Article

Subscriber access provided by UNIV NEW ORLEANS

Catalytic Asymmetric Conjugate Addition and Sulfenylation of Diarylthiazolidin-2,4-Diones

Lihui Jiao, Liwei Bu, Xinyi Ye, Xiaowei Zhao, and Zhiyong Jiang

J. Org. Chem., Just Accepted Manuscript • DOI: 10.1021/acs.joc.6b01637 • Publication Date (Web): 23 Sep 2016

Downloaded from http://pubs.acs.org on September 24, 2016

Just Accepted

"Just Accepted" manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides "Just Accepted" as a free service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. "Just Accepted" manuscripts appear in full in PDF format accompanied by an HTML abstract. "Just Accepted" manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are accessible to all readers and citable by the Digital Object Identifier (DOI®). "Just Accepted" is an optional service offered to authors. Therefore, the "Just Accepted" Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the "Just Accepted" Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these "Just Accepted" manuscripts.



The Journal of Organic Chemistry is published by the American Chemical Society. 1155 Sixteenth Street N.W., Washington, DC 20036

Published by American Chemical Society. Copyright © American Chemical Society. However, no copyright claim is made to original U.S. Government works, or works produced by employees of any Commonwealth realm Crown government in the course of their duties.

Catalytic Asymmetric Conjugate Addition and Sulfenylation of

Diarylthiazolidin-2,4-Diones

Lihui Jiao,^{†,¶} Liwei Bu,^{†,¶} Xinyi Ye,[‡] Xiaowei Zhao,[†] and Zhiyong Jiang^{*,†}

[†]Key Laboratory of Natural Medicine and Immuno-Engineering of Henan Province, Henan University, Kaifeng, Henan, P. R. China, 475004 [‡]Division of Chemistry and Biological Chemistry, Nanyang Technological University, 21 Nanyang Link, 637371, Singapore

E-mail: chmjzy@henu.edu.cn

Abstract:

This work reports the first application of diarylthiazolidin-2,4-diones as nucleophiles in asymmetric catalysis. By utilizing chiral amino acid-based (thio)urea-tertiary amines as the catalysts, asymmetric conjugate addition to nitroolefins and sulfenylation to *N*-(sulfanyl)-succinimides of diarylthiazolidin-2,4-diones have been established successively. Two series of biologically important 5-aryl-5-substituted thiazolidin-2,4-diones were obtained in high enantio- and diastereoselectivities (up to >99% ee and >19:1 dr). The enantio-enriched adducts were found to show satisfactory anticancer activities against three different cancer cell lines using the MTT assay. All the successes depended on the development of a general and expedient synthetic strategy to provide diverse 5*H*-thiazolidin-2,4-diones.

Keywords: Asymmetric organocatalysis; Thiazolidinediones; Diarylthiazolidin-2,4-diones; Conjugate addition; Sulfenylation

INTRODUCTION

Thiazolidinediones¹, also known as glitazones² are important heterocyclic compounds for the treatment of type II diabetes mellitus. More complex compounds such as 5-aryl-5-substituted thiazolidin-2,4-diones bearing a fully substituted stereogenic center on the 5-position are also important as they are promising drug candidates as inhibitors of lactamase^{3a} and aldose reductase,^{3b} agrochemical fungicides^{3c} and angiotensin-II receptor antagonists^{3d} (compounds I-IV, Figure 1). To our knowledge, the asymmetric synthesis of chiral 5-aryl-5-substituted thiazolidin-2,4-diones has not been established yet.

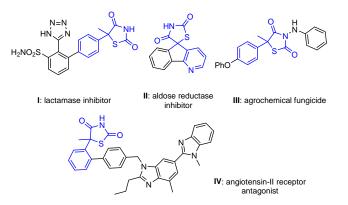


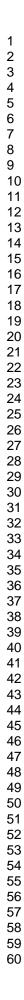
Figure 1. Representative examples of bioactive compounds.

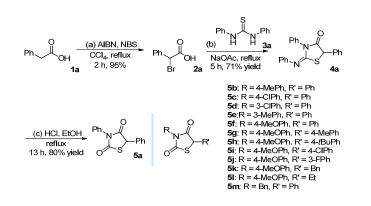
Structurally, catalytic asymmetric reaction using the enolizable 5-aryl-substituted thiazolidin-2,4-diones as nucleophiles would provide the most direct approach to the desired chiral 5-aryl-5-substituted thiazolidin-2,4-diones. Wheeler's synthetic method of making diarylthiazolidin-2,4-diones (*N*-aryl-5-aryl-substituted thiazolidin-2,4-diones) could be dated back to more than a century ago.⁴ From then on, no reports of these heterocyclic molecules in catalytic reaction were further pursued.⁵ In this context, the development of asymmetric reaction using diarylthiazolidin-2,4-diones as reagents remains highly desirable and challenging, given their less known chemical reactivity.

In recent years, we have invested great efforts in developing organocatalytic asymmetric strategies to access biologically important chiral molecules with hetero-quaternary (thia⁶ and oxa^7) stereogenic centers. As an extension of these works, we herein report the first catalytic asymmetric reaction of diarylthiazolidin-2,4-diones, including conjugate addition to nitroolefins and sulfenylation to *N*-(sulfanyl)-succinimides, thus leading to the desired chiral 5-aryl-5-substituted thiazolidin-2,4-diones with high stereoselectivity. To this end, a general and efficient synthetic method of 5*H*-thiazolidin-2,4-diones is reported for the first time.

RESULTS AND DISCUSSION

Reported synthetic protocols^{4,8} for 5-aryl-substituted thiazolidin-2,4-diones lack the product scope thus re-igniting our interest in this work. As shown in Scheme 1, the representative diphenylthiazolidin-2,4-dione **5a** could be prepared from commercially available phenylacetic acid **1a** through a simple three-step process. Treatment of AIBN and NBS could transform **1a** to α -bromo carboxylic acid **2a** in 95% yield. Subsequently **2a** was condensed with diphenyl thiourea **3a** in the presence of NaOAc to form 2-(phenylimino)-4-thiazolidinone **4a** in 71% yield. After hydrolysis with HCl, diphenylthiazolidin-2,4-dione **5a** was obtained satisfactorily. Noteworthy is that this methodology is versatile and suitable to provide a series of *5H*-thiazolidin-2,4-diones with diverse *N*-functionalized groups and 5-aryl, benzyl and alkyl substituents (**5b-m**).⁹





Scheme 1. Representative synthesis of 5H-thiazolin-2,4-diones.

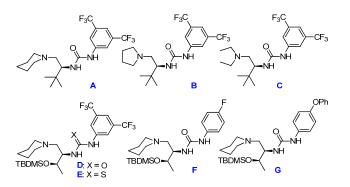


Figure 2. Structures of catalysts A-G.

