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Naphthoquinone-based chalcone hybrids and derivatives: synthesis and potent activity against cancer cell lines†

Guilherme A. M. Jardim,^a Tiago T. Guimarães,^b Maria do Carmo F. R. Pinto,^c Bruno C. Cavalcanti,^d Kaio M. de Farias,^d Claudia Pessoa,^{de} Claudia C. Gatto,^f Divya K. Nair,^g Irishi N. N. Namboothiri^{*g} and Eufrânio N. da Silva Júnior^{*a}

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Novel naphthoquinone-based chalcones were prepared from the reaction between 3-bromo-nor- β -lapachone and amino-chalcones. Lapachone derivatives are also described here. All the substances were evaluated against cancer and normal cell lines and several compounds demonstrated potent antitumor activity.

1. Introduction

The population growth and aging associated with some risk factors have increased the incidence of new cases of cancer and related deaths in developed countries and in developing countries.¹ In a four-year period, the estimated number increased from 12.7 million new cancer cases with 7.6 million cancer-related deaths in 2008, to 14.1 million new cancer cases and 8.2 million cancer-related deaths in 2012. The projection for the year 2030, estimates 27 million new cancer cases with 17 million cancer-related deaths.^{2,3}

Naphthoquinone containing natural products belong to an important class of naturally occurring secondary metabolites found in the bignoniaceae family.⁴ Among these, naphthoquinoidal compounds obtained from natural products, such as lapachol and lawsone (2-hydroxy-1,4-naphthoquinone), have received considerable attention because of their antitumor potential.^{5–8} 1,4- and 1,2-naphthoquinones, and for instance, dehydro- α -lapachone, an important anti-vascular agent,⁹ and β -lapachone, a potent antitumoral compound, have also been used as prototypes for the development of new drugs.¹⁰

Generally, quinones are able to provoke apoptosis and act as topoisomerase inhibitors *via* DNA intercalation. Their toxicity can also be explained by inducing oxidative stress through reactive oxygen species (ROS) generation.¹¹ Recently, described by Bolognesi and coworkers, quinones have attracted attention due to their activity being intrinsically related by a multitarget mechanism.¹²

In recent years, lapachones were employed as key substrates for the synthesis of complex diazaazulenones,¹³ spirolactones,¹⁴ oxazoles¹⁵ and other potentially bioactive heterocyclic compounds.¹⁵ Our research group has explored the potential of quinones against cancer, mainly, *via* structural modification of the nor- β -lapachone, particularly, the C-ring and redox centre modification¹⁶ since these moieties are deeply related with the generation of ROS (Scheme 1).

Earlier, we reported the synthesis of nor- β -lapachone-based 1,2,3-triazole and 3-arylamino derivatives with potent antitumor activity *via* C-ring modification of nor- β -lapachone and molecular hybridization¹⁹ with the junction of 1,2,3-triazole groups (Scheme 1).^{17,18} Recently, we described a derivative of nor- β -lapachone coupled benzothiadiazole (Scheme 1) with potent activity against twenty cancer cell lines and low cytotoxicity against three normal cells.²⁰ These results were very promising and the mechanism of action of this substance in tumor cells is being currently investigated in our laboratories and it will be reported in due course.

Using redox centre modification, imidazoles and an oxirane derivative were obtained from nor- β -lapachone and β -lapachone (Scheme 1), with antimycobacterial²¹ and trypanocidal²² activities, indicating the importance of this approach. The C-ring modification in the α -lapachone was also accomplished and thio-derivatives with antitumor activities and 1,2,3-triazoles with leishmanicidal activity were recently reported.^{23,24}

Finally, following our program to develop new bioactive molecules, particularly, new nor- β -lapachones with activity

^aInstitute of Exact Sciences, Department of Chemistry, Federal University of Minas Gerais, CEP 31270-901, Belo Horizonte-MG, Brazil. E-mail: eufranio@ufmg.br; Fax: +55 31 34095700; Tel: +55 31 34095720

^bInstituto Nacional de Câncer, Hospital do Câncer – Unidade I – Seção de Medicina Nuclear, 20230-130, Rio de Janeiro, RJ, Brazil

^cNúcleo de Pesquisas de Produtos Naturais, UFRJ, 21944-971, Rio de Janeiro, RJ, Brazil

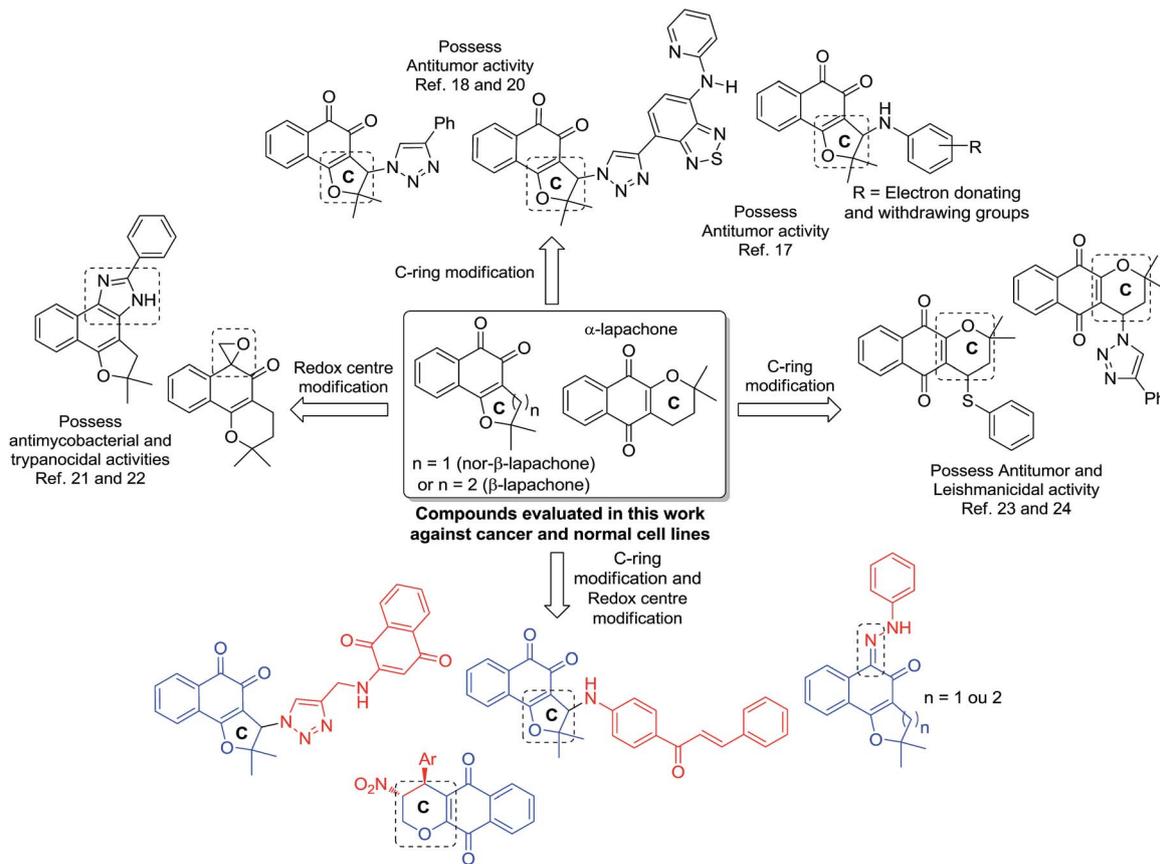
^dDepartamento de Fisiologia e Farmacologia, UFC, 60430-270, Fortaleza, CE, Brazil

^eFiocruz – Ceará, 60180-900, Fortaleza, CE, Brazil

^fInstitute of Chemistry, University of Brasília, CEP 70904970, Brasília-DF, Brazil

^gDepartment of Chemistry, Indian Institute of Technology Bombay, Mumbai 400 076, India. E-mail: irishi@iitb.ac.in; Fax: +91-22-2576-7152; Tel: +91-22-2576-7196

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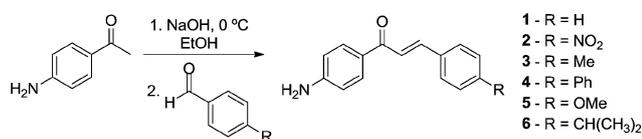
Scheme 1 Structural modification of the prototype lapachones.

against cancer cell lines and low cytotoxicity against normal cells, the synthesis of quinone-based chalcones, is described herein for the first time. The strategies of molecular hybridization and C-ring modification were used because of chalcone's known antitumor activity.²⁵ Redox centre modification in lapachones to obtain hydrazone derivatives and C-ring modification in α -lapachone in addition to nor- β -lapachone coupled with *para*-quinones were also accomplished. All the substances were evaluated against cancer and normal cell lines.

