 mL), dried over Na₂SO₄, filtered, and evaporated to dryness. The crude product was purified by column chromatographically (silica; petroleum ether/ethyl acetate; 10:1-5:1 v/v) to provide 5a in 25% yield (22 mg). mp 158.7-160.9 °C. $^1\!H$ NMR (300 MHz, $CDCl_3$) δ 10.47 (s, 2H), 7.11 (d, J = 4.2 Hz, 2H), 7.00 (s, 2H), 6.83 (d, J = 3.9 Hz, 2H), 2.38 (s, 3H), 2.11 (s, 6H); ¹³C NMR (126 MHz, CDCl3) δ 184.00, 154.60, 151.55, 140.19, 138.56, 136.06, 131.47, 128.78, 128.67, 120.34, 21.17, 120.15. HRMS calcd. for C₂₀H₁₇BF₂N₂O₂Na⁺ [M+Na]⁺ 389.1243, found 389.1246.

5b (R = Me, Ar = mesityl) was prepared in 23% yield (20 mg) from **1a** and **2b**. mp 83.5-85.7 °C. ¹H NMR (500 MHz, CDCl₃) δ 6.98 (s, 2H), 6.94 (d, J = 4.4 Hz, 2H), 6.72 (d, J = 4.4 Hz, 2H), 2.82 (s, 6H), 2.37 (s, 3H), 2.09 (s, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 192.53, 154.02, 153.15, 139.68, 138.09, 136.00, 130.54, 129.46, 128.46, 122.85, 29.68, 21.15, 20.06. HRMS calcd. for C₂₂H₂₁BFN₂O₂⁺ [M-F]⁺ 375.1674, found 375.1678.

5c (R = Ph, Ar = mesityl) was prepared in 41% yield (53 mg) from 1a and 2d. mp 149.3-151.8 °C. ¹H NMR (300 MHz, CDCl₃) δ7.91 (d, J = 7.1 Hz, 4H), 7.61-7.56 (m, 2H), 7.48-7.43 (m, 4H), 7.02 (s, 2H), 6.80 (d, J = 3.8 Hz, 2H), 6.63 (d, J = 3.8 Hz, 2H), 2.39 (s, 3H), 2.20 (s, 6H); 13 C NMR (101 MHz, CDCl₃) δ 187.99, 154.22, 151.23, 139.53, 137.31, 136.38, 136.35, 133.91, 130.30, 130.22, 129.44, 129.06, 128.46, 121.80, 21.21, 20.26. HRMS calcd. for C₃₂H₂₆BF₂N₂O₂ [M+H]⁺: 519.2050, found 519.2059.

5d (R = 4-methoxyphenyl, Ar = mesityl) was prepared in 45% yield (65 mg)) from 1a and 2e. mp 133.7-135.9 °C. ¹H NMR (300 MHz, d_6 -DMSO) δ 7.76 (d, J = 8.5 Hz, 4H), 7.11 (d, J = 9.7 Hz, 4H), 7.07 (s, 2H), 6.89 (d, J = 4.0 Hz, 2H), 6.82 (d, J = 3.9 Hz, 2H), 3.85 (s, 6H), 2.37 (s, 3H), 2.16 (s, 6H); ¹³C NMR (126 MHz, $CDCl_3$) δ 186.85, 164.29, 154.67, 139.37, 136.37, 132.71, 132.25, 130.09, 129.38, 128.39, 122.65, 121.08, 113.94, 113.74, 55.55, 21.18, 20.22. HRMS calcd. for C₃₄H₂₉BFN₂O₄⁺ [M-F]⁺ 559.2199, found 559.2198.

