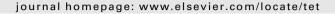
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Selective β -oxidation of α -sulfanyl amides

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ABSTRACT

A selective β -oxidation of a series of α -sulfanyl amides to the corresponding β -oxo- α -sulfanyl amides is described. This selective efficient oxidation of an unfunctionalised methyl or methylene group occurs under mild conditions, involving three sequential transformations conducted without isolation of the intermediates. Critically neither the sulfur nor the reactive α -CH bond is affected in the overall process. © 2011 Elsevier Ltd. All rights reserved.

1. Introduction

The selective oxygenation of unactivated C–H bonds is challenging and has attracted much attention. Alkane oxidation is normally achieved using oxidants such as hydrogen peroxide or molecular oxygen in the presence of a transition metal catalyst. ^{1–4} For substrates, which contain a sulfide, selective oxidation at carbon over the sulfur centre is extremely difficult to achieve.

β-Oxo esters and amides are very useful compounds having widespread biological applications. For example, 5,6-dihydro-2*H*-pyran-2-ones **1**, which contain the β-keto- α -sulfanyl ester functionality, have been investigated as inhibitors of HIV protease, which is essential for viral replication. ^{5,6} Westwood et al. reported the activity of α -cyano- β -oxoamide **2** and its derivatives as an inhibitor of an enzyme that is involved in the de novo pyrimidine biosynthesis. ⁷ Antirheumatic oxindoles contain a β -ketoamide functionality in which the acidic enolic OH group is essential for cyclooxygenase inhibitory activity; ⁸ Tenidap **3** is one of the most potent oxindoles in the treatment of rheumatoid and osteoarthritis.

The preparation of β -oxo esters/amides does not usually involve direct oxidation at the β -carbon. Oxidation of a nonactivated methyl or methylene group is challenging, 9 and almost invariably involves attack by free radicals, resulting in regiocontrol and selectivity problems. β -Keto amides and esters may be prepared by various methods where the β -carbon is already at the same level of oxidation as a carbonyl carbon. 10,11

We recently reported a highly efficient and stereoselective transformation of α -sulfanyl amides to the corresponding α -sulfanyl- β -chloroacrylamide derivatives on treatment with NCS. ¹² Chemoselective and stereoselective oxidations of the β -chloroacrylamides to the sulfoxide and sulfone levels of oxidation have extended the scope of this methodology, ^{13,14} and the dipolarophilic and dienophilic behaviour of the β -chloroacrylamides has also been described. ^{15–17} The synthetic potential of the β -chloroacrylamides as Michael acceptors in nucleophilic addition/substitution reactions has also been investigated, including addition of morpholine to yield β -enaminoamides. ¹⁸ In early experiments, partial hydrolysis was seen on purification of the morpholine adducts by chromatography on silica gel, hence showing that the β -enaminoamides could be hydrolysed (Scheme 1).

Scheme 1.

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Herein, the optimisation of the hydrolysis of the β -enaminoamides to the corresponding β -hydroxyacrylamides is described. Having established that the β -enaminoamides could be efficiently hydrolysed, a 'one-pot' oxidation from the α -sulfanyl amides to the β -hydroxyacrylamides was subsequently developed (Scheme 2).

2. Results and discussion

The nucleophilic addition of morpholine to a range of β -chloroacrylamides has been described. The resulting β -morpholinoacrylamides contain an enamine functionality, and during chromatographic purification with a number of these β -morpholinoacrylamides, partial or complete hydrolysis of the acid labile enamine group was observed (Scheme 3). For example, partial hydrolysis of the β -morpholinoacrylamide **4a** on silica gel led to a 3:1 mixture of the keto and enol tautomers of **5a**, while hydrolysis of **4b** and **4c** led to the enol tautomers **5b** and **5c** exclusively.

The β-hydroxyacrylamides **5d**, **7a**, **9a**–**9c** were isolated, following extraction into dichloromethane and concentration, as single stereoisomers (tentatively assigned as E), while a second set of minor signals (\sim 7%), which were present in the 1 H NMR spectrum of **9c**, and which persisted on recrystallisation, were tentatively assigned as the other stereoisomer. The β-hydroxyacrylamides **5d**, **7a**, **9a**–**9b** were clean by 1 H and 13 C NMR spectroscopy, and further purification was not required.

The β -hydroxyacrylamides may exist as the E- or Z-isomers. It is likely that the E-isomer is favoured due to a stabilising intramolecular hydrogen bond between the enol OH and the amide carbonyl; however, at this stage there is no definitive structural evidence for the stereochemistry. The enol form predominates in most instances as indicated by the signal for the β -hydrogen at 7.10–7.99 ppm and the signal for the OH group at 13.52–15.70 ppm in the 1 H NMR spectra.

In the room temperature ${}^{1}H$ NMR spectra of **9a**—**9c**, the signals for the β -hydrogens were evident as broad singlets. To establish if the broadening of the β -hydrogen signal arises from the interconversion of the E and Z isomers on the NMR timescale or if coupling to the adjacent hydroxyl group is responsible, the ${}^{1}H$ NMR spectrum of **9a** was recorded at 220 K on a 500 MHz NMR spectrometer in CDCl₃; both signals for the β -hydrogen and the

Scheme 3.

As formation of β-oxygenated acrylamides by hydrolysis was an interesting and potentially useful transformation, the conditions were optimised using the β -morpholinoacrylamide **4d**. Use of silica gel in 1:1 ethanol/water or 0.1 M hydrochloric acid in hexane, water or toluene led to partial hydrolysis on stirring at room temperature for 16 h. However, the use of 0.1 M hydrochloric acid in acetone at room temperature resulted in complete hydrolysis of **4d** within 30 min. Hydrolysis of the β-morpholinosulfinylacrylamide **6a** and a number of β-morpholinosulfonylacrylamides (8a-8c) was also achieved under these conditions (Table 1), highlighting that the transformation can be achieved equally efficiently at the sulfide, sulfoxide and sulfone levels of oxidation. Notably, exposure of the α -sulfanyl- β -chloroacrylamides to aqueous HCl does not result in hydrolysis to form the enol under the same reaction conditions; therefore the sequential morpholine addition followed by enamine hydrolysis is necessary.

