This article was downloaded by: [Nipissing University] On: 04 October 2014, At: 18:15 Publisher: Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Bioscience, Biotechnology, and Biochemistry

Publication details, including instructions for authors and subscription information: <u>http://www.tandfonline.com/loi/tbbb20</u>

Enantioselective Synthesis of Ancepsenolide and its Analogs

Katsuki TAKAI^a & Ryozo IRIYE^a

^a Department of Bioscience and Biotechnology, Faculty of Agriculture, Shinshu University (United Graduate School of Gifu University) Published online: 22 May 2014.

To cite this article: Katsuki TAKAI & Ryozo IRIYE (2001) Enantioselective Synthesis of Ancepsenolide and its Analogs, Bioscience, Biotechnology, and Biochemistry, 65:8, 1903-1906, DOI: <u>10.1271/bbb.65.1903</u>

To link to this article: http://dx.doi.org/10.1271/bbb.65.1903

PLEASE SCROLL DOWN FOR ARTICLE

Taylor & Francis makes every effort to ensure the accuracy of all the information (the "Content") contained in the publications on our platform. However, Taylor & Francis, our agents, and our licensors make no representations or warranties whatsoever as to the accuracy, completeness, or suitability for any purpose of the Content. Any opinions and views expressed in this publication are the opinions and views of the authors, and are not the views of or endorsed by Taylor & Francis. The accuracy of the Content should not be relied upon and should be independently verified with primary sources of information. Taylor and Francis shall not be liable for any losses, actions, claims, proceedings, demands, costs, expenses, damages, and other liabilities whatsoever or howsoever caused arising directly or indirectly in connection with, in relation to or arising out of the use of the Content.

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden. Terms & Conditions of access and use can be found at http://www.tandfonline.com/page/terms-and-conditions

Note

Enantioselective Synthesis of Ancepsenolide and its Analogs

Katsuki TAKAI and Ryozo IRIYE[†]

Department of Bioscience and Biotechnology, Faculty of Agriculture, Shinshu University (United Graduate School of Gifu University), 8304 Minamiminowa, Kamiina, Nagano 399-4598, Japan

Received February 2, 2001; Accepted March 12, 2001

Ancepsenolide (1a-s) and the enantiomer (1a-r) were respectively synthesized from (S)- and (R)-2-[(R)-O-MEM-mandeloyloxy]propanal (3a-s and 3a-r) and diisopropyl hexadecanedioate (5). The analogs (1b, 2a and 2b) were synthesized by a similar method.

Key words: ancepsenolide; aldol reaction; optical resolution; butenolide

Novel lipids with one or two butenolide moieties have been isolated from gorgonians.¹⁻⁶⁾ Although they are considered to be defense substances of these organisms,^{1,6,7)} their activity is not clear. Ancepsenolide (1a-s) is one component and has been isolated from *Pterogorgia anceps*,¹⁻³⁾ *P. citrina*⁵⁾ and *P.* guadalupensis.⁶⁾ Ancepsenolide (1a-r) has two (S)-4methyl-2-buten-4-olide moieties at both terminal positions of dodecane (see the Scheme). The presence of a butenolide moiety with (S)-configuration relates it to Annonaceous acetogenins⁷⁻¹⁰ in structural classification. Many acetogenins are known for their potent biological activities as cytotoxic, antitumorial, antimalarial, microbicidal, immunosuppressive, antifeeding and pesticidal substances, and their (S)-4methyl-2-buten-4-olide group has been reported to be important for these activities.7-11) Although many papers on the synthesis of ancepsenolide (1a-s) have been published,^{12,13)} a simple synthesis was investigated in order to elucidate the biological activity of ancepsenolide (1a-s) and related compounds. We have already reported that the γ -acyloxy- β -hydroxy carbonyl moiety obtained by aldol reaction could be transformed into a γ -hydroxy- α , β -unsaturated carbonyl moiety by treating with AcONa.¹⁴⁻¹⁶⁾ A modification of this method was used to synthesize the (R,R)- and (S,S)-forms of an epsenolide (1a) and its derivatives (1b, 2a and 2b).

