Organic & Biomolecular Chemistry



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Cite this: DOI: 10.1039/d0ob00910e

Efficient synthesis of a galectin inhibitor clinical candidate (TD139) using a Payne rearrangement/ azidation reaction cascade[†]

Jacob St-Gelais, Vincent Denavit and Denis Giguère 🕩 *

Received 1st May 2020, Accepted 7th May 2020 DOI: 10.1039/d0ob00910e Selective galectin inhibitors are valuable research tools and could also be used as drug candidates. In that context, TD139, a thiodigalactoside galectin-3 inhibitor, is currently being evaluated clinically for the treatment of idiopathic pulmonary fibrosis. Herein, we describe a new strategy for the preparation of TD139. Starting from inexpensive levoglucosan, we used a rarely employed reaction cascade: Payne rearrangement/azidation process leading to 3-azido-galactopyranose. The latter intermediate was efficiently converted into TD139 in a few simple and practical steps.

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Introduction

Galectins are proteins that bind to galactoside residues and their natural ligands are any glycoconjugates with a non-reducing galactopyranoside terminus.¹ Galectins have the ability to regulate numerous biological processes, including neoplastic transformation, tumor cell survival processes, angiogenesis, tumor metastasis, and cell homeostasis.² Among the 15 mammalian members, galectins contain a conserved carbohydrate recognition domain (CRD) of about 135 amino acids. Sharing a consensus amino acid sequence for the CRD makes it difficult to prepare selective galectin inhibitors for biological investigations. Over the years, the scientific community has directed efforts in the synthesis of potent and optimized galectin inhibitors. These works have been summarized in reviews by the groups of Pieters,³ Kiss,⁴ Nilsson,⁵ Mayo⁶ and ours.⁷ Small molecular weight glycomimetics are rationally designed to bind the CRD and inhibit the galectin target. As such, TD139 1⁸ (Fig. 1) is a ditriazolylthiodigalactoside that was designed as one of the most potent antagonist of galectin-3.9 Compound 1 showed a K_d of 14 nM to galectin-3, as determined using a competitive fluorescence anisotropy assay.¹⁰ Developed by Galecto Inc., TD139 passed a Ib/IIa phase clinical trial in idiopathic pulmonary fibrosis patients (IPF) and is currently in a randomized, double-blind, placebo-controlled phase IIb trial in subjects with IPF investigating its efficacy and safety (see http://www.clinicaltrials.gov).11 A particular

Québec City, Qc, Canada G1V 0A6. E-mail: denis.giguere@chm.ulaval.ca

feature of thiodigalactoside analogues is their resistance to glycosidase *in vivo*. Combined with the long and difficult synthetic route to access bis-(3-azido-3-deoxy- β -D-galactopyranosyl)-sulfane core,¹² there is a major need for a fast and efficient synthesis of TD139.

Fig. 1 shows, in retrosynthetic format, the known synthetic strategy leading to TD139 1, a 3,3'-bis-(4-aryltriazol-1-yl) thiodigalactoside. The C_2 symmetry of 1 allowed for a general twodirectional strategy to install triazole moieties through click chemistry. Then, bis-(2,4,6-tri-O-acetyl-3-azido-3-deoxy-β-Dgalactopyranosyl)-sulfane 2 is accessible from dimerization of 3-azido-3-deoxy-galactopyranoside bromide 3. Currently, two distinct approaches were reported for the synthesis of 3-azido-3-deoxy-galactose. The first one was initially reported by Lemieux¹³ and involved a nucleophilic displacement of a gulofuranose triflate derivative 4. This approach began with expensive gulofuranose, but the latter compound can be accessed from inexpensive glucose diacetonide 5 in a five-step protocol.14 The second approach was described more recently by the group of Nilsson and involved the introduction of the 3-azidofunctionality via nucleophilic azidation at C-3 of a gulopyranoside triflate intermediate 6.15 This compound arises from C-3 inversion and functionalization of galactose 7. Because of the instability of triflate intermediates, this method was later on improved by using more stable imidazylate and tosylate intermediates.¹⁶ Nevertheless, a more convenient preparation of 3-azido-3-deoxy-galactose is much needed for enabling large scale synthesis of thiodigalactoside galectin inhibitors and for a rapid access to active pharmaceutical ingredients. We aimed to develop a new synthetic route to 1,2,4,6-tetra-O-acetyl-3azido-3-deoxy-D-galactopyranose that could operate on large scale and be initiated from inexpensive starting material. The intended synthetic pathway was designed to develop a practical

Département de Chimie, 1045 av. De la Médecine, Université Laval, GlycoNet,

[†]Electronic supplementary information (ESI) available. CCDC 1956891. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/ d0ob00910e