 Table 1

 Hydrolysis of β-morpholinoacrylamides

Starting material	R	R^1	n	Product	% Yield
4d	Ph	p-Tol	0	5d	86ª
6a	Bn	$4-F-C_6H_4$	1	7a	71 ^a
8a	Bn	Bn	2	9a	95ª
8b	Bn	$4-F-C_6H_4$	2	9b	72 ^a
8c	Bn	p-Tol	2	9c	54 ^b

^a Crude yield, further purification was not required.

hydroxyl proton split into clearly resolved doublets with a coupling constant of 11.0 Hz. Thus, the broadening of the signal for the β -hydrogen when the 1 H NMR spectrum is recorded at room temperature is attributed to coupling to the exchanging hydroxyl proton, and is not due to rapid E–Z interconversion.

Since the chlorination, nucleophilic substitution and hydrolysis conditions are intrinsically compatible, we decided to explore the possibility of a 'one-pot' oxidation sequence. To that end, the conditions for the transformation of the α -sulfanyl amide **10d** to the corresponding β-hydroxyacrylamide **5d** without isolation of the intermediates were subsequently developed; treatment of the sulfide 10d with 2.1 equiv of NCS in toluene under reflux for 2 h gave quantitative transformation to the β -chloroacrylamide **11d**. The oil bath was removed and the reaction mixture was allowed to cool to room temperature before 2.5 equiv morpholine was added to the stirring solution. The morpholine substitution reaction was complete within 5 min by TLC analysis and the product was cooled to 0 °C, enabling most of the succinimide and morpholine hydrochloride by-products to be removed by filtration prior to evaporation of the toluene. The sample was subsequently redissolved in acetone and 1 equiv of 0.1 M hydrochloric acid was added. Following stirring at room temperature for 5 min and extraction into dichloromethane, the β -hydroxyacrylamide **5d** was isolated. This simple, rapid procedure involving only filtration, evaporation and extraction gave a very clean product within a total reaction time of 2.5 h in 84% yield over three steps (Scheme 4). When the transformations were conducted in a stepwise manner, the yields of the oxidative

b Isolated yield after recrystallisation from dichloromethane/hexane.

β-chlorination, morpholine enamide formation and enamine hydrolysis are 80%, 67% and 86%, respectively, leading to an overall yield of 46%, highlighting the advantage of the telescoped process.

Investigation of the efficiency of the sequential oxidation sequence with a range of thio, amide and β -alkyl substituents was undertaken to establish the scope of this synthetic method. Table 2 summarises the results of these experiments.

Table 2 $\beta\textsc{-Oxidation}$ of $\alpha\textsc{-sulfanyl}$ amides—telescoped process without isolation of intermediates

$$\begin{array}{c} \text{SR}^1 \\ \text{NR}^2 \text{R}^3 \end{array} \begin{array}{c} \text{1. NCS, toluene, } \Delta, \, t_1 \\ \text{2. 2.5 eq. morpholine, rt, } t_2 \\ \text{3. 0.1 M HCl, acetone, rt} \end{array} \begin{array}{c} \text{SR}^1 \\ \text{OH O} \end{array}$$

Sulfide	R ¹	R ²	R ³	R ⁴	t ₁ ^a	t ₂ ^b	Product	% Yield ^c
					(h)	(h)		
10d	Ph	Tol	Н	Н	2	0.1	5d	84 ^d
10e	Ph	i-Pr	Н	Н	2	0.1	5e	88
10f	n-Bu	Tol	Н	Н	1.5	2	5f	60 ^e
10g	Ph	Н	Н	Н	1.5	0.1	5g	81
10a	Ph	Me	Me	Н	2	16	5a	41 ^f
10b	Ph	(S)-CH(CH ₃)Ph	Н	Et	2	23	5b	67
10h	Ph	Tol	Н	Ph	2	60 ^g	5h	<17
10i	n-Bu	Bn	Н	Н	1.5	0.1	5i	83
11j ^h	$(CH_2)_2OH$	Ph	Н	Me	—h	22	5j	24 ^e

- ^a Reaction with NCS was complete within this time.
- b Reaction with morpholine was complete within this time.
- $^{\rm c}$ Yield of β -hydroxyacrylamide based on starting sulfide.
- ^d Isolated yield following trituration using water and hexane.
- ^e Isolated yield following chromatographic purification.
- f Isolated as a mixture of the enol and keto tautomers.
- g Heating at reflux was required for reaction completion.
- ^h Compound **5j** was prepared from the β-chloroacrylamide **11j** and not the sulfide. Dichloromethane was used as the solvent in the reaction with morpholine.

As is evident from Table 2, the limiting step in the reaction sequence is nucleophilic substitution of the chloride with morpholine, with the reaction time for this step varying from 5 min for **10d** to 60 h for **10h**. In most instances, the β -hydroxyacrylamides were isolated in good yields as clean products, although purification by trituration or chromatography was necessary for **5d**, **5f** and **5j**. The reactivity of the β -carbon, with the rate of morpholine substitution and yield of β -hydroxyacrylamide highest for the primary and secondary β -chloroacrylamides. The reduced yield observed for the β -hydroxyacrylamide **5a** derived from tertiary propanamide is presumably due to conformational effects, ^{12,19} while steric effects account for the lower yields of the extended chain acrylamides **5b** and **5h**.