To explore the reactivity of 5*H*-thiazolidin-2,4-diones, we first attempted catalytic asymmetric conjugate addition to the most commonly used electrophiles, i.e. nitroolefins. The reaction between diphenylthiazolidin-2,4-dione **5a** and nitroolefin **6a** was chosen as the model reaction (Table 1). Our recent works have revealed that *L*-amino acid-based urea-tertiary amines as efficient bifunctional Brønsted base catalysts could be conveniently prepared.^{7d-f} Therefore, *L-tert*-leucine-based urea-tertiary amine **A** (Figure 2) was first screened as the catalyst (entry 1). It was found that the reaction worked smoothly in toluene at 25 °C, and the desired conjugate adduct **7a** was obtained in 77% yield after 24 hours. While enantio- and diastereoselectivity were poor, the good reactivity encouraged us to further examine the asymmetric reaction with catalysts **B** and **C**, containing pyrrolidine and

diethylamine as the tertiary amine moiety respectively (entries 2–3). Catalysts **B** and **C** did not give improved results. Next, we examined catalyst **D** with *L*-threonine as the chiral skeleton and *tert*-butyldimethylsilyl (TBDMS) as the alcohol protecting group, and ee value of **7a** was increased to 30% (entry 4). We also observed that the analogous thiourea **E** slightly improved the enantioselectivity, but the reaction became sluggish (Table 1, entry 5). Effect of the substituent of urea was then investigated (catalysts **F** and **G**, entries 6–7). It was detected that catalyst **G** with a 4-PhO-Ph urea moiety could further increase the enantioselectivity (entry 7), indicating that different substituents of urea affected the H-bond interactions between urea and the substrate (see Figure 3) thus leading to distinct stereoselective outcomes.

Ph.N	O I↓_Ph S	+ Ph_NO2		atalyst A-G 10 mol %)			
	5a	6a		7a		'a	
entry	cat.	solvent	<i>t</i> (h)	yield $(\%)^b$	$ee (\%)^c$	dr ^c	
1	Α	toluene	24	77	18/6	48:52	
2	В	toluene	24	49	15/4	57:43	
3	С	toluene	24	81	17/4	47:53	
4	D	toluene	24	62	30/12	52:48	
5	Ε	toluene	24	27	32/13	46:54	
6	F	toluene	24	73	32/13	52:48	
7	G	toluene	24	71	36/27	53:47	
8	G	THF	24	76	23/17	51:49	
9	G	Et ₂ O	24	87	27/8	54:46	
10	G	CH_2Cl_2	24	82	35/15	48:52	
11	G	<i>m</i> -xylene	24	85	60/9	64:36	
12	G	<i>m</i> -xylene	48	89	77/7	78:22	
13	G	<i>m</i> -xylene	48	64	81/1	82:18	
14^d	G	<i>m</i> -xylene	60	91	87/11	83:17	
15^{e}	G	<i>m</i> -xylene	60	89	84/9	86:14	
16 ^f	G	<i>m</i> -xylene	60	92	89/4	90:10	

Table 1. Screening Studies^a

^{*a*}The reaction was carried out with 0.05 mmol of **5a**, 0.06 mmol of **6a**, and 0.005 mmol of catalyst in 0.5 mL solvent. Entries 1–11, T = 25 °C; entry 12, T = 0 °C; entries 13–16, T = -20 °C. ^{*b*}Isolated yield. ^{*c*}Determined by HPLC methods. ^{*d*}25 mg of 4 Å molecular sieves were used. ^{*e*} NaCl (10 mol%) was used. ^{*f*}Both 25 mg of 4 Å molecular sieves and 10 mol% NaCl were utilized.

In the presence of 10 mol% of catalyst **G**, a range of solvents, including THF, ether, dichloromethane and *m*-xylene, were evaluated (Table 1, entries 8–11), and *m*-xylene was the best, providing **7a** in 85% yield with 60% ee and 64:36 dr (entry 11). When the temperature was decreased, both enantio- and diastereoselectivity were improved (Table 1, entries 12–13); at -20 °C, **7a** with 81% ee and 82:18 dr was attained (entry 13). The effect of additive was examined (entries 14–16). Molecular sieves (4 Å; 10 mg) boosted the enantioselectivity but gave similar diastereoselectivity (entry 14).^{7b} NaCl was shown to be effective in enhancing diastereoselectivity (entry 15). The combination of 4 Å molecular sieves and NaCl optimally provided **7a** in 92% yield with 89% ee and 90:10 dr (entry 16).

Subsequently, we anticipated to improve the stereoselectivity by modifying the *N*-substituent from *N*-phenyl of **5a** to other aryl groups. As shown in Table 2, the introduction of 4-MePh (**5b**) and 4-ClPh (**5c**) on the *N*-position led to similar enantio- and diastereoselectivities (Table 2, entries 1–2). Surprisingly, no reaction was observed for **5d** (3-ClPh) (entry 3). The best results were obtained with 4-MeOPh (**5e**) substituent on the *N*-position, and the corresponding product **7f** was attained in 90% yield with 98% ee and 90:10 dr (entry 5). These results indicated that the *N*-substituted aryl groups are pivotal for modulating the reactivity and stereoselectivity.

Ar N	Ph	+ Ph	√∕~ _{NC}	catalyst 0 (10 mol % 2 <i>m</i> -xylene, -20 60 h) Ar	N Ph	10 ₂
	5		6a			7	
	entry	5	7	yield $(\%)^b$	$ee (\%)^{c}$	dr^c	
-	1	5b	7b	87	87	92:8	
	2	5c	7c	83	85	87:13	
	3	5d	7d	N.R.	N.A.	N.A.	
	4	5e	7e	71	87	90:10	
_	5	5f	7f	90	98	90:10	

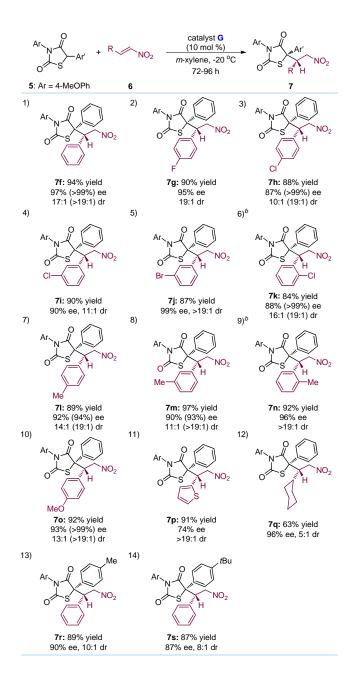
Table 2. Investigation on the effect of N-aryl groups of 5^a

^{*a*}The reaction was carried out with 0.05 mmol of **5**, 0.06 mmol of **6a**, 0.005 mmol of catalyst **G**, 0.005 mmol of NaCl and 25 mg of 4 Å molecular sieves in 0.5 mL *m*-xylene. ^{*b*}Isolated yield. ^{*c*}Determined by HPLC methods. N.R. = no reaction.

With the optimized reaction conditions in hand, the reaction scope was expanded (Table 3). First, we evaluated the conjugate addition of **5e** with a variety of nitroolefins **6** in the presence of 10 mol% of catalyst **G** at -20 °C in *m*-xylene solvent and employing 4Å molecular sieves and NaCl as additives (Table 3, entries 1–12). The corresponding conjugate adducts **7f-q** were obtained in 63–97% yield with 74% to 99% ee and 5:1 to >19:1 dr within 72–96 hours. 2-Thienyl nitroolefin (**7p**, Table 3, entry 11) was found to suppress enantioselectivity. With 15 mol% of catalyst **G**, enantiomeric pure adduct **7n** was obtained (Table 3, entry 9). Next, diarylthiazolidin-2,4-diones with 4-MePh (**5f**) and 4-*t*BuPh (**5g**) on the 5-position were subjected to conjugate addition reaction with **6a**, affording adducts **7r-s** with excellent enantioselectivities and slightly lower but satisfactory diastereoselectivities (Table 3, entries 13–14). Unfortunately, 5-benzyl and ethyl-substituted as well as *N*-benzyl-substituted thiazolidin-2,4-diones (**5j-I**) were unreactive under the established reaction conditions. The absolute configurations of conjugate adducts **7** were assigned based on *X*-ray crystallographic