2. Results and discussion

Chemistry

To obtain the first class of compounds, nor- β -lapachone-based chalcones, initially amine-chalcones were prepared in good yields (Scheme 2). The syntheses of chalcones were easily accomplished by condensing 4'-aminoacetophenone with different aldehydes in the presence of sodium hydroxide in



Scheme 2 Synthesis of the chalcone intermediates 1–6.

ethanol. Compounds 1–6 were prepared with electron-donating (methyl, phenyl, methoxy and isopropyl groups) and an electron-withdrawing group (nitro) in addition to the unsubstituted, 1. All the substances are known in the literature.^{26–28}

Using the Hooker oxidation methodology²⁹ nor-lapachol 8 was prepared from lapachol 7 and used to obtain the classic intermediate 3-bromo-nor- β -lapachone 9 in quantitative yield. Compound 9 was reacted with the previously prepared amine-chalcones 1–6 to provide nor- β -lapachone-based chalcones 10–15 (Scheme 3).

New compounds 10–15 were well characterized by ¹H and ¹³C NMR and high resolution mass spectrometry and all the data are in accordance with the proposed structures. It was observed in the ¹H NMR spectrum of chalcone derivatives that the respective doublet signals with coupling constant, $J = 15.6$ Hz indicated the presence of the *trans*-olefin of the quinone coupled chalcone. Another important observation is a singlet at around δ 4.90–5.00 corresponding to the hydrogen attached to the C-3 in the chalcone derivatives. It is more shielded, compared to its counterpart in the bromo derivative 9, which is observed at δ 5.40. In addition, the ¹³C NMR spectra showed all the expected signals. The structure of quinone-based chalcone 14 was reconfirmed by X-ray crystallography (Fig. 1).

Recently, the second class of substances was reported by us (Scheme 4).³⁰ Lawsonia was reacted with Morita–Baylis–Hillman acetates of nitroalkenes in the presence of a quinone-

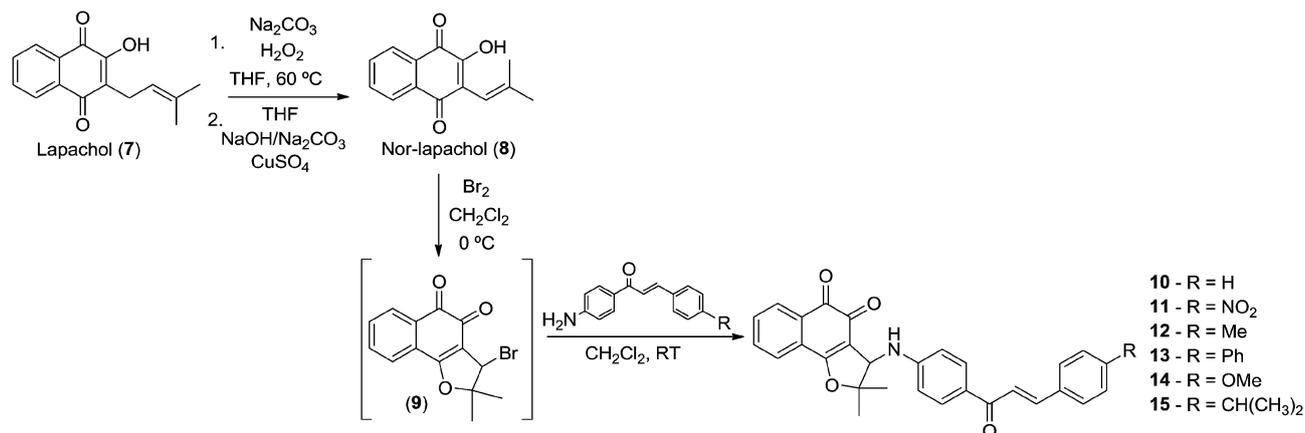
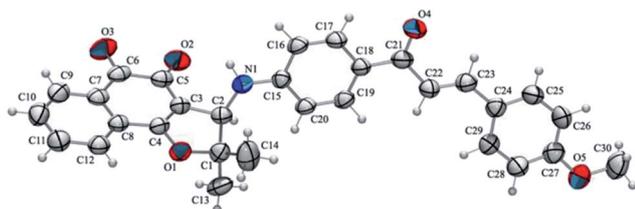
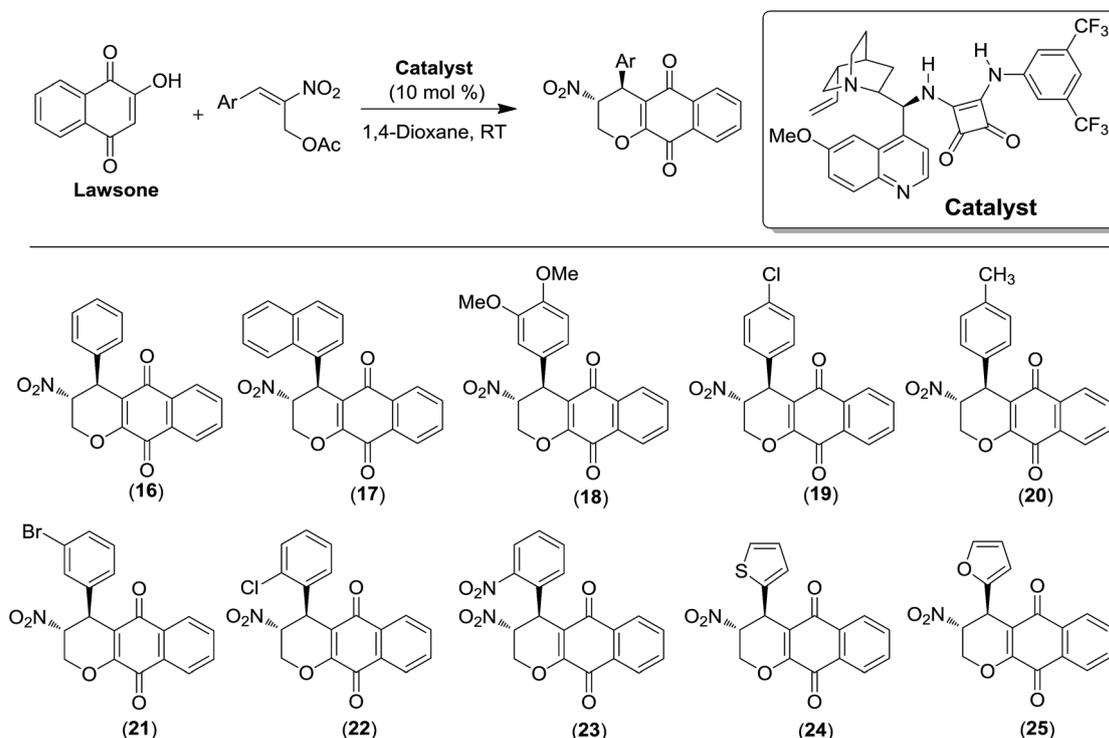
Scheme 3 Nor- β -lapachone-based chalcones obtained from lapachol 7.

Fig. 1 An ORTEP-3 representation (50% probability displacement ellipsoids) of the asymmetric unit of compound 14.

squaramide organocatalyst to provide enantioenriched disubstituted α -lapachones. This methodology allowed us to access asymmetric lapachones that are C-ring modified in a single step. In general, the products were obtained in high yields with excellent diastereo- and enantioselectivities.³⁰ The catalyst shown in Scheme 4 was the most effective.

The compounds 16–25 assayed against cancer cell lines here were previously described.³⁰

In view of the ease of access to lapachone derivatives by selective chemical transformation of lapachol 7, we invested to study the corresponding hydrazone derivatives. In addition, the hydrazones of the β -lapachone 26, 3-iodo- β -lapachone 27, 3-

Scheme 4 α -Lapachone derivatives 16–25.