Furthermore, the rate of the nucleophilic addition is dependent on the stereochemistry of the β -chloroacrylamide. Thus, treatment of the tertiary propanamide **10a** with NCS led to a mixture of the E and Z isomers of the β -chloroacrylamide **11a**. Upon addition of morpholine, both the E and Z isomers reacted to give the corresponding β -morpholinoacrylamide **4a**, and subsequent hydrolysis yielded the β -hydroxyacrylamide **5a**. Reaction of the extended chain α -thioamides **10b** and **10h** with NCS also led to a mixture of the E and E E and E E E and E E or the E and E and E E and E E and E E and E E or the E sulfanyl amides E E E or the E and E E or the E or the E and E E or the E and E E or the E

The β -hydroxyacrylamides **5b**, **5d**–**5j** were isolated exclusively as their enol tautomers, while the *N*,*N*-dimethyl- β -oxopropanamide **5a** was isolated as a 3:1 mixture of the keto and enol tautomers; this

was the only instance that there was evidence for the existence of the keto tautomer. β -Keto esters and amides normally adopt the conformation that allows stabilising hydrogen bond formation between the β-OH and the carbonyl oxygen. The hydrogen bond between the amide NH and sulfur holds the primary and secondary propanamides in a rigid s-cis conformation favouring hydrogen bonding between the β-OH and the carbonyl oxygen in these β-hydroxyacrylamides. In the tertiary amides, there is no NH–S hydrogen bond thereby allowing the β-chloroacrylamide to adopt either the s-cis or s-trans conformation. 12 In the s-trans conformation, hydrogen bonding between the carbonyl oxygen and the hydrogen of the OH group is not possible and therefore this compound would exist in the keto form. However, in the s-cis conformation the tertiary β -hydroxyacrylamide **5a** could exist in the enol form (Fig. 1). Apparently, **5a** exists in both *s-cis* and *s-trans* conformations (possibly in equilibrium) as evidence for both the keto and enol forms was observed in the ¹H NMR spectrum.

Fig. 1. Keto and enol tautomers of 5a.

3. Conclusion

The transformation of secondary propanamides into the corresponding β -hydroxypropenamides in a telescoped process without isolation of the intermediates proceeds under mild conditions, without the use of metal catalysis, in good yields of 60–88% and in a very short time ($\sim\!2.5$ h) for the overall three-step process. In effect, this transformation oxidises the unactivated β -methyl group of the α -sulfanyl amide to the aldehyde level, while the products exist predominantly as the enol tautomer. In most instances, product purification is not required. This 'one-pot' oxidation is also possible for primary, tertiary and extended chain amides, albeit in lower yields than those obtained for the secondary propanamides. The synthetically powerful oxidative functionalisation of the β -carbon without affecting the sulfur centre provides scope for further reaction at this carbon, including olefination or asymmetric 1,2-addition.

4. Experimental

4.1. General

All solvents were distilled prior to use as follows: dichloromethane was distilled from phosphorous pentoxide and ethyl acetate was distilled from potassium carbonate, ethanol and methanol were distilled from magnesium in the presence of iodine. Acetone was distilled from potassium permanganate and toluene was distilled from sodium and stored over 4 Å molecular sieves. *N,N*-Dimethylformamide was stored overnight over calcium hydride, then distilled and stored over 4 Å molecular sieves. Organic phases were dried using anhydrous magnesium sulfate.

 1 H (500 MHz) and 13 C (125.8 MHz) NMR spectra were recorded on a Bruker (AV500) NMR spectrometer. 1 H (300 MHz) and 13 C (75.5 MHz) NMR spectra were recorded on a Bruker AV300 NMR spectrometer. 1 H (270 MHz) and 13 C (67.8 MHz) NMR spectra were recorded on a Jeol GSX 270 NMR spectrometer. All spectra were recorded at room temperature (\sim 20 $^{\circ}$ C) in deuterated chloroform

(CDCl₃) unless otherwise stated using tetramethylsilane (TMS) as an internal standard. Chemical shifts are expressed in parts per million (ppm) and coupling constants in Hertz (Hz).

Elemental analyses were performed by the Microanalysis Laboratory, National University of Ireland, Cork, using a Exeter Analytical 240 elemental analyzer. Melting points were carried out on a Uni-Melt Thomas Hoover Capillary melting point apparatus. Mass spectra were recorded on a Kratos Profile HV-4 double focussing high resolution mass spectrometer (EI), a Waters/Micromass LCT Premier Time of Flight spectrometer (ESI) and a Waters/Micromass Quattro Micro triple quadrupole spectrometer (ESI). Infrared spectra were recorded as potassium bromide (KBr) discs for solids or thin films on sodium chloride plates for oils on a Perkin—Elmer Paragon 1000 FT-IR spectrometer.

Thin layer chromatography (TLC) was carried out on precoated silica gel plates (Merck 60 PF₂₅₄). Column chromatography was performed using Merck silica gel 60. Visualisation was achieved by UV (254 nm) light detection, iodine staining, vanillin staining and ceric sulfate staining.