Synthesis of BODIPY 6. To a solution of 3-formylBODIPY 4a (0.1 mmol, 33 mg) in N,N-dimethylformanide(DNF, 39RL) was added diethylamine (0.21 mmol, 2.1 equiv). The mixture was stirred at room temperature for the 2 h under air. Upon completion, the reaction mixture was poured into dichloromethane (30 mL), washed three times with water (50 mL), dried over Na₂SO₄, filtered, and evaporated to dryness. product was purified The crude bv column chromatographically (silica; petroleum ether/dichloromethane; 2:1 v/v) to provide 6 in 64% yield (26 mg). mp 212.6-213.4 °C. ¹H NMR (500 MHz, CDCl₃) δ 10.20 (s, 1H), 7.04 (d, J = 3.8 Hz, 1H), 6.93 (s, 2H), 6.71 (d, J = 5.2 Hz, 1H), 6.36 (d, J = 5.3 Hz, 1H), 5.98 (d, J = 3.8 Hz, 1H), 3.91 (s, 4H), 2.34 (s, 3H), 2.08 (s, 6H), 1.42 (t, J = 7.1 Hz, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 183.66, 163.02, 140.51, 138.25, 138.20, 137.40, 136.22, 135.47, 130.20, 128.19, 127.36, 117.92, 117.41, 115.35, 47.32, 21.12, 20.06, 13.77. HRMS calcd. for $C_{23}H_{26}BFN_3O^+$ [M-F]⁺ 390.2147, found 390.2149.

Synthesis of BODIPY 7. To a mixture of 4a (33 mg, 0.1 mmol) and pyrrole (1 mL, 16 mmol) was added TFA (0.02 mL, 0.2 mmol). The reaction mixture was stirred at room temperature for 5 min and was guenched by adding 30 mL aqueous solution of NaOH (0.2 M). The reaction mixture was extracted with CH₂Cl₂ (30 mL) and the organic layer was washed with water (50 mL), dried over anhydrous Na₂SO₄, filtered, and evaporated to dryness. The residue was purified through (silica, column chromatography CH_2Cl_2) to afford dipyrromethane intermediate 8, which was dissolved in 20 mL CH₂Cl₂ and directly used for the subsequent oxidation with DDQ (2,3-dichloro-5,6-dicyano-1,4-benzoquinone, 40 mg, 0.2 mmol) at room temperature for 1 h. The resultant mixture was further treated with triethylamine (1 mL, 7.2 mmol) for 20 min, and was complexed with boron trifluoride etherate (3 mL, 23.9 mmol) for 2 h at room temperature. The solvent was removed under vacuum and the residue was purified by column chromatography on silica gel (ethyl acetate/petroleum ether = 1:5, v/v) to give 7 as a red solid in 78% yield (40 mg). mp 233.4-235.2 °C. ¹H NMR (300 MHz, CDCl₃) δ 7.95 (s, 2H), 7.93 (s, 1H), 7.01 (s, 4H), 6.82 (d, J = 3.8 Hz, 1H), 6.74 (d, J = 3.6 Hz, 1H), 6.59 (d, J = 3.7 Hz, 1H), 6.52 (d, J = 3.6 Hz, 3H), 2.39 (s, 3H), 2.17 (s, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 148.46, 147.59, 146.66, 145.34, 139.33, 136.54, 136.41, 136.29, 136.21, 135.15, 132.15, 131.82, 129.35, 128.45, 128.39, 121.83, 120.53, 118.51, 21.18, 20.07. HRMS calcd. For C₂₇H₂₂B₂F₃N₄⁺ [M-F]⁺ 481.1977, found 481.1979.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- (a) L. D. Lavis and R. T. Raines, ACS Chem. Biol., 2014, 9, 855; (b) H. Zhu, J. Fan, J. Du and X. Peng, Acc. Chem. Res., 2016, 49, 2115; (c) L. He, B. Dong, Y. Liu and W. Lin, Chem. Soc. Rev., 2016, 45, 6449.
- (a) A. Loudet and K. Burgess, *Chem. Rev.*, 2007, **107**, 4891; (b) R. Ziessel, G. Ulrich and A. Harriman, *New J. Chem.*, 2007, **31**, 496; (c) G. Ulrich, R. Ziessel and A. Harriman, *Angew. Chem. Int. Ed.*, 2008, **47**, 1184.
