Organic & **Biomolecular Chemistry**

PAPER

Check for updates

Cite this: DOI: 10.1039/c9ob00736a

Synthesis of amino acid derivatives of 5-alkoxy-3,4-dihalo-2(5H)-furanones and their preliminary bioactivity investigation as linkers*

Shi-He Luo, ^[] ^{a,b} Kai Yang, *^{a,c} Jian-Yun Lin,^a Juan-Juan Gao,^d Xin-Yan Wu^a and Zhao-Yang Wang 匝 *^{a,b}

A series of amino acid derivatives are successfully synthesized via a metal-free C-N coupling reaction of 5-alkoxy-3,4-dihalo-2(5H)-furanones and amino acids. Their structures are well characterized with ¹H NMR, ¹³C NMR, ESI-MS and elemental analysis. As potential linkers of the 2(5*H*)-furanone unit with other drug moieties containing a hydroxyl or amino group, the effect of amino acids is investigated by comparison with other 2(5H)-furanone compounds by constructing C-O/C-S bonds. The preliminary results of the biological activity assay by the MTT method on a series of cancer cell lines in vitro reveal that the introduction of amino acids basically has no toxic effect. This can lead to these 2(5H)-furanone derivatives being further well-linked with other bioactive moieties with amino or hydroxy groups as expected. Thus, the biological activity assay gives a direction for the design of bioactive 2(5H)-furanones based on these amino acid linkers.

Received 31st March 2019, Accepted 2nd May 2019 DOI: 10.1039/c9ob00736a

rsc li/obc

Published on 02 May 2019. Downloaded by UNIV OF LOUISIANA AT LAFAYETTE on 5/10/2019 10:14:53 AM

Introduction

Amino acids are very important living substances in organisms and the basic units of enzymes and proteins.¹ Due to the presence of both amino and carboxyl groups, amino acids are also one of the most versatile building blocks used in many fields, such as organic synthesis,² drug design,³⁻⁵ molecular recognition,⁶⁻⁸ supramolecular chemistry,^{9,10} functional materials,^{11,12} and so on.¹³ Importantly, the fact that amino acids are used as key linkers between the structural molecule and bioactive unit has been widely utilized.^{14–19}

Molecules possessing a 2(5H)-furanone moiety, a kind of α,β -unsaturated lactone substructure, are frequently found in natural products.²⁰⁻²² They have received considerable interest

because of their significant biological activities,²³⁻²⁵ such as antifungal,^{26,27} antibacterial,²⁸ antiviral,^{29,30} and anticancer.^{31,32} Therefore, the synthesis of different 2(5H)-furanones and their applications have drawn much attention in recent years.³³⁻³⁹ Being interested in the chemistry of 2(5H)-furanones, we also reported some studies,40-47 and investigated the bioactivities of partial 2(5H)-furanone derivatives.48-50

View Article Online

Nowadays, there are reports on the role of amino acids as linking units.^{13,19,51,52} However, it is still necessary to investigate whether the amino acid unit is a safe linker without any impact on the bioactivity of 2(5H)-furanones when amino acids are combined with the 2(5H)-furanone moiety by C-N coupling to expand the functionalization of 2(5H)-furanones. Thus, in order to make full use of two synthons both 2(5H)-fur-

^aSchool of Chemistry and Environment, South China Normal University,

Key Laboratory of Theoretical Chemistry of Environment, Ministry of Education, Guangzhou Key Laboratory of Analytical Chemistry for Biomedicine,

Guangzhou 510006, P. R. China. E-mail: wangzy@scnu.edu.cn

^bSchool of Chemistry and Chemical Engineering, Key Laboratory of Functional Molecular Engineering of Guangdong Province, South China University of Technology, Guangzhou 510641, P. R. China

^cCollege of Pharmacy, Gannan Medical University, Ganzhou, Jiangxi province, 341000, P. R. China. E-mail: kai_yangyang@126.com

[†]Electronic supplementary information (ESI) available: Synthesis procedures for contrasting compounds O1-O27, S1-S16 and SO1-SO16 and ¹H NMR, ¹³C NMR and ESI-MS spectra of all new compounds N1-N16. See DOI: 10.1039/ c9ob00736a



Scheme 1 Synthetic route to target compounds N1-N16

^dCollege of Sports and Rehabilitation, Gannan Medical University, Ganzhou, Jiangxi province, 341000, P. R. China

Paper

anones and amino acids,⁵³ and make clear how they work together especially in a small molecular system, a series of amino acid derivatives were synthesized with 5-alkoxy-3,4-dihalo-2(5H)-furanones and amino acids, including nonnatural amino acids, *via* a metal-free C–N coupling reaction (Scheme 1). And the effect of amino acids as linkers on bioactivity was investigated by comparison with other 5-substituted-3-halo-2(5H)-furanone derivatives.

Experimental

Materials and methods

¹H and ¹³C NMR spectra were obtained in CDCl_3 or $\text{DMSO-}d_6$ on a Varian DRX-400 MHz spectrometer and tetramethylsilane (TMS) was used as an internal standard. Elemental analysis was performed on a Thermo FlashEA TM 112 elemental analyzer. Mass spectra (MS) were recorded on a Thermo LCQ DECA XP MAX mass spectrometer.

All reagents and solvents were commercially available and used as received. Using furfural, ethanol and methanol as starting materials, 5-substituted-3,4-dihalo-2(5*H*)-furanones **1a–1d** were prepared according to the literature.^{44–50}

General procedure for syntheses of compounds N1-N16

Amino acid 2 (4 mmol), KOH (4.48 mmol) and C_2H_5OH (5 mL) were added into a round-bottomed flask equipped with a magnetic stirring bar. After the mixture was dissolved under stirring, 5-substituted 3,4-dihalo-2(5*H*)-furanones 1 (2 mmol) in CH₂Cl₂ (5 mL) were dropped into the above system. The mixed solution was stirred at room temperature under an N_2 atmosphere for 24 h.

After the solvent was removed, the residue was dissolved in CH_2Cl_2 , and the pH value was adjusted to 3–4 using 15% HCl. The organic layers were washed with saturated NH_4Cl solution three times and then dried over anhydrous $MgSO_4$. The concentration under vacuum gave a crude product, which was purified by column chromatography on silica gel with gradient eluents of petroleum ether and ethyl acetate to afford pure compounds.

Biochemical approach

The cell viability of the compounds was determined by measuring the ability of cells to transform MTT into a purple formazan dye.^{54,55} All compounds were dissolved in DMSO with stock solution at 10 mM. The cells were seeded in 96-well tissue culture plates at 5×10^3 cells per well and incubated in an incubator at 37 °C and 5% CO₂ for 24 h. Then, the cells were incubated with the tested compounds in a concentration range of 4–300 mM, ensuring an equal volume of 200 µL across the wells of the plate. These plates were incubated at 37 °C in a 5% CO₂ incubator for 72 h. After incubation, 20 µL of MTT solution (5 mg mL⁻¹) was added across the plate and further incubated for 4 h. Finally, the medium was aspirated and replaced with a 150 µL per well of DMSO to dissolve the formazan salt formed. The color intensity of the formazan

solution, which reflects the cell growth conditions, was measured by using a microplate spectrophotometer at 490 nm.

Characterization data for compounds N1-N16

N-(4-Bromo-2-methoxy-5-oxo-2,5-dihydrofuran-3-yl)-*N*methylglycine (N1). Brown sticky liquid; yield: 222 mg (79%); ¹H NMR (400 MHz, CDCl₃-TMS), δ, ppm: 3.26 (s, 3H, NCH₃), 3.46 (s, 3H, OCH₃), 4.34 (s, 2H, CH₂), 5.77 (s, 1H, CH), 6.61 (b, 1H, COOH); ¹³C NMR (100 MHz, CDCl₃-TMS), δ, ppm: 40.5, 53.9, 54.5, 74.4, 98.7, 158.2, 168.9, 172.2; ESI-MS, *m*/*z* (%): calcd for C₈H₉BrNO₅[−] ([M − H][−]): 277.97 (100.0%), 279.97 (97.3%), found: 277.79 (100.0%), 279.79 (96.1%); anal. calcd for C₈H₁₀BrNO₅: C 34.31, H 3.60, N 5.00, found: C 34.43, H 3.52, N 5.02.