Fig. 1 Retrosynthetic analysis of TD139 1.

alternative to known reported methods.^{13,15,16} We explored the possibility of generating crucial intermediate 3 *via* cleavage of the 1,6-anhydro core from 1,6-anhydro-3-azido-3-deoxy-galactopyranose derivative 8. The requisite azide moiety was introduced using a rarely employed reaction cascade: Payne rearrangement/azidation process.¹⁷ Levoglucosan 9 is an ideal starting material since the 1,6-anhydro core avoided the preliminary protection of O-6 and anomeric positions, and can easily lead to scalable 3-azido-3-deoxy-galactose derivatives *via* simple experimental protocols.

Cascade processes generate *in situ* a series of reactive intermediates that undergo consecutive transformations. They can offer cost saving in terms of reagents and solvents, as well as time and effort. Cascade processes can drastically shorten a synthetic route and thus reducing the number of manipulations. Because of their many advantages, these reactions have found applications in the synthesis of useful pharmaceuticals and other fine chemicals.¹⁸

Finally, this novel synthetic process could provide a rapid access to 3-azido-3-deoxy-galactose analogues, crucial component to probe the active site of glycosyltransferases,¹⁹ as inhibitors of bacterial adhesins,²⁰ and to generate novel aminoglycoside antibiotics.²¹

Results and discussion

The synthesis of 1,6-anhydro-3-azido-3-deoxy- β -D-galactopyranose **13** from levoglucosan **9** is summarized in Scheme **1**. Monotoluenesulfonylation of levoglucosan **9** afforded known 1,6-anhydro-4-*O-p*-tolylsulfonyl- β -D-glucopyranose **10**²² and the latter compound was treated under basic condition to generate 1,6:3,4-dianhydro- β -D-galactopyranose **11** in quantitative yield.²³ An epoxide migration (known as Payne rearrangement)²⁴ of compound **11** lead to the requisite 1,6:2,3-dianhy-

dro- β -D-gulopyranose 12, an ideal precursor for azidation reactions. Efforts towards this end are presented in Table 1. Accordingly, compound 11 was treated with sodium hydride in N,N-dimethylformamide and allowed to come to equilibrium for 6 h prior addition of sodium azide followed by heating at 120 °C for 24 h (entry 1). As propose by the group of Fraser-Reid, in an aprotic solvent, the gulose form should predominate because of a possible chelation between a sodium cation, the O-3 alkoxide, the endocyclic oxygen, and the 1,6-anhydro bridge.²⁵ The ¹H NMR of the crude reaction mixture after treatment with sodium hydride revealed formation of compound 12 as major isomer $(11/12 \approx 1:9)$. Unexpectedly, we only isolated an unprecedented dimeric by-product 15 in 72% yield. In order to improve the azidation step, we added 5 equivalents of ammonium chloride (entry 2) and compound 13 was isolated in 65% yield, along with 7% of compound 14. According to the Fürst-Plattner rule, nucleophilic attack occurs on the C-3 oxirane carbon of compound 12 (leading to 13) and on the C-4 carbon of compound 11 (leading to 14).²⁶ The obtained selectivity can be explain using steric and electronic reasons. Hence, an incoming nucleophile suffered from 1,3-diaxial interaction with the C-2 hydroxyl group for compound 11 and the partial positive charge at C-3 of the oxirane carbon was stabilized by nearby acetal for compound 12.²⁷ Further optimisation using other additives (Na₂SO₄: entry 3; Na₂HPO₄: entry 4; NH₄Br: entry 5) failed, but lowering the temperature to 100 °C for 66 hours (entry 6) provided the desired 1,6-anhydro-3-azido-3deoxy-\beta-D-galactopyranose 13 in 81% yield, along with 9% of compound 14 as inseparable mixture using standard flash column chromatography. Finally, the latter reaction was even perform on gram scale with no apparent formation of byproduct 15.

A proposed mechanism for the formation compound **15** is shown in Fig. 2 and presumably involves an intermolecular epoxide opening followed by an intramolecular attack from the

Paper

Organic & Biomolecular Chemistry



Scheme 1 Synthesis of 1,6-anhydro-3-azido-3-deoxy- β -D-galactopyranose 13 from levoglucosan 9.





^a Yields refer to isolated pure products after flash column chromatography. ^b The reaction was perform on gram scale.