The starting compound, (S)-(-)-2-[(R)-O-MEM-mandeloyloxy] propanal (**3a-s**), was prepared with sodium (*R*)-*O*-MEM-mandelate by the methods described previously.^{15,16)} Aldehyde **3a-s** was treated

with the dienolate of diisopropyl hexadecanedioate (5) to form the hydroxyester (7a-s). Hydroxyester 7a-s was transformed into 8a-s by treating with AcO-Na (44% yield), since 7a-s was unstable. 8a-s was transformed into (S, S)-ancepsenolide (1a-s) by treating with NaH,¹⁷⁾ and then by successive purification by TLC and recrystallization (24% yield). Similarly, the (R, R)-form compound (1a-r) was synthesized from 3a-r and 5 in a 13% yield.

(R,R)- and (S,S)-bis-4-pentyl-2-buten-4-olide derivatives with a dodecamethylene (**1b-r** and **1b-s**) were respectively prepared from the aldehydes (**3b-r** and **3b-s**) and **5** via **7b-r** and **7b-s**, and respectively transformed to **8b-r** and **8b-s** in 36.9% and 49.7% yields.

(R,R)- and (S,S)-bis-4-methyl-2-buten-4-olide derivatives with an octamethylene (**2a-r** and **2a-s**) were similarly synthesized from the respective aldehydes (**3a-r** and **3a-s**) and diisopropyl dodecanedioate (**6**) via **9a-r** and **9a-s** and then transformed into **10a-r** and **10a-s**.

(R, R)- and (S, S)-4-pentyl-2-buten-4-olide derivatives with an octamethylene (**2b-r** and **2b-s**) were also respectively synthesized from **3b-r** and **3b-s**, and **6** in a similar manner described above (see the Scheme).

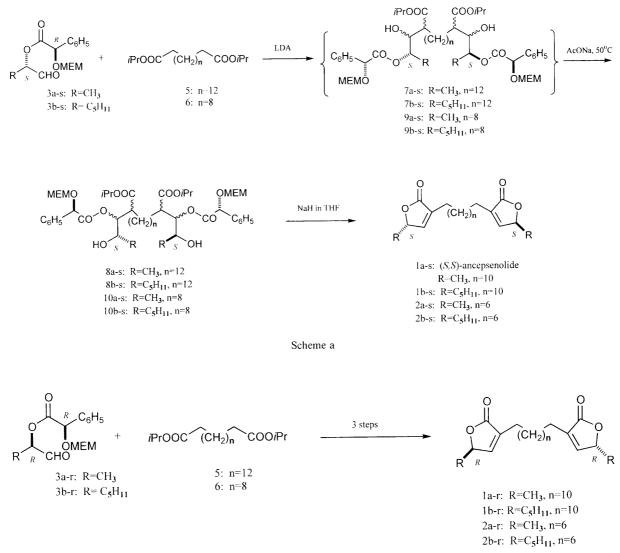
Experimental

NMR spectra were measured with a Bruker DRX-500 spectrometer (¹H at 500.13 MHz, ¹³C at 125.77 MHz) with the Bruker pulse program and TMS as an internal standard. Mass spectra were taken with a JEOL JMS-700 spectrometer using the JEOL data processing system, and optical rotation was measured with a JASCO DIP-1000 polarimeter. NMR spectral data resulting from similar structures and protective groups are not repeatedly described.

(R)-(-)- and (S)-(-)-2-[(R)-O-MEM-mandeloyloxy]alkanal (3a-r and 3a-s, and 3b-r and 3b-s). (R)-(-)- and (S)-(-)-2-[(R)-O-MEM-mandeloylox-

[†] To whom correspondence should be addressed. Tel: +81-265-77-1604; Fax: +81-265-77-1629; E-mail: kn53278@gipmc.shinshuu.ac.jp

Abbreviations: MEM, 2-methoxyethoxymethyl; AcONa, sodium acetate; DMF, N, N-dimethylformamide; HMPA, hexamethyl-phosphoric triamide





y]alkanal (**3a-r** and **3a-s**, and **3b-r** and **3b-s**) were prepared by the method described previously.^{15,16)} The mixture of **3a-r** and **3a-s** was prepared by adding an ether solution of an excess amount of 2bromopropanal to a suspension of sodium (R)-O-MEM-mandelate in DMF and HMPA.