- (a) T. Kowada, H. Maeda and K. Kikuchi, *Chem. Soc. Rev.*, 2015, **44**, 4953; (b) W. Zhang, W. Sheng, C. Yu, Y. Wei, H. Wang, E. Hao and L. Jiao, *Chem. Commun.*, 2017, **53**, 5318; (c) Y. V. Zatsikha, N. O. Didukh, D. Nemez, A. C. Schlachter, P.-L. Karsenti, Y. P. Kovtun, P. D. Harvey and V. N. Nemykin, *Chem. Commun.*, 2017, **53**, 7612; (d) G. Xu, Q. Yan, X. Lv, Y. Zhu, K. Xin, B. Shi, R. Wang, J. Chen, W. Gao, P. Shi, C. Fan, C. Zhao and H. Tian, *Angew. Chem., Int. Ed.*, 2018, **57**, 3626.
- (a) N. Boens, V. Leen and W. Dehaen, *Chem. Soc. Rev.*, 2012, 41, 1130; (b) S. Kolemen and E. U. Akkaya, Coord. *Chem. Rev.*, 2018, 354, 121; (c) L. Niu, Y. Guan, Y. Chen, L. Wu, C. Tung and Q. Yang, *J. Am. Chem. Soc.* 2012, 134, 18928; (d) A. M. Christianson and F. P. Gabbaï, *Chem. Commun.*, 2017, 53, 2471.
- 5 (a) A. Kamkaew, S. H. Lim, H. B. Lee, L. V. Kiew, L. Y. Chung and K. Burgess, *Chem. Soc. Rev.*, 2013, **42**, 77; (b) J. Zhao, K. Xu, W. Yang, Z. Wang and F. Zhong, *Chem. Soc. Rev.*, 2015, **44**, 8904; (c) A. Turksoy, D. Yildiz and E. U. Akkaya, Coord. *Chem. Rev.*, 2019, **379**, 47; (d) N. Kiseleva, M. A. Filatov, M. Oldenburg, D. Busko, M. Jakoby, I. A. Howard, B. S. Richards, M. O. Senge, S. M. Borisov and A. Turshatov, *Chem. Commun.*, 2018, **54**, 1607.
- H. Klfout, A. Stewart, M. Elkhalifa and H. He, ACS Appl. Mater. Interfaces, 2017, 9, 39873.
- 7 (a) H. Lu, J. Mack, Y. Yang and Z. Shen, *Chem. Soc. Rev.*, 2014, **43**, 4778; (b) Y.
 Ni and J. Wu, *Org. Biomol. Chem.*, 2014, **12**, 3774; (c) N. Boens, B. Verbelen and W. Dehaen, Eur. *J. Org. Chem.*, 2015, 6577; (d) V. Lakshmi, M. R. Rao and M. Ravikanth, *Org. Biomol. Chem.*, 2015, **13**, 2501.
- (a) X. Kong, F. Su, L. Zhang, J. Yaron, F. Lee, Z. Shi, Y. Tian and D. R. Meldrum, *Angew. Chem., Int. Ed.*, 2015, 54, 12053; (b) J. Zhang, M. Yang, C. Li, N. Dorh, F. Xie, F. Luo, A. Tiwari and H. Liu, *J. Mater. Chem. B*, 2015, 3, 2173; (c) J. Ahrens, B. Cordes, R. Wicht, B. Wolfram and M. Bröring, *Chem. Eur. J.*, 2016, 22, 10320; (d) X. Zheng, W. Du, L. Gai, X. Xiao, Z. Li, L. Xu, Y. Tian, M. Kira and H. Lu, *Chem. Commun.* 2018, 54, 8834; (e) B. Liu, N. Novikova, M. C. Simpson, M. S. M. Timmer, B. L. Stocker, T. Sohnel, D. C. Ware and P. J. Brothers, *Org. Biomol. Chem.*, 2016, 14, 5205; (f) Z. Zhou, J. Zhou, L. Gai, A. Yuan and Z. Shen, *Chem. Commun.*, 2017, 53, 6621; (g) N. Zhao, S. Xuan, B. Byrd, F. R. Fronczek, K. M. Smith and M. G. H. Vicente, *Org. Biomol. Chem.*, 2016, 14, 6184; (h) W. Wang, M. M. Lorion, O. Martinazzoli and L. Ackermann, *Angew. Chem. Int. Ed.*, 2018, 57, 10554.