analysis of a single crystal of **7f**.¹⁰

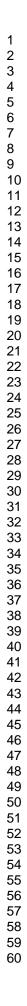


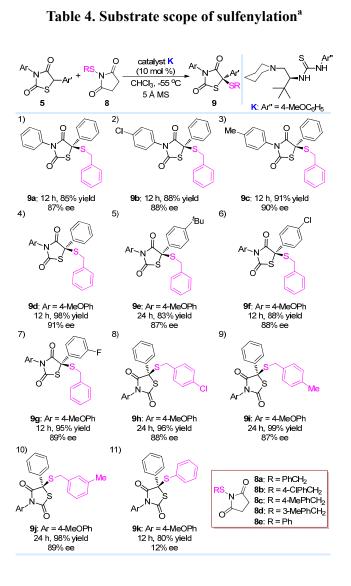


^{*a*}The reaction was carried out with 0.1 mmol of **5**, 0.12 mmol of **6**, 0.01 mmol of **G**, 0.01 mmol of **NaCl** and 50 mg of 4 Å molecular sieves in 1.0 mL *m*-xylene. Yields of isolated products are presented. The dr was determined by ¹H NMR analysis. Ee values were

 determined by HPLC using chiral stationary phase. The data in parentheses were obtained after a single recrystallization. $^{b}15$ mol% of catalyst **G** was utilized.

In recent years, asymmetric sulfenylation^{6a-b,11} has been demonstrated as one of the most efficient strategies to build optically active sulfur-containing compounds. As an continuation of the success in conjugate addition, we were subsequently engaged in surveying sulfenylation of diarylthiazolidin-2,4-diones, to facilitate the first asymmetric synthesis of 5-sulfur-5-aryl-disubstituted thiazolidin-2,4-diones and 5-sulfone-5-aryl-disubstituted thiazolidin-2,4-diones, which were tested as inhibitors for farnesyl-protein transferase.¹² Under the established reaction conditions towards conjugate addition, we attempted sulfenylation of diphenylthiazolidin-2,4-dione 5a using N-(benzylthio)succinimide 8a as the sulfenylating reagent. However, the reaction became very sluggish, indicating that amino acid-based urea-tertiary amine catalyst is not optimal for sulfenylation. We next screened amino acid-based thiourea-tertiary amines, which is another class of important bifunctional Brønsted base catalysts.¹³ We were pleased to find that the reaction worked, and the sulferylated adduct 9a was isolated in 85% yield with 87% ee after 12 hours when employing *L-tert*-leucine-based thiourea-tertiary amine K as the catalyst and 5 Å molecular sieves as the additive in CHCl₃ at -55 °C (Table 4, entry 1). Moreover, 5e was found to yield higher enantioselectivity (entry 4).9 The substituents on aromatic rings at 5-position of thiazolidin-2,4-diones (5g-i) and of N-(benzylthio)succinimides (8b-d) did not affect the reactivity and enantioselectivity (entries 5-10). However for N-(arylthio)succinimides, only 12% ee of 9k was obtained when using N-(phenylthio)succinimide 8e as the suferylating reagent. (entry 11).





^{*a*}The reaction was carried out with 0.1 mmol of **5**, 0.2 mmol of **8**, 0.01 mmol of **K**, and 10 mg of 5Å molecular sieves in 0.5 mL CHCl₃. Yields of isolated products are presented. Ee values were determined by HPLC using chiral stationary phase.

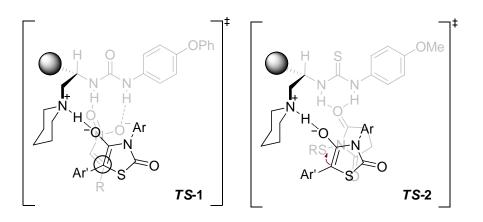


Figure 3. Plausible transition states of two reactions.

On the basis of our previous investigations,⁷ the plausible transition states of two reactions were proposed (Figure 3). First, the enolates of diarylthiazolidin-2,4-diones were generated after protonation and would bind to the R_3NH^+ arm of the catalysts G or K. Two N-H bonds of (thio)urea unit could activate the LUMO of nitroolefins (TS-1)or N-(sulfanyl)-succinimides (TS-2) through two H-bonding interactions. The conjugate adducts 7 and sulferylated adducts 9 were thus obtainable with the observed absolute configurations after nucleophilic addition. The used additives, such as 4 Å and 5 Å molecular sieves as well as NaCl, should affect the solution environment to increase the free energy differences between the transition states, thus leading to slightly improved enantio- and diastereoselectivity in two transformations.

 Table 5. IC₅₀ values of chiral conjugate adducts 7 on the growth of human cancer cell

 lines^a

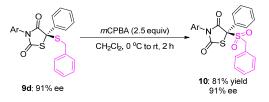
compound	7f	7g	7h	7j	71	7m	7n
H22	21	55	20	25	18	40	9.9
HCT116	36	94	45	73	60	94	30
K562	29	>100	>100	>100	>100	>100	45

^{*a*}Values are means of three experiments each done in duplicate. IC_{50} values are described in μ M.

To demonstrate the utility of the methodology, we endeavored to evaluate the biological activities of adducts. A variety of chiral 5-aryl-5-substituted thiazolidin-2,4-diones, including **7f-h**, **7j** and **7l**, were subjected to cytotoxic activity measurements for three human cancer cell lines employing the MTT assay. A summary of the IC50 values is shown in Table 5. The

analogues **7f**, **7h**, **7j** and **7l** showed inhibitory effects on the H22 with IC₅₀ values of 18 to 25 μ M. Moreover, **7n** gave a lower IC₅₀ value of 9.9 μ M. While **7n** and **7f** presented weaker inhibitory activity (IC₅₀ = 30 μ M, 29 μ M) on the HCT116 and K562 respectively, they were still overall the most effective compounds. The distinct cytotoxicities suggested that the cancer cell lines exhibited different sensitivities to chiral 5-aryl-5-substituted thiazolidinediones.

We also attempted transformation of adducts **9** to verify the synthetic utility of the method. Using *m*CPBA in dichloromethane, the oxidation of the sulfenylated adduct **9d** was performed as shown in Scheme 2. After 2 hours when the reaction completed, the sulfone **10** could be readily achieved in 81% yield with no loss of enantiomeric purity. The absolute configurations of sulfenylated adducts **9** could be assigned based on *X*-ray crystallographic analysis of a single crystal of sulfone **10**.¹⁰



Scheme 2. Transformation of adducts 9.

CONCLUSION

In summary, we have established the pioneer work of employing diarylthiazolidin-2,4-diones as nucleophiles in asymmetric synthesis. By utilizing an *L*-amino acid-based tertiary amine as a bifunctional Brønsted base catalyst, asymmetric conjugate addition of diarylthiazolidin-2,4-diones to nitroolefins afforded a series of chiral 5-aryl-5-substituted thiazolidin-2,4-diones, which structurally feature two contiguous

thia-quaternary and tertiary stereogenic centers, with high enantio- and diastereoselectivities (up to >99% ee and >19:1 dr). Several conjugate adducts have been observed to show potential anticancer activities. Moreover, a highly enantioselective sulfenylation of diarylthiazolidin-2,4-diones to *N*-(sulfanyl)-succinimides has been developed, leading to chiral 5-sulfur-5-aryl-disubstituted thiazolidin-2,4-diones and 5-sulfone-5-aryl-disubstituted thiazolidin-2,4-diones. Given our devised expedient synthetic approach to various *5H*-thiazolidin-2,4-diones with highly tunable *N*-substituents, we anticipate that such novel nucleophilic reagents will find application in more kinds of reaction requiring access to diverse chiral 5,5-disubstituted thiazolidinediones with potentially positive biological and pharmaceutical activities.