N-(4-Bromo-2-ethoxy-5-oxo-2,5-dihydrofuran-3-yl)-*N*-methylglycine (N2). Brown sticky liquid; yield: 208 mg (71%); ¹H NMR (400 MHz, DMSO- d_6), δ, ppm: 1.13 (t, *J* = 8.0 Hz, 3H, CH₃), 3.00–3.34 (m, 4H, NCH₃, COOH), 3.61–3.70 (m, 2H, OCH₂), 4.08–4.44 (m, 2H, CH₂), 6.14 (s, 1H, CH), 13.02 (b, 1H); ¹³C NMR (100 MHz, DMSO- d_6), δ, ppm: 15.1, 40.2, 53.9, 63.8, 71.9, 98.0, 159.3, 168.3, 170.4; ESI-MS, *m/z* (%): calcd for C₉H₁₁BrNO₅⁻ ([M – H]⁻): 291.98 (100.0%), 293.98 (97.3%), found: 291.74 (100.0%), 293.74 (94.7%); anal. calcd for C₉H₁₂BrNO₅: C 36.76, H 4.11, N 4.76, found: C 36.82, H 4.02, N 4.83.

N-(4-Chloro-2-methoxy-5-oxo-2,5-dihydrofuran-3-yl)-*N*methylglycine (N3). Brown sticky liquid; yield: 157 mg (67%); ¹H NMR (400 MHz, CDCl₃-TMS), *δ*, ppm: 3.24 (3H, s, NCH₃), 3.48 (s, 3H, OCH₃), 4.31 (s, 2H, CH₂), 5.74 (s, 1H, CH), 7.46 (b, 1H, COOH); ¹³C NMR (100 MHz, CDCl₃-TMS), *δ*, ppm: 40.2, 53.4, 54.5, 88.8, 97.7, 155.3, 168.2, 172.6; ESI-MS, *m*/*z* (%): calcd for C₈H₉ClNO₅⁻ ([M – H]⁻): 234.02 (100.0%), 236.01 (32.0%), found: 233.71 (100.0%), 235.71 (35.0%); anal. calcd for C₈H₁₀ClNO₅: C 40.78, H 4.28, N 5.94, found: C 40.63, H 4.25, N 5.82.

N-(4-Chloro-2-ethoxy-5-oxo-2,5-dihydrofuran-3-yl)-*N*-methylglycine (N4). Brown sticky liquid; yield: 139 mg (56%); ¹H NMR (400 MHz, CDCl₃-TMS), δ, ppm: 1.27 (t, *J* = 8.0 Hz, 3H, CH₃), 3.27 (s, 3H, NCH₃), 3.69–3.91 (m, 2H, OCH₂), 4.24–4.39 (m, 2H, CH₂), 5.81 (s, 1H, CH), 8.74 (b, 1H, COOH); ¹³C NMR (100 MHz, CDCl₃-TMS), δ, ppm: 14.7, 40.2, 53.6, 64.5, 88.3, 97.4, 156.2, 169.1, 172.4; ESI-MS, *m*/*z* (%): calcd for C₉H₁₁ClNO₅[−] ([M − H][−]): 248.03 (100.0%), 250.03 (32.0%), found: 248.46 (100.0%), 250.46 (27.8%); anal. calcd for C₉H₁₂ClNO₅: C 43.30, H 4.85, N, 5.61, found: C 43.36, H 4.82, N 5.55.

2-(4-Bromo-2-methoxy-5-oxo-2,5-dihydrofuran-3-yl)amino-2phenylacetic acid (N5). Brown sticky liquid; yield: 153 mg (45%); ¹H NMR (400 MHz, CDCl₃-TMS), δ , ppm: 3.41 (s, 3H, OCH₃), 5.35–6.44 (m, 3H, NH, COOH, CH), 5.75 (s, 1H, CH), 7.38–7.42 (m, 5H, ArH); ¹³C NMR (100 MHz, CDCl₃-TMS), δ , ppm: 58.6, 65.6, 81.5, 98.1, 127.0, 129.3, 129.4, 135.4, 153.2, 167.7, 172.1; ESI-MS, *m/z* (%): calcd for C₁₃H₁₃BrNO₅⁺ ([M + H]⁺): 342.00 (100.0%), 344.00 (97.3%), found: 341.64 (100%), 343.64 (93.2%); anal. calcd for C₁₃H₁₂BrNO₅: C 45.64, H 3.54, N 4.09, found: C 45.64, H 3.74, N 4.19. **2-(4-Bromo-2-ethoxy-5-oxo-2,5-dihydrofuran-3-yl)amino-2phenylacetic acid (N6).** Brown sticky liquid; yield: 159 mg (45%); ¹H NMR (400 MHz, CDCl₃-TMS), *δ*, ppm: 1.27 (t, *J* = 8.0 Hz, 3H, CH₃), 3.62–3.79 (m, 2H, OCH₂), 5.20–5.80 (m, 2H, CH), 6.29 (b, 1H, N–H), 6.37 (b, 1H, COOH), 7.35–7.41 (m, 5H, ArH); ¹³C NMR (100 MHz, CDCl₃-TMS), *δ*, ppm: 15.0, 58.7, 65.3, 83.3, 97.5, 127.0, 129.3, 129.5, 135.0, 152.4, 167.4, 172.9; ESI-MS, *m*/*z* (%): calcd for C₁₄H₁₅BrNO₅⁺ ([M + H]⁺): 356.01 (100.0%), 358.01 (97.3%), found: 355.74 (97.3%), 357.74 (100%); anal. calcd for C₁₄H₁₄BrNO₅: C 47.21, H 3.96, N 3.92, found: C 46.91, H 4.06, N 3.93.

2-(4-Chloro-2-methoxy-5-oxo-2,5-dihydrofuran-3-yl)amino-2phenylacetic acid (N7). Brown sticky liquid; yield: 112 mg (38%); ¹H NMR (400 MHz, CDCl₃-TMS), *δ*, ppm: 3.43 (s, 3H, OCH₃), 5.27–5.74 (m, 4H, NH, COOH, 2CH), 7.32–7.46 (m, 5H, ArH); ¹³C NMR (100 MHz, CDCl₃-TMS), *δ*, ppm: 58.6, 64.0, 85.1, 96.4, 127.0, 129.3, 129.4, 132.2, 149.0, 163.5, 172.7; ESI-MS, *m*/*z* (%): calcd for $C_{13}H_{13}ClNO_5^+$ ([M + H]⁺): 298.05 (100.0%), 300.04 (32.0%), found: 297.76 (100.0%), 300.05 (43.0%); anal. calcd for $C_{13}H_{12}ClNO_5$: C 52.45, H 4.06, N 4.71, found: C 52.35, H 4.10, N 4.82.

2-(4-Chloro-2-ethoxy-5-oxo-2,5-dihydrofuran-3-yl)amino-2phenylacetic acid (N8). Brown sticky liquid; yield: 127 mg (41%); ¹H NMR (400 MHz, CDCl₃-TMS), *δ*, ppm: 1.26 (t, *J* = 8.0 Hz, 3H, CH₃), 3.57–3.82 (m, 2H, CH₂), 5.13–5.79 (m, 3H, NH, 2CH), 6.96 (b, 1H, COOH), 7.31–7.41 (m, 5H, ArH); ¹³C NMR (100 MHz, CDCl₃-TMS), *δ*, ppm: 15.0, 58.9, 65.3, 83.8, 97.2, 127.0, 129.2, 129.4, 135.5, 150.1, 167.5, 172.9; ESI-MS, *m/z* (%): calcd for C₁₄H₁₅ClNO₅⁺ ([M + H]⁺): 312.06 (100.0%), 314.06 (32.0%), found: 311.68 (100.0%), 313.68 (48.6%); anal. calcd for C₁₄H₁₄ClNO₅: C 53.94, H 4.53, N 4.49, found: C 54.05, H 4.46, N 4.51.

4-(4-Bromo-2-methoxy-5-oxo-2,5-dihydrofuran-3-yl)aminobutanoic acid (N9). Yellow sticky liquid; yield: 157 mg (54%); ¹H NMR (400 MHz, CDCl₃-TMS), *δ*, ppm: 1.63–1.70 (m, 2H, CH₂), 1.96–2.01 (m, 2H, CH₂), 3.51 (s, 3H, OCH₃), 3.55 (b, 1H, COOH), 3.62–3.90 (m, 2H, NCH₂), 5.33 (s, 1H, NH), 5.73 (s, 1H, CH); ¹³C NMR (100 MHz, CDCl₃-TMS), *δ*, ppm: 25.2, 30.5, 43.2, 65.6, 85.0, 98.3, 149.5, 167.8, 178.0; ESI-MS, *m/z* (%): calcd for C₉H₁₃BrNO₅⁺ ([M + H]⁺): 294.00 (100.0%), 296.00 (97.3%), found: 293.71 (100%), 295.72 (94.8%); anal. calcd for C₉H₁₂BrNO₅: C 36.76, H 4.11, N 4.76, found: C 36.75, H 4.38, N 4.84.