- 9 (a) K. Krumova and G. Cosa, J. Am. Chem. Soc., 2010, 132, 17560; (b) L. A. Juárez, A. M. Costero, M. Parra, S. Gil, F. Sancenón and R. Martínez-Máñez, Chem. Commun., 2015, 51, 1725; (c) L. Jiao, C. Yu, J. Li, Z. Wang, M. Wu and E. Hao, J. Org. Chem., 2009, 74, 7525; (d) C. Yu, L. Jiao, H. Yin, J. Zhou, W. Pang, Y. Wu, Z. Wang, G. Yang and E. Hao, *Eur. J. Org. Chem.*, 2011, 5460; (e) S. Madhu, M. R. Rao, M. S. Shaikh and M. Ravikanth, *Inorg. Chem.*, 2011, 50, 4392; (f) S. Zhu, J. Zhang, G. Vegesna, A. Tiwari, F. Luo, M. Zeller, R. Luck, H. Li, S. Green and H. Liu, *RSC Adv.*, 2012, 2, 404; (g) S. Zhu, N. Dorh, J. Zhang, G. Vegeson, H. Li, F. Luo, A. Tiwari, and H. Liu, *Chem. Commun.*, 2011, 47, 3508; (h) E. Palao-Utiel, L. Montalvillo-Jiménez, I. Esnal, R. Prieto-Montero, A. R. Agarrabeitia, I. García-Moreno, J. Bañuelos, I. López-Arbeloa, S. de la Moya and M. J. Ortiz, *Dyes Pigments*, 2017, 141, 286; (i) G. Sathyamoorthi, L. T. Wolford, A. M. Haag and J. H. Boyer, *Heteroat. Chem.*, 1994, 5, 245.
- (a) Mirri, D. C. Schoenmakers, P. H. J. Kouwer, P. Veranič, I. Musevič and B. Štefane, *ChemistryOpen*, 2016, 5, 450; (b) S. Tamaru, L. Yu, W. J. Youngblood, K. Muthukumaran, M.Taniguchi and J. S. Lindsey, *J. Org. Chem.*, 2004, 69, 765.
- (a) B. Verbelen, V. Leen, L. Wang, N. Boens and W. Dehaen, *Chem. Commun.* 2012, **48**, 9129; (b) V. Leen, V. Z. Gonzalvo, W. M. Deborggraeve, N. Boens and
 W. Dehaen, *Chem. Commun*, 2010, **46**, 4908; (c) M. Zhang, E. Hao, J. Zhou, C.
 Yu, G. Bai, F. Wang and L. Jiao, *Org. Biomol. Chem.* 2012, **10**, 2139; (d) X. Zhou,
 C. Yu, Z. Feng, Y. Yu, J. Wang, E. Hao, Y. Wei, X. Mu and L. Jiao, *Org. Lett.*, 2015,
 17, 4632.
- 12 (a) B. Verbelen, S. Boodts, J. Hofkens, N. Boens and W. Dehaen, Angew. Chem. Int. Ed., 2015, 54, 4612; (b) B. Verbelen, L. Cunha Dias Rezenda, S. Boodts, J. Jacobs, L. Van Meervelt, J. Hofkens and W. Dehaen, Chem. Eur. J., 2015, 21, 12667; (c) X. Zhou, Q. Wu, Y. Yu, C. Yu, E. Hao, Y. Wei, X. Mu and L. Jiao, Org.

Lett., 2016, **18**, 736; (d) Y. Yu, L. Jiao, J. Wang, H. Wang, C. Yu, E. Hao and N. Boens, *Chem. Commun.*, 2017, **53**, 581; (e) H. Zhang, Xo Ghers, Common. Zhou, D. Wu, J. You, *Chem. Commun.*, 2018, **54**, 3219; (f) F. Lv, Y. Yu, E. Hao, C. Yu, H. Wang, L. Jiao and N. Boens, *Chem. Commun.*, 2018, **54**, 9059; (g) F. Lv, B. Tang, E. Hao, Q. Liu, H. Wang and L. Jiao, *Chem. Commun.*, 2019, **55**, 1639; (h) F. Ma, L. Zhou, Q. Liu, C. Li and Y. Xie, *Org. Lett.*, 2019, **21**, 733; (i) B. Tang, F. Lv, K. K. Chen, L. Jiao, Q. Liu, H. Wang and E. Hao, *Chem. Commun.*, 2019, **55**, 4691.

