



Pergamon

Discovery of Orally Bioavailable NK₁ Receptor Antagonists

C. Genicot,^{a,*} B. Christophe,^b P. Collart,^d M. Gillard,^b L. Goossens,^c J.-P. Hénichart,^c
M.-A. Lassoie,^a F. Moureau,^a M. Neuwels,^a J.-M. Nicolas,^d P. Pasau,^a
L. Quéré,^a T. Ryckmans,^a F. Stiernet,^c T. Taverne^c
and B. J. Van Keulen^a

^aChemistry Department, UCB Pharma, Chemin du Foriest, B-1420, Braine-l'Alleud, Belgium

^bIn vitro Pharmacology, UCB Pharma, Chemin du Foriest, B-1420, Braine-l'Alleud, Belgium

^cA/R Pharmacology, UCB Pharma, Chemin du Foriest, B-1420, Braine-l'Alleud, Belgium

^dIn vitro DMPK, R&D UCB Pharma, Chemin du Foriest, B-1420, Braine-l'Alleud, Belgium

^eInstitut de Chimie Pharmaceutique A. Lespagnol, Université de Lille II,
3 rue du Professeur Laguesse, F-59006, Lille, France

Received 23 July 2002; revised 30 September 2002; accepted 29 October 2002

Abstract—Benzyloxyphenethylpiperazines are a new class of high affinity NK₁ receptor antagonists. Oral bioavailability and selectivity can be fine tuned by the nature of the substituents on the basic nitrogen atom. Addition of substituents with a carboxylic acid group led to very selective and orally active NK₁ antagonists free of interaction with L-type calcium channels.

© 2003 Elsevier Science Ltd. All rights reserved.

The endogenous undecapeptide Substance P (SP) elicits a wide range of physiological responses both centrally and peripherally. Through binding to NK₁ receptors, SP has been implicated in the induction of neurogenic inflammation, in the transmission of nociceptive stimulus, in the control of vomiting reflex and in affective disorders.¹ Thus, NK₁ receptor antagonists are potentially useful for the treatment of a wide range of clinical diseases such as asthma, chronic pain, emesis, anxiety and depression. The disclosure by Pfizer of the first two non peptide NK₁ antagonists, CP-96,345 and CP-99,994 (**1**)² (Fig. 1), has spurred intensive research in this field. During the last few years, several other NK₁ receptor antagonists have been reported including benzyloxy-aminopiperidines such as L-733,060 **2**³ (Fig. 1). However, clinical development of the early NK₁ antagonists has been hampered by poor bioavailability and/or cardiovascular side effects resulting from interaction with L-type calcium channels. Subsequent efforts have been devoted to the conception of NK₁ antagonists with improved in vivo activity and reduced interaction with calcium channels such as MK-869 **3**⁴ and CP-122,721 **5**.⁵

To date, only two NK₁ antagonists have reached phase III clinical trials. A prodrug of MK-869 **4**⁶ is currently under development for the treatment of emesis whereas CP-122,721 has shown robust antidepressant effect in humans and is now undergoing further evaluation.

Herein, we would like to report our own efforts directed towards the discovery of orally bioavailable NK₁ antagonists free of interaction with calcium channels. Our starting point **6** suffered from several drawbacks