4-(4-Bromo-2-ethoxy-5-oxo-2,5-dihydrofuran-3-yl)aminobutanoic acid (N10). Yellow sticky liquid; yield: 187 mg (61%); ¹H NMR (400 MHz, CDCl₃-TMS), *δ*, ppm: 1.28 (t, J = 8.0 Hz, 3H, CH₃), 1.90–2.56 (m, 5H, COOH, 2CH₂), 3.70–3.90 (m, 4H, OCH₂, NCH₂), 5.36 (s, 1H, CH), 5.77 (s, 1H, NH); ¹³C NMR (100 MHz, CDCl₃-TMS), *δ*, ppm: 15.0, 25.2, 30.5, 43.2, 65.6, 85.0, 98.3, 149.5, 167.8, 178.0; ESI-MS, *m/z* (%): calcd for C₁₀H₁₅BrNO₅⁺ ([M + H]⁺): 308.01 (100.0%), 310.01 (97.3%), found: 307.74 (100.0%), 309.74 (94.8%); anal. calcd for C₁₀H₁₄BrNO₅: C 38.98, H 4.58, N 4.55, found: C 39.00, H 4.48, N 4.62.

4-(4-Chloro-2-methoxy-5-oxo-2,5-dihydrofuran-3-yl)aminobutanoic acid (N11). Brown liquid; yield: 136 mg (55%); ¹H NMR (400 MHz, CDCl₃-TMS), δ, ppm: 1.93–1.99 (m, 2H, CH₂), 2.46–2.50 (m, 2H, CH₂), 3.50–3.60 (m, 5H, NCH₂, OCH₃), 5.19–5.35 (m, 2H, CH, NH), 5.67 (s, 1H, COOH); ¹³C NMR (100 MHz, CDCl₃-TMS), δ, ppm: 25.3, 30.8, 43.0, 55.2, 83.0, 97.5, 155.5, 166.3, 177.9; ESI-MS, m/z (%): calcd for C₉H₁₃ClNO₅⁺ ([M + H]⁺): 250.05 (100.0%), 252.04 (32.0%), found: 249.80 (100.0%), 251.80 (30.4%); anal. calcd for C₉H₁₂ClNO₅: C 43.30, H 4.85, N 5.61, found: C 43.45, H 5.06, N 5.51.

4-(4-Chloro-2-ethoxy-5-oxo-2,5-dihydrofuran-3-yl)amino-butanoic acid (N12). Brown liquid; yield: 183 mg (70%); ¹H NMR (400 MHz, CDCl₃-TMS), *δ*, ppm: 1.29 (3H, t, *J* = 8.0 Hz, CH₃), 1.95–2.25 (m, 2H, CH₂), 2.50 (t, *J* = 8.0 Hz, 2H, CH₂), 3.51–3.61 (m, 2H, NCH₂), 3.70–3.91 (m, 2H, OCH₂), 4.63 (b, 1H, COOH), 5.31 (s, 1H, NH), 5.74 (s, 1H, CH); ¹³C NMR (100 MHz, CDCl₃-TMS), *δ*, ppm: 14.9, 25.3, 30.9, 43.0, 64.9, 82.5, 96.8, 156.5, 163.1, 178.3; ESI-MS, *m*/*z* (%): calcd for C₁₀H₁₃ClNO₅⁻ ([M - H]⁻): 262.05 (100.0%), 264.05 (32.0%), found: 261.80 (100.0%), 263.75 (42.1%); anal. calcd for C₁₀H₁₄ClNO₅: C 45.55, H 5.35, N 5.31, found: C 45.45, H 5.36, N 5.30.

6-(**4**-Bromo-2-methoxy-5-oxo-2,5-dihydrofuran-3-yl)aminohexanoic acid (N13). Yellow sticky liquid; yield: 244 mg (76%); ¹H NMR (400 MHz, CDCl₃-TMS), δ, ppm: 1.43–1.48 (m, 2H, CH₂), 1.63–1.71 (m, 4H, 2CH₂), 2.40 (t, J = 8.0 Hz, 2H, CH₂), 3.32–3.47 (m, 2H, NCH₂), 3.50 (s, 3H, OCH₃), 4.94 (s, 1H, COOH), 5.35 (b, 1H, NH), 5.72 (s, 1H, CH); ¹³C NMR (100 MHz, CDCl₃-TMS), δ, ppm: 24.0, 25.7, 30.1, 33.6, 43.5, 54.8, 84.6, 98.4, 157.1, 163.4, 178.9; ESI-MS, *m*/*z* (%): calcd for C₁₁H₁₇BrNO₅⁺ ([M + H]⁺): 322.03 (100.0%), 324.03 (97.3%), found: 321.65 (100%), 323.65 (88.7%); anal. calcd for C₁₁H₁₆BrNO₅: C 41.01, H 5.01, N 4.35, found: C 40.92, H 5.16, N 4.45.

6-(4-Bromo-2-ethoxy-5-oxo-2,5-dihydrofuran-3-yl)amino-hexanoic acid (N14). Yellow sticky liquid; yield: 267 mg (80%); ¹H NMR (400 MHz, CDCl₃-TMS), *δ*, ppm: 1.28 (t, *J* = 8.0 Hz, 3H, CH₃), 1.34–1.47 (m, 2H, CH₂), 1.61–1.70 (m, 4H, 2CH₂), 2.33–2.41 (m, 2H, CH₂), 3.36–3.60 (m, 2H, NCH₂), 3.70–3.89 (m, 2H, OCH₂), 5.36 (s, 1H, CH), 5.78 (b, 1H, NH), 6.88 (b, 1H, COOH); ¹³C NMR (100 MHz, CDCl₃-TMS), *δ*, ppm: 15.0, 24.0, 25.7, 30.2, 33.6, 40.7, 64.5, 84.7, 97.6, 138.8, 163.5, 178.8; ESI-MS, *m*/*z* (%): calcd for C₁₂H₁₉BrNO₅⁺ ([M + H]⁺): 336.04 (100.0%), 338.04 (97.3%), found: 335.73 (100%), 337.75 (94.5%); anal. calcd for C₁₂H₁₈BrNO₅: C 42.87, H 5.40, N 4.17, found: C 43.02, H 5.45, N 4.20.

6-(4-Chloro-2-methoxy-5-oxo-2,5-dihydrofuran-3-yl)aminohexanoic acid (N15). Brown liquid; yield: 214 mg (78%); ¹H NMR (400 MHz, CDCl₃-TMS), δ, ppm: 1.26–1.36 (m, 2H, CH₂), 1.40–1.55 (m, 2H, CH₂), 1.62–1.70 (m, 2H, CH₂), 2.34–2.41 (m, 2H, CH₂), 3.37–3.47 (m, 2H, NCH₂), 3.50 (s, 3H, OCH₃), 5.26 (b, 1H, NH), 5.32 (s, 1H, CH), 5.69 (b, 1H, COOH); ¹³C NMR (100 MHz, CDCl₃-TMS), δ, ppm: 24.0, 25.7, 30.2, 33.7, 43.4, 55.0, 84.3, 97.6, 156.0, 167.8, 179.2; ESI-MS, *m*/*z* (%): calcd for C₁₁H₁₇ClNO₅⁺ ([M + H]⁺): 278.08 (100.0%), 280.08 (32.0%), found: 277.78 (100%), 279.78 (30.2%); anal. calcd for C₁₁H₁₆ClNO₅: C 47.58, H 5.81, N 5.04, found: C 47.62, H 5.86, N 5.05. **6-(4-Chloro-2-ethoxy-5-oxo-2,5-dihydrofuran-3-yl)amino-hexanoic acid (N16).** Brown liquid; yield: 228 mg (79%); ¹H NMR (400 MHz, CDCl₃-TMS), *δ*, ppm: 1.25 (t, *J* = 8.0 MHz, 3H, CH₃), 1.31–1.44 (m, 2H, CH₂), 1.58–1.65 (m, 4H, 2CH₂), 2.31–2.37 (m, 2H, CH₂), 3.31–3.55 (m, 2H, NCH₂), 3.67–3.86 (m, 2H, OCH₂), 5.31 (s, 1H, CH), 5.39 (b, 1H, NH), 5.72 (b, 1H, COOH); ¹³C NMR (100 MHz, CDCl₃-TMS), *δ*, ppm: 14.9, 24.0, 25.7, 30.2, 33.7, 43.4, 64.6, 85.5, 96.9, 156.0, 167.8, 179.2; ESI-MS, *m/z* (%): calcd for C₁₂H₁₉ClNO₅⁺ ([M + H]⁺): 292.09 (100.0%), 294.09 (32.0%), found: 291.84 (100%); 293.84 (25.8%); anal. calcd for C₁₂H₁₈ClNO₅: C 49.41, H 6.22, N 4.80, found: C 49.42, H 6.42, N 4.75.