4.2. *Z*-3-Hydroxy-*N*-(4-methylphenyl)-2-(phenylsulfanyl) propenamide 5d

4.2.1. Hydrolysis of β -morpholinopropenamide. Aqueous HCl (3.1 mL, 0.1 M, 0.31 mmol) was added to a stirred solution of N-(4-methylphenyl)-3-morpholino-2-(phenylsulfanyl)propenamide 4d (0.10 g, 0.28 mmol) in acetone (4 mL). Further acetone (2-3 mL) was added to redissolve the propenamide 4d if it came out of solution. After stirring at room temperature for 5 min, the reaction was complete (by TLC analysis). CH₂Cl₂ (10 mL) was added, the phases were separated and the organic layer was washed with brine (2×5 mL), dried and evaporated to give 5d (76 mg, 86%) as a light pink, crystalline solid, which was essentially pure by ¹H NMR spectroscopy; mp 61–63 °C. Found C, 67.47; H, 5.28; N, 5.01; S, 11.20. C₁₆H₁₅NO₂S requires C, 67.34; H, 5.30; N, 4.91; S, 11.24; UV λ_{max}/nm 274, 245, 207; ϵ/l $dm^{-3} mol^{-1} cm^{-1}$ 16,180, 17,470, 25,770; ν_{max}/cm^{-1} (KBr) 3355 (br NH, OH), 1628, 1594 (CO α , β -unsaturated amide); $\delta_{\rm H}$ (270 MHz, CDCl₃) 2.29 (3H, s, ArCH₃), 7.09–7.32 (9H, m, ArH), 7.74 (1H, d, J 11, CHOH=), 8.30 (1H, br s, NH), 14.21 (1H, d, J 11, =CHOH); $\delta_{\rm C}$ (67.8 MHz, CDCl₃) 20.8 (ArCH₃), 96.4 (SC=), 120.2, 125.4, 125.8, 129.3, 129.8 (aromatic CH), 133.9, 134.8, 136.5 (aromatic C), 169.0 (CO amide), 172.0 (CHOH=); MS m/z 285 (M⁺, 32%), 196 (89), 107 (100), 91 (100).

4.2.2. One pot synthesis. NCS (0.31 g, 2.3 mmol) was added in one portion to a solution of **10d** (0.30 g, 1.11 mmol) in toluene (6 mL) and the reaction flask was submerged rapidly in an oil bath preheated to 130 °C. After 2 h the reaction was complete (by TLC analysis) and the reaction mixture was removed from the oil bath and cooled to room temperature. Morpholine (243 µL, 2.78 mmol) was added generating fumes (probably hydrogen chloride gas). TLC analysis after 5 min showed complete reaction. Filtration to remove the by-products (succinimide and morpholine hydrochloride) followed by evaporation of the toluene gave the β-morpholinopropenamide 4d (436 mg, quant.), which was dissolved in acetone (3 mL). HCl (0.1 M, 11 mL, 1.11 mmol) was added followed by acetone (3 mL) to redissolve the reactants. After stirring for 10 min at room temperature the reaction was complete (by TLC analysis) and CH₂Cl₂ (10 mL) was added. The aqueous layer was washed with CH₂Cl₂ (2×5 mL) and the combined organic layers were washed with brine $(2\times10 \text{ mL})$, dried and evaporated to give **5d** (0.30 mg)94%) as a dark pink, crystalline solid. The compound was essentially pure by ¹H NMR spectroscopy at this stage, however trituration with water/hexane (99:1) gave 5d (0.27 g, 84%) as a light pink, crystalline solid. The spectroscopic details were identical to those outlined above.

4.3. *N*-(4-Fluorophenyl)-3-hydroxy-2-(benzylsulfinyl) propenamide 7a

Agueous hydrochloric acid (8.0 mL, 0.1 M, 0.80 mmol) was added to a solution of **6a** (0.16 g. 0.40 mmol) in acetone (5 mL). Following stirring at room temperature for 15 min, TLC analysis indicated that the reaction was complete. CH₂Cl₂ (10 mL) was added to the reaction mixture, the phases were separated and the organic layer was washed with brine (2×5 mL), dried, filtered and concentrated at reduced pressure to give the product 7a (0.09 g, 71%) as a sticky yellow solid, as a single isomer; $\nu_{\text{max}}/\text{cm}^{-1}$ (KBr) 3368 (br, OH and NH), 2924 (CH), 1620 (CO), 1572, 1508, 1371 (CN stretch), 1013 (SO); δ_{H} (300 MHz, CDCl₃) 4.30 (2H, s, SCH₂), 6.99–7.07 (2H, m, ArH), 7.10 [1H, s, C(3)H=], 7.16–7.39 (5H, m, ArH), 7.42-7.49 (2H, m, ArH), 10.14 (1H, br s, NH), 13.52 (1H, br s, OH); δ_c (75.5 MHz, CDCl₃) 60.9 (CH₂, SCH₂), 104.8 [C, C(2)S], 115.7 [CH, d, ${}^{2}J_{CF}$ 23, aromatic C(3')H], 122.5 [CH, d, ${}^{3}J_{CF}$ 8, aromatic C(2')H], 128.88, 128.93, 129.1 (3×CH, 3×aromatic CH), 132.5, 132.6 (2×C, $2 \times \text{aromatic } C$), 159.8 [C, d, ${}^{1}J_{CF}$ 245, aromatic C(4')], 165.7 [CH, C(3)H=], 167.0 (C, CO); HRMS (ES+): exact mass calculated for C₁₆H₁₅NO₃SF [M+H]⁺ 320.0757. Found 320.0764; m/z (ES⁻) 318.0 $\{[(C_{16}H_{14}NO_3SF)-H^-], 100\%\}.$