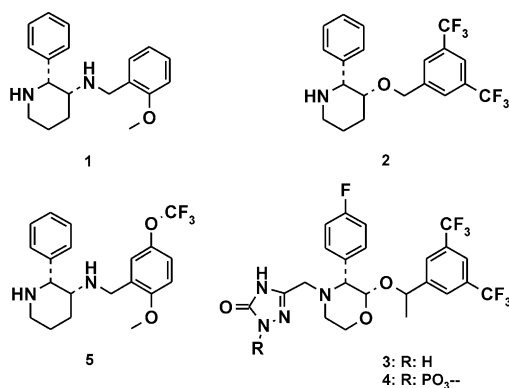
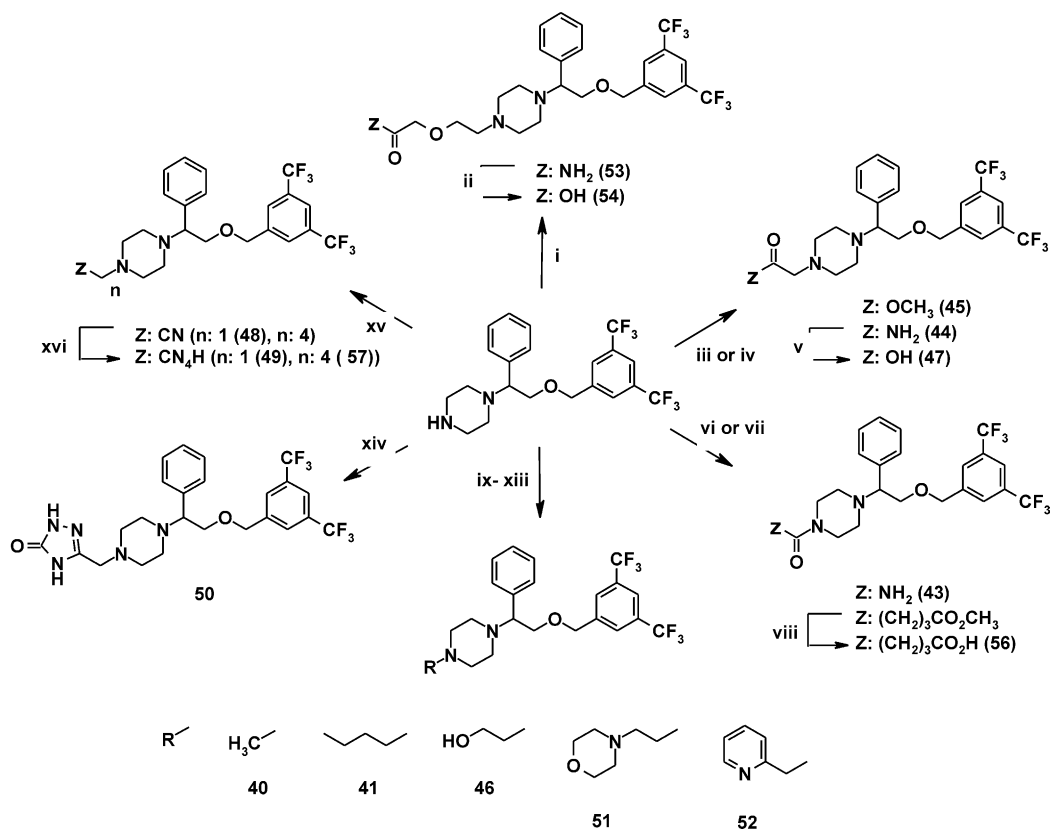


Figure 1. Selected NK₁ antagonists.

*Corresponding author: Tel.: +32-2-386-2797; fax: +32-2-386-2704;
e-mail: christophe.genicot@ucb-group.com



Scheme 2. (i) $\text{Cl}(\text{CH}_2)_2\text{OCH}_2\text{CONH}_2$, K_2CO_3 , DMF; (ii) HCl 1 N, reflux; (iii) $\text{BrCH}_2\text{CONH}_2$, Na_2CO_3 , MeOH, reflux; (iv) $\text{BrCH}_2\text{CO}_2\text{Me}$, K_2CO_3 , DMF; (v) HCl 1 N, reflux; (vi) Me_3SiNCO , THF; (vii) $\text{ClCO}(\text{CH}_2)_3\text{CO}_2\text{Me}$, $\text{CH}_2\text{Cl}_2/\text{NaOH}$ aq; (viii) KOH , MeOH, rt; (ix) HCHO , NaBH_3CN , AcOH, MeOH; (x) $\text{Br}(\text{CH}_2)_3\text{CH}_3$, KI , NaHCO_3 , EtOH; (xi) $\text{Cl}(\text{CH}_2)_2\text{OH}$, Na_2CO_3 , MeOH, reflux; (xii) $\text{Cl}(\text{CH}_2)_2\text{morpholine}$, NaI , NaHCO_3 , EtOH; (xiii) 2-pyridinecarboxaldehyde, NaBH_3CN , AcOH, MeOH; (xiv) ClCH_2CN , $\text{H}_2\text{NNHCO}_2\text{CH}_3$, MeOH then K_2CO_3 , DMF, 140°C ; (xv) $\text{Br}(\text{CH}_2)_4\text{CN}$, K_2CO_3 , DMF; (xvi) Me_3SiN_3 , Bu_3SnO , toluene, reflux.