(*R*)-(-)-2-[(*R*)-*O*-MEM-mandeloyloxy]propanal (**3a-r**): 10.3% yield, $[\alpha]_D = -52.5^\circ$, *c* 0.10 (EtOH). **3a-s**: 9.1% yield, $[\alpha]_D = -72.8^\circ$, *c* 0.10 (EtOH). **3a-r** was separated with a flash column (silica gel, hexane:EtOAc = 4:1), and **3a-s**, by preparative TLC (hexane:EtOAc = 5:3, 5 developments) from the residual mixture. **3a-r**: NMR δ_H (CDCl₃): 1.39 (3H, d, J = 7.2 Hz, CH₃), 3.37 (3H, s, MEM), 3.45–3.57 (2H, m, MEM), 3.69 (1H, ddd, J = 3.7, 5.5, 11.2 Hz, MEM), 3.81 (1H, ddd, J = 3.7, 5.5, 11.0 Hz, MEM), 4.81 (1H, d, J = 7.1 Hz, MEM), 4.90 (1H, d, J = 7.1 Hz, MEM), 5.09 (1H, q, J = 7.1 Hz, CH), 5.32 (1H, s, CH), 7.28–7.43 (3H, m, Ph), 7.49 (2H, d, J = 7.2 Hz, Ph), 9.33 (1H, s, CHO). NMR δ_C : 14.20, 58.98, 67.63, 71.64, 75.21, 94.23, 127.49, 128.77, 128.99, 135.80, 170.39, 197.94. HR-MS m/z: calcd. for C₁₅H₂₀O₆ (M⁺), 296.1260; found, 296.1243. **3a-s**: NMR $\delta_{\rm H}$ (CDCl₃): 1.31 (3H, d, J= 7.1 Hz, CH₃), 3.37 (3H, s, MEM), 3.45–3.58 (2H, m, MEM), 3.67 (1H, ddd, J=0.5, 5.6, 11.0 Hz, MEM), 3.80 (1H, ddd, J=3.8, 5.3, 11.3 Hz, MEM), 4.82 (1H, d, J=7.0 Hz, MEM), 4.89 (1H, d, J=7.0 Hz, MEM), 5.09 (1H, q, J=6.9 Hz, CH), 5.34 (1H, s, CH), 7.29–7.42 (3H, m, Ph), 7.48 (2H, d, J=6.0 Hz, Ph), 9.54 (1H, s, CHO). NMR $\delta_{\rm C}$: 14.00, 58.98, 67.67, 71.62, 75.29, 94.32, 127.43, 128.74, 128.94, 135.72, 170.39, 197.99. HR-MS m/z: calcd. for C₁₅H₂₀O₆ (M⁺), 296.1260; found, 296.1260.

(*R*)-(-)- and (*S*)-(-)-2-[(*R*)-*O*-MEM-mandeloyloxy]heptanal (**3b-r** and **3b-s**) were identified with (*R*)-(+)- and (*S*)-(+)-2-[(*S*)-*O*-MEM-mandeloyloxy]heptanal¹⁶) on the NMR spectra. (*R*)-(-)-2-[(*R*)-*O*-MEM-mandeloyloxy]heptanal (**3b-r**): $[\alpha]_{\rm D} =$ -26.7°, *c* 0.25 (EtOH); and the (*S*)-(-)-form (**3b-s**): $[\alpha]_{\rm D} = -53.0^{\circ}, c \ 0.17$ (EtOH).