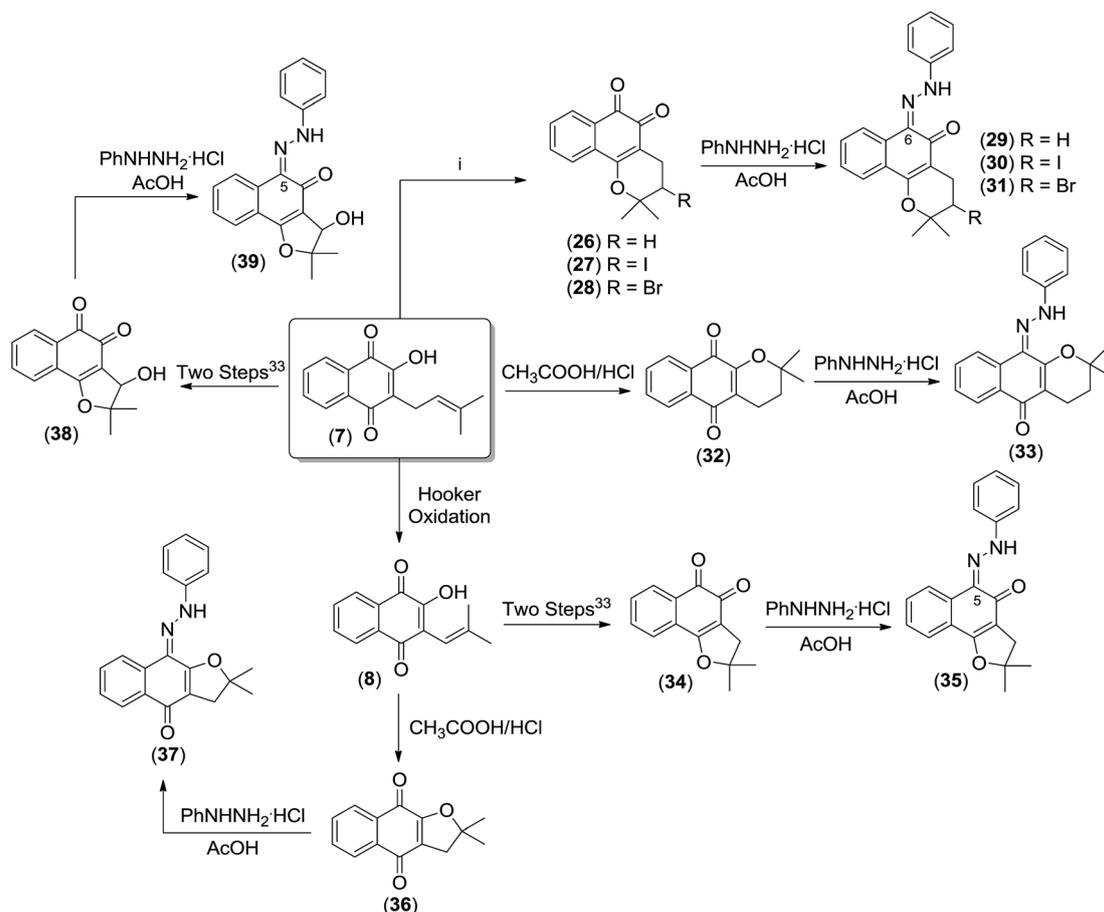
bromo- β -lapachone **28**, α -lapachone **32**, nor- β -lapachone **34**, nor- α -lapachone **36** and 3-hydroxy-nor- β -lapachone **38**, were obtained in high yields. In fact, the hydrazones of naturally occurring quinones have been considered for a long time as simple derivatives to confirm the identity of quinones. To the best of our knowledge, the use of hydrazones from quinones of the lapachol's group was not considered for major antitumor studies until now.

Initially, β -lapachone **26**, its derivatives **27** and **28** and α -lapachone **32**, were prepared from lapachol **7**, by reaction with sulfuric acid, iodine, bromine and $\text{CH}_3\text{COOH}/\text{HCl}$, respectively, as previously reported.^{31,32} Nor-lapachol **8** was used to obtain 3-hydroxy-nor- β -lapachone **38**, nor- β -lapachone **34** and nor- α -lapachone **36**.³³ The hydrazone derivatives were prepared according to the methodology previously published with minor modifications.³⁴ The reaction of the appropriate quinones with phenylhydrazine hydrochloride in acetic acid yielded the respective derivatives **29–31**, **33**, **35**, **37** and **39** in excellent yields (Scheme 5).

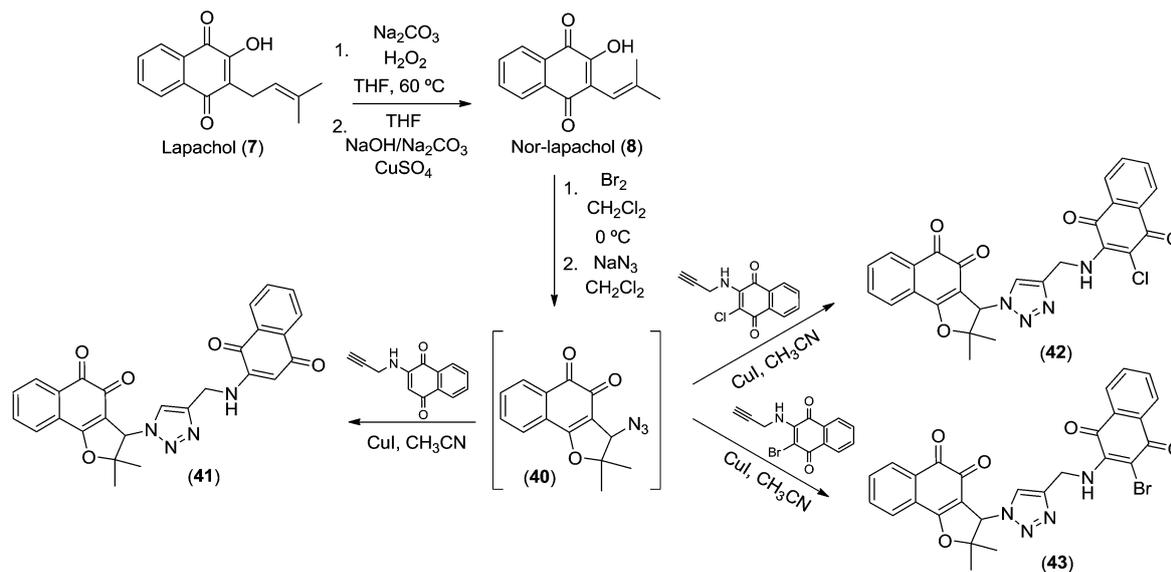
The spectroscopic data of hydrazone derivatives are in accordance with the proposed structures. The formation of these compounds followed the regioselective pattern previously observed and it is possible to conclude that the benzylic carbonyl group formed the hydrazone in all the cases.³⁴ This is

evident from ^1H NMR data that the *peri*-hydrogen of the naphthalene ring appeared at δ 7.55 to 7.14 ppm, due to the anisotropic hydrazo group at carbon C-6. In addition, the ^{13}C NMR spectra of **30**, **35** and **39**, for instance, showed similar chemical shifts for the carbonyls, at δ 179.1, 178.2 and 177.0 ppm, respectively, which is consistent with carbonyl at carbon C-6 (for **30**) and C-5 (for **35** and **39**). A common feature observed for compounds **30**, **35** and **39** in the ^1H NMR spectra using CDCl_3 as solvent is the presence of sharp singlet signals in the downfield region (δ 15.5–16.5 ppm) attributed to one N–H, which could form an internal hydrogen bond with the carbonyl of the *ortho*-hydrazone moiety, a strong H-bond acceptor.

α -Lapachone **32** gave the regioselective *para*-hydrazone **33**, which is confirmed by the presence in its ^1H NMR spectrum of an isolated downfield doublet in the aromatic region, attributed to one naphthalene *peri*-hydrogen. Thus, as in the case of the above *ortho*-hydrazones, the anisotropic effect of the hydrazine moiety on the *peri*-hydrogen of naphthalene is also observed in the case of *para*-hydrazones. On the other hand, in comparison to the original methylene hydrogens of the pyran ring in **32**, the ^1H NMR of the corresponding ones of **33**, shows only negligible effects, indicating that the hydrazo group has little influence over the pyran ring.



Scheme 5 Synthesis of the hydrazone derivatives **29–31**, **33**, **35**, **37** and **39**. (i) To obtain compound **26**: H_2SO_4 , **27**: I_2 , Py, CH_2Cl_2 , 0°C , **28**: Br_2 , CH_2Cl_2 .



Scheme 6 Lapachones 41–43 obtained from lapachol (7).

Recently, the trypanocidal activity of nor- β -lapachone-derived 1,2,3-triazole-conjugated aminoquinones was described.³⁵ These compounds present *ortho*- and *para*-carbonyl moieties that are able to generate ROS and these can be considered as important structures for evaluation against cancer cell lines because the activity of quinoidal molecules is deeply related with ROS generation as previously discussed.^{11,12} Bearing in mind the importance of these substances, we considered the compounds 41–43, obtained from lapachol 7 (Scheme 6) for antitumor studies. Nor-lapachol 8 was cyclized to 3-bromo-nor- β -lapachone that was used to prepare compound 40. The azido derivative 40 was used to synthesize 41–43 and all spectroscopy data were in accordance with the literature.³⁵

3. Biology

All the substances described (Schemes 2–5), were evaluated *in vitro* using the MTT assay against four cancer cell lines: HL-60 (human promyelocytic leukemia), OVCAR-8 (human ovarian adenocarcinoma), SF295 (human glioblastoma) and HCT-116 (human colon carcinoma). Doxorubicin was used as the positive control (Table 1).³⁶ The selectivity of the compounds toward a normal proliferating cell line was investigated using the MTT assay with human peripheral blood mononuclear cells (PBMC) after 72 h of drug exposure.

Our strategy was based on the modification of the prototypes β -lapachone 26, α -lapachone 32, nor- β -lapachone 34 and nor- α -lapachone 36. Recently, 1,2,3-triazole-, arylamino- and thio-substituted α -lapachones were synthesized and evaluated against eleven cancer cell lines.^{24,35,36} Most of them were considered moderately active and several compounds were considered highly active ($\text{IC}_{50} < 2 \mu\text{M}$). Following this previous work, herein, the antitumor activities of asymmetric α -lapachone C-ring modified 16–25 were evaluated.