EXPERIMENTAL SECTION

General information

General Procedures and Methods

Experiments involving moisture and/or air sensitive components were performed under a positive pressure of nitrogen in oven-dried glassware equipped with a rubber septum inlet. Dried solvents and liquid reagents were transferred by oven-dried syringes or hypodermic syringe cooled to ambient temperature in a desiccator. Reactions mixtures were stirred in 10 mL sample vial with Teflon-coated magnetic stirring bars unless otherwise stated. Moisture in non-volatile reagents/compounds was removed in high *vacuo* by means of an oil pump and subsequent purging with nitrogen. Solvents were removed in vacuo under ~30 mmHg and heated with a water bath at 30-35 °C using rotary evaporator with aspirator. The condenser was cooled with running water at 0 °C.

All experiments were monitored by analytical thin layer chromatography (TLC). TLC was performed on pre-coated plates. After elution, plate was visualized under UV illumination at

254 nm for UV active material. Further visualization was achieved by staining $KMnO_4$, ceric molybdate, or anisaldehyde solution. For those using the aqueous stains, the TLC plates were heated on a hot plate.

Columns for flash chromatography (FC) contained silica gel 200-300 mesh. Columns were packed as slurry of *silica gel* in petroleum ether and equilibrated solution using the appropriate solvent system. The elution was assisted by applying pressure of about **2** atm with an air pump.

Instrumentations

Proton nuclear magnetic resonance (¹H NMR) and carbon NMR (¹³C NMR) were recorded in CDCl₃ otherwise stated. ¹H (300 MHz) and ¹³C (75 MHz) were performed on (300 MHz) spectrometers. Chemical shifts are reported in parts per million (ppm), using the residual solvent signal as an internal standard: CDCl₃ (¹H NMR: δ 7.26, singlet; ¹³C NMR: δ 77.0, triplet). Multiplicities were given as: *s* (singlet), *d* (doublet), *t* (triplet), *q* (quartet), *quintet*, *m* (multiplets), *dd* (doublet of doublets), *dt* (doublet of triplets), and *br* (broad). Coupling constants (*J*) were recorded in Hertz (Hz). The number of proton atoms (*n*) for a given resonance was indicated by *n*H. The number of carbon atoms (*n*) for a given resonance was indicated by *n*C. HRMS (analyzer: TOF) was reported in units of mass of charge ratio (m/z). Optical rotations were recorded on a polarimeter with a sodium lamp of wavelength 589 nm and reported as follows; $[\alpha]_{\lambda}^{T^{\circ}C}$ (*c* = g/100 mL, solvent). Melting points were determined on a microscopic melting point apparatus.

Enantiomeric excesses were determined by chiral High Performance Liquid Chromatography (HPLC) analysis. UV detection was monitored at 254 nm. HPLC samples were dissolved in HPLC grade isopropanol (IPA) unless otherwise stated.

<u>Materials</u>

All commercial reagents were purchased with the highest purity grade. They were used without further purification unless specified. All solvents used, mainly petroleum ether (PE) and ethyl acetate (EtOAc) were distilled. Anhydrous DCM and MeCN were freshly distilled

from CaH₂ and stored under N₂ atmosphere. THF, Et₂O, *m*-xylene and toluene were freshly distilled from sodium/benzophenone before use. Anhydrous methanol and etanol were distilled from Mg. All compounds synthesized were stored in a -20 °C freezer and light-sensitive compounds were protected with aluminium foil.

General procedure for the synthesis of 2

A flame-dried 50 mL two-necked round bottomed flask equipped with a reflux condenser, a teflon-coated magnetic stirring bar, a rubber septum, and an inlet adapter with three-way stopcock was charged with **1** (2.77 mmol), NBS (540 mg, 3.05 mmol), and CCl₄ (5.5 mL). To the solution was added AIBN (23 mg, 0.14 mmol). The mixture was heated at reflux for 2 hours, then diluted with hexanes and filtered through a pad of celite. The filtrate was concentrated under reduced pressure. The residue was purified by *silica gel* column chromatography (hexanes-Et₂O = 2:1) to afford **2** as a white solid.

General procedure for the synthesis of 4.

A mixture of **2** (white solid, 2.0 mmol), thiourea (2.0 mmol), sodium acetate (2.0 mmol) and ethanol (10 mL) was stirred under reflux for 5 hours, and concentrated *in vacuo*. The residue was neutralized with saturated aqueous sodium bicarbonate, and Et_2O (10 mL) with hexane (50 mL) were then added. The mixture was stirred at room temperature for 15 minutes, and the imino compounds were collected by filtration.

General procedure for the synthesis of 5

A mixture of **4** (0.1 mmol), 4 *N* HCl (1.0 mL) and ethanol (5.0 mL) was stirred under reflux for 13 hours. The reaction mixture was concentrated *in vacuo*. The residue was diluted with water, neutralized with saturated aqueous sodium bicarbonate and extracted with chloroform. The organic layer was then washed with brine, dried with anhydrous magnesium sulphate and concentrated in vacuo to give the title compounds.

General procedure for the synthesis of 7.

5-Argio-3-(4-methoxyphenyl)thiazolidine-2,4-dione **5** (0.1 mmol, 1.0 equiv), nitroolefin **6** (0.12 mmol, 1.2 equiv), **G** (0.01 mmol, 0.1 equiv), NaCl (0.1 equiv) and 4 Å molecular sieves (50 mg) were dissolved in *m*-xylene (1.0 mL). The reaction mixture was stirred at -20 °C for

72–96 hours and monitored by TLC. Upon complete consumption of **5**, the reaction mixture was concentrated under reduced pressure. The crude material was subsequently purified by flash column chromatography on *silica gel* with PE/EtOAc mixture (20:1–5:1 ratio, the crude material was completely dissolved in CH₂Cl₂/PE before loaded on *silica gel*). After removing the solvent in *vacuo*, the product **7** could be obtained.

Procedure for the synthesis of 9

5-Argio-3-(4-methoxyphenyl)thiazolidine-2,4-dione **5** (0.1 mmol, 1.0 equiv), **8** (0.2 mmol, 2 equiv), **K** (0.01 mmol, 0.1 equiv), and 5 Å molecular sieves (10 mg) were dissolved in CHCl₃ (0.5 mL). The reaction mixture was stirred at -55 °C for 12–24 hours and monitored by TLC. Upon complete consumption of **9**, the reaction mixture was concentrated under reduced pressure, the recovered crude material was subsequently purified by flash column chromatography on silica gel with PE/EtOAc mixture (20:1–5:1 ratio, the crude material was completely dissolved in CH₂Cl₂/PE before loaded on *silica gel*). After removing the solvent in *vacuo*, the product **9** could be obtained.

General procedure for the synthesis of 10

A solution of 9d (210 mg, 0.5 mmol) in dichloromethane (5 mL) was cooled to 0 °C and *m*CPBA (215 mg, 1.25 mmol) was added. After stirring for 10 min, the solution was warmed to room temperature, ant then stirred for two hours. The solvent was removed under vacuum, and the residue was purified by column chromatography on *silica gel* to give 11 as a white solid.

Gerneral methods for procedure of the biological studies

H22, HCT116 and K562 cells were seeded at a density of 4000–5000 cells in 96-well plates. Compounds were added 24 hours after seeding. After 2 days in culture, the MTT stock solution (5 mg/mL in PBS) was added to each well and incubated at 37 °C for 4 hours. The medium was removed carefully, and dimethyl sulfoxide was added to each well to dissolve formazan. The absorbance of each well at 490 nm was measured by using a BioTek microplate reader.