Diisopropyl 2,15-bis $\{(2S)-2-hydroxy-1-f(R)-O-$ *MEM-mandeloyloxy]propy*} hexadecanedioate(8a-s). **3a-s** (183.9 mg, 0.62 mM) was added to a THF solution of the dienolate of diisopropyl hexadecanedioate [which had been prepared from diisopropyl hexadecanedioate (5), 100.0 mg, 0.27 mM and LDA (0.62 mM) at -78° C for 1.5 hr in dry THF] at -78° C, and the mixture was stirred for 40 min at -78° C to give 7a-s (139.1 mg, quantitative yield). NMR $\delta_{\rm H}$ (CDCl₃): 3.6-3.8 (CH-OH), 4.7-4.9 (CH-Omandeloyl). NMR $\delta_{\rm C}$: 67.05, 70.00, 70.28, 71.05 (C–OH), 73.18, 74.40, 75.68 (CH–Omandeloyl). HR-MS m/z: calcd. for $C_{52}H_{74}O_{14}$ (M⁺-2H₂O-2H₂), 922.5079; found, 922.5058. Since 7a-s was unstable, it was transformed to 8a-s. 7a-s (139.1 mg, 0.14 mm) was treated with an excess amount of AcONa (1.0 g) in a mixed solvent of DMF:HMPA = 3:1 at 50° C for 5 hr, and the crude products (100.2 mg) were separated by preparative TLC (hexane:EtOAc = 1:2) to give 8a-s (61.6 mg, 44.4% yield). NMR $\delta_{\rm H}$ (CDCl₃): 1.09–1.36 $(20H, m, CH_2), 1.25 (12H, d, J=7.1 Hz, -CH_3),$ 1.43 (6H, d, J=6.6 Hz, CH₃), 1.49–1.70 (8H, m, $HO-C-CH_2\&OCO-C-CH_2$, 2.35-2.50 (2H, m, CO-CH₂), 3.36 (3H, s, MEM), 3.43-3.54 (4H, m, MEM), 3.54-3.70 (2H, m, MEM), 3.70-3.83 (2H, m, MEM), 3.7-4.0 (2H, m, CH-OH), 4.72-4.82 (2H, m, $J = 7.1 \, \text{Hz},$ CH-Omandeloyl), 4.78 (2H, d. J=6.7 Hz, MEM), 4.87 (2H, d, J=6.6 Hz, MEM), 4.91-5.10 (4H, m, CH of isopropyl), 5.21 (2H, d, J=7.1 Hz, CO-CH(OMEM)Ph), 7.30-7.43 (6H, m, Ph), 7.43–7.49 (4H, m, Ph). NMR δ_c : 69.27, 69.53 (CH-OH), 77.17 (CH-Omandeloyl). HR-MS m/z: calcd. for $C_{52}H_{74}O_{14}$ (M⁺-2H₂O-2H₂), 922.5079; found, 922.5079.

(S, S)-(+)-Ancepsenolide (1a-s). 8a-s (61.6 mg, 0.064 mm) in dry THF (2 ml) was added to a stirred suspension of NaH (a 60% dispersion in mineral oil, 11.5 mg (0.19 mM)) in THF (1 ml) at 0°C, and the mixture stirred for 1 hr at room temperature.¹⁷⁾ The reaction mixture was poured into an aqueous NH₄Cl solution (2 ml) and ether (10 ml). The organic materials were extracted with ether and EtOAc, and successively washed with 2 N HCl, sat. NaHCO₃ and sat. NaCl. After drying over MgSO₄, the solvents were evaporated to give crude products (38.1 mg) which were purified with silica gel TLC (benzene) and successive crystallization (hexane) to give optically active (S,S)-(+)-ancepsenolide (1a-s, 5.5 mg, 23.8% yield, $[\alpha]_{\rm D} = +20.0^{\circ}, c \ 0.133 \ ({\rm CHCl}_3))$. The NMR spectra of 1a-s agreed with those of ancepsenolide. 1a-s. NMR $\delta_{\rm H}$ (CDCl₃): 1.07–1.38 (16H, m, CH₂), 1.43 $(6H, d, J=6.8 \text{ Hz}, CH_3), 1.49-1.69 (4H, m, =$ C-CH₂-CH₂), 2.26 (4H, t, J=7.7 Hz, $=C-CH_2$), 4.99 (2H, dq, J=1.6 Hz, COOCH), 6.98 (2H, d, J= 1.3 Hz, = CH). NMR $\delta_{\rm C}$: 19.24, 25.20, 27.43, 29.19,

29.30, 29.50, 29.57, 77.27, 134.37, 148.85, 173.89. HR-MS m/z: calcd. for C₂₂H₃₄O₄ (M⁺), 362.2457; found, 362.2454.