Results and discussion

Chemical synthesis by metal-free C-N coupling reaction

Based on our previous work,^{40–47} the reaction conditions were optimized firstly. Using 2 mmol 3,4-dibromo-5-ethoxy-2(5*H*)-furanone (**1b**) and 6-aminohexanoic acid (**2d**)⁵⁶ as representative reactants, the reaction parameters, such as catalysts, bases, additives and temperature, were examined. The obtained results are listed in Table 1.

It is well known that the formation of a C_{sp2} -N bond between a C_{sp2} -X bond and N–H is usually catalyzed by transition metal catalysts,^{57–60} especially the inexpensive Cu(i) salts.^{61–64} Thus, under the same conditions, we initially investigated the effect of the available Cu(i) salt. Using three sets of test comparisons, it can be found that CuI (0.015 mmol) does not work well as a catalyst in this reaction system irrespective of the base (Table 1, entries 1–6). And obviously, among the selected bases, KOH (1.12 equiv. of amino acid 2d) is the best and the isolated yield of the target compound, 6-(4-bromo-2ethoxy-5-oxo-2,5-dihydrofuran-3-yl)aminohexanoic acid (N14), is 71% (entry 6). However, mixing the additive of KI into the base system is not effective irrespective of the proportion it is

Table 1 Optimization of the reaction conditions^a

H ₃ CH		о ⁺ H ₂ N 2d	KOH tOH, r.t.	HO HO H ₃ CH ₂ C	$ \begin{array}{c} H \\ N \\ 0 \\ 0 \\ 0 \end{array} \begin{array}{c} Br \\ 0 \\ 0 \\ 0 \\ 0 \end{array} $
Entry	2d : 1b	Base (equiv.)	Cat.	Temp. (°C)	$\operatorname{Yield}^{b}(\%)$
1	1.5	K_3PO_4 (1.12)	CuI	r.t.	45
2	1.5	$K_{3}PO_{4}(1.12)$	_	r.t.	57
3	1.5	t BuOK (1.12)	CuI	r.t.	47
4	1.5	t BuOK (1.12)	_	r.t.	39
5	1.5	KOH (1.12)	CuI	r.t.	66
6	1.5	KOH (1.12)	_	r.t.	71
7	1.5	KOH + KI (1.12, 1:1)	_	r.t.	58
8	1.5	KOH + KI(1.12, 1:2)	_	r.t.	30
9	1.5	KOH + KI(1.12, 2:1)	_	r.t.	49
10	1.5	KOH (1.12)	_	40	45
11	1.5	KOH (1.12)	_	60	45
12	2.0	KOH (1.12)	_	r.t.	80

 a Using C_2H_5OH/CH_2Cl_2 as a solvent, the reaction was carried out for 24 h. b Isolated yield.

used in (entries 7–9). In addition, we did not notice any significant advantages when altering the reaction temperature (entries 10 and 11 *vs.* entry 6). Fortunately, when the dosage ratio of **2d** with **1b** is increased to 2 equiv., the desired product **N14** can be obtained in 80% yield (Table 1, entry 12). Thus, using C₂H₅OH/CH₂Cl₂ as a solvent and KOH (4.48 mmol) as a base, we successfully achieved the reaction of **1b** (2 mmol) and **2d** (4 mmol) at room temperature for 24 h to obtain a satisfactory yield.

Under the optimized reaction conditions, four typical amino acids were used in the transformation to establish the scope and generality of this protocol (Table 2). Generally, irrespective of whether bromine or chlorine is used in 5-alkoxyl-3,4-dihalo-2(5H)-furanones 1, the reaction can work well. What's more, the type of amino acid also has some effects on the yield. In most cases, on using secondary amine sarcosine as the substrate, the yield of products is higher than that when using primary amines (e.g. phenylglycine and 4-aminobutyric acid, N1-N4 vs. N5-N11). This means that the secondary amine is beneficial for this reaction. Furthermore, the yield of products is increased with the increase in the alkyl chain length of primary amines, especially for 6-aminohexanoic acid (N13-N16). This is because, the longer the chain, the greater the flexibility and the smaller the steric resistance. In addition, due to the large steric hindrance of the benzene ring, though the yield of products obtained by phenylglycine can be over 38% (N5–N8), it is obviously lower than that of others.

The structures of all amino acid derivatives have been systematically characterized by ¹H NMR, ¹³C NMR, ESI-MS and elemental analysis. In the ¹H NMR spectra, there is a single peak between 5.27–6.14 ppm, and the chemical shift belongs to the C–H of the furanone ring. The significant chemical shift of NH and COOH in the amino acid unit usually appears near

Table 2
Substrate
scope
of
various
5-substituted
3,4-dihalo-2(5H)

furanones 1 and amino acids $2^{a,b}$ $2^{a,b}$



 a Using KOH as a base and $\rm C_2H_5OH/CH_2Cl_2$ as a solvent at room temperature. b Isolated yield.

5.35 ppm and 6.88 ppm, respectively. The characteristic chemical shift of the furanone ring and amino acid moiety also can be found in ¹³C NMR spectra. Furthermore, the ESI-MS results are consistent with the data of their corresponding theoretical value. Thus, all structural testing results confirm that the metal-free C–N coupling reaction between 5-substituted 3,4dihalo-2(5*H*)-furanones and amino acids is carried out indeed.

There have been many research studies to make the C–N bond construction reaction more eco-friendly.^{64–66} Particularly, due to high cost and environmental pollution in transition metal catalysis, the development of metal-free methodologies in the C–N coupling reaction is becoming popular.^{67–69} Therefore, our design for the formation of the C_{sp2}–N bond is practical and allows for the successful combination of amino acids with the 2(5*H*)-furanone unit.

Inhibition activities on various anticancer cells in vitro

In order to investigate the potential bioactive compounds with no/less cytotoxicity, other 2(5H)-furanone compounds **O1–O27**,⁷⁰ **S1–S16**,⁴⁶ and **SO1–SO16**⁴⁷ were selected as contrasting substances (their structures can be seen in Scheme 2) by constructing C–O/C–S bonds at the same site of 5-substituted 3,4-dihalo-2(5H)-furanones (their corresponding synthesis procedures are described in the ESI†); the effect of amino acids as potential linkers in the 2(5H)-furanone unit with other drug moieties containing amino or hydroxy groups was investigated through the biological activity assay by the MTT method on a series of cancer cell lines *in vitro*, such as C6 (rat glioma cells),

Scheme 2 2(5H)-Furanone derivatives synthesized previously.

View Article Online

EC-1 (human esophageal carcinoma cells), MDA-MB-231 (estrogen receptor-negative human breast cells), MCF-7 (estrogen receptor-positive human breast cells), HepG2 (human hepatoma cells), CNE-1 (nasopharyngeal carcinoma cells) and A549 (human lung adenocarcinoma cells).^{54,55} Using cisplatin as the positive control, the antitumor activities of the above-mentioned 2(5H)-furanone derivatives are summarized in Table 3.

It can be seen that, **N1–N16**, the serial amino acid derivatives of 5-alkoxy-3,4-dihalo-2(5*H*)-furanones basically have no obvious antiproliferative activity, so the tumor cells can grow without exerting cytotoxicity from the target compounds. However, most of the **O1–O27** furanone derivatives have certain cytotoxic activity on HepG2 hepatocellular carcinoma cells, especially for **O5** and **O10** both containing a trifluoroethyl group. For **S1–S16** and **SO1–SO16** series, most derivatives of furanones have certain antiproliferative activity on MDA-MB-231 breast cancer cells, C6 brain glioma cells and MCF-7 breast cancer cells. Therefore, the preliminary results reveal that the introduction of amino acids as safe linkers basically has no toxic effect as anticipated.

At the same time, due to the presence of an amino acid structural unit in **N1–N16** series, they can be expected to be further studied in the field of cell compatibility. And in combination with other bioactive results, it is possible to introduce a fluorine-containing group into **N1–N16** series *via* esterification with trifluoroethanol to develop the potential 2(5*H*)-furanone drugs. Studies on these aspects are in progress.