3,5-Diphenylthiazolidine-2,4-dione (5a),^{9d} White solid; Mp 169.2–171.0 °C; 216.3 mg (1

mmol), 80% yield; ¹H NMR (300 MHz, CDCl₃) δ 7.60–7.39 (m, 8H), 7.31 (dd, *J* = 10.0, 3.1, 2H), 5.47 (s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 172.2, 170.4, 134.2, 132.8, 129.5, 129.4 (two peaks), 129.3, 128.3, 127.3, 53.0; HRMS (ESI) m/z 270.0588 (M+H⁺), Calcd for C₁₅H₁₂NO₂S 270.0589.

5-Phenyl-3-(*p*-tolyl)thiazolidine-2,4-dione (5b), White solid; Mp 187.0–188.2 °C; 235.6 mg (1 mmol), 83% yield; ¹H NMR (300 MHz, CDCl₃) δ 7.57–7.38 (m, 5H), 7.28 (t, *J* = 11.7, 2H), 7.16 (d, *J* = 7.8, 2H), 5.43 (s, 1H), 2.40 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 172.3, 170.5, 139.5, 134.3, 130.2, 130.1, 129.3, 129.2, 128.3, 127.0, 53.0, 21.3; HRMS (ESI) m/z 284.0746 (M+H⁺), Calcd for C₁₆H₁₄NO₂S 284.0745.

3-(4-Chlorophenyl)-5-phenylthiazolidine-2,4-dione (5c), White solid; Mp 133.5–134.7 °C; 251.5 mg (1 mmol), 83% yield; ¹H NMR (300 MHz, CDCl₃) δ 7.50–7.31 (m, 6H), 7.16 (d, *J* = 8.9, 3H), 5.37 (s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 171.9, 170.0, 135.3, 133.8, 131.2, 129.7, 129.4, 129.4, 128.6, 128.2, 53.1; HRMS (ESI) m/z 304.0201 (M+H⁺), Calcd for C₁₅H₁₁CINO₂S 304.0199.

3-(3-Chlorophenyl)-5-phenylthiazolidine-2,4-dione (5d), White solid; Mp 125.0–126.3 °C; 227.3 mg (1 mmol), 75% yield; ¹H NMR (300 MHz, CDCl₃) δ 7.55–7.39 (m, 7H), 7.32 (s, 1H), 7.24–7.16 (m, 1H), 5.41 (s, 1H); ¹³C NMR (75 MHz, DMSO) δ 173.0, 170.8, 135.7, 134.3, 132.6, 130.4, 130.1, 129.7, 129.5, 129.4, 129.3, 125.7, 53.1; HRMS (ESI) m/z 304.0197 (M+H⁺), Calcd for C₁₅H₁₁ClNO₂S 304.0199.

5-Phenyl-3-(m-tolyl)thiazolidine-2,4-dione (5e), White solid; Mp 154.3–155.3 °C; 224.4 mg (1 mmol), 79% yield; ¹H NMR (300 MHz, DMSO) δ 6.58 (d, *J* = 6.9, 2H), 6.43 (t, *J* = 8.1, 4H), 6.31 (d, *J* = 7.4, 1H), 6.22 (d, *J* = 12.2, 2H), 4.96 (s, 1H), 1.36 (s, 3H); ¹³C NMR (75 MHz, DMSO) δ 173.2, 171.0, 139.4, 135.8, 133.8, 130.3, 129.6, 129.5, 129.3, 128.8, 125.6, 53.0, 21.2; HRMS (ESI) m/z 284.0743 (M+H⁺), Calcd for C₁₆H₁₄NO₂S 284.0745.

3-(4-Methoxyphenyl)-5-phenylthiazolidine-2,4-dione (5f), White solid; Mp 169.7–171.0 °C; 243.2 mg (1 mmol), 81% yield; ¹H NMR (300 MHz, CDCl₃) δ 7.60–7.35 (m, 5H), 7.23–7.12 (m, 2H), 6.99 (d, *J* = 9.0, 2H), 5.42 (s, 1H), 3.83 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 172.4, 170.6, 160.0, 134.3, 129.4, 129.3, 128.5, 128.3, 125.4, 114.8, 55.6, 53.0; HRMS (ESI) m/z

 $300.0695 (M+H^+)$, Calcd for C₁₆H₁₄NO₃S 300.0694.

3-(4-Methoxyphenyl)-5-(*p*-tolyl)thiazolidine-2,4-dione (5g), White solid; Mp 162.3–163.5 ^oC; 266.1 mg (1 mmol), 85% yield; ¹H NMR (300 MHz, CDCl₃) δ 7.44 (d, *J* = 8.1, 2H), 7.37–7.15 (m, 4H), 7.14–6.85 (m, 2H), 5.47 (s, 1H), 3.91 (s, 3H), 2.46 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 172.6, 170.7, 160.0, 139.3, 131.2, 130.0, 128.5, 128.1, 125.4, 114.7, 55.6, 52.8, 21.2; HRMS (ESI) m/z 314.0852 (M+H⁺), Calcd for C₁₇H₁₆NO₃S 314.0851.

5-(4-(*tert***-Butyl)phenyl)-3-(4-methoxyphenyl)thiazolidine-2,4-dione (5h)**, White solid; Mp 182.9–184.1 °C; 287.6 mg (1 mmol), 81% yield; ¹H NMR (300 MHz, CDCl₃) δ 7.43 (q, *J* = 8.6, 4H), 7.25–7.12 (m, 2H), 7.05–6.87 (m, 2H), 5.41 (s, 1H), 3.83 (s, 3H), 1.33 (s, 9H); ¹³C NMR (75 MHz, CDCl₃) δ 172.7, 170.8, 160.0, 152.4, 131.2, 128.6, 127.9, 126.4, 125.4, 114.7, 55.6, 52.7, 34.7, 31.3; HRMS (ESI) m/z 356.1321 (M+H⁺), Calcd for C₂₀H₂₂NO₃S 356.1320.

5-(4-Chlorophenyl)-3-(4-methoxyphenyl)thiazolidine-2,4-dione (5i), White solid; Mp 169.2–170.4 °C; 259.8 mg (1 mmol), 78% yield; ¹H NMR (300 MHz, CDCl₃) δ 7.42 (s, 4H), 7.17 (d, *J* = 8.5, 2H), 6.99 (d, *J* = 8.4, 2H), 5.40 (s, 1H), 3.83 (s, 3H); ¹³C NMR (75 MHz, *D*₆-acetone) δ 172.3, 170.1, 160.0, 134.6, 134.2, 130.6, 129.1, 126.2, 114.3, 55.0, 51.9; HRMS (ESI) m/z 334.0306 (M+H⁺), Calcd for C₁₆H₁₃ClNO₃S 334.0305.

5-(3-Fluorophenyl)-3-(4-methoxyphenyl)thiazolidine-2,4-dione (5j), White solid; Mp 139.7–141.3 °C; 256.8 mg (1 mmol), 81% yield; ¹H NMR (300 MHz, CDCl₃) δ 7.44 (dd, J = 13.8, 7.9, 2H), 7.25 – 7.10 (m, 4H), 7.02 (d, J = 8.9, 2H), 5.44 (s, 1H), 3.86 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 171.8, 170.1, 164.7, 161.4, 160.1, 136.4, 136.3, 131.0, 130.9, 128.4, 125.2, 124.1, 124.1, 116.5, 116.3, 115.6, 115.3, 114.8, 55.6, 52.4; HRMS (ESI) m/z 318.0601 (M+H⁺), Calcd for C₁₆H₁₃FNO₃S 318.0600.