4.4. N-Benzyl-Z-3-hydroxy-2-(benzylsulfonyl)propenamide

The title compound was prepared as described for 7a using 8a (0.14 g, 0.36 mmol) in acetone (10 mL) and hydrochloric acid (7.1 mL, 0.1 M, 0.71 mmol). Following stirring at room temperature for 60 min, TLC analysis indicated that the reaction had gone to completion. Dichloromethane (15 mL) was added to the reaction mixture, the phases were separated and the organic layer was washed with brine (2×10 mL), dried, filtered and concentrated at reduced pressure to give 9a (0.11 g, 95%) as an off-white solid, mp 113–114 °C; $\nu_{\text{max}}/\text{cm}^{-1}$ (KBr) 3434 (OH), 3350 (NH), 2923 (CH), 1619 (CO), 1552 (NH bend), 1453 (CN stretch), 1360 (asymmetric SO₂ stretch), 1148 (symmetric SO₂ stretch); $\delta_{\rm H}$ (400 MHz, CDCl₃) 4.24 (2H, s, SCH₂), 4.28 (2H, d, J 6.0, NHCH₂), 7.14-7.20 (2H, m, ArH), 7.23–7.41 (8H, m, ArH), 7.55 (1H, br s, NH), 7.88 [1H, br s, C(3)H=1], 15.70 (1H, br s, OH); $\delta_{\rm H}$ (500 MHz, CDCl₃, 220 K) 4.23 (2H, d, J 4.7, SCH₂), 4.32 (2H, s, NHCH₂), 7.19 (2H, d, J 7.2, ArH), 7.30-7.52 (9H, m, NH and ArH), 7.93 [1H, d, J 11.0, C(3)H=], 15.88 (1H, d, J 11.0, OH); δ_c (75.5 MHz, CDCl₃) 43.2 (CH₂, NHCH₂), 63.9 (CH₂, SCH₂), 105.1 [C, C(2)S], 127.6, 127.9 (2×CH, 2×aromatic CH), 128.0 (C, aromatic C), 128.9, 129.0, 129.3, 130.9 (4×CH, 4×aromatic CH), 136.5 (C, aromatic C), 167.2 (C, CO), 176.1 [CH, C(3)H=]; HRMS (ES⁺): exact mass calculated for C₁₇H₁₈NO₄S [M+H]⁺ 332.0957. Found 332.0947; *m*/*z* (ES^+) 332.0 {[($C_{17}H_{18}NO_3S$)+ H^+], 48%}.

4.5. *N*-(4-Fluorophenyl)-3-hydroxy-2-(benzylsulfonyl) propenamide 9b

This was synthesised as outlined for **7a** using **8b** (0.10 g, 0.3 mmol) in acetone (5 mL) and hydrochloric acid (10.0 mL, 0.1 M, 10.0 mmol). TLC analysis showed the reaction to be complete after 40 min and following the work-up, **9b** (0.06 g, 72%) was obtained as a white solid and as a single isomer, mp 113–114 °C; $\nu_{\rm max}/{\rm cm}^{-1}$ (KBr) 3429 (OH), 3320 (NH), 2923 (CH), 1618 (CO), 1567 (NH bend), 1507, 1367 (asymmetric SO₂ stretch), 1143 (symmetric SO₂ stretch); $\delta_{\rm H}$ (400 MHz, CDCl₃) 4.34 (2H, s, SCH₂), 6.95–7.03 (2H, m, Ar*H*), 7.18–7.36 (7H, m, Ar*H*), 7.99 [1H, br s, C(3)*H*=], 8.91 (1H, br s, N*H*), 15.16 (1H, br s, O*H*); $\delta_{\rm C}$ (75.5 MHz, CDCl₃) 64.4 (CH₂, SCH₂), 106.1 [C, C(2)S], 115.7 [CH, d, $^2J_{\rm CF}$ 23, aromatic C(3')H], 122.9 [CH, d, $^3J_{\rm CF}$ 8,

aromatic C(2')H], 127.8 (C, aromatic C), 129.1, 129.5, 130.9 (3×CH, 3×aromatic CH), 131.6 (C, aromatic C), 160.1 [C, d, $^1J_{CF}$ 245, aromatic C(4')], 165.3 (C, CO), 175.9 [CH, C(3)H=]; HRMS (ES $^-$): exact mass calculated for $C_{16}H_{13}NO_4SF$ [M $^-$ H] $^-$ 334.0549. Found 334.0560; m/z (ES $^-$) 334.0 {[($C_{16}H_{13}NO_4SF$) $^-$ H $^-$], 100%}.

4.6. 3-Hydroxy-*N*-(4-methylphenyl)-2-(benzylsulfonyl) propenamide 9c

The title compound was prepared as described for 7a using 8c (0.09 g, 0.2 mmol) in acetone (5 mL) and hydrochloric acid (9.0 mL, 0.1 M, 0.9 mmol). Following stirring for 30 min, TLC analysis indicated that the reaction had gone to completion and the crude product **9c** was obtained as an off-white solid after the work-up. After recrystallisation from dichloromethane/hexane, 9c was isolated as a white solid (0.04 g, 54%), mp 126–127 °C; $\nu_{\text{max}}/\text{cm}^{-1}$ (KBr) 3439 (OH), 3314 (NH), 2984 (CH), 1630 (CO), 1607, 1558 (NH bend), 1509, 1408 (CN stretch), 1324 (asymmetric SO₂ stretch), 1125 (symmetric SO₂ stretch); $\delta_{\rm H}$ (400 MHz, CDCl₃) 2.33 (3H, s, ArCH₃), 4.33 (2H, s, SCH₂), 7.11 (2H, d, J 8.4, ArH), 7.16 (2H, d, J 8.4, ArH), 7.22–7.40 (5H, m, ArH), 7.94 [1H, s, C(3)H=], 8.92 (1H, br s, NH), 15.36 (1H, br s, OH); δ_c (75.5 MHz, CDCl₃) 20.9 (CH₃, ArCH₃), 64.2 (CH₂, SCH₂), 105.9 [C, C(2)S], 121.2 (CH, aromatic CH), 127.8 (C, aromatic C), 129.1, 129.5 (signal for $2 \times CH$), 130.9 ($3 \times CH$, $3 \times aromatic$ CH), 133.0, 135.5 (2×C, 2×aromatic C), 165.3 (C, CO), 176.0 [CH, C(3) H=]; HRMS (ES⁺): exact mass calculated for $C_{17}H_{18}NO_4S$ [M+H]⁺ 332.0957. Found 332.0962; m/z (ES⁻) 330.1 {[($C_{17}H_{17}NO_3S$)+ H^+],

A second set of signals (\sim 7%) were also present in the 1 H NMR spectrum and were tentatively assigned to the stereoisomer: δ_{H} (400 MHz, CDCl₃) 4.37 (2H, s), 7.88 (1H, br d), 9.45 (1H, br s).