Conclusions and outlook

A series of amino acid derivatives are synthesized with 5-alkoxy-3,4-dihalo-2(5*H*)-furanones and amino acids *via* a metal-free C–N coupling reaction as expected after the optimal reaction conditions are established. As linkers, the effect of amino acids has been investigated by comparison with other 2(5H)-furanone compounds by constructing C–O/C–S bonds. The results of the biological activity assay reveal that the amino acids in these 2(5H)-furanone derivatives exhibiting good biocompatibility can be good linkers as expected. What's more, a fluorine-containing group is beneficial for improving the bioactivity of 2(5H)-furanone drugs. The introduction of fluorine-containing groups into 2(5H)-furanones will be our future work.

Importantly, as linkers of potential 2(5H)-furanone drugs, amino acids can provide a well-reactive site into 2(5H)-furanone derivatives by reacting with the carboxyl group in our amino acid linking unit under mild conditions. Particularly, some amino acids with long chains may improve the tolerance of 2(5H)-furanone substrates for the introduction of some complex, sensitive and sterically hindered bioactive moieties with amino or hydroxy groups.^{71–73} Thus, the introduction of amino acids as linkers will provide a new effective approach for the development of 2(5H)-furanone derivative drugs with good biocompatibility.

Published on 02 May 2019. Downloaded by UNIV OF LOUISIANA AT LAFAYETTE on 5/10/2019 10:14:53 AM



Table 3 Inhibitory effects of target molecules N1–N16, contrasting substances and cisplatin against the growth of the selected cell lines

	$IC_{50}/\mu M$ (Max co	IC ₅₀ /μM (Max concentration: 300 μM)						
No.	HepG2	MDA-MB-231	C6	MCF-7	A549	EC-1	CNE-1	
N1	>300	>300	>300	>300	>300	>300	66.69 ± 2.25	
N2	>300	>300	>300	>300	>300	>300	>300	
N3	>300	>300	>300	>300	>300	>300	>300	
N4	>300	>300	>300	>300	>300	>300	>300	
N5	>300	>300	>300	>300	>300	284.98 ± 20.15	>300	
N6	>300	>300	>300	>300	>300	>300	>300	
N/	>300	>300	>300	$104.6/ \pm 8.50$	>300	>300	>300	
IN8 NO	>300	>300	>300	>300	>300	>300	>300	
N9 N10	>300	>300	>300	>300	>300	>300	>300	
N11	>300	>300	>300	>300	>300	>300	>300	
N12	>300	>300	>300	>300	>300	>300	>300	
N13	>300	>300	>300	>300	>300	>300	>300	
N14	>300	>300	>300	>300	>300	>300	>300	
N15	>300	>300	>300	>300	>300	>300	>300	
N16	>300	>300	>300	>300	>300	>300	>300	
01	196.08 ± 8.46	>300	>300	239.43 ± 12.43	299.79 ± 8.33	223.73 ± 12.06	>300	
O2	190.05 ± 3.85	>300	>300	>300	>300	>300	299.13 ± 3.03	
O3	152.40 ± 2.28	161.59 ± 33.99	>300	>300	>300	>300	230.58 ± 3.54	
04	151.04 ± 3.26	>300	>300	>300	>300	>300	>300	
05	40.22 ± 0.57	127.06 ± 1.25	73.11 ± 0.61	73.53 ± 1.57	127.38 ± 0.76	286.20 ± 12.94	>300	
06	>300	>300	266.23 ± 6.85	>300	$2/1.60 \pm /.83$	>300	220.98 ± 12.11	
0/	127.48 ± 2.55	>300	>300	>300	>300	>300	>300	
08	208.05 ± 4.28 149.64 ± 6.47	>300	>300	>300	>300	229.70 ± 7.49	274.53 ± 2.00	
09	149.04 ± 0.47	2300	> 300	2300	> 500	> 300	> 300	
O10	41.12 ± 0.94	123.49 ± 1.96	58.47 ± 0.52	19.67 ± 0.37	108.36 ± 1.09	44.68 ± 0.68	>300	
011	120.68 ± 4.39	>300	279.32 ± 6.40	>300	>300	>300	>300	
012	107.51 ± 2.41	>300	>300	>300	>300	284.35 ± 10.33	256.45 ± 6.08	
013	195.15 ± 7.25	>300	>300	223.30 ± 10.27	150.25 ± 8.62	281.50 ± 16.25	295.63 ± 7.74	
014	292.53 ± 3.82	257.21 ± 9.05	>300	>300	>300	>300	>300	
015	110.13 ± 2.91	>300	>300	>300	>300	>300	>300	
010	>300	>300	>300	>300	>300	>300	>300	
018	>300	>300	>300	>300	>300	>300	203.88 ± 3.24	
019	186 63 + 3 68	288 69 + 3 81	>300	193 52 + 3 58	186 87+ 3 96	246 71 + 5 86	>300	
020	>300	>300	>300	>300	>300	159.90 + 3.95	>300	
O21	>300	181.45 ± 1.93	109.13 ± 2.39	145.12 ± 1.23	182.53 ± 3.50	>300	185.65 ± 2.58	
O22	>300	>300	>300	>300	>300	>300	>300	
O23	>300	>300	269.43 ± 5.91	>300	>300	>300	>300	
O24	>300	297.26 ± 7.76	>300	299.34 ± 9.65	>300	293.68 ± 7.9	154.90 ± 2.82	
O25	>300	233.76 ± 10.25	>300	>300	>300	>300	>300	
O26	416.03 ± 4.1	221.64 ± 2.6	231.35 ± 2.24	192.44 ± 3.80	212.16 ± 4.57	197.19 ± 7.37	108.72 ± 1.57	
O27	85.79 ± 3.04	215.44 ± 6.57	158.83 ± 1.21	130.73 ± 2.20	152.05 ± 1.23	>300	>300	
S1	>300	>300	175.32 ± 1.41	252.41 ± 8.31	>300	224.25 ± 8.04	>300	
S2	>300	>300	176.68 ± 4.81	165.10 ± 4.34	218.16 ± 5.70	185.17 ± 10.56	>300	
S3	184.70 ± 1.27	180.77 ± 2.77	138.88 ± 2.18	181.02 ± 3.18	296.80 ± 2.70	229.75 ± 9.35	>300	
S4	>300	116.69 ± 2.38	84.52 ± 1.44	86.19 ± 2.84	$143./3 \pm 5.42$	$248./9 \pm 3./3$	>300	
55	>300	>300	>300	>300	>300	>300	>300	
50	239.97 ± 4.31	249.08 ± 5.48	147.08 ± 3.32	196.05 ± 9.83 175.27 ± 5.75	262.89 ± 7.39	100.80 ± 9.25 110 E2 ± 2 E6	>300	
57	133.90 ± 1.03	216.70 ± 2.17 176.20 ± 2.62	207.33 ± 3.30 170 15 ± 1 50	173.37 ± 3.73	202.98 ± 3.39	112.33 ± 3.30 250.70 ± 9.92	>300	
50	>300	170.29 ± 3.02 167.66 ± 1.48	178.13 ± 1.38 165.51 ± 2.64	187.09 ± 4.02	230 93 + 3 95	250.79 ± 3.83 169 95 + 3.08	2300 190 79 + 3 73	
S10	231.03 ± 1.44	21458 ± 342	166.01 ± 2.04	107.50 ± 4.15 184 67 + 3 13	230.33 ± 3.93 239 34 + 1 98	>300	>300	
S11	>300	197.28 ± 4.72	162.49 ± 3.17	211.02 ± 7.35	>300	224 22 + 12 31	>300	
S12	224.00 + 3.05	215.34 + 3.43	157.99 ± 2.07	130.12 ± 5.53	>300	144.78 ± 11.34	>300	
S13	>300	168.41 ± 4.70	152.69 ± 2.97	178.06 ± 6.30	268.61 ± 7.82	205.52 ± 4.64	>300	
S14	>300	>300	158.05 ± 2.64	>300	>300	>300	>300	
S15	>300	185.54 ± 6.72	146.55 ± 1.72	191.93 ± 11.57	256.50 ± 8.67	207.351 ± 10.57	>300	
S16	251.28 ± 2.72	295.29 ± 8.66	167.40 ± 3.88	175.21 ± 5.34	258.95 ± 2.16	187.75 ± 11.82	>300	
SO1	>300	184.71 ± 4.58	198.43 ± 4.76	181.12 ± 7.73	>300	>300	>300	
SO2	>300	>300	128.90 ± 3.52	168.99 ± 2.74	>300	>300	>300	
SO3	246.52 ± 3.72	142.23 ± 1.94	115.98 ± 2.15	119.12 ± 1.69	173.78 ± 2.67	>300	>300	
SO4	>300	175.28 ± 3.36	129.44 ± 6.93	134.66 ± 5.44	282.91 ± 9.71	>300	>300	
SO2	>300	216.38 ± 3.32	101.25 ± 3.43	216.78 ± 7.16	>300	>300	>300	