The selected compounds have different patterns of substitution with an aryl ring substituted by electron withdrawing (Cl and NO_2) and donating (OMe and CH_3) groups in different positions, in addition to furan and thiophene rings. Unfortunately, in this study our strategy has failed and none of the compounds presented antitumor activity with IC_{50} values $> 12.1 \mu\text{M}$ (Table 1). At present, several modifications of the asymmetric lapachones, for instance the insertion of 1,2,3-triazoles, are being currently investigated and will be reported in due course.

Recently, redox centre modification in quinones was successfully employed to obtain antitumor compounds.¹⁶ Imine derivatives obtained from β -lapachone 26 presented potent antitumor activity.³⁷ In this context, we conducted the synthesis of hydrazone derivatives from β -lapachone 26, their iodine and bromine derivatives 27 and 28, α -lapachone 32, nor- β -lapachone 34, nor- α -lapachone 36 and 3-hydroxy-nor- β -lapachone 38, all the substances from the lapachol group. The derivatives 29–31, 33, 35, 37 and 39 were evaluated against four cancer cell lines. In the same manner as the asymmetric quinones, the hydrazones were not active against all the lineages evaluated (Table 1). In comparison of these structures with the imines, previously obtained from β -lapachone 26,³⁷ the simple insertion of the NH-bond was enough to provoke the suppression of the activity. As suggested by Burton and coworkers,³⁷ the change in the redox centre does not affect the overall activity and selectivity of the lapachone imine derivatives and it was described as an alternative in the search for new anticancer agents considering the high cytotoxicity of the β -lapachone 26 in normal cell lines. On the basis of this principle, we considered the redox centre modification of lapachone derivatives as an important strategy to obtain novel antitumor drugs despite the lack of activity of the compounds described herein.

The third group of compounds was strategically planned by C-ring modification¹⁶ of nor- β -lapachone 34 and molecular hybridization¹⁹ with chalcone moieties (Scheme 7).

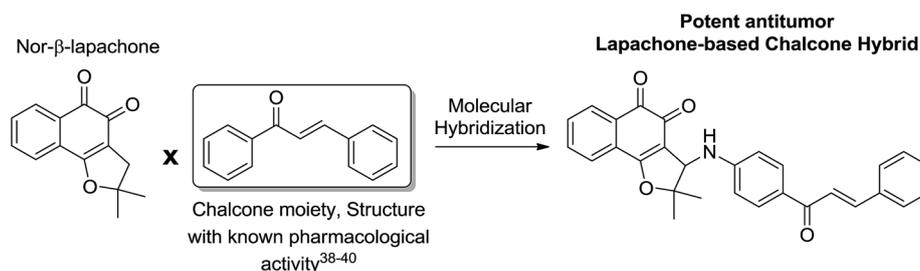
Table 1 Cytotoxic activities expressed by IC₅₀ μM (95% CI) for compounds 1–6, 10–25, 29–31, 33, 35, 37, 39 and 41–43 and for the positive control doxorubicin and the precursors lapachone^a

Compounds	HL-60	HCT-116	OVCAR-8	SF295	PBMC
1	6.40 (5.24–7.84)	14.1 (12.86–15.59)	>22.4	>22.4	20.2 (19.54–22.00)
2	1.75 (0.89–3.46)	10.96 (9.99–12.01)	>18.65	>18.65	13.83 (13.54–14.54)
3	0.46 (0.08–1.94)	15.35 (12.73–18.47)	>21.08	>21.08	>21.08
4	>16.71	>16.71	>16.71	>16.71	>16.71
5	10.50 (8.53–12.99)	>19.75	>19.75	>19.75	>19.75
6	10.0 (12.52–17.53)	>18.85	>18.85	>18.85	>18.85
10	0.04 (0.02–0.04)	0.71 (0.62–0.82)	1.53 (1.33–1.75)	0.37 (0.26–0.53)	2.20 (1.82–2.64)
11	0.20 (0.16–0.26)	2.22 (1.94–2.57)	4.35 (3.78–4.81)	2.89 (2.50–3.39)	6.51 (5.80–6.88)
12	0.10 (0.04–0.21)	1.94 (1.77–2.15)	2.13 (1.81–2.52)	4.57 (4.14–5.05)	7.90 (7.66–8.26)
13	2.20 (1.84–2.60)	>9.5	>9.5	>9.5	>9.5
14	0.10 (0.08–0.12)	1.54 (0.91–1.89)	0.64 (0.33–1.08)	1.08 (0.56–1.46)	1.33 (0.85–1.73)
15	0.14 (0.10–0.18)	1.56 (1.18–2.23)	2.54 (1.99–2.91)	1.62 (1.30–2.13)	2.28 (1.77–2.64)
16	>14.92	>14.92	>14.92	>14.92	>14.92
17	>12.98	>12.98	>12.98	>12.98	>12.98
18	>12.65	>12.65	>12.65	>12.65	>12.65
19	>13.54	>13.54	>13.54	>13.54	>13.54
20	>14.32	>14.32	>14.32	>14.32	>14.32
21	>12.10	>12.10	>12.10	>12.10	>12.10
22	>13.54	>13.54	>13.54	>13.54	>13.54
23	>13.15	>13.15	>13.15	>13.15	>13.15
24	>14.66	>14.66	>14.66	>14.66	>14.66
25	>15.38	>15.38	>15.38	>15.38	>15.38
29	>15.05	>15.05	>15.05	>15.05	>15.05
30	>10.91	>10.91	>10.91	>10.91	>10.91
31	>12.19	>12.19	>12.19	>12.19	>12.19
33	>15.05	>15.05	>15.05	>15.05	>15.05
35	>15.71	>15.71	>15.71	>15.71	>15.71
37	>15.71	>15.71	>15.71	>15.71	>15.71
39	>14.96	>14.96	>14.96	>14.96	>14.96
41	0.45 (0.39–0.54)	0.58 (0.52–0.66)	1.33 (1.12–1.58)	1.29 (1.14–1.43)	0.27 (0.14–0.37)
42	1.76 (1.43–2.17)	1.90 (1.65–2.21)	3.32 (2.82–4.06)	1.80 (1.55–2.10)	1.10 (0.83–1.18)
43	1.75 (1.37–2.25)	2.32 (2.25–2.72)	4.36 (3.83–4.78)	2.21 (1.52–2.79)	1.00 (0.87–1.07)
Nor-β-lapachone	1.75, 1.59–1.83	1.41, 1.28–1.54	1.25, 1.07–1.43	1.58, 1.31–1.88	>21.90
β-Lapachone	1.65, 1.49–1.78	0.97, 0.81–1.04	1.16, 0.97–1.25	0.91, 0.74–1.11	>20.60
α-Lapachone	8.18, 7.55–8.92	19.11, 18.75–19.88	14.79, 14.42–18.15	18.77, 18.15–19.03	>20.66
Doxorubicin	0.03, 0.01–0.05	0.02, 0.01–0.03	0.09, 0.04–0.12	0.48, 0.37–0.52	0.37, 0.30–0.43

^a Results obtained by nonlinear regression for all assayed cell lines from three independent experiments (deviations in parenthesis).

Chalcones are recognized by their biological activities, for instance, antitumor,³⁸ leishmanicidal³⁹ and antimalarial.⁴⁰ Considering the potential of the chalcones, naphthoquinoidal compounds 10–15 were prepared and evaluated against cancer and normal cell lines (Table 1). The selectivity index was considered to establish the potential of these structures (Table 2).

For the insertion of chalcone in the naphthoquinoidal moiety, compounds 1–6 were prepared. The structures 1–3 were also evaluated and showed activity against HL-60, HCT-116 and OVCAR-8 with IC₅₀ values in the range of 0.46 to 15.35 μM. Compound 4 was not active and the structures 5 and 6 presented activity only for HL-60 with IC₅₀ values = 10.5 and 10.0 μM, respectively.



Scheme 7 Strategy used to obtain new antitumor naphthoquinoid compounds.

Table 2 Selectivity index for the most active compounds [ratio between the cytotoxicities, expressed as IC_{50} (μM), against PBMC and four cancer cell lines]. Nd = not determined

Compounds	PBMC vs. HL 60	PBMC vs. HCT-116	PBMC vs. OVCAR-8	PBMC vs. SF295
1	3.1	1.4	0.9	0.9
2	7.9	1.2	0.7	0.7
3	45.8	1.3	1.0	1.0
10	55.0	3.0	1.4	5.9
11	32.5	2.9	1.4	2.2
12	79.0	4.0	3.7	1.7
13	4.3	1.0	1.0	1.0
14	13.3	0.8	2.0	1.2
15	16.2	1.4	0.8	1.4
41	0.6	0.4	0.2	0.2
42	0.6	0.5	0.3	0.6
43	0.5	0.4	0.2	0.4
Doxorubicin	12.0	18.5	4.1	0.7

After evaluating nor- β -lapachone based chalcone **10–15**, the data confirmed the success of our approach. As previously described,⁸ the compounds were classified according to their activity as highly active ($IC_{50} < 2 \mu M$), moderately active ($2 \mu M < IC_{50} < 10 \mu M$), or inactive ($IC_{50} > 10 \mu M$). The structures **10–15** were considered highly active against HL-60 and HCT-116 with IC_{50} values between 0.04 to 2.2 μM . Compounds **3**, **10–12**, **14** and **15** were considered very promising against HL-60 after assaying these structures against peripheral blood mononuclear cells (PBMC). The selectivity index (SI), which is an excellent parameter to determine the efficiency of the compounds in cancer cells with low cytotoxicity in normal cell lines is expressed by the ratio of cytotoxicities between normal cell (PBMC) and different cancer cell lines. For compounds **3**, **10**, **11**, **12**, **14** and **15** the values of SI (considering HL-60) were 45.8, 55.0, 32.5, 79, 13.3 and 16.2 (Table 2), respectively. For doxorubicin, the drug used clinically against several types of cancer, the SI value is 12.0. In comparison with the positive control, doxorubicin, nor- β -lapachone-based chalcone **12** is more active and this structure could be considered as an important prototype for further studies. To the best of our knowledge, **12** can be considered as one of the most active substances for HL-60 cancer cell line with a high selectivity index obtained from lapachol **7** already reported.