4.7. 3-Hydroxy-N-i-propyl-2-(phenylsulfanyl)propenamide 5e

This was prepared following the procedure described for **5d** using **10e** (0.30 g, 1.35 mmol), NCS (0.38 g, 2.83 mmol) and toluene (6 mL) with a reaction time of 1.5 h, followed by morpholine (0.29 mL, 3.36 mmol) with a reaction time of 5 min. Hydrolysis using acetone (4 mL) and HCl (14 mL, 0.1 M, 1.4 mmol) gave **5e** (0.19 g, 60%) as a pink oil. The propenamide was judged to be analytically pure; $\nu_{\text{max}}/\text{cm}^{-1}$ (film) 3380 (br NH, OH), 1613 (CO α,β-unsaturated amide); δ_{H} (270 MHz, CDCl₃) 1.06 [6H, d, J 7, NHCH(CH₃)₂], 4.07 (1H, symm m, J 7, NCH), 6.31–6.41 (1H, br m, NH), 7.13–7.31 (5H, m, ArH), 7.65 (1H, d, J 7, CHOH=), 14.52 (1H, br d, CHOH=); δ_{C} (67.8 MHz, CDCl₃) 22.9 [NHCH(CH₃)₂], 41.8 (NCH), 96.4 (SC=), 125.7, 126.4, 129.3 (aromatic CH), 137.6 (quaternary aromatic C), 170.2, 171.6 (CO amide and CHOH=); MS m/z 237 (M⁺, 78%), 208 (30, M⁺–CHO), 178 (68), 120 (100), 105 (49). Found (HRMS, EI) M⁺ 237.08249. C₁₂H₁₅NO₂S requires m/z 237.08235.

4.8. 3-Hydroxy-*N*-(4-methylphenyl)-2-(*n*-butylsulfanyl) propenamide 5f

This was prepared following the procedure described for **5d** using **10f** (0.25 g, 1.0 mmol), NCS (0.28 mg, 2.09 mmol) and toluene (5 mL) with a reaction time of 2 h, followed by morpholine (0.22 mL, 2.49 mmol) with a reaction time of 2 h. Hydrolysis using acetone (5 mL) and HCl (0.1 M, 10 mL, 1 mmol) gave crude 3-hydroxypropenamide **5f** (0.22 mg, 82%) as a yellow oil. Purification by chromatography using ethyl acetate/hexane (4:96) as eluent gave **5f** (0.16 g, 60%) as a light pink oil. Found C, 63.42; H, 7.31; N, 5.42; S, 12.17. $C_{14}H_{19}NO_2S$ requires C, 63.37; H, 7.22; N, 5.28; S, 12.09; $\nu_{\text{max}}/\text{cm}^{-1}$ (film) 3330 (br NH, OH), 1625, 1595 (CO amide); δ_{H} (270 MHz, CDCl₃) 0.91 [3H, t, *J* 7, C(4')*H*₃], 1.34–1.45 [2H, m, C(3') *H*₂], 1.47–1.62 [2H, m, C(2')*H*₂], 2.33 (3H, s, ArC*H*₃), 2.52 (2H, t, *J* 7, SC*H*₂), 7.15–7.45 (4H, ABq, *J* 8, ArH), 7.60 (1H, d, *J* 12, CHOH=), 8.70

(1H, br s, N*H*), 13.81 (1H, d, *J* 12, CHO*H*=); δ_C (67.8 MHz, CDCl₃) 13.6 [*C*(4′)H₃], 20.9 (Ar*C*H₃), 21.7 [*C*(3′)H₂], 31.1 [*C*(2′)H₂], 36.9 [*C*(1′)H₂], 98.7 (S*C*=), 120.4, 129.6 (aromatic *C*H), 134.3, 134.7 (aromatic *C*), 169.5 (CO amide), 170.1 (CHOH=); MS m/z 265 (M⁺, 18%), 107 (100), 91 (8, [Tol]⁺).

4.9. Z-3-Hydroxy-2-(phenylsulfanyl)propenamide 5g

This was prepared following the procedure described for **5d** using **10g** (0.30 g, 1.66 mmol), NCS (0.47 g, 3.5 mmol) and toluene (6 mL) with a reaction time of 2 h, followed by morpholine (0.36 mL, 4.15 mmol) with a reaction time of 10 min. Hydrolysis using acetone (4 mL) and HCl (1 M, 2 mL, 2 mmol) gave **5g** (0.21 g, 81%) as a light pink, crystalline solid; mp 58–60 °C. Found C 55.65; H, 4.84; N, 7.33; S, 16.08. C₉H₉NO₂S requires C, 55.37; H, 4.65; N, 7.17; S, 16.42; $\nu_{\text{max}}/\text{cm}^{-1}$ (KBr) 3421 (br NH, OH), 1654, 1604 (CO α , β -unsaturated amide); δ_{H} (270 MHz, CDCl₃) 6.11 (1H, br s, NH), 6.48 (1H, br s, NH), 7.14–7.48 (5H, m, ArH), 7.70 (1H, br s, CH=), (1H, br d, COH=); δ_{C} (67.8 MHz, CDCl₃) 95.2 (SC=), 124.7, 125.7, 129.4 (aromatic CH), 136.7 (aromatic C), 172.6 (aromatic CH), 173.7 (CO); MS m/z 195 (M⁺, 80%), 178 (71, M⁺–OH), 121 (100, [PhS=C]⁺).

4.10. N,N-Dimethyl-3-oxo-2-(phenylsulfanyl)propanamide 5a

Note: This compound is judged to be a mixture of keto and enol tautomers.