No.	$IC_{50}/\mu M$ (Max concentration: 300 μM)							
	HepG2	MDA-MB-231	C6	MCF-7	A549	EC-1	CNE-1	
SO6	295.97 ± 4.38	86.19 ± 1.95	67.87 ± 1.32	>300	>300	199.58 ± 10.07	154.22 ± 6.66	
SO7	>300	174.93 ± 3.96	96.39 ± 1.21	122.02 ± 4.57	>300	>300	>300	
SO8	>300	183.81 ± 6.59	106.69 ± 2.66	112.87 ± 3.65	>300	>300	>300	
SO9	266.74 ± 2.41	145.58 ± 2.23	49.80 ± 0.97	83.24 ± 2.68	253.47 ± 3.74	>300	>300	
SO10	>300	216.27 ± 3.60	117.68 ± 3.70	150.63 ± 6.98	>300	>300	>300	
SO11	>300	283.00 ± 5.54	>300	262.22 ± 10.25	>300	>300	>300	
SO12	>300	198.36 ± 5.51	105.17 ± 2.12	124.86 ± 1.81	255.38 ± 1.52	>300	>300	
SO13	>300	159.93 ± 2.74	153.07 ± 3.29	123.13 ± 3.62	240.55 ± 4.87	>300	>300	
SO14	>300	131.58 ± 4.22	92.70 ± 3.62	>300	>300	293.03 ± 11.18	186.76 ± 7.86	
SO15	>300	161.48 ± 1.97	108.28 ± 2.73	117.14 ± 4.25	235.38 ± 7.38	>300	>300	
SO16	>300	147.13 ± 4.65	65.93 ± 2.73	86.30 ± 2.18	216.27 ± 8.55	170.43 ± 2.80	123.23 ± 2.67	
Cisplatin	93.1 ± 0.9	17.8 ± 0.9	18.7 ± 0.5	76.9 ± 0.9	ND	ND	19.1 ± 0.5	

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

We gratefully acknowledge financial support from the Guangdong Natural Science Foundation (No. 2014A030313429), the Guangzhou Science and Technology Project Scientific Special (General Items, No. 201607010251), the Open Fund of the Key Laboratory of Functional Molecular Engineering of Guangdong Province in SCUT (No. 2017kf01), and the Guangdong Provincial Science and Technology Project (No. 2017A010103016).

Notes and references

- 1 D. M. M. Jaradat, Thirteen decades of peptide synthesis: Key developments in solid phase peptide synthesis and amide bond formation utilized in peptide ligation, *Amino Acids*, 2018, **50**, 39.
- 2 F. Agostini, J.-S. Vçller, B. Koksch, C. G. Acevedo-Rocha, V. Kubyshkin and N. Budisa, Biocatalysis with unnatural amino acids: Enzymology meets xenobiology, *Angew. Chem., Int. Ed.*, 2017, **56**, 9680.
- 3 R. A. Jones, Y. Thillier, S. S. Panda, N. R. Rosario, C. D. Hall and A. R. Katritzky, Synthesis and characterization of glucosamine-NSAID bioconjugates, *Org. Biomol. Chem.*, 2014, **12**, 8325.
- 4 Z. Meng, B. Yu, G. Han, M. Liu, B. Shan, G. Dong, Z. Miao, N. Jia, Z. Tan and B. Li, Chlorin p₆-based water-soluble amino acid derivatives as potent photosensitizers for photodynamic therapy, *J. Med. Chem.*, 2016, **59**, 4999.
- 5 D. M. Suyoga Vardhan, C. S. Shantharam, R. Suhas and D. Channe Gowda, Synthesis and evaluation of novel ureido/thioureido derivatives of amino acid conjugated 2,3dichlorophenyl piperazine as highly potent antiglycating agents, *J. Saudi Chem. Soc.*, 2017, **21**, S248.
- 6 D. Haeussler, A.-C. Schulz-Fincke, A.-M. Beckmann, A. Keils, E. Gilberg, M. Mangold, J. Bajorath, M. Stirnberg,

T. Steinmetzer and M. Guetschow, A fluorescent-labeled phosphono bisbenzguanidine as an activity-based probe for matriptase, *Chem. – Eur. J.*, 2017, **23**, 5205.

- 7 G. F. Li, W. L. Feng, N. Corrigan, C. Boyer, X. Wang and J. T. Xu, Precise synthesis of poly(*N*-acryloyl amino acid) through photoinduced living polymerization, *Polym. Chem.*, 2018, 9, 2733.
- 8 T. Smidlehner, A. Kurutos, J. Slade, R. Belužić, D. L. Ang, A. Rodger and I. Piantanida, Versatile click cyanine amino acid conjugates showing one-atom-influenced recognition of DNA/RNA secondary structure and mitochondrial localisation in living cells, *Eur. J. Org. Chem.*, 2018, 1682.
- 9 Y. M. Liu, L. Y. Zhao, R. R. Xing, T. F. Jiao, W. X. Song and X. H. Yan, Covalent assembly of amphiphilic bola-amino acids into robust and biodegradable nanoparticles for *in vitro* photothermal therapy, *Chem. – Asian J.*, 2018, **13**, 3526.
- 10 S. R. Nelli, R. D. Chakravarthy, M. Mohiuddin and H.-C. Lin, The role of amino acids on supramolecular coassembly of naphthalenediimide-pyrene based hydrogelators, *RSC Adv.*, 2018, 8, 14753.
- 11 K. S. Yu, M. M. Lin, H. J. Lee, K.-S. Tae, B.-S. Kang, J. H. Lee, N. S. Lee, Y. G. Jeong, S.-Y. Han and D. K. Kim, Receptor-meditated endocytosis by hyaluronic acid@superpara-magnetic nanovetor for targeting of CD44overexpressing tumor cells, *Nanomaterials*, 2016, 6, 149.
- 12 J. Malakootikhah, A. H. Rezayan, B. Negahdari, S. Nasseri and H. Rastegar, Glucose reinforced Fe₃O₄@cellulose mediated amino acid: Reusable magnetic glycolnanoparticles with enhanced bacteria capture efficiency, *Carbohydr. Polym.*, 2017, **170**, 190.
- 13 L. Momtazi, H. H. Soensteby, D. A. Dartt, J. R. Eidet and O. Nilsen, Bioactive titaminates from molecular layer deposition, *RSC Adv.*, 2017, 7, 20900.
- 14 A. D. Tiwari, S. S. Panda, A. S. Girgis, S. Sahu, R. F. George, A. M. Srour, B. L. Starza, A. M. Asiri, C. D. Hall and A. R. Katritzky, Microwave assisted synthesis and QSAR study of novel NSAID acetaminophen conjugates with amino acid linkers, *Org. Biomol. Chem.*, 2014, **12**, 7238.
- 15 S. A. Dingsdag, B. C. Yap, N. Hunter and M. J. Crossley, Amino acid-linked porphyrin-nitroimidazole antibiotics

targeting porphyromonas gingivalis, *Org. Biomol. Chem.*, 2015, 13, 98.