The (R, R)-(-)-isomer (1a-r) was also synthesized from the aldehyde (3a-r) and 5 in a 14% yield by a procedure similar to that just described. The structure was identified with that of 1a-s by their ¹H- and ¹³C-NMR spectra. $[\alpha]_D = -21.7^\circ$, c 0.273 (CHCl₃).

(S,S)-2,2'-dodecamethylene-bis(2-(R,R)and nonen-4-olide) (1b-r and 1b-s). 3b-r (173 mg, 0.50 mm) was stirred with the lithium enolate of diisopropyl hexadecanedioate [which had been prepared from 5 (80 mg, 0.022 mM) and LDA at -78° C for 1.5 hr in dry THF] at -78° C for 40 min in dry THF to give 7b-r (237.5 mg, 0.22 mM, quantitative yield). 7b-r (237.5 mg, 0.22 mM) was transformed into diisopropyl 2,15-bis{(2R)-2-hydroxy-1-[(R)-O-MEM-mandeloyloxy]heptyl}hexadecanedioate (8b-r, 85.9 mg, 0.08 mM, 36.1% yield. HR-MS m/z: calcd. for $C_{57}H_{90}O_{13}$ $(M^+-CH_3OCH_2CH_2O-H_2O),$ 982.6381; found, 982.6390) by a method similar to that just described. 8b-r (85.9 mg, 0.08mM) was transformed into bis-butenolide (1b-r, 4.7 mg, 0.01 mM, 12.4% yield). 1b-r: $[\alpha]_D = -19.1^\circ$, c 0.27 (CHCl₃). NMR $\delta_{\rm H}$ (CDCl₃): 0.89 (6H, t, J = 6.6 Hz, CH₃), 1.18–1.38 (24H, m, CH₂), 1.38–1.50 (4H, m, CH₂), 1.50–1.59 (4H, m, CH₂), 1.59–1.67 (2H, m, C_5 -H& C_{24} -H), 1.67-1.78 (2H, m, C_5 -H& C_{24} -H), 2.27 (4H, t, J = 7.5 Hz, $C_9 \& C_{20}$), 4.88 (2H, t, J = 5.6Hz, COOCH), 6.98 (2H, s, = CH). NMR $\delta_{\rm C}$: 13.94. 22.46, 24.69, 25.27, 27.48, 29.20, 29.32, 29.52. 29.58, 31.54, 33.57, 81.24, 134.57, 147.81, 174.00. HR-MS m/z: calcd. for C₃₀H₄₈O₄ (M⁺-H₂), 472.3553; found, 472.3569.

1b-s was also synthesized from **3b-s** (220.0 mg, 0.062 mm) and lithium enolate of diisopropyl hexadecanedioate (5) [which had been prepared from 5 (100.0 mg, 0.027 mM) and LDA at -78° C for 1.5 hr in dry THF] at -78° C for 40 min in dry THF to give 7b-s (300.8 mg, 0.028 mM, quantitative yield). 7b-s (300.8 mg, 0.028 mM) was transformed into diisopropyl 2,15-bis{(2S)-2-hydroxy-1-[(R)-O-MEMmandeloyloxy]heptyl}hexadecanedioate (8b-s, 122.8 mg, 0.11 mm) by a method similar to that just described (40.3% yield), and 8b-s (122.8 mg, 0.11 mM) was then transformed into (S, S)-bis-4pentyl-2-butenolide (1b-s, 16.3 mg, 0.034 mM, 30.1% yield). **1b-s**: $[\alpha]_D = +21.3^\circ$, c 1.08 (CHCl₃). The ¹Hand ¹³C-NMR spectra of 1b-s agreed with those of 1b-r.