5-Benzyl-3-(4-methoxyphenyl)thiazolidine-2,4-dione (5k), White solid; Mp 138.2–139.6 °C; 247.3 mg (1 mmol), 79% yield; ¹H NMR (300 MHz, CDCl₃) δ 7.47–7.20 (m, 5H), 6.96 (s, 4H), 4.60 (dd, *J* = 8.2, 3.4, 1H), 3.80 (s, 3H), 3.53–3.26 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 173.5, 170.6, 160.0, 135.3, 129.6, 128.7, 128.5, 127.7, 125.2, 114.7, 55.5, 51.0, 38.6; HRMS (ESI) m/z 314.0852 (M+H⁺), Calcd for C₁₇H₁₆NO₃S 314.0851.

5-Ethyl-3-(4-methoxyphenyl)thiazolidine-2,4-dione (5l),^{9a} White solid; Mp 87.8-88.4 °C;

195.8 mg (1 mmol), 78% yield; ¹H NMR (300 MHz, CDCl₃) δ 7.15 (d, J = 8.6, 2H), 6.99 (d, J = 8.6, 2H), 4.33 (dd, J = 8.0, 4.1, 1H), 3.82 (s, 3H), 2.38–1.95 (m, 2H), 1.12 (t, J = 7.3, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 174.2, 171.1, 159.9, 128.5, 125.3, 114.7, 55.5, 50.9, 26.4, 10.7; HRMS (ESI) m/z 252.0695 (M+H⁺), Calcd for C₁₂H₁₄NO₃S 252.0694.

3-Benzyl-5-phenylthiazolidine-2,4-dione (5m), White solid; Mp 148.8–150.3 °C; 189.6 mg (1 mmol), 67% yield; ¹H NMR (300 MHz, DMSO) δ 7.73–7.03 (m, 10H), 5.96 (s, 1H), 4.76 (s, 2H); ¹³C NMR (75 MHz, DMSO) δ 173.6, 171.4, 136.0, 135.6, 129.5, 129.2, 129.1, 129.1, 128.3, 128.0, 52.8, 45.3; HRMS (ESI) m/z 284.0746 (M+H⁺), Calcd for C₁₆H₁₄NO₂S 284.0745.

(*S*)-5-((*S*)-2-Nitro-1-phenylethyl)-3,5-diphenylthiazolidine-2,4-dione (7a), White solid; Mp 139.4–140.2 °C; 89% ee; dr = 9:1; 36.4 mg (0.1 mmol), 87% yield; $[\alpha]_{D}^{22}$ +27.3 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.05 (d, *J* = 7.5, 2H), 7.70–7.28 (m, 11H), 6.51 (d, *J* = 7.2, 2H), 5.08 – 4.74 (m, 2H), 4.44–4.40 (m, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 172.7, 167.4, 134.3, 132.4, 132.2, 131.6, 130.1, 129.9, 129.6, 129.4, 129.3, 128.9, 128.0, 127.1, 75.7, 69.0, 53.0; HRMS (ESI) m/z 419.1064 (M+H⁺), Calcd for C₂₃H₁₉N₂O₄S 419.1066. The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 25.8 min (minor, major diastereomer), 28.7 min (minor diastereomer), 33.5 min (major, major diastereomer).

(S)-5-((S)-2-Nitro-1-phenylethyl)-5-phenyl-3-(p-tolyl)thiazolidine-2,4-dione. (7b), White solid; Mp 153.3–154.5 °C; 86% ee; dr = 7:1; 35.1 mg (0.1 mmol), 81% yield; $[\alpha]_{D}^{22}$ +24.6 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.05 (d, *J* = 7.4, 2H), 7.66–7.34 (m, 8H), 7.11 (d, *J* = 7.6, 2H), 6.38 (d, *J* = 7.5, 2H), 4.99 – 4.73 (m, 2H), 4.43–4.40 (m, 1H), 2.31 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 172.8, 167.5, 139.7, 134.3, 132.4, 130.1, 129.9, 129.8, 129.6, 129.5, 128.8, 128.0, 126.8, 75.7, 69.0, 53.0, 21.2; HRMS (ESI) m/z 433.1223 (M+H⁺), Calcd for C₂₄H₂₁N₂O₄S 433.1222. The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 21.2

min (minor diastereomer), 32.9 min (minor, major diastereomer), 37.7 min (major, major diastereomer), 42.3 min (minor diastereomer).

(S)-3-(4-Chlorophenyl)-5-((S)-2-nitro-1-phenylethyl)-5-phenylthiazolidine-2,4-dione. (7c), White solid; Mp 144.6–146.0 °C; 82% ee; dr = 6:1; 42.2 mg (0.1 mmol), 94% yield; $[\alpha]_{D}^{22}$ +21.2 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, DMSO) δ 8.01 (d, *J* = 7.2, 2H), 7.56–7.44 (m, 10H), 6.57 (d, *J* = 8.2, 2H), 5.31–5.21 (m, 1H), 4.84 (d, *J* = 11.1, 1H), 4.66 (d, *J* = 13.2, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 172.5, 167.1, 135.5, 134.0, 132.4, 130.5, 130.0, 130.0 129.9, 129.6, 129.5, 128.9, 128.4, 128.0, 75.6, 69.1, 53.0; HRMS (ESI) m/z 453.0673 (M+H⁺), Calcd for C₂₃H₁₈ClN₂O₄S 453.0676. The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 12.9 min (minor, major diastereomer), 17.4 min (minor diastereomer), 29.4 min (major, major diastereomer).

(S)-5-((S)-2-Nitro-1-phenylethyl)-5-phenyl-3-(m-tolyl)thiazolidine-2,4-dione.(7e), White solid; Mp 227.8–228.2 °C; 87% ee; dr = 9:1; 30.8 mg (0.1 mmol), 71% yield; $[\alpha]_D^{22}$ +30.5 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.05 (d, *J* = 7.3, 2H), 7.54–7.26 (m, 8H), 7.23–7.01 (m, 2H), 6.50–6.04 (m, 2H), 5.05–4.69 (m, 2H), 4.48–4.40 (m, 1H), 2.26 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 172.8, 167.4, 139.5, 134.3, 132.5, 132.0, 130.3, 130.1, 129.9, 129.6, 129.5, 129.0, 128.9, 128.0, 127.7, 124.1, 75.7, 69.1, 53.0, 21.1; HRMS (ESI) m/z 433.1223 (M+H⁺), Calcd for C₂₄H₂₁N₂O₄S 433.1222. The ee was determined by HPLC analysis. Nu-Analytical INA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 24.7 min (minor, major diastereomer), 29.3 min (minor diastereomer), 31.3 min (minor diastereomer), 34.4 min (major, major diastereomer).

(*S*)-3-(4-Methoxyphenyl)-5-((*S*)-2-nitro-1-phenylethyl)-5-phenylthiazolidine-2,4-dione (7f), White solid; Mp 144.6–146.0 °C; 97% ee; dr = 16:1 (after a single recrystallization, ee > 99%, dr >19:1); 42.2 mg (0.1 mmol), 94% yield; $[\alpha]_{D}^{22}$ +21.2 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.06 (d, *J* = 7.0,2H), 7.54–7.41 (m, 8H), 6.81 (d, *J* = 8.9, 2H), 6.41 (d, *J* = 8.9, 2H), 5.02–4.75 (m, 2H), 4.43–4.40 (m, 1H), 3.76 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 172.9, 167.7, 160.1, 134.3, 132.5, 130.1, 130.0, 129.6, 129.5, 128.9, 128.3, 128.1, 124.6, 114.6, 75.7, 68.9, 55.5, 53.0; HRMS (ESI) m/z 449.1176 (M+H⁺), Calcd for C₂₄H₂₁N₂O₅S 449.1171. The

ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 29.6 min (minor, major diastereomer), 37.6 min (minor diastereomer), 51.7 min (major, major diastereomer), 56.8 min (minor diastereomer).