Another important characteristic that points to the success of the strategy herein employed was the increase of the activity of nor- β -lapachone **34** after the insertion of the chalcone moieties (IC_{50} values for **34** in the range of 1.25–1.75 μM), Table 1. The hybrid compounds were also more active than the non-hybridized chalcones **1–4**.

Finally, compound **13** was not active against HCT-116, OVCAR-8 and SF295, but this molecule presented selective activity for HL-60 with IC_{50} value = 2.20 μM . In terms of selectivity index this structure was not promising, but further modifications in this prototype are under investigation in our research group to develop drugs with selectivity against a certain type of cancer. Compounds **14** and **15** were highly active

against all the cancer cell lines evaluated, but these compounds were more cytotoxic than the other derivatives of this class.

In general terms, lapachone-based chalcone hybrids presented important activity against cancer cell lines with low cytotoxicity in normal cancer cell lines. This manuscript represents the first report on this class of compounds with promising activity, and based on our experience with lapachone derivatives, the compounds herein described deserve further subsequent studies.

The last class of compounds herein evaluated has been chosen because of the intrinsic ability of these structures to generate ROS. In fact, the compounds **41–43** were considered highly active ($IC_{50} < 2 \mu M$) against all the cancer lineages evaluated with some exceptions. The main problem observed for these compounds was the high cytotoxicity observed in normal cell lines (PBMC). For instance, compound **41** was highly active against HL-60 cell lines with $IC_{50} = 0.45 \mu M$, but it presented IC_{50} value = 0.27 μM for normal cells. In general, **41–43** are more cytotoxic for normal cell than cancer cell lines.

Lately, microencapsulation aimed controlled release is an important strategy to increase the efficiency of antitumor drugs.⁴¹ Recent studies using β -lapachone were described to try to diminish the cytotoxicity of this important antitumor quinone.⁴² Considering the promising results observed for **41–43**, in four types of cancer cell lineage, studies aiming controlled delivery systems, could be an interesting possibility to solve the problem of cytotoxicity.

4. Conclusions

We evaluated thirty-two compounds against four cancer cell lines and peripheral blood mononuclear cells (PBMC). These compounds were obtained by C-ring and redox centre modifications based on the prototypes nor- β -lapachone, α -lapachone, nor- α -lapachone and β -lapachone analogues. Fourteen compounds were discovered as important substances with potent antitumor activity. In general, all the compounds were highly active ($IC_{50} < 2 \mu M$) and the substances were more active than doxorubicin, a clinically used drug against several types of cancer. Compound **12** presented good selectivity for HL-60 cell line with high selectivity index even better than doxorubicin the positive control. This compound presents a promising profile for further experimental investigations in this study.

5. Experimental section

Melting points were obtained on a Thomas Hoover apparatus and are uncorrected. Analytical grade solvents were used. Column chromatography was performed on silica gel (Acros Organics 0.035–0.070 mm, pore diameter *ca.* 6 nm). ¹H and ¹³C NMR were recorded at room temperature using a Bruker AVANCE DRX 200 and Varian Mercury 400 and 500 MHz spectrometers in the solvents indicated with TMS as an internal reference. Chemical shifts (δ) are given in ppm. Electron-impact mass spectra (70 eV) were obtained using a VG Autospec apparatus (Micromass, Manchester, UK). The main fragments were described as a relation between atomic mass units and the

charge (m/z) and the relative abundance as a percentage of the base peak intensity. Lapachol (**5**) (2-hydroxy-3-(3'-methyl-2'-butenyl)-1,4-naphthoquinone) was extracted from the heartwood of *Tabebuia* sp. (Tecoma) and purified by a series of recrystallizations in an appropriate solvent. From this quinone, nor-lapachol (2-hydroxy-3-(2'-methyl-propenyl)-1,4-naphthoquinone) was obtained by the Hooker oxidation method.²⁸

General procedure to prepare the quinone-based chalcones

An excess of bromine (2 mL) was added to a cooled solution of nor-lapachol (228 mg, 1 mmol) in 25 mL of dichloromethane. The brominated intermediate **9** was obtained as an orange solid. After the removal of excess bromine, a solution of the respective chalcone (1 mmol) in 25 mL of dichloromethane was added and stirred overnight. The reaction mixture was concentrated under reduced pressure and the residue was purified by column chromatography in silica gel by eluting with an increasing polarity gradient mixture of hexane and ethyl acetate.

(E)-3-(4-Cinnamoylphenylamino)-2,2-dimethyl-2,3-dihydronaphtho[1,2-*b*]furan-4,5-dione (10). The compound **10** was obtained as a red solid (157 mg, 0.35 mmol, 35% yield); mp 196–199 °C. ¹H NMR (500 MHz, CDCl₃) δ 7.98 (d, 1H, $J = 7.4$ Hz), 7.88 (d, 2H, $J = 8.4$ Hz), 7.69 (dt, 3H, $J = 14.8, 11.6$ Hz), 7.59 (t, 3H, $J = 7.4$ Hz), 7.49 (d, 1H, $J = 15.6$ Hz), 7.39–7.35 (m, 3H), 6.61 (d, 2H, $J = 8.7$ Hz), 4.91 (d, 1H, $J = 7.3$ Hz), 1.69 (s, 3H), 1.57 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 187.8, 180.7, 175.2, 170.0, 151.3, 142.9, 135.3, 134.6, 132.7, 131.1, 131.0, 129.9, 129.5, 128.8, 128.2, 128.0, 127.2, 125.2, 121.9, 114.5, 112.2, 96.7, 60.8, 27.4, 21.8. EI/MS (m/z) [$M + Na$]⁺: 472.1511. Calcd for [C₂₉H₂₃NO₄Na]⁺: 472.1519.

(E)-2,2-Dimethyl-3-(4-(3-(4-nitrophenyl)acryloyl)phenylamino)-2,3-dihydronaphtho[1,2-*b*]furan-4,5-dione (11). The compound **11** was obtained as a brown solid (144 mg, 0.3 mmol, 30% yield); mp 224–227 °C. ¹H NMR (400 MHz, acetone) δ 8.39–8.28 (m, 3H), 8.17 (d, 1H, $J = 9.1$ Hz), 8.15–8.01 (m, 3H), 7.92 (d, 1H, $J = 15.7$ Hz), 7.82–7.75 (m, 2H), 7.01 (d, 2H, $J = 8.5$ Hz), 6.84 (d, 2H, $J = 8.8$ Hz), 5.06 (s, 1H), 1.76 (s, 3H), 1.60 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 189.2, 183.9, 180.6, 171.1, 151.6, 148.2, 141.6, 139.8, 134.7, 132.7, 131.2, 131.2, 129.6, 128.7, 127.6, 127.1, 125.8, 125.1, 124.1, 114.3, 112.2, 96.4, 60.8, 27.4, 21.8. EI/MS (m/z) [$M + Na$]⁺: 517.1378. Calcd for [C₂₉H₂₂N₂NaO₆]⁺: 517.1370.

(E)-2,2-Dimethyl-3-(4-(3-*p*-tolylacryloyl)phenylamino)-2,3-dihydronaphtho[1,2-*b*]furan-4,5-dione (12). The compound **12** was obtained as a red solid (153 mg, 0.3 mmol, 33% yield); mp 217–220 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.14 (d, 1H, $J = 7.3$ Hz), 7.96 (d, 2H, $J = 8.6$ Hz), 7.76 (d, 1H, $J = 16.0$ Hz), 7.74–7.62 (m, 3H), 7.53 (d, 2H, $J = 8.1$ Hz), 7.50 (d, 1H, $J = 15.9$ Hz), 7.21 (d, 2H, $J = 7.8$ Hz), 6.62 (d, 2H, $J = 8.7$ Hz), 4.95 (s, 1H), 2.39 (s, 3H), 1.71 (s, 3H), 1.58 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 187.9, 180.7, 175.3, 170.1, 151.2, 143.2, 140.4, 134.6, 132.7, 132.5, 131.1, 131.0, 129.6, 128.3, 128.1, 127.2, 125.2, 120.9, 114.4, 112.1, 96.7, 60.7, 27.4, 21.8, 21.5. EI/MS (m/z) [$M + Na$]⁺: 486.1669. Calcd for [C₃₀H₂₅NNaO₄]⁺: 486.1676.