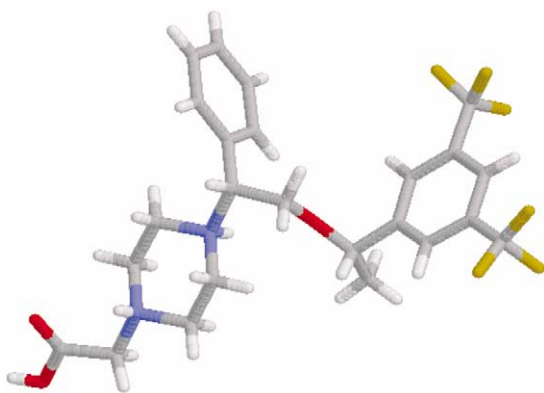


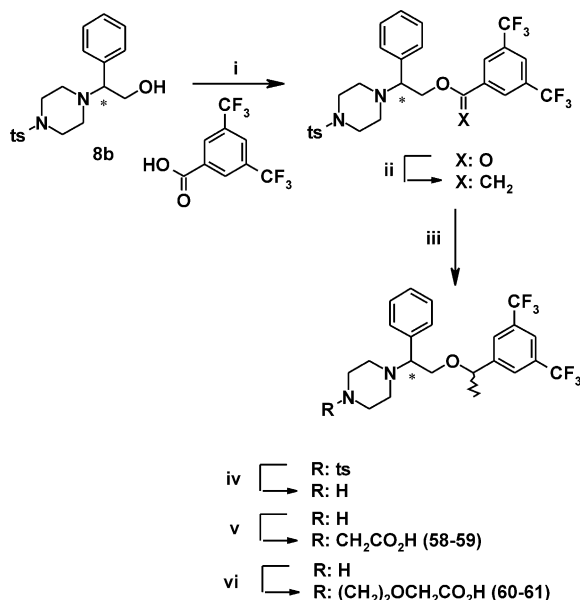
Figure 3. X-ray diffraction of **59a**.

atom in this position gave a compound with much weaker affinity than the unsubstituted analogue. Substitution in the *meta* or *ortho* position was well tolerated. Compound **36** with an α -naphthyl has similar affinity to **7** whereas the one with a β -naphthyl **37** has lower affinity. Replacement of phenyl with a heteroaryl nucleus revealed that thiophene but not furan could be used as a surrogate for phenyl. We then turned our attention to the introduction of substituents on the basic nitrogen of the piperazine nucleus (Table 3).

Introduction of alkyl groups (CH_3 , $(\text{CH}_2)_3\text{CH}_3$) did not affect the affinity for the NK_1 receptor. However, we did

observe subtle variations when the alkyl side chain contained polar functions such as alcohols and amides. Hence, compound **44** with an acetamide side chain displays a lower affinity than the parent compound whereas compounds with an ethanolamine side chain (**46**) or a longer amide (**53**) show similar affinity to **7**. The difference between **44** and **53** could lie in the pK_a of the basic nitrogen. pK_a values for compounds **44** and **53** (6.2 and 7.4, respectively) indicate that the nitrogen atom of **53** is half protonated at pH 7.4 whereas **44** is not. A protonated nitrogen seems to be favorable but not necessary for the interaction with the NK_1 receptor. This assumption is supported by the observation that acylation of the nitrogen further reduces the affinity. However, since compound **43** (urea) retains a reasonable affinity, the protonated nitrogen is not involved in an ionic interaction with the receptor but would rather act as a hydrogen bond donor. The interaction with the receptor is not stereoselective since we did not notice any difference between the two enantiomers **44a** and **44b**. Compounds with acidic functions also have a diminished binding affinity in comparison to simple alkyl derivatives.

The incorporation of a methyl group in the benzylic position next to the oxygen atom led to the creation of a second chiral center. The four were prepared and tested separately. The absolute configuration of the carbon atom next to the piperazine nucleus has only a



Scheme 3. (i) $(\text{EtO})_3\text{P(O)CH}_2\text{COOEt}$, NaH, THF; (ii) Cp_2TiMe_2 , toluene/THF, rt; (iii) H_2 , $\text{RhCl(PPH}_3\text{)}_3$, EtOH; (iv) HCl aq 60 °C; (v) $\text{BrCH}_2\text{CONH}_2$, Na_2CO_3 , MeOH, reflux then HCl 1 N reflux; (vi) $\text{Cl(CH}_2\text{)}_2\text{OCH}_2\text{CONH}_2$, K_2CO_3 , DMF, then HCl 1 N, reflux.