This was prepared following the procedure described for 5d using 10a (0.20 g, 1.0 mmol), NCS (0.28 mg, 2.06 mmol) and toluene (4 mL) with a reaction time of 2 h, followed by morpholine (0.22 mL, 2.5 mmol) with a reaction time of 16 h. Hydrolysis using acetone (6 mL) and HCl (10 mL, 0.1 M, 1 mmol) gave a mixture of products. Purification by chromatography using ethyl acetate/hexane (25:75) as eluent gave N,N-dimethyl-3-oxo-2-(phenylthio) propanamide 5a (0.90 g, 41%) as a colourless oil, which is unstable at room temperature. The estimated ratio of keto to enol tautomeric forms is 3:1; $\nu_{\text{max}}/\text{cm}^{-1}$ (film) 3272 (br NH), 1721 (CO aldehyde), 1645 (CO amide); $\delta_{\rm H}$ (270 MHz, CDCl₃) 3.00 (ArCH₃), 3.13, 3.18 [6H, $3\times$ s, N(CH₃)₂ enol/keto forms], 4.33 (<1H, d, I 5, CHS keto form), 7.13–7.57 (5H, m, ArH), 7.71 (<1H, s, CHOH=enol form), 9.62 (<1H, d, J 5, CHO keto form); δ_{C} (67.8 MHz, CDCl₃) 35.8, 37.2, 38.2 [N(CH₃)₂], 57.6 (CHS), 129.2, 129.3, 133.3 (aromatic CH), 136.0 (aromatic C), 165.2, 175.7, 192.2 (CO amide, C-3 enol/keto forms); MS m/z 223 (M⁺, 5%), 194 (8, M⁺–CHO), 72 (100, [CON(CH₃)₂]⁺).

4.11. *Z*-3-Hydroxy-(1/*S*)-*N*-1/-phenylethyl-2-(phenylsulfanyl)-2-pentenamide 5b

This was prepared following the procedure described for 5d using **10b** (0.20 g, 0.64 mmol), NCS (0.18 g, 1.34 mmol) and toluene (4 mL) with a reaction time of 2 h, followed by morpholine (0.14 mL, 1.6 mmol) with a reaction time of 23 h. Acetone (6 mL) and HCl (0.1 M, 6.5 mL, 0.65 mmol) were used for hydrolysis giving a crude reaction mixture (190 mg). Purification by chromatography using ethyl acetate/hexane (10:90) as eluent gave **5b** (95 mg, 67%) (R_f 0.6 using ethyl acetate/hexane (25:75) as eluent) as a colourless oil; $[\alpha]_D^{20}$ 13.48 (*c* 7, ethanol); $\nu_{\text{max}}/\text{cm}^{-1}$ (film) 3380 (br NH, OH), 1581, 1518 (CO α,β-unsaturated amide); δ_H (270 MHz, CDCl₃) 1.09 $[3H, t, J, 8, C(5)H_3], 1.35 [3H, d, J, 8, C(2')H_3], 2.64 [1H, q, J, 8, C(4)H_2],$ 5.00-5.13 [1H, dq, J 8, 8, C(1')H], 7.09-7.30 (11H, m, ArH, NH), enolic OH seen at δ >10 ppm; δ_C (67.8 MHz, CDCl₃) 11.0 [C(5)H₃], 22.2 $[C(2')H_3]$, 27.2 $[C(4)H_2]$, 48.9 [C(1')H], 90.0 (SC=), 125.3, 125.8, 126.1, 127.2, 128.6, 129.2 (aromatic CH), 137.2, 143.1 (aromatic C), 171.3 (CO amide), 187.5 (COH=); MS m/z 327 (M $^+$, 100%), 271 (5), 120 (15, [NHCHCH₃Ph]⁺), 105 (28, [CHCH₃Ph]⁺). Found (HRMS, EI) M⁺ 327.13609. C₁₉H₂₁NO₂S requires *m*/*z* 327.12930.

4.12. 3-Hydroxy-*N*-(4-methylphenyl)-3-phenyl-2-(phenylsulfanyl)propenamide 5h

This was prepared following the procedure described for **5d** using **10h** (0.30 g, 0.86 mmol), NCS (0.24 g, 1.76 mmol) and toluene (6 mL) with a reaction time of 2 h, followed by morpholine (0.19 mL, 2.15 mmol) with a reaction time of 60 h (by TLC analysis), including heating at reflux for 1 h. Hydrolysis using acetone (7 mL) and HCl (0.1 M, 9 mL, 0.9 mmol) in a reaction time of 24 h gave a crude reaction mixture (198 mg) as an oil. Purification by chromatography using ethyl acetate/hexane (5:95) as eluent gave **5h**; $\nu_{\text{max}}/\text{cm}^{-1}$ (film) 3339 (br NH, OH), 1678 (CO α , β -unsaturated amide), δ_{H} (270 MHz, CDCl₃) 2.31 (3H, s, ArCH₃), 7.09–7.62 (14H, m, ArH), 8.85 (1H, br s, NH), the signal for the enolic proton was seen at δ_{H} >11.

4.13. N-Benzyl-3-hydroxy-2-(n-butylsulfanyl)propenamide 5i

This was prepared following the procedure described for **5d** using **10i** (0.28 g, 1.12 mmol), NCS (0.31 g, 2.34 mmol) and toluene (6 mL) with a reaction time of 1.5 h, followed by morpholine (0.25 mL, 2.8 mmol), with a reaction time of 5 min. Hydrolysis using acetone (6 mL) and HCl (0.1 M, 11 mL, 1.1 mmol) gave **5i** (0.21 g, 83%), as a pink oil; $\nu_{\text{max}}/\text{cm}^{-1}$ (film) 3369 (br NH, OH), 1618 (CO α,β-unsaturated amide); δ_{H} (270 MHz, CDCl₃) 0.86 [3H, t, *J* 7, C(4')H₃], 1.27–1.38 [2H, m, C(3')H₂],1.39–1.65 [2H, m, C(2')H₂], 2.39–2.45 [2H, m, C(1')H₂], 4.51 (2H, d, *J* 6, CH₂Ph), 7.25–7.35 (5H, m, Ar*H*), 7.52 (1H, d, *J* 10, CH=), 11.30 (1H, d, *J* 11, COH=); δ_{C} (67.8 MHz, CDCl₃) 14.0 [C(4')H₃], 22.0 [C(3')H₂], 31.6 [C(2')H₂], 36.9 [C(1')H₂], 44.2 (CH₂Ph), 98.6 (SC=), 128.1, 128.8 (aromatic CH, 2 signals for 3 carbons), 138.1 (aromatic *C*), 169.8 (CH=), 171.5 (CO); MS m/z 265 (M⁺, 20%), 237 (22, M⁺–CO), 208 (12), 178 (18). Found (HRMS, EI), M⁺ 265.11326. C₁₄H₁₉NO₂S requires m/z 265.11365.