(E)-3-(4-(3-(Biphenyl-4-yl)acryloyl)phenylamino)-2,2-dimethyl-2,3-dihydronaphtho[1,2-*b*]furan-4,5-dione (13). The compound **13** was obtained as an orange solid (157 mg, 0.3 mmol, 30% yield); mp 248–252 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.12 (dd, 1H, $J = 7.4, 1.3$ Hz), 7.98 (d, 2H, $J = 8.7$ Hz), 7.82 (d, 1H, $J = 15.6$ Hz), 7.75–7.73 (m, 1H), 7.73–7.69 (m, 3H), 7.69–7.67 (m, 1H), 7.67–7.61 (m, 5H), 7.58 (d, 1H, $J = 15.6$ Hz), 7.50–7.44 (m, 2H), 7.41–7.36 (m, 1H), 6.64 (d, 2H, $J = 8.8$ Hz), 1.72 (s, 3H), 1.59 (s, 3H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ 186.4, 181.0, 175.3, 168.5, 152.8, 141.9, 141.7, 139.7, 135.5, 135.4, 134.7, 133.1, 132.9, 131.6, 131.4, 130.6, 129.7, 129.5, 127.6, 127.5, 127.2, 126.6, 125.0, 122.7, 115.5, 96.2, 59.7, 27.1, 21.8. EI/MS (m/z) [$M + Na$]⁺: 548.1836. Calcd for [C₃₅H₂₇NNaO₄]⁺: 548.1832.

(E)-3-(4-(3-(4-Methoxyphenyl)acryloyl)phenylamino)-2,2-dimethyl-2,3-dihydronaphtho[1,2-*b*]furan-4,5-dione (14). The compound **14** was obtained as a red solid (191 mg, 0.4 mmol, 40% yield); mp 230–232 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.06 (d, 1H, $J = 7.4$ Hz), 7.90 (d, 2H, $J = 8.7$ Hz), 7.74–7.59 (m, 4H), 7.54 (d, 2H, $J = 8.7$ Hz), 7.38 (d, 1H, $J = 15.6$ Hz), 6.89 (d, 2H, $J = 8.7$ Hz), 6.59 (d, 2H, $J = 8.8$ Hz), 4.91 (d, 1H, $J = 7.2$ Hz), 4.60 (d, 1H, $J = 7.2$ Hz), 3.82 (s, 3H), 1.68 (s, 3H), 1.55 (s, 3H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ 187.9, 180.7, 175.2, 170.0, 151.3, 151.2, 143.1, 134.6, 132.9, 132.7, 131.1, 131.0, 129.5, 128.4, 128.1, 127.1, 126.9, 125.1, 120.9, 114.4, 113.9, 112.1, 96.7, 60.7, 27.4, 21.7. EI/MS (m/z) [$M + Na$]⁺: 502.1624. Calcd for [C₃₀H₂₅NO₅Na]⁺: 502.1630.

(E)-3-(4-(3-(4-Isopropylphenyl)acryloyl)phenylamino)-2,2-dimethyl-2,3-dihydronaphtho[1,2-*b*]furan-4,5-dione (15). The compound **15** was obtained as an orange solid (171 mg, 0.3 mmol, 35% yield); mp 212–214 °C. ¹H NMR (400 MHz, CDCl₃) δ: 8.10 (d, 1H, $J = 7.5$ Hz), 7.94 (d, 2H, $J = 8.6$ Hz), 7.77–7.71 (m, 2H), 7.69–7.64 (m, 1H), 7.56 (d, 2H, $J = 8.1$ Hz), 2.97–2.92 (m, 1H), 7.27 (t, 3H, $J = 4.1$ Hz), 6.62 (d, 2H, $J = 8.7$ Hz), 4.94 (d, 1H, $J = 7.2$ Hz), 4.59 (d, 1H, $J = 7.2$ Hz), 2.97–2.92 (m, 1H), 1.71 (s, 3H), 1.58 (s, 3H), 1.28 (s, 3H), 1.26 (s, 3H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ: 187.9, 180.7, 175.2, 170.0, 151.3, 151.8, 143.1, 134.6, 132.9, 132.7, 131.1, 130.9, 129.5, 128.4, 128.1, 127.1, 127.0, 126.9, 125.1, 120.9, 114.4, 113.8, 112.1, 96.7, 60.7, 34.1, 27.4, 23.4, 21.7. EI/MS (m/z) [$M + Na$]⁺: 514.1979. Calcd for [C₃₂H₂₉NO₄Na]⁺: 514.1994.

General procedure to prepare the hydrazone-derivatives

Phenylhydrazine hydrochloride (288 mg, 2 mmol) was added all at once to a solution of respective quinone (1 mmol) in acetic acid (15 mL). The mixture was stirred at reflux and monitored by TLC until complete consumption of the quinone. The residue was poured into water and the precipitate was filtered and washed with plenty of water. After dried, the crude hydrazones were purified by column chromatography in silica gel by eluting with an increasing polarity gradient mixture of hexane and ethyl acetate.

3-Iodo-2,2-dimethyl-6-(2-phenylhydrazone)-3,4-dihydro-2*H*-benzo[*h*]chromen-5(6*H*)-one (30). The compound **30** was obtained as a red solid (435 mg, 0.9 mmol, 95% yield); mp 157–158 °C. ¹H NMR (200 MHz, CDCl₃) δ 16.11 (s, 1H), 8.25 (d, 1H, $J = 8.1$ Hz), 7.80 (d, 1H, $J = 7.8$ Hz), 7.43–7.22 (m, 6H), 7.08–7.01

(m, 1H), 4.29 (dd, 1H, $J = 8.9, 5.7$ Hz), 3.44 (dd, 1H, $J = 5.5, 7.3$ Hz), 3.22 (dd, 1H, $J = 8.6, 8.0$ Hz) 1.59 (s, 3H), 1.50 (s, 3H). ^{13}C NMR (50 MHz, CDCl_3) δ 178.0, 158.7, 142.6, 133.0, 129.4, 127.6, 125.9, 124.9, 123.7, 122.5, 121.6, 116.2, 110.6, 79.4, 30.9, 29.9, 27.3, 24.1. EI/MS (m/z) [$\text{M} + \text{Na}$] $^+$: 481.0371. Calcd for [$\text{C}_{21}\text{H}_{19}\text{IN}_2\text{O}_2\text{Na}$] $^+$: 481.0389.

3-Bromo-2,2-dimethyl-6-(2-phenylhydrazono)-3,4-dihydro-2H-benzo[h]chromen-5(6H)-one (31). The compound 31 was obtained as an orange solid (369 mg, 0.9 mmol, 90% yield); mp 187–189 °C. ^1H NMR (200 MHz, CDCl_3) δ 16.17 (s, 1H), 8.31 (d, 1H, $J = 7.9$ Hz), 7.87 (d, 1H, $J = 7.8$ Hz), 7.58–7.33 (m, 6H), 7.12 (m, 1H), 4.22 (dd, 1H, $J = 8.1, 5.7$ Hz), 3.29 (dd, 1H, $J = 5.5, 12.2$ Hz), 3.03 (dd, 1H, $J = 8.4, 9.4$ Hz), 1.61 (s, 3H), 1.52 (s, 3H). ^{13}C NMR (50 MHz, CDCl_3) δ 178.3, 158.5, 142.6, 133.0, 129.4, 127.7, 125.9, 124.9, 123.6, 122.6, 121.6, 116.2, 109.8, 79.4, 51.3, 28.3, 26.6, 22.4. EI/MS (m/z) [$\text{M} + \text{Na}$] $^+$: 433.0522. Calcd for [$\text{C}_{21}\text{H}_{19}\text{BrN}_2\text{O}_2\text{Na}$] $^+$: 433.0527.

2,2-Dimethyl-5-(2-phenylhydrazono)-2,3-dihydronaphtho[1,2-*b*]furan-4(5H)-one (35). The compound 35 was obtained as an orange solid (334 mg, 0.9 mmol, 95% yield); mp 186–188 °C. ^1H NMR (200 MHz, CDCl_3) δ 16.30 (s, 1H), 8.42 (d, 1H, $J = 7.8$ Hz), 7.74 (d, 1H, $J = 7.8$ Hz), 7.64–7.32 (m, 6H), 7.15 (t, 1H, $J = 7.2$ Hz), 3.02 (s, 2H), 1.60 (s, 6H). ^{13}C NMR (50 MHz, CDCl_3) δ 177.3, 165.9, 142.7, 135.0, 129.7, 129.4, 129.3, 125.9, 124.6, 123.0, 122.4, 120.4, 116.0, 113.6, 91.6, 39.5, 28.5. Mass spectra (relative intensity): 318 (M^+ , 100), 303 (10), 226 (85), 213 (27), 130 (20). Elemental analysis calcd for $\text{C}_{20}\text{H}_{18}\text{N}_2\text{O}_2$ (318.36): C = 75.45, H = 7.55, N = 8.79. Found: C = 75.52, H = 7.60, N = 8.33.