Table 1. Affinities^a of compounds **9–23** for the NK₁ receptor

Compd	R ¹	R ²	R ³	R ⁴	pK _i NK ₁
7	H	CF ₃	H	CF ₃	8.8
9	H	H	H	H	+
10	OCH ₃	H	H	H	+
11	OCF ₃	H	H	H	++
12	Cl	H	H	H	++
13	H	Cl	H	H	6.4
14	H	Cl	Cl	H	7.0
15	H	Cl	H	Cl	8.5
16	H	Br	H	Br	8.3
17	H	F	H	F	6.7
18	H	F	H	CF ₃	7.9
19	H	CH ₃	H	CH ₃	7.6
20	H	C(CH ₃) ₃	H	C(CH ₃) ₃	7.0
21	CF ₃	H	CF ₃	H	7.8
22	H	OCH ₃	OCH ₃	OCH ₃	6.9
23	OCH ₃	Br	H	Br	8.1

^aDisplacement of [³H]-substance P from human NK₁ receptors expressed in CHO cells (*n* = 3); +: <20% inhibition at 10^{−5} M; ++: 20–50% inhibition at 10^{−5} M.

marginal influence on the affinity (Table 4). However, it appears that the absolute configuration of the carbon atom bearing the methyl group has a marked effect on the affinity¹⁰ with the (*S*) enantiomers being much more potent than the non-methylated derivatives while the (*R*) enantiomers are essentially inactive. The gain in affinity may be ascribed to a conformational effect and/or to an additional hydrophobic interaction with the NK₁ receptor.

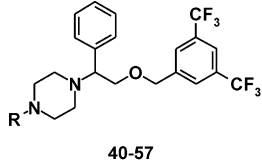
Table 2. Affinities of compounds **24–39** for the NK₁ receptor^a

Compd	R ¹	R ²	R ³	pK _i NK ₁
7	H	H	H	8.8
24	F	H	H	8.3
25	Cl	H	H	8.8
26	CH ₃	H	H	9.0
27	OCH ₃	H	H	8.2
28	H	F	H	8.7
29	H	Cl	H	8.8
30	H	CH ₃	H	8.5
31	H	OCH ₃	H	9.0
32	H	H	F	7.1
33	H	H	OCH ₃	7.3
34	F	F	H	8.5
35	H	Cl	Cl	8.2
36	CH = CH	CH = CH	H	8.8
37	H	CH = CH	CH = CH	7.5
38	—	—	—	8.1
39	—	—	—	6.8

^aDisplacement of [³H]-substance P from human NK₁ receptors expressed in CHO cells (*n* = 3).





In parallel to the binding assay, physicochemical and ADME properties of a selected set of compounds were investigated in order to detect possible flaws that might impair oral activity such as poor aqueous solubility, low membrane permeation and high first pass effect (Table 5). Solubility appears to be strongly influenced by the nature of the substituents on the basic nitrogen. Introduction of a butyl group on **7** (see **41**) markedly reduced solubility. Oddly enough, solubility at physiological pH was still low when an acetamide group was anchored to **7** (see **44**). This is likely to be due to the reduced pK_a value of the basic nitrogen. Compounds with acidic functions (**47**, **54**, **57**, **60b**) showed improved solubility with respect to **7**. The ability of the compounds to cross the intestinal membrane was evaluated with Caco-2 cells. Papp values (> 10^{−6} cm/s) indicate that permeability is not an issue for the compounds tested. Metabolic stability, assessed using guinea pig microsomes, varied widely. Addition of an acidic function considerably increases the metabolic stability of these compounds. On the other hand, it has already been shown that the introduction of a methyl group in the benzyloxy position is favorable.¹¹ As far as potential cardiovascular effects were concerned, we found that compounds with acidic side chains did not interact significantly with L-type calcium channels. Based on in vitro screening, compound **60b** was the most interesting.

In vivo NK₁ antagonist activity of the selected compounds after oral administration was evaluated in the guinea-pig (Table 6). In agreement with the in vitro results, **60b** was the most potent compound to inhibit SP induced pulmonary edema. This compound displayed also a much longer duration of action being still active after 4 h at 1 mg/kg oral dose.