(*R*,*R*)- and (*S*,*S*)-2,2'-octamethylene-bis(2-penten-4-olide) (2a-s and 2a-r). 3a-s (103.7 mg, 0.35 mM) and the lithium enolate of isopropyl dodecanedioate (6, 50.0 mg, 0.16 mM) were reacted to give 9a-s (149.7 mg, 0.35 mM, quantitative yield. HR-MS m/z: calcd. for C₄₈H₆₉O₁₄ (M⁺-2H₂O+H), 869.4687; found, 869.4648) which was transformed into **10a-s** (34.6 mg, 23.1% rough yield. HR-MS m/z: calcd. for C₄₅H₆₅O₁₃ (M⁺-CH₃OCH₂CH₂O-H₂O), 813.4425; found, 813.4457). **10a-s** (34.6 mg, 3.8×10^{-2} mM) similarly yielded optically active **2a-s** (5.0 mg, 42.8% yield, $[\alpha]_D = +26.4^\circ$, *c* 0.33 (CHCl₃)). **2a-s**: NMR $\delta_{\rm H}$ (CDCl₃): 1.20–1.37 (8H, m, -CH₂), 1.41 (6H, d, J = 6.8 Hz, CH₃), 1.48–1.62 (4H, m, = C-CH₂-CH₂), 2.26 (4H, dt, J = 7.7, 1.5 Hz, = C-CH₂), 5.00 (2H, qq, J = 6.8, 1.6 Hz, CO-O-CH), 6.99 (2H, d, J = 1.5 Hz, C = CH). NMR $\delta_{\rm C}$: 19.23, 25.17, 27.39, 29.10, 29.12, 77.42, 134.29, 148.93, 173.88. HR-MS m/z: calcd. for C₁₈H₂₆O₄ (M⁺), 306.1831; found, 306.1862.

Bis-(4*R*)-4-methyl-2-butenolide (2a-r) was similarly synthesized from 3a-r and 6 in a 5.5% yield. $[\alpha]_D = -22.9^\circ$, c 0.14 (CHCl₃). 2a-r was identified with 2a-s by their NMR spectra.

(R,R)- and (S,S)-octamethylene-bis(2-nonen-4olide) (2b-s and 2b-r). 2b-s was similarly prepared by the reaction of **3b-s** and **6** via **9b-s** in a 17.4% yield. $[\alpha]_{\rm D} = +10.4^{\circ}, \ c \ 0.40 \ ({\rm CHCl}_3). \ {\rm NMR} \ \delta_{\rm H} \ ({\rm CDCl}_3):$ 0.89 (6H, t, J=6.8 Hz, CH₃), 1.23–1.37 (16H, m, CH₂), 1.37-1.49 (4H, m, O-CH-CH₂-CH₂), 1.54 $(4H, t, = C-CH_2-CH_2), 1.58-1.67 (2H, m,$ O-CH-CH-H), 1.67-1.77 (2H, m, O-CH-CH-H), 2.26 (4H, t, J = 7.6 Hz, $= C-CH_2$), 4.89 (2H, t, J =5.7 Hz, CO-O-CH), 6.99 (2H, d, J=1.4 Hz, C= CH). NMR $\delta_{\rm C}$: 13.94, 22.46, 24.70, 25.24, 27.45, 29.13, 29.15, 31.54, 33.57, 81.26, 134.49, 147.89, 173.98. HR-MS m/z: calcd. for C₂₆H₄₂O₄ (M⁺), 418.3083; found, 418.3056. The (4R)-isomer (2b-r) was similarly prepared (6.5% yield). $[\alpha]_D = -9.5^\circ$, c 0.40 (CHCl₃). 2b-r was identified with the authentic sample of 2b-s by their NMR spectra.