2,2-Dimethyl-9-(2-phenylhydrazono)-2,3-dihydronaphtho[2,3-*b*]furan-4(9H)-one (37). The compound 37 was obtained as an orange solid (270 mg, 0.8 mmol, 85% yield); mp 165–167 °C. ^1H NMR (200 MHz, CDCl_3) δ 10.86 (s, 1H), 8.26 (d, 1H, $J = 7.7$ Hz), 8.16 (d, 1H, $J = 7.4$ Hz), 7.51–7.28 (m, 6H), 7.04–7.01 (m, 1H), 2.92 (s, 2H), 1.61 (s, 6H). ^{13}C NMR (50 MHz, CDCl_3) δ 181.1, 157.7, 142.7, 133.8, 130.8, 130.5, 129.5, 127.2, 125.3, 123.2, 122.7, 116.2, 114.5, 93.5, 38.8, 28.7. EI/MS (m/z) [$\text{M} + \text{Na}$] $^+$: 341.1258. Calcd for [$\text{C}_{20}\text{H}_{18}\text{N}_2\text{O}_2\text{Na}$] $^+$: 341.1266.

3-Hydroxy-2,2-dimethyl-5-(2-phenylhydrazono)-2,3-dihydronaphtho[1,2-*b*]furan-4(5H)-one (39). The compound 39 was obtained as a red solid (300 mg, 0.9 mmol, 90% yield); mp 197–198 °C. ^1H NMR (200 MHz, CDCl_3) δ 16.11 (s, 1H), 8.34 (d, 1H, $J = 8.2$ Hz), 7.74 (d, 1H, $J = 7.8$ Hz), 7.58–7.32 (m, 6H), 7.15 (t, 1H, $J = 14.1$ Hz), 5.12 (s, 1H), 1.67 (s, 3H), 1.48 (s, 3H). ^{13}C NMR (50 MHz, CDCl_3) δ 178.0, 167.8, 142.7, 135.9, 130.9, 129.7, 129.5, 126.3, 125.2, 123.9, 122.8, 120.3, 116.4, 116.2, 94.7, 27.0, 21.0. EI/MS (m/z) [$\text{M} + \text{Na}$] $^+$: 357.1208. Calcd for [$\text{C}_{20}\text{H}_{18}\text{N}_2\text{O}_3\text{Na}$] $^+$: 357.1215.

Crystallographic data for compound 14

X-ray data collection was accomplished on a Bruker CCD SMART APEX II single crystal diffractometer with Mo $K\alpha$ radiation (0.71073 Å) at 296 K. The data were processed with SAINT⁴³ and were corrected for absorption using SADABS.⁴⁴ The structures were solved by direct methods using SHELXS-97 and subsequent Fourier-difference map analyses yielded the positions of the nonhydrogen atoms, the refinement was performed

using SHELXL-97.⁴⁵ Molecular graphics, ORTEP-3,⁴⁶ software was used to prepare material for publication, WinGX-Routine.^{47†}

Crystal data for compound 14

$\text{C}_{30}\text{H}_{25}\text{NO}_5$; $M = 479.51$; monoclinic, space group $C2/c$; $a = 37.446(2)$ Å, $b = 10.070(7)$ Å, $c = 13.198(10)$ Å; $\alpha = \gamma = 90^\circ$, $\beta = 101.015(5)^\circ$; $V = 4885.3(6)$ Å³; $Z = 8$; $D_c = 1.304$ g cm⁻³; $F(000) = 2016$; red prism, size $0.39 \times 0.22 \times 0.11$ mm; 4944 independent measured reflections, refinement based on F^2 to give $R_1[F^2 > 4\sigma(F^2)] = 0.055$; $w_2 = 0.106$ for 17 276 observed reflections, and 393 parameters.

Cytotoxicity against cancer and normal cell lines

Compounds (0.001–5 $\mu\text{g mL}^{-1}$) were tested for cytotoxic activity against several human cancer cell lines obtained from the National Cancer Institute, NCI (Bethesda, MD). Peripheral blood mononuclear cells (PBMC) were isolated from heparinized blood from healthy, non-smoker donors who had not taken any medication at least 15 days prior to sampling by a standard method of density-gradient centrifugation on Histopaque-1077 (Sigma Aldrich Co. – St. Louis, MO/USA). All cancer cell lines and PBMC were maintained in RPMI 1640 medium. All culture media were supplemented with 20% (PBMC) or 10% (HL-60, HCT-116, OVCAR-8 and SF295) fetal bovine serum, 2 mM L-glutamine, 100 IU mL⁻¹ penicillin, 100 $\mu\text{g mL}^{-1}$ streptomycin at 37 °C with 5% CO₂. PBMC cultures were also supplemented with 2% phytohaemagglutinin. In cytotoxicity experiments, cells were plated in 96-well plates (1 $\times 10^6$ cells per well for leukemia HL-60 cells and 1 $\times 10^6$ cells per well for solid tumors cell lines and PBMC). All the tested compounds were dissolved with DMSO. The final concentration of DMSO in the culture medium was kept constant (0.1%, v/v). Doxorubicin (0.001–1.10 μM) was used as the positive control, and negative control groups received the same amount of vehicle (DMSO). The cell viability was determined by reduction of the yellow dye 3-(4,5-dimethyl-2-thiazol)-2,5-diphenyl-2H-tetrazolium bromide (MTT) to a blue formazan product as described by Mosmann.⁴⁸ At the end of the incubation time (72 h), the plates were centrifuged and the medium was replaced by fresh medium (200 μL) containing 0.5 mg mL⁻¹ MTT. Three hours later, the MTT formazan product was dissolved in DMSO (150 μL) and the absorbance was measured using a multiplate reader (Spectra Count, Packard, Ontario, Canada). Drug effect was quantified as the percentage of control absorbance of the reduced dye at 550 nm. All cell treatments were carried out with three replicates.