(*S*)-5-((*S*)-1-(4-Fluorophenyl)-2-nitroethyl)-3-(4-methoxyphenyl)-5-phenylthiazolidine-2, 4-dione (7g), White solid; Mp 97.3–98.9 °C; 95% ee; dr > 19:1; 41.9 mg (0.1 mmol), 90% yield; $[\alpha]_{p}^{22}$ +20.6 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.03 (dd, *J* = 8.1, 1.3, 2H), 7.68–7.41 (m, 5H), 7.11 (t, *J* = 8.6, 2H), 6.94–6.76 (m, 2H), 6.49 (d, *J* = 8.9, 2H), 4.95–4.68 (m, 2H), 4.41 (dd, *J* = 12.8, 3.5, 1H), 3.77 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 172.9, 167.4, 163.2 (d, 1JC-F = 248.3 Hz), 160.2, 134.2, 131.9 (d, 1JC-F = 8.2 Hz), 129.9, 129.6, 128.1, 127.9, 124.5, 116.0, 115.8, 114.7, 75.7, 68.8, 55.5, 52.2;HRMS (ESI) m/z 467.1072 (M+H⁺), Calcd for C₂₄H₂₀FN₂O₅S 467.1077. The ee was determined by HPLC analysis. Nu-Analytical INA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 41.6 min (minor diastereomer), 43.3 min (minor, major diastereomer), 48.8 min (major, major diastereomer), 70.0 min (minor diastereomer).

(*S*)-5-((*S*)-1-(4-Chlorophenyl)-2-nitroethyl)-3-(4-methoxyphenyl)-5-phenylthiazolidine-2, 4-dione (7h), White solid; Mp 163.1–164.6 °C; 87% ee; dr = 11:1 (after a single recrystallization, ee > 99%, dr >19:1); 42.4 mg (0.1 mmol), 88% yield; $[\alpha]_{D}^{22}$ +19.9 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.05 (dd, *J* = 8.1, 1.4, 2H), 7.64–7.19 (m, 7H), 6.96–6.76 (m, 2H), 6.56–6.43 (m, 2H), 5.00–4.73 (m, 2H), 4.44 (dd, *J* = 12.6, 3.2, 1H), 3.80 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 172.8, 167.3, 160.2, 135.7, 134.1, 131.4, 130.9, 129.9, 129.6, 129.1, 128.0, 124.5, 114.7, 75.5, 68.7, 55.5, 52.4; HRMS (ESI) m/z 483.0778 (M+H⁺), Calcd for C₂₄H₂₀ClN₂O₅S 483.0781. The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; tR = 30.1 min (minor diastereomer), 39.4 min (minor, major diastereomer), 53.4 min (major, major diastereomer), 65.8 min (minor diastereomer).

(*S*)-5-((*S*)-1-(3-Chlorophenyl)-2-nitroethyl)-3,5-diphenylthiazolidine-2,4-dione (7i), White solid; Mp 74.1–75.7 °C; 90% ee; dr = 19:1; 43.4 mg (0.1 mmol), 90% yield; $[\alpha]_{D}^{22}$ +21.2 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.27 (dd, *J* = 8.1, 1.5, 2H), 7.97–7.42 (m, 7H), 7.21–7.02 (m, 2H), 6.90–6.67 (m, 2H), 5.37–4.88 (m, 2H), 4.66 (dd, *J* = 12.8, 3.2, 1H), 4.03 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 172.8, 167.2, 160.2, 134.9, 134.0, 133.4, 132.7, 130.3, 130.0, 129.6, 128.3, 128.2, 128.0, 124.5, 122.8, 114.7, 75.3, 68.5, 55.5, 52.4; HRMS (ESI) m/z 483.0781 (M+H⁺), Calcd for C₂₄H₂₀ClN₂O₅S 483.0782. The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK IE (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 24.6 min (minor, major diastereomer), 31.4 min (minor diastereomer), 35.9 min (minor diastereomer), 41.4 min (major, major diastereomer).

(*S*)-5-((*S*)-1-(3-Bromophenyl)-2-nitroethyl)-3-(4-methoxyphenyl)-5-phenylthiazolidine-2, 4-dione (7j), White solid; Mp 116.7–118.1 °C; 99% ee; dr > 19:1; 45.8 mg (0.1 mmol), 87% yield; $[\alpha]_{p}^{22}$ +20.8 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.27 (dd, *J* = 8.1, 1.5, 2H), 8.00–7.63 (m, 6H), 7.54 (dd, *J* = 13.1, 5.2, 1H), 7.22–6.96 (m, 2H), 6.93–6.51 (m, 2H), 5.17–4.95 (m, 2H), 4.66 (dd, *J* = 12.8, 3.2, 1H), 4.03 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 172.8, 167.2, 160.2, 134.9, 134.0, 133.4, 132.7, 130.3, 130.0, 129.6, 128.3, 128.2, 128.0, 124.5, 122.8, 114.7, 75.3, 68.5, 55.5, 52.4; HRMS (ESI) m/z 527.0285 (M+H⁺), Calcd for C₂₄H₂₀BrN₂O₅S 527.0276. The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK IE (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/5; flow rate 0.5 mL/min; 25 °C; 254 nm t_R = 30.4 min (minor diastereomer), 32.8 min (major, major diastereomer), 55.6 min (minor, major diastereomer), 62.6 min (minor diastereomer).

(*S*)-5-((*S*)-1-(2-Chlorophenyl)-2-nitroethyl)-3-(4-methoxyphenyl)-5-phenylthiazolidine-2, 4-dione (7k), White solid; Mp 156.3–157.1 °C; 88% ee; dr = 16:1 (after a single recrystallization, ee > 99%, dr >19:1); 40.5 mg (0.1 mmol), 84% yield, $[\alpha]_{D}^{22}$ +23.2 (*c* 1.00, CHCl₃);¹H NMR (300 MHz, CDCl₃) δ 8.17–7.98 (m, 2H), 7.67–7.30 (m, 7H), 6.81 (d, *J* = 8.9, 2H), 6.46 (d, *J* = 8.9, 2H), 5.66 (dd, *J* = 11.1, 4.0, 1H), 4.93–4.68 (m, 1H), 4.58 (dd, *J* = 13.2, 4.1, 1H), 3.76 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 172.2, 167.5, 160.2, 137.0, 134.7, 131.7, 130.9, 130.5, 129.9, 129.4, 128.2, 127.2, 124.8, 114.6, 134.9, 68.4, 55.5, 52.9, 47.5; HRMS (ESI) m/z 483.0781 (M+H⁺), Calcd for C₂₄H₂₀ClN₂O₅S 483.0780. The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 13.7 min (minor, major diastereomer), 14.7 min (major, major diastereomer), 22.8 min (minor diastereomer).