References

- Ciereszko, L. S., Sifford, D. H., and Weinheimer, A. J., Chemistry of coelenterates I. Occurrence of terpenoid compounds in gorgonians. *Ann. N.Y. Acad. Sci.*, **90**, 917–919 (1960).
- Schmitz, F. J., Kraus, K. W., Ciereszko, L. S., Sifford, D. H., and Weinheimer, A. J., Ancepsenolide: A novel bisbutenolide of marine origin. Chemistry of coelenterates V. *Tetrahedron Letters*, 1, 97-104 (1966).
- Schmitz, F. J., Lorance, E. D., and Ciereszko, L. S., Chemistry of Coelenterates XII. J. Org. Chem., 34, 1989–1990 (1969).

- Guo, Y., Gavagunin, M., Mollo, E., Trivellone, E., and Cimino, G., Three new butenolide lipids from the Caribbean gorgonian Pterogorgia anceps. J. Nat. Prod., 62, 1194-1196 (1999).
- Rodríguez, A. D. and Ramírez, C., Further butenolides from the Caribbean octocoral, *Pterogorgia citrina*. J. Nat. Prod., 57, 339-347 (1994).
- Schmitz, F. J. and Lorance, E. D., Chemistry of coelenterates XXI. Lactones from the gorgonian *Pterogorgia guadalupensis*. J. Org. Chem., 36, 719–721 (1971).
- Rupprecht, J. K., Hui, Y., and McLaughlin, J. L., Annonaceous acetogenins: a review. J. Nat. Prod., 53, 237-278 (1990).
- Fang, X., Rieser, M. J., Gu, Z., Zhao, G., and McLaughlin, J. L., Annaceous acetogenins: An update review. *Phytochem. Anal.*, 4, 27-48 (1993).
- Cavé, A., Figadère, B., Laurens, A., and Cortes, D., Acetogenins from Annonaceae. In "Progress in the Chemistry of Natural Products," eds. Herz, W., Kirby, G. W., Moore, R. E., Steglich, W., and Tamm, Ch., Springer, Wien, 70, pp. 81-288 (1997).
- Alali, F. Q., Liu, X., and McLaughlin, J. L., Annoceous acetogenins: Recent progress. J. Nat. Prod., 62, 504–540 (1999).
- Hoppen, S., Emide, U., Friedrich, T., Grubert, L., and Koert, U., Natural-product hybrids: Design, synthesis, and biological evaluation of quinone-annonaceous acetogenins. *Angew. Chem. Int. Ed.*, 39, 2099-2102 (2000).
- 12) Podraza, K. F. and Sneden, A. T., A short synthesis of ancepsenolide. J. Nat. Prod., 48, 792-795 (1985).
- Trost, B. M., Müller, T. J. J., and Martinez, J., Ruthenium catalyzed synthesis of butenolides and pentenolide via contra-electronic α-alkylation of hydroxyalkynoates. J. Am. Chem. Soc., 117, 1888–1899 (1995).
- 14) Iriye, R., Nakamura, A., and Takeshita, M., Synthesis of 2-(2-hydroxyalkylidene)cyclopentanones and their bio-antimutagenic activity. *Biosci. Biotechnol. Biochem.*, 59, 401-407 (1995).
- 15) Iriye, R., Takai, K., and Noguchi, M., The synthesis of (*R*)-1-(2-oxocyclopentyliden)-2-alkanols and the (*S*)-forms, and their bio-antimutagenic activity against UV-induced *Escherichia coli* WP2 B/r *Trp⁻*. *Bioorg. Med. Chem. Lett.*, 7, 199–202 (1997).
- 16) Noguchi, M., Takai, K., Takei, Y., and Iriye, R., Synthesis of (*R*)- and (*S*)-13-hydroxy-10-oxo-*trans*-11-octadecenoic acid and their antimutagenic activity. *Biocontrol Sci.*, 5, 91–95 (2000).
- 17) van Delft, F. L., Timmers, C. M., van der Marel, G. A., and van Boom, J. H., Preparation of 2-oxazolidinones by intramolecular nucleophilic substitution. *Synthesis*, 450–454 (1997).

1906