Table 3. Affinities of compounds **40–57** for the NK₁ receptor^a


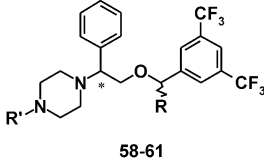
40–57

G1–G4

Compd	R	pK _i NK ₁
7	H	8.8
40	CH ₃	8.8
41	(CH ₂) ₃ CH ₃	8.6
42	CO ₂ CH ₂ CH ₃	7.2
43	CONH ₂	7.7
44a(R)	CH ₂ CONH ₂	8.0
44b(S)	CH ₂ CONH ₂	8.1
45	CH ₂ CO ₂ CH ₃	7.4
46	CH ₂ CH ₂ OH	8.7
47	CH ₂ CO ₂ H	7.8
48	CH ₂ CN	8.1
49	CH ₂ G1	7.2
50	CH ₂ G2	7.8
51	(CH ₂) ₂ G3	7.8
52	CH ₂ G4	8.7
53	(CH ₂) ₂ OCH ₂ CONH ₂	8.5
54	(CH ₂) ₂ OCH ₂ CO ₂ H	7.8
55	(CH ₂) ₄ CO ₂ H	8.0
56	CO(CH ₂) ₃ CO ₂ H	+
57	(CH ₂) ₄ G1	8.2

^aDisplacement of [³H]-substance P from human NK₁ receptors expressed in CHO cells (*n* = 3).

Table 4. Affinities of compounds **59–61** for the NK₁ receptor^a


58–61

Compd	R	R'	pK _i NK ₁
47	H	CH ₂ CO ₂ H	7.8
58a(R,R)	CH ₃	CH ₂ CO ₂ H	+
58b(S,S)	CH ₃	CH ₂ CO ₂ H	8.9
59a(R,S)	CH ₃	CH ₂ CO ₂ H	8.5
59b(S,R)	CH ₃	CH ₂ CO ₂ H	+
54	H	(CH ₂) ₂ OCH ₂ CO ₂ H	7.8
60a(R,R)	CH ₃	(CH ₂) ₂ OCH ₂ CO ₂ H	+
60b(S,S)	CH ₃	(CH ₂) ₂ OCH ₂ CO ₂ H	8.9
61a(R,S)	CH ₃	(CH ₂) ₂ OCH ₂ CO ₂ H	8.6
61b(S,R)	CH ₃	(CH ₂) ₂ OCH ₂ CO ₂ H	+

^aDisplacement of [³H]-substance P from human NK₁ receptors expressed in CHO cells (*n* = 3).

In conclusion, we have described a series of benzyloxyphenethylpiperazines which are potent NK₁ antagonists. We found that the addition of an acidic function increases both solubility and metabolic stability of the compounds and that this translated in vivo into a better oral activity and longer duration of action. Pleasingly, acidic groups also reduced interaction with L-type calcium channels and gave much more selective com-

Table 5. Physicochemical/in vitro ADME properties and calcium channel blocking effect of selected compounds

Cpds	Log D ^a	S ^b	Papps ^c	Clm ^d	pD'2 ^c
6	5.20	0.0006	—	—	—
7	2.41	0.2	2.1	54	5.38
41	4.09	0.001	5.2	170	4.60
44	3.26	0.002	7.9	50	4.27
47	2.35	1.3	9.6	<10	<4.0
54	2.17	>2.0	4.2	<10	<4.0
57	2.71	0.2	2.6	<10	<4.0
60b	2.64	>2.0	8.7	<10	<4.0

^aLog D_{oct/w}, buffer PBS, pH: 7.4.

^bS: solubility (mg/mL, pH: 7.4).

^cPapps, caco-2 permeability (×10^{−6} cm/s).

^dClint, intrinsic clearance (μL/min/mg protein).

^eDetermined on isolated rat aorta [inhibition of contraction induced by KCl (100 mM)].

Table 6. Effect of orally administered compounds on SP-induced pulmonary oedema in guinea-pig¹²

Cpds	Dose ^a	30 min ^b	60 min ^b	120 min ^b	240 min ^b
41	10	62 ^c	—	—	58 ^c
43	10	15 ^c	47 ^c	26 ^c	—
47	10	84 ^c	76 ^c	68 ^c	53 ^c
54	10	53 ^c	61 ^c	54 ^c	29 ^c
57	10	65 ^c	99 ^c	65 ^c	20 ^c
60b	1	77 ^c	75 ^c	—	70 ^c

^amg/kg.