- 16 R. W. Jinadasa, Z. Zhou, M. G. H. Vicente and K. M. Smith, Syntheses and cellular investigations of di-aspartate and aspartate-lysine chlorin e₆ conjugates, *Org. Biomol. Chem.*, 2016, 14, 1049.
- 17 S. Udhayakumar, K. G. Shankar, S. Sowndarya, S. Venkatesh, C. Muralidharan and C. Rose, L-Arginine intercedes bio-crosslinking of a collagen-chitosan 3D-hybrid scaffold for tissue engineering and regeneration: in silico, *in vitro*, and *in vivo* studies, *RSC Adv.*, 2017, 7, 25070.
- 18 A. S. Skwarecki, K. Skarbek, D. Martynow, M. Serocki, I. Bylińska, M. J. Milewska and S. Milewski, Molecular umbrellas modulate the selective toxicity of polyene macrolide antifungals, *Bioconjugate Chem.*, 2018, 29, 1454.
- 19 A. M. K. Sweed, M. O. Senge, S. M. S. Atta, D. S. Farrag, A.-R. H. Abdel-Rahman and Y. M. Shaker, Synthesis of amphiphilic meso-tetrasubstituted porphyrin-L-amino acid and -heterocyclic conjugates based on m-THPP, *J. Porphyrins Phthalocyanines*, 2018, 22, 997.
- 20 J. Luo, H.-F. Wang, X. Han, L.-W. Xu, J. Kwiatkowski, K.-W. Huang and Y.-X. Lu, The direct asymmetric vinylogous aldol reaction of furanones with α-ketoesters: Access to chiral γ-butenolides and glycerol derivatives, *Angew. Chem., Int. Ed.*, 2011, **50**, 1861.
- 21 P. Yang, M. Yao, J. Li, Y. Li and A. Li, Total synthesis of rubriflordilactone B, *Angew. Chem., Int. Ed.*, 2016, 55, 6964.
- 22 Q. A. Castillo, J. Triana, J. L. Eiroa, L. Calcul, E. Rivera, L. Wojtas, J. MPadron, L. Boberieth, M. Keramane, E. Abel-Santos, L. A. Baez and E. A. Germosen, Ent-labdane diterpenoids from the aerial parts of eupatorium obtusissmum, *J. Nat. Prod.*, 2016, **79**, 907.
- 23 C.-K. Peng, T. Zeng, X.-J. Xu, Y.-Q. Chang, W. Hou, K. Lu, H. Lin, P.-H. Sun, J. Lin and W.-M. Chen, Novel 4-(4-substituted amidobenzyl)furan-2(5*H*)-one derivatives as topoisomerase I inhibitors, *Eur. J. Med. Chem.*, 2017, **127**, 187.
- 24 M.-X. Wei, J. Zhang, F.-L. Ma, M. Li, J.-Y. Yu, W. Luo and X.-Q. Li, Synthesis and biological activities of dithiocarbamates containing 2(5*H*)-furanone-piperazine, *Eur. J. Med. Chem.*, 2018, 155, 165.
- 25 Z.-H. Wang, Y. You, Y.-Z. Chen, X.-Y. Xu and W.-C. Yuan, An asymmetric organocatalytic vinylogous Mannich reaction of 3-methyl-5-arylfuran-2(3*H*)-ones with *N*,-(2-pyridinesulfonyl) imines: enantioselective synthesis of δ-amino γ , γ -disubstituted butenolides, *Org. Biomol. Chem.*, 2018, **16**, 1636.
- 26 K. Williams, A. J. Szwalbe, N. P. Mulholland, J. L. Vincent, A. M. Bailey, C. L. Willis, T. J. Simpson and R. J. Cox, Heterologous production of fungal maleidrides reveals the cryptic cyclization involved in their biosynthesis, *Angew. Chem., Int. Ed.*, 2016, 55, 6784.
- 27 S. Senthilkumar, S. Valdomir, D. Ganapathy, Y. Zhang and L. F. Tietze, Enantioselective total synthesis of the fungal metabolite blennolide D and the enantiomers of blennolide E and F, *Org. Lett.*, 2018, **20**, 2186.

- 28 M. J. Byrne, N. R. Lees, L.-C. Han, M. W. van der Kamp, A. J. Mulholland, J. E. M. Stach, C. L. Willis and P. R. Race, The catalytic mechanism of a natural Diels-Alderase revealed in molecular detail, *J. Am. Chem. Soc.*, 2016, 138, 6095.
- 29 H. Zhang, K.-K. Zhu, Y.-S. Han, C. Luo, M. A. Wainberg and J.-M. Yue, Flueggether A and virosinine A, anti-HIV alkaloids from flueggea virosa, *Org. Lett.*, 2015, 17, 6274.
- 30 C.-C. Yuan, B. Du, H.-P. Deng, Y. Man and B. Liu, Total syntheses of Sarcandrolide J and Shizukaol D: Lindenane sesquiterpenoid [4+2] dimers, *Angew. Chem., Int. Ed.*, 2017, 56, 637.
- 31 F.-M. Xi, S.-G. Ma, Y.-B. Liu, L. Li and S.-S. Yu, Artaboterpenoids A and B, bisabolene-derived sesquiterpenoids from artabotrys hexapetalus, *Org. Lett.*, 2016, **18**, 3374.
- 32 H. R. Khatri, B. Bhattarai, W. Kaplan, Z.-Z. Li, M. J. C. Long, Y. Aye and P. Nagorny, Modular total synthesis and cell-based anticancer activity evaluation of ouabagenin and other cardiotonic steroids with varying degrees of oxygenation, *J. Am. Chem. Soc.*, 2019, 141, 4849.
- 33 A. DeBono, B. Capuano and P. J. Scammells, Progress toward the development of noscapine and derivatives as anticancer agents, *J. Med. Chem.*, 2015, **58**, 5699.
- 34 Z. Z. Li, H. Su, W. W. Yu, X. J. Li, H. Cheng, M. Y. Liu, X. F. Pang and X. Z. Zou, Design, synthesis and anticancer activities of novel otobain derivatives, *Org. Biomol. Chem.*, 2016, 14, 277.
- 35 N. Grimblat, T. S. Kaufman and A. M. Sarotti, Computational chemistry driven solution to rubriflordilactone B, *Org. Lett.*, 2016, **18**, 6420.
- 36 X. Li, P. H.-Y. Cheong and R. G. Carter, Schinortriterpenoids: A case study in synthetic design, *Angew. Chem., Int. Ed.*, 2017, **56**, 1704.
- 37 X. Wu, T. Iwata, A. Scharf, T. Qin, K. D. Reichl and J. A. Porco Jr., Asymmetric synthesis of gonytolide A: Strategic use of an aryl halide blocking group for oxidative coupling, *J. Am. Chem. Soc.*, 2018, **140**, 5969.
- 38 K. Tan, H. Yan, P. B. Lu, Y. H. Liu, R. G. Ji, Z. X. Liu, Y.-M. Li, F.-C. Yu and Y. H. Shen, Access to multisubstituted 2(5*H*)-furanones using hydrogen bonding-promoted ring-closing metathesis and polyamine workup, *J. Org. Chem.*, 2019, 84, 3419.
- 39 B. M. Trost, E. Gnanamani, C. A. Kalnmals, C.-I. J. Hung and J. S. Tracy, Direct enantio- and diastereoselective vinylogous addition of butenolides to chromones catalyzed by Zn-pro phenol, *J. Am. Chem. Soc.*, 2019, **141**, 1489.
- 40 Y.-H. Tan, J.-X. Li, F.-L. Xue, J. Qi and Z.-Y. Wang, Concise synthesis of chiral 2(5*H*)-furanone derivatives possessing 1,2,3-triazole moiety *via* one-pot approach, *Tetrahedron*, 2012, 68, 2827.
- 41 J.-P. Huo, J.-C. Luo, W. Wu, J.-F. Xiong, G.-Z. Mo and Z.-Y. Wang, Synthesis and characterization of fluorescent brightening agents with chiral 2(5*H*)-furanone and bis-1,2,3-triazole structure, *Ind. Eng. Chem. Res.*, 2013, **52**, 11850.