- 13 J. M. Ganley, M. Christensen, Y. Lam, Z. Peng, A. R. Angeles and C. S. Yeung, Org. Lett., 2018, 20, 5752.
- 14 W. Pang, X. Zhang, J. Zhou, C. Yu, E. Hao and L. Jiao, *Chem. Commun.*, 2012, **48**, 5437.
- C. Ripoll, C. Cheng, E. Garcia-Fernandez, J. Li, A. Orte, H. Do, L. Jiao, D. Robinson, L. Crovetto, J. A. González-Vera, E. M. Talavera, J. M. Alvarez-Pez, N. Boens and M. J. Ruedas-Rama, *Eur. J. Org. Chem.* 2018, 2561.
- (a) Y. Liu, J. Zhao, A. Iagatti, L. Bussotti, P. Foggi, E. Castellucci, M. Di Donato and K. Han, J. Phys. Chem. C., 2018, 122, 2502; (b) N. Epelde-Elezcano, E. Palao, H. Manzano, A. Prieto-Castañeda, A. R. Agarrabeitia, A. Tabero, A. Villanueva, S. de la Moya, I, López-Arbeloa, V. Martínez-Martínez and M. J. Ortiz, Chem. Eur. J., 2017, 23, 4837.
- (a) M. Bröring, R. Krüger, S. Link, C. Kleeberg, S. Köhler, X. Xie, B. Ventura and L. Flamigni, *Chem. Eur. J.*, 2008, **14**, 2976; (b) B. Ventura, G. Marconi, M. Bröring, R. Krüger and L. Flamigni, *New J. Chem.*, 2009, **33**, 428.
- (a) Y. Cakmak, S. Kolemen, S. Duman, Y. Dede, Y. Dolen, B. Kilic, Z. Kostereli, L. T. Yildirim, A. L. Dogan, D. Guc and E. U. Akkaya, *Angew. Chem., Int. Ed.*, 2011, **50**, 11937; (b) S. Duman, Y. Cakmak, S. Kolemen, E. U. Akkaya and Y. Dede, *J. Org. Chem.*, 2012, **77**, 4516; (c) T. Ozdemir, J. L. Bila, F. Sozmen, L. T. Yildirim and E. U. Akkaya, *Org. Lett.*, 2016, **18**, 4821.
- 19 J. Olmsted, J. Phys. Chem., 1979, 83, 2581.
- 20 (a) R. C. Benson and H. A. Kues, *Phys. Med. Biol.*, 1978, **23**, 159; (b) J. R. Lakowicz, *Principles of Fluorescence Spectroscopy*, 3rd ed.; Springer: New York, 2006.
- 21 SAINT V 6.01 (NT) Software for the CCD Detector System, Bruker Analytical Xray Systems, Madison, WI 1999.
- 22 G. M. Sheldrick, SHELXS-90, Program for the Solution of Crystal Structure, University of Göttingen, Germany, 1990.
- 23 SHELXL-97, Program for the Refinement of Crystal Structure, University of Göttingen, Germany, 1997.
- 24 SHELXTL 5.10 (PC/NT-Version), *Program library for Structure Solution and Molecular Graphics*, Bruker Analytical X-ray Systems, Madison, WI, 1998.
- 25 (a) F. M. Menger and H. Lu. *Chem. Commun.*, 2006, **42**, 3235; (b) Y. S. Hon, C. F. Lee, R. J. Chen and P. H. Szu, *Tetrahedron*, 2001, **57**, 599; (c) C. Mukai, J. W. Cho, L. J. Kim, M. Kido and M. Hanaoka, *Tetrahedron*, 1991, **47**, 3007; (d) B. Wang, P. Li, F. Yu, P. Song, X. Sun, S. Yang, Z. Lou and K. Han, *Chem. Commun.*, 2013, **49**, 1014.