4.14. *N*-Phenyl-2-[2'-(hydroxyethyl)sulfanyl]-3-hydroxy-2-butenamide 5j

This was prepared following the procedure described for **5d** using **11j**-*Z* (85 mg, 0.31 mmol), morpholine (68 μ L, 0.78 mmol) and CH₂Cl₂ (2 mL) with a reaction time of 22 h. Acetone (4 mL) and HCl (0.1 M, 3 mL, 3 mmol) were used for the hydrolysis giving a crude mixture (50 mg). Purification by preparative thin layer chromatography using ethyl acetate/DCM/hexane (25:5:70) gave **5j**

(16 mg, 24%) as a red oil; $\nu_{\text{max}}/\text{cm}^{-1}$ (film) 3314 (br NH, OH), 1682, 1597 (CO α,β-unsaturated amide); δ_{H} (270 MHz, CDCl₃) 2.39 [3H, s, C(4)H₃], 2.74 (2H, dd, J 6, 6, CH₂S), 3.80 (2H, dd, J 6, 6, CH₂O), 7.13–7.59 (5H, m, Ar*H*), 9.35 (1H, br s, N*H*). The enolic OH was seen in one sample at >10 ppm; MS m/z 253 (M⁺, 17%), 209 (9), 107(48), 93 (100, [NH₂Ph]⁺).

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References and notes

- Hudlicky, M. Oxidations in Organic Chemistry; American Chemical Society: Washington, DC, 1990.
- 2. Thomas, J. M.; Raja, R.; Sankar, G.; Bell, R. G. Acc. Chem. Res. 2001, 34, 191–200.
- Mizuno, N.; Nozaki, C.; Kiyoto, I.; Misono, M. J. Am. Chem. Soc. 1998, 120, 9267–9272.
- 4. Hayashi, T.; Kishida, A.; Mizuno, N. Chem. Commun. 2000, 381-382.
- 5. Hahn, H. G.; Nam, K. D.; Mah, H.; Lee, J. J. J. Org. Chem. 1996, 61, 3894-3896.
- Hamilton, H. W.; Tait, B. D.; Gajda, C.; Hagen, S. E.; Ferguson, D.; Lunney, E. A.; Pavlovsky, A.; Tummino, P. J. Bioorg. Med. Chem. Lett. 1996, 6, 719–724.
- 7. Kuo, E. A.; Hambleton, P. T.; Kay, D. P.; Evans, P. L.; Matharu, S. S.; Little, E.; McDowall, N.; Jones, C. B.; Hedgecock, C. J.; Yea, C. M.; Chan, A. W.; Hairsine, P. W.; Ager, I. R.; Tully, W. R.; Williamson, R. A.; Westwood, R. J. Med. Chem. 1996, 39, 4608–4621.
- Robinson, R. P.; Reiter, L. A.; Barth, W. E.; Campeta, A. M.; Cooper, K.; Cronin, B. J.; Destito, R.; Donahue, K. M.; Falkner, F. C.; Fiese, E. F.; Johnson, D. L.; Kuperman, A. V.; Liston, T. E.; Malloy, D.; Martin, J. J.; Mitchell, D. Y.; Rusek, F. W.; Shamblin, S. L.; Wright, C. F. *I. Med. Chem.* 1996, 39, 10–18.
- Haines, A. H. Methods for the Oxidation of Organic Compounds; Academic Press: London, 1985.
- Kanner, C. B.: Pandit, U. K. Tetrahedron 1982, 38, 3597-3604.
- Abram, T. S.; Boeshagen, H.; Butler, J. E.; Cuthbert, N. J.; Francis, H. P.; Gardiner, P. J.; Hartwig, W.; Kluender, H. C.; Meier, H. Bioorg. Med. Chem. Lett. 1993, 3, 1517–1522.
- Murphy, M.; Lynch, D.; Schaeffer, M.; Kissane, M.; Chopra, J.; O'Brien, E.; Ford, A.; Ferguson, G.; Maguire, A. R. Org. Biomol. Chem. 2007, 5, 1228–1241.
- Kissane, M.; Lynch, D.; Chopra, J.; Lawrence, S. E.; Maguire, A. R. Tetrahedron: Asymmetry 2008, 19, 1256–1273.
- Kissane, M.; Lawrence, S. E.; Maguire, A. R. Tetrahedron: Asymmetry 2010, 21, 871–884.
- Kissane, M.; Lawrence, S. E.; Maguire, A. R. Org. Biomol. Chem. 2010, 8, 2735–2748.
- 16. Kissane, M.; Lawrence, S. E.; Maguire, A. R. Tetrahedron 2010, 66, 4564-4572.
- Kissane, M.; Lynch, D.; Chopra, J.; Lawrence, S. E.; Maguire, A. R. Org. Biomol. Chem. 2010, 8, 5602–5613.
- Kissane, M.; Murphy, M.; O'Brien, E.; Chopra, J.; Murphy, L.; Collins, S. G.; Lawrence, S. E.; Maguire, A. R. Org. Biomol. Chem. 2011, 9, 2452–2472.
- Kissane, M.; Murphy, M.; Lynch, D.; Ford, A.; Maguire, A. R. Tetrahedron 2008, 64, 7639–7649.