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Notes and references

- B. D. Smith, G. L. Smith, A. Hurria, G. N. Hortobagyi and T. A. Buchholz, *J. Clin. Oncol.*, 2009, **27**, 2758–2765.
- P. Boyle and B. Levin, *World Cancer Report, 2008*, IARC Press, Lyon, 2008, p. 510.
- J. Ferlay, I. Soerjomataram, M. Ervik, R. Dikshit, S. Eser, C. Mathers, M. Rebelo, D. M. Parkin, D. Forman and F. Bray, *GLOBOCAN 2012 v1.0, Cancer Incidence and Mortality Worldwide: IARC CancerBase No. 11 [Internet]*, International Agency for Research on Cancer, Lyon, France, 2013, <http://globocan.iarc.fr>, accessed 10 April 2014.
- A. Reichstein, S. Vortherms, S. Bannwitz, J. Tentrop, H. Prinz and K. Müller, *J. Med. Chem.*, 2012, **55**, 7273–7284.
- E. N. da Silva Júnior, M. C. B. V. Souza, A. V. Pinto, M. C. F. R. Pinto, V. F. Ferreira, R. F. S. Menna-Barreto, R. S. F. Silva, D. V. Teixeira, C. A. de Simone and S. L. de Castro, *Eur. J. Med. Chem.*, 2008, **43**, 1774–1780.
- M. L. Bolognesi, N. Calonghi, C. Mangano, L. Masotti and C. Melchiorre, *J. Med. Chem.*, 2008, **51**, 5463–5467.
- E. N. da Silva Júnior, C. F. de Deus, B. C. Cavalcanti, C. Pessoa, L. V. Costa-Lotufo, R. C. Montenegro, M. O. de Moraes, M. C. F. R. Pinto, C. A. de Simone, V. F. Ferreira, M. O. F. Goulart, C. K. Z. Andrade and A. V. Pinto, *J. Med. Chem.*, 2010, **53**, 504–508.
- E. Pérez-Sacau, R. G. Díaz-Peñate, A. Estévez-Braun, A. G. Ravelo, J. M. Garcia-Castellano, L. Pardo and M. Campillo, *J. Med. Chem.*, 2007, **50**, 696–706.
- I. Garkavtsev, V. P. Chauhan, H. K. Wong, A. Mukhopadhyay, M. A. Glicksman, R. T. Peterson and R. K. Jaina, *Proc. Natl. Acad. Sci. U. S. A.*, 2011, **108**, 11596–11601.
- C. J. Li, Y. Z. Li, A. V. Pinto and A. B. Pardee, *Proc. Natl. Acad. Sci. U. S. A.*, 1999, **96**, 13369–13374.
- R. P. Verma, *Anti-Cancer Agents Med. Chem.*, 2006, **6**, 489–499.
- F. Prati, E. Uliassi and M. L. Bolognesi, *Med. Chem. Commun.*, 2014, **5**, 853–861.
- F. S. Emery, M. C. F. R. Pinto, C. A. de Simone, V. R. S. Malta, E. N. da Silva Júnior and A. V. Pinto, *Synlett*, 2010, **13**, 1931–1934.
- E. N. da Silva Júnior, C. A. de Simone, A. C. B. de Souza, C. N. Pinto, T. T. Guimarães, M. C. F. R. Pinto and A. V. Pinto, *Tetrahedron Lett.*, 2009, **50**, 1550–1553.
- A. V. Pinto, C. Neves Pinto, M. C. F. R. Pinto, R. M. Santa Rita, C. Pezzella and S. L. de Castro, *Arzneim.-Forsch.*, 1997, **47**, 74–79.
- S. L. de Castro, F. S. Emery and E. N. da Silva Júnior, *Eur. J. Med. Chem.*, 2013, **69**, 678–700.
- E. N. da Silva Júnior, C. F. de Deus, B. C. Cavalcanti, C. Pessoa, L. V. Costa-Lotufo, R. C. Montenegro, M. O. de Moraes, M. C. F. R. Pinto, C. A. de Simone, V. F. Ferreira, M. O. F. Goulart, C. K. Z. Andrade and A. V. Pinto, *J. Med. Chem.*, 2010, **53**, 504–508.
- E. N. da Silva Júnior, M. A. B. F. de Moura, A. V. Pinto, M. C. F. R. Pinto, M. C. B. V. de Souza, A. J. Araújo, C. Pessoa, L. V. Costa-Lotufo, R. C. Montenegro, M. O. de Moraes, V. F. Ferreira and M. O. F. Goulart, *J. Braz. Chem. Soc.*, 2009, **20**, 635–643.
- C. Viegas Júnior, A. C. Danuello, V. S. Bolzani, E. J. Barreiro and C. A. M. Fraga, *Curr. Med. Chem.*, 2007, **14**, 1829–1852.
- E. H. G. da Cruz, P. H. P. R. Carvalho, J. R. Corrêa, D. A. C. Silva, E. B. T. Diogo, J. D. de Souza Filho, B. C. Cavalcanti, C. Pessoa, H. C. B. de Oliveira, B. C. Guido, D. A. da Silva Filho, B. A. D. Neto and E. N. da Silva Júnior, *New J. Chem.*, 2014, **38**, 2569–2580.
- K. C. G. Moura, P. F. Carneiro, M. C. F. R. Pinto, J. A. da Silva, V. R. S. Malta, C. A. de Simone, G. G. Dias, G. A. M. Jardim, J. Cantos, T. S. Coelho, P. E. A. da Silva and E. N. da Silva Júnior, *Bioorg. Med. Chem.*, 2012, **20**, 6482–6488.
- V. F. Ferreira, A. Jorqueira, A. M. T. Souza, M. N. da Silva, M. C. B. V. de Souza, R. M. Gouvêa, C. R. Rodrigues, A. V. Pinto, H. C. Castro, D. O. Santos, H. P. Araújo and S. C. Bourguignon, *Bioorg. Med. Chem.*, 2006, **14**, 5459–5466.
- E. H. G. da Cruz, C. M. B. Hussene, G. G. Dias, E. B. T. Diogo, I. M. M. de Melo, B. L. Rodrigues, M. G. da Silva, W. O. Valença, C. A. Camara, R. N. de Oliveira, Y. G. de Paiva, M. O. F. Goulart, B. C. Cavalcanti, C. Pessoa and E. N. da Silva Júnior, *Bioorg. Med. Chem.*, 2014, **22**, 1608–1619.
- T. T. Guimarães, M. C. F. R. Pinto, J. S. Lanza, M. N. Melo, R. L. do Monte-Neto, I. M. M. de Melo, E. B. T. Diogo, V. F. Ferreira, C. A. Camara, W. O. Valença, R. N. de Oliveira, F. Frézard and E. N. da Silva Júnior, *Eur. J. Med. Chem.*, 2013, **63**, 523–530.
- Y. Zhang, B. Srinivasan, C. Xing and J. Lü, *Anticancer Res.*, 2012, **32**, 3689–3698.
- R. T. Blickenstaff, W. R. Hanson, S. Reddy and R. Witt, *Bioorg. Med. Chem.*, 1995, **3**, 917–922.
- V. Tomar, G. Bhattacharjee, Kamaluddin, S. Rajakumar, K. Srivastava and S. K. Puri, *Eur. J. Med. Chem.*, 2010, **45**, 745–751.
- F. L. Ansari, S. Umbreen, L. Hussain, T. Makhmoor, S. A. Nawaz, M. A. Lodhi, S. N. Khan, F. Shaheen, M. I. Choudhary and Atta-ur-Rahman, *Chem. Biodiversity*, 2005, **2**, 487–496.
- L. F. Fieser and M. Fieser, *J. Am. Chem. Soc.*, 1948, **70**, 3215.
- D. K. Nair, R. F. S. Menna-Barreto, E. N. da Silva Júnior, S. M. Mobin and I. N. N. Namboothiri, *Chem. Commun.*, 2014, **50**, 6973–6976.
- J. S. Sun, A. H. Geiser and B. Frydman, *Tetrahedron Lett.*, 1998, **39**, 8221–8224.
- C. Salas, R. A. Tapia, K. Ciudad, V. Armstrong, M. Orellana, U. Kemmerling, J. Ferreira, J. D. Maya and A. Morello, *Bioorg. Med. Chem.*, 2008, **16**, 668–674.
- A. V. Pinto, M. C. F. R. Pinto and C. G. T. de Oliveira, *An. Acad. Bras. Cienc.*, 1982, **54**, 108–114.
- C. E. M. Carvalho, V. F. Ferreira, A. V. Pinto, M. C. F. R. Pinto and W. Harrison, *Dyes Pigm.*, 2002, **52**, 209–214.
- E. B. T. Diogo, G. G. Dias, B. L. Rodrigues, T. T. Guimarães, W. O. Valença, C. A. Camara, R. N. de Oliveira, M. G. da Silva, V. F. Ferreira, Y. G. de Paiva, M. O. F. Goulart, R. F. S. Menna-Barreto, S. L. de Castro and E. N. da Silva Júnior, *Bioorg. Med. Chem.*, 2013, **21**, 6337–6348.

- 36 E. N. da Silva Júnior, M. C. B. V. de Souza, A. V. Pinto, M. C. F. R. Pinto, M. O. F. Goulart, F. W. A. Barros, C. Pessoa, L. V. Costa-Lotufo, R. C. Montenegro, M. O. de Moraes and V. F. Ferreira, *Bioorg. Med. Chem.*, 2007, **15**, 7035–7041.
- 37 P. H. Di Chenna, V. Benedetti-Doctorovich, R. F. Baggio, M. T. Garland and G. Burton, *J. Med. Chem.*, 2001, **44**, 2486–2489.
- 38 E. Winter, P. D. Neuenfeldt, L. D. Chiaradia-Delatorre, C. Gauthier, R. A. Yunes, R. J. Nunes, T. B. Creczynski-Pasa and A. D. Pietro, *J. Med. Chem.*, 2014, **57**, 2930–2941.
- 39 R. Shivahare, V. Korthikunta, H. Chandasana, M. K. Suthar, P. Agnihotri, P. Vishwakarma, T. K. Chaitanya, P. Kancharla, T. Khaliq, S. Gupta, R. S. Bhatta, J. V. Pratap, J. K. Saxena, S. Gupta and N. Tadigoppula, *J. Med. Chem.*, 2014, **57**, 3342–3357.
- 40 F. J. Smit and D. D. N'Da, *Bioorg. Med. Chem.*, 2014, **22**, 1128–1138.
- 41 E. Blanco, E. A. Bey, C. Khemtong, S. G. Yang, J. Setti-Guthi, H. Chen, C. W. Kessinger, K. A. Carnevale, W. G. Bornmann, D. A. Boothman and J. Gao, *Cancer Res.*, 2010, **70**, 3896–3904.
- 42 S. Y. Jeong, S. J. Park, S. M. Yoon, J. Jung, H. N. Woo, S. L. Yi, S. Y. Song, H. J. Park, C. Kim, J. S. Lee, J. S. Lee and E. K. Choi, *J. Controlled Release*, 2009, **139**, 239–245.
- 43 *SMART and SAINT, Area Detector Control Integration Software*, Bruker Analytical X-ray Instruments, Inc., Madison Wisconsin, USA, 1999.
- 44 G. M. Sheldrick, *SADABS, Program for Empirical Absorption Correction of Area Detector Data*, University of Göttingen, Germany, 1997.
- 45 G. M. Sheldrick, *Acta Crystallogr., Sect. A: Found. Crystallogr.*, 2008, **64**, 112.
- 46 L. J. Farrugia, *J. Appl. Crystallogr.*, 1997, **30**, 565.
- 47 L. J. Farrugia, *J. Appl. Crystallogr.*, 2012, **45**, 849–854.
- 48 T. J. Mosmann, *Immunol. Methods*, 1983, **65**, 55–63.