^bTime of compound preadministration (before SP-induced extravasation).

^c% of inhibition.

pounds. Indeed, **60b** has been screened at 10 μM against a variety of G-protein coupled receptor and ion channels and no significant interactions have been identified.

Acknowledgements

The authors wish to thank A. Debeuckelaere, N. Price, F. Coloretti, J.-C. Hayez, V. Verbois, N. Pirlot and C. Delaunoy for their skillfull synthetic assistance and L. Ellens for physicochemical property measurements.

References and Notes

- Hökfelt, T.; Pernow, P.; Wharen, J. *J. Int. Med.* **2001**, *249*, 27.
- Desai, M. C.; Lefkowitz, S. L.; Thadeio, P. F.; Longo, K. P.; Snider, R. M. *J. Med. Chem.* **1992**, *35*, 4911.
- Harrison, T.; Williams, B. J.; Swain, C. J.; Ball, R. G. *Bioorg. Med. Chem. Lett.* **1994**, *31*, 2545.
- Hale, J. J.; Mills, S. G.; MacCoss, M.; Finke, P. E.; Cascieri, M. A.; Sadowski, S.; Ber, E.; Chicchi, G. G.; Kurtz, M.; Metzger, J.; Eiermann, G.; Tsou, N. N.; Tattersall, F. D.; Rupniak, N. M.; Williams, A. R.; Rycroft, W.; Hargreave, R.; MacIntyre, D. E. *J. Med. Chem.* **1998**, *41*, 4607.
- Rosen, T. J.; Coffman, K. J.; McLean, S.; Crawford, R. T.; Bryce, D. K.; Gohda, Y.; Tsuchiya, M.; Nagahisa, A.; Nakane, M.; Lowe, J. A., III. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 281.
- Hale, J. J.; Mills, S. G.; MacCoss, M.; Dorn, C. P.; Finke, P. E.; Budhu, R. J.; Reamer, R. A.; Huskey, S. E.; Luffer-Atlas, D.; Dean, B. J.; McGowan, E. M.; Feeney, W. P.; Chiu,

- S. H.; Cascieri, M. A.; Chicchi, G. G.; Kurtz, M. M.; Sadowski, S.; Ber, E.; Tattersall, F. D.; Rupniak, N. M.; Williams, A. R.; Rycroft, W.; Hargreaves, R.; Metzger, J. M.; MacIntyre, D. E. *J. Med. Chem.* **2000**, *43*, 1234.
7. Stevenson, G. I.; Huscroft, I.; MacLeod, A. M.; Swain, C. J.; Cascieri, M. A.; Chicchi, G. G.; Graham, M. I.; Harrison, T.; Kelleher, F. J.; Kurtz, M.; Ladduwahetty, T.; Merchant, K. J.; Metzger, J. M.; MacIntyre, D. E.; Sadowski, S.; Sohal, B.; Owens, A. P. *J. Med. Chem.* **1998**, *41*, 4623.
8. Petasis, N. A.; Goodman, A.; Zavialov, I. A. *Tetrahedron* **1997**, *48*, 16463.
9. Petasis, N. A.; Lu, S.-P. *Tetrahedron Lett.* **1995**, *36*, 2393.
10. Owens, A. P.; Williams, B. J.; Harrison, T.; Swain, C. J.; Baker, R.; Sadowski, S.; Cascieri, M. A. *Biomed. Chem. Lett.* **1995**, *5*, 2761.
11. Swain, C. J.; Baker, B. J.; Cascieri, M. A.; Chicchi, G.; Forrest, M.; Herbert, R.; Keown, L.; Ladduwahetty, T.; Luell, S.; MacIntyre, D. E.; Metzger, J.; Morton, S.; Owens, A. P.; Sadowski, S.; Watt, A. P. *Bioorg. Med. Chem. Lett.* **1997**, *8*, 2959.
12. Murai, M.; Morimoto, H.; Maeda, Y.; Kiyotoh, S.; Nishikawa, M.; Fujii, T. *J. Pharmacol. Exp. Ther.* **1992**, *262*, 403.