- 42 J.-P. Huo, G.-H. Deng, W. Wu, J.-F. Xiong, M.-L. Zhong and Z.-Y. Wang, Electrophoretic deposition polymerization of diacetylenes with tunable structure, *Macromol. Rapid Commun.*, 2013, 34, 1779.
- 43 Y.-H. Tan, J.-X. Li, J.-P. Huo, F.-L. Xue and Z.-Y. Wang, Synthesis of 2(5*H*)-furanone derivatives with symmetrical and unsymmetrical bis-1,2,3-triazole structure, *Synth. Commun.*, 2014, 44, 2974.
- 44 J. Shi, X.-D. Tang, Y.-C. Wu, H.-N. Li, L.-J. Song and Z.-Y. Wang, Palladium-catalyzed desulfitative arylation of 5-alkoxy-3,4-dibromo-2(5*H*)-furanone with sodium arylsulfinates, *Eur. J. Org. Chem.*, 2015, 1193.
- 45 J. Shi, X.-D. Tang, Y.-C. Wu, J.-F. Fang, L. Cao, X.-Y. Chen and Z.-Y. Wang, A radical coupling reaction of DMSO with sodium arylsulfinates in air: Mild utilization of DMSO as C_1 resource for the synthesis of arylsulfonyl dibromomethane, *RSC Adv.*, 2016, **6**, 25651.
- 46 L. Cao, S.-H. Luo, H.-Q. Wu, L.-Q. Chen, K. Jiang, Z.-F. Hao and Z.-Y. Wang, Copper(1)-catalyzed alkyl- and arylsulfenylation of 3,4-dihalo-2(5*H*)-furanones (X=Br, Cl) with sulfoxides under mild conditions, *Adv. Synth. Catal.*, 2017, 359, 2961.
- 47 L. Cao, J.-X. Li, H.-Q. Wu, K. Jiang, Z.-F. Hao, S.-H. Luo and Z.-Y. Wang, Metal-free sulfonylation of 3,4-dihalo-2(5*H*)-furanones (X = Cl, Br) with sodium sulfinates under air atmosphere in aqueous media *via* a radical pathway, *ACS Sustainable Chem. Eng.*, 2018, 6, 4147.
- 48 Y.-Q. Mo, Z.-Y. Wang, W.-J. Mei, J.-H. Fu, Y.-H. Tan and S.-H. Luo, Reaction of 5-alkoxy-3,4-dihalo-2(5*H*)-furanones with secondary amines: Expected versus unanticipated products and their preliminary bioactivity investigations, *Monatsh. Chem.*, 2012, 143, 443.
- 49 Y.-C. Wu, S.-H. Luo, W.-J. Mei, L. Cao, H.-Q. Wu and Z.-Y. Wang, Synthesis and biological evaluation of 4-biphenylamino-3-halo-2(5*H*)-furanones as potential anticancer agents, *Eur. J. Med. Chem.*, 2017, **139**, 84.
- 50 Y.-C. Wu, L. Cao, W.-J. Mei, H.-Q. Wu, S.-H. Luo, H.-Y. Zhan and Z.-Y. Wang, Bis-2(5*H*)-furanone derivatives as new anticancer agents: Design, synthesis, biological evaluation, and mechanism studies, *Chem. Biol. Drug Des.*, 2018, **92**, 1232.
- 51 S. Bagmare, A. D. Gunjal and V. A. Kumar, Investigation of the effect of amino acid chirality in the internucleoside linker on DNA:DNA and DNA:RNA duplex stability, *Tetrahedron*, 2015, **71**, 2442–2449.
- 52 Q. Wang and J. Holst, L-type amino acid transport and cancer: targeting the mTORC1 pathway to inhibit neoplasia, *Am. J. Cancer Res.*, 2015, **5**, 1281.
- 53 P. Peng, J.-F. Xiong, G.-Z. Mo, J.-L. Zheng, R.-H. Chen, X.-Y. Chen and Z.-Y. Wang, A concise synthesis of benzimidazoles *via* the microwave-assisted one-pot batch reaction of amino acids up to a 10 g scale, *Amino Acids*, 2014, 46, 2427.
- 54 C.-W. Bi, C.-X. Zhang, Y.-H. Li, S. Tang, S.-G. Wang, R.-G. Shao, H.-G. Fu, F. Su and D.-Q. Song, Synthesis and biological evaluation of sophoridinol derivatives as a novel family of potential anticancer agents, *ACS Med. Chem. Lett.*, 2014, 5, 1225.

- 55 Y.-N. Liu, J.-J. Wang and Y.-T. Ji, Design, synthesis, and biological evaluation of 1-methyl-1,4-dihydroindeno- [1,2-c]pyrazole analogues as potential anticancer agents targeting tubulin colchicine binding site, *J. Med. Chem.*, 2016, **59**, 5341.
- 56 T. Szekely, O. Roy, E. Dériaud, A. Job, R. Lo-Man, C. Leclerc and C. Taillefumier, Design, synthesis, and immunological evaluation of a multicomponent construct based on a glycotripeptoid core comprising B and T cell epitopes and a toll-like receptor 7 agonist that elicits potent immune responses, *J. Med. Chem.*, 2018, **61**, 9568.
- 57 D. Maiti, B. P. Fors, J. L. Henderson, Y. Nakamura and S. L. Buchwald, Palladium-catalyzed coupling of functionalized primary and secondary amines with aryl and heteroaryl halides: two ligands suffice in most cases, *Chem. Sci.*, 2011, 2, 57.
- 58 M. J. D. Pires, D. L. Poeira, S. I. Purificacao and M. M. B. Marques, Synthesis of substituted 4-, 5-, 6-, and 7-azaindoles from aminopyridines *via* a cascade C-N crosscoupling/Heck reaction, *Org. Lett.*, 2016, **18**, 3250.
- 59 L. Ouyang and W.-Q. Wu, Recent advancements in palladium-catalyzed reactions involving molecular oxygen, *Curr. Opin. Green Sustain. Chem.*, 2017, 7, 46.
- 60 L. Ouyang, J.-Z. Huang, J.-X. Li, C.-Q. Qi, W.-Q. Wu and H.-F. Jiang, Palladium-catalyzed oxidative amination of homoallylic alcohols: Sequentially installing carbonyl and amino groups along an alkyl chain, *Chem. Commun.*, 2017, 53, 10422.
- 61 K. Yang, Y.-T. Qiu, Z. Li, Z.-Y. Wang and S. Jiang, Ligands for copper-catalyzed C-N bond forming reactions with 1 mol% CuBr as catalyst, *J. Org. Chem.*, 2011, **76**, 3151.
- 62 C. Thomas, M. Wu and K. L. Billingsley, Aminationoxidation strategy for the copper-catalyzed synthesis of monoarylamines, *J. Org. Chem.*, 2016, **81**, 330.
- 63 X. Gao, L. Y. Tang, L. H. Huang, Z. S. Huang, Y. F. Ma and G. Wu, Oxidative aminoarylselenation of maleimides *via* copper-catalyzed four-component cross-coupling, *Org. Lett.*, 2019, **21**, 745.
- 64 L. Ouyang, J.-X. Li, J. Zheng, J.-Z. Huang, C.-R. Qi, W.-Q. Wu and H.-F. Jiang, Access to α -amino acid asters through palladium-catalyzed oxidative amination of vinyl ethers with hydrogen peroxide as the oxidant and oxygen source, *Angew. Chem., Int. Ed.*, 2017, **56**, 15926.
- 65 J. W. Yuan, S. N. Liu, Y. M. Xiao, P. Mao, L. R. Yang and L. B. Qu, Palladium-catalyzed oxidative amidation of quinoxalin-2(1*H*)-ones with acetonitrile: a highly efficient strategy toward 3-amidated quinoxalin-2(1*H*)-ones, *Org. Biomol. Chem.*, 2019, **17**, 876.
- 66 J. J. Zhang, Q. L. Qian, Y. Wang, B. B. A. Bediako, M. Cui, G. Y. Yang and B. X. Han, Synthesis of acetamides using CO₂, methanol, H₂ and amines, *Green Chem.*, 2019, 21, 233.
- 67 X. Q. Yu, S. N. Yang, Y. Zhang, M. J. Guo, Y. Yamamoto and M. Bao, Intermolecular amidation of quinoline N-oxides with arylsulfonamides under metal-free conditions, *Org. Lett.*, 2017, **19**, 6088.

- 68 K. Shanmugam, M. Karthick and G. Thirumanavelan, Basepromoted amidation and esterification of imidazolium salts *via* acyl C-C bond cleavage: an access to aromatic amides and esters, *J. Org. Chem.*, 2018, **84**, 738.
- 69 S. R. Manne, J. Chandra and B. Mandal, Synthesis of *o*-nitroarylamines *via* ipso nucleophilic substitution of sulfonic acids, *Org. Lett.*, 2019, 21, 636.
- 70 H.-Q. Wu, S.-H. Luo, L. Cao, H.-N. Shi, B.-W. Wang and Z.-Y. Wang, DABCO-mediated C-O bond formation from C_{sp2}-halogen bond-containing compounds and alkyl alcohols, *Asian J. Org. Chem.*, 2018, 7, 2479.
- 71 Y. Ohsedo, M. Oono, K. Saruhashi, H. Watanabe and N. Miyamoto, Thixotropic stiff hydrogels from a new class of oleoyl-D-glucamine-based low-molecular-weight gelators, *RSC Adv.*, 2017, 7, 41686.

- 72 L. Ma, Y. Chen, X. Wang, M. Xiong, Y. Sun, X. Zhang and Y. Zhao, Design, characterization, and *in vitro* antiproliferative efficacy of gemcitabine conjugates based on carboxymethyl glucan, *Bioorg. Med. Chem. Lett.*, 2018, 28, 2920.
- 73 P. E. Brandish, A. Palmieri, S. Antonenko, M. Beaumont, L. Benso, M. Cancilla, M. Cheng, L. Fayadat-Dilman, G. Feng, I. Figueroa, J. Firdos, R. Garbaccio, L. Garvin-Queen, D. Gately, P. Geda, C. Haines, S. Hseih, D. Hodges, J. Kern, N. Knudsen, K. Kwasnjuk, L. Liang, H. Ma, A. Manibusan, P. L. Miller, L. Y. Moy, Y. Qu, S. Shah, J. S. Shin, P. Stivers, Y. Sun, D. Tomazela, H. C. Woo, D. Zaller, S. Zhang, Y. Zhang and M. Zielstorff, Development of anti-CD74 antibody-drug conjugates to target glucocorticoids to immune cells, *Bioconjugate Chem.*, 2018, 29, 2357.