(S)-3-(4-methoxyphenyl)-5-((S)-2-nitro-1-(p-tolyl)ethyl)-5-phenylthiazolidine-2,4-dione

(71), White solid; Mp 159.5–161.1 °C; 92% ee; dr = 14:1 (after a single recrystallization, ee = 94%, dr = 19:1); 41.1 mg (0.1 mmol), 89% yield; $[\alpha]_{D}^{22}$ +24.0 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.09–7.85 (m, 2H), 7.59–7.26 (m, 5H), 7.28–7.09 (m, 2H), 6.78 (d, *J* = 9.0, 2H), 6.41 (t, *J* = 6.0, 2H), 4.95–4.66 (m, 2H), 4.37–4.34 (m, 1H), 3.75 (s, 3H), 2.33 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 173.0, 167.8, 160.1, 139.5, 134.4, 129.9, 129.8, 129.5, 129.3, 128.3, 128.1, 124.7, 114.5, 75.8, 69.1, 55.5, 52.8, 21.2; HRMS (ESI) m/z 463.1321 (M+H⁺), Calcd for C₂₅H₂₃N₂O₅S 463.1328. The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK IE (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R=46.6 min (minor, major diastereomer), 50.4 min (minor diastereomer), 69.0 min (major, major diastereomer), 82.3 min (minor diastereomer).

(*S*)-3-(4-Methoxyphenyl)-5-((*S*)-2-nitro-1-(m-tolyl)ethyl)-5-phenylthiazolidine-2,4-dione (7m), White solid; Mp 143.6–145.2 °C; 90% ee; dr = 11:1 (after a single recrystallization, ee = 93%, dr >19:1); 44.8 mg (0.1 mmol), 97% yield; $[\alpha]_{D}^{22}$ +25.0 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.01 (d, *J* = 7.0, 2H), 7.59–7.38 (m, 3H), 7.40–7.11 (m, 4H), 6.78 (d, *J* = 8.9, 2H), 6.39 (d, *J* = 8.9, 2H), 4.98–4.69 (m, 2H), 4.41–4.34 (m, 1H), 3.72 (s, 3H), 2.33 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 172.9, 167.8, 160.1, 138.6, 134.4, 132.4, 130.9, 130.2, 129.8, 129.5, 128.7, 128.3, 128.2, 126.8, 124.7, 114.5, 75.7, 69.0, 55.5, 53.0, 21.5; HRMS (ESI) m/z 463.1318 (M+H⁺), Calcd for C₂₅H₂₃N₂O₅S 463.1328. The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 22.9 min (minor,

major diastereomer), 33.2 min (minor diastereomer), 38.9 min (major, major diastereomer), 44.5 min (minor diastereomer).

(*S*)-3-(4-Methoxyphenyl)-5-((*S*)-2-nitro-1-(o-tolyl)ethyl)-5-phenylthiazolidine-2,4-dione (7n), White solid; Mp 142.9–144.0 °C; 99% ee; dr > 19:1; 42.5 mg (0.1 mmol), 92% yield; $[\alpha]_{D}^{22}$ +20.4 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.13–7.86 (m, 3H), 7.61–7.46 (m, 3H), 7.36–7.27 (m, 3H), 6.89–6.67 (m, 2H), 6.65–6.09 (m, 2H), 5.24 (dd, *J* = 11.3, 3.8, 1H), 4.87 (dd, *J* = 13.0, 11.3, 1H), 4.48 (dd, *J* = 13.1, 3.8, 1H), 3.74 (s, 3H), 2.59 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 173.0, 167.8, 160.1, 139.6, 135.1, 131.8, 129.8, 129.5, 129.4, 129.2, 128.5, 128.3, 128.2, 129.5, 114.6, 68.9, 55.5, 52.9, 47.2, 20.1; HRMS (ESI) m/z 485.1148 (M+Na⁺), Calcd for C₂₅H₂₃N₂O₅S 485.1147. The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK IE (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 27.1 min (major, major diastereomer), 31.6 min (minor, major diastereomer), 55.6 min (minor diastereomer).

(*S*)-3-(4-Methoxyphenyl)-5-((*S*)-1-(4-methoxyphenyl)-2-nitroethyl)-5-phenylthiazolidine-2,4-dione (70), White solid; Mp 81.0–82.3 °C; 93% ee; dr = 12:1 (after a single recrystallization, ee > 99%, dr >19:1); 44.0 mg (0.1 mmol), 92% yield; $[\alpha]_{D}^{22}$ +21.7 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.04 (dd, *J* = 8.1, 1.2, 2H), 7.65–7.36 (m, 5H), 7.06–6.70 (m, 4H), 6.66–6.27 (m, 2H), 4.85–4.79 (m, 2H), 4.40–4.36 (m, 1H), 3.82 (s, 3H), 3.76 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 173.0, 167.8, 160.4, 160.1, 134.4, 131.2,129.8, 129.6, 129.5, 128.3, 128.0, 124.7, 124.1, 114.2, 75.8, 69.2, 55.5, 55.4, 52.4; HRMS (ESI) m/z 479.1270 (M+H⁺), Calcd for C₂₅H₂₃N₂O₆S 479.1277. The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK ID-3 (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 36.7 min (minor, major diastereomer), 46.4 min (minor diastereomer), 52.3 min (major, major diastereomer), 65.5 min (minor diastereomer).

(*S*)-3-(4-Methoxyphenyl)-5-((*R*)-2-nitro-1-(thiophen-2-yl)ethyl)-5-phenylthiazolidine-2,4dione (7p), White solid; Mp 105.3–106.6 °C; 74% ee; dr > 19:1; 41.3 mg (0.1 mmol), 91% yield; $[\alpha]_D^{22}$ +18.4 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.17–7.98 (m, 2H), 7.62–7.37 (m, 4H), 7.34–7.20 (m, 1H), 7.09 (dd, *J* = 5.1, 3.6, 1H), 6.96–6.78 (m, 2H), 6.67–6.43 (m, 2H), 5.28 (dd, *J* = 11.3, 3.5, 1H), 4.74 (dd, *J* = 12.9, 11.4, 1H), 4.43 (dd, *J* = 13.0, 3.5, 1H), 3.79 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 172.9, 167.8, 160.2, 134.6, 133.8, 130.3, 130.0, 129.6, 128.3, 128.0, 127.2, 126.9, 124.7, 114.7, 68.9, 55.5, 49.5; HRMS (ESI) m/z 455.0727 (M+H⁺), Calcd for C₂₂H₁₉N₂O₅S₂ 455.0735. The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 13.7 min (minor diastereomer), 22.8 min (minor diastereomer), 31.2 min (minor, major diastereomer), 35.3 min (major, major diastereomer).

(*S*)-5-((*S*)-1-Cyclohexyl-2-nitroethyl)-3-(4-methoxyphenyl)-5-phenylthiazolidine-2,4-dion e (7q), White solid; Mp 126.0–127.2 °C; 96% ee; dr = 5:1; 28.6 mg (0.1 mmol), 63% yield; $[\alpha]_D^{22}$ –25.1 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.97–7.80 (m, 2H), 7.54–7.34 (m, 3H), 7.13–6.88 (m, 4H), 4.33–4.15 (m, 2H), 3.81 (s, 3H), 3.70–3.47 (m, 1H), 1.98–1.57 (m, 6H), 1.29–1.10 (m, 4H); ¹³C NMR (75 MHz, CDCl₃) δ 174.5, 169.2, 160.1, 135.8, 129.7, 129.4, 128.3, 127.8, 125.1, 114.8, 73.9, 69.1, 55.6, 50.5, 40.5, 33.8, 30.0, 27.1, 26.6, 25.8; HRMS (ESI) m/z 455.1649 (M+H⁺), Calcd for C₂₄H₂₇N₂O₅S 455.1641. The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 12.8 min (minor diastereomer), 17.4 min (major, major diastereomer), 18.9 min (minor, major diastereomer), 22.