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Fig. 2 Proposed mechanism for the formation of dimeric by-product **15** and X-ray derived ORTEP of compound **15** showing 50% thermal ellipsoid probability, carbon (gray), oxygen (red), hydrogen (white).

cis-vicinal alkoxide. Also, an X-ray crystallographic analysis of **15** confirmed its dimeric nature unambiguously (see X-ray derived ORTEP in Fig. 2).²⁸

With the ongoing objective of shortening the number of steps to prepare valuable 3-azido-3-deoxy-galactose, we performed our key step starting from mono-tosylate **10** using aqueous sodium hydroxide as base (Scheme 2).²⁹ To our delight, compound **13** and **14** were isolated in 79% yield (**13/14** = 4 : 1, ~93% per step). Presumably, this green process avoided organic solvent and involved the formation of the 3,4-anhydro analogue followed by Payne rearrangement and azidation. To the best of our knowledge, this is the first example of an



Scheme 2 Development of a green reaction sequence: epoxide formation/Payne rearrangement/azidation, leading to 1,6-anhydro-3azido-3-deoxy- β -D-galactopyranose 13.

epoxide formation/Payne rearrangement/azidation reaction sequence.

Encouraged by the successful synthesis of 3-azido-3-deoxy- β -p-galactopyranose derivative from levoglucosan, we maintained our efforts towards the preparation of TD139 **1** (Scheme 3). Thus, triethylsilyl triflate-catalyzed acetolysis of a mixture of **13** and **14** furnished a separable mixture of 3-azidogalactose **16** (77%) and 4-azido-glucose **17** (19%). Then, the galactosyl bromide **3** was slowly generated in 74% yield using TiBr₄ from intermediate **16**. Alternatively, intermediate **3** was generated from compound **11**. Accordingly, acetylation of the crude reaction mixture of **18** and **8** (1:10) in 81% yield over 2 steps. With compound **8** in hand, a direct bromolysis with TiBr₄ afforded in 97% yield a mixture of galactosyl bromide **3**



Scheme 3 Rapid synthesis of TD139 1 from 1,6:3,4-dianhydro- β -D-galactopyranose 11.

and 3-azido-galactose 16 (3/16 = 3:2). Subsequently, we used a similar strategy to the group of Nilsson for the preparation of compound $1.^{\rm 30}$ Briefly, a base promoted $S_{\rm N}2$ substitution of galactosyl bromide 3 with triisopropylsilanethiol afforded triisopropylsilyl β -thio-galactoside **19** in 76% yield.³¹ The dimeric nature of the thiodigalactoside core was achieved by treating compounds 3 and 19 under tetrabutylammonium fluoride to generate bis-(2,4,6-tri-O-acetyl-3-azido-3-deoxy-β-Dgalactopyranosyl)-sulfane 2 in 75% yield. Finally, triazole installation with known alkyne 20 preceded global deprotection, allowing the preparation of TD139 1 in 65% over 2 steps. The synthesis of compound 1 was described by the group of Nilsson in 11 steps (6% global yield) starting from known intermediate phenyl 4,6-O-benzylidene-1-thio-β-Dglucopyranoside.^{9,12} Using the present strategy, compound 1 was prepared in only 8 steps and 21% yield from known tosylate 10. It is also important to point out that our strategy avoided the use of triflate intermediates,^{9,12} unsuitable in large scale synthesis of active pharmaceutical ingredients.

Conclusions

TD139 is a galectin-3 inhibitor and is currently being evaluated clinically for the treatment of idiopathic pulmonary fibrosis. An efficient synthesis of TD139 **1** from levoglucosan was described. To the best of our knowledge, this is the shortest route to 3-azido-3-deoxy-galactose, a crucial intermediate for the preparation of bis- $(2,4,6-\text{tri-}O-\text{acetyl-}3-\text{azido-}3-\text{deoxy-}\beta-\text{p-}galactopyranosyl)$ -sulfane. In that manner, we used a rarely employed reaction cascade: Payne rearrangement/azidation, leading to 1,6-anhydro-3-azido-3-deoxy- β -p-galactopyranose. In the course of this study, we also isolated an unprecedented

dimeric by-product **15**. We are currently exploring the chemistry and biology of this novel C_2 -symmetrical compound. Finally, the strategy described herein allowed the preparation of galactose derivatives functionalized at C-3 that could provide useful options for the synthesis of other galectin inhibitors.

Conflicts of interest

A related provisional patent has been filed with the title "Synthesis of 3-azido-3-deoxy-D-galactopyranose" by Jacob St-Gelais, Vincent Denavit and Denis Giguère (USPTO 62/861,476).

Acknowledgements

This work was supported by the Natural Sciences and Engineering Research Council of Canada (NSERC) and Université Laval. J. St-G. thanks the Fonds de Recherche du Québec-Nature et Technologies for a postgraduate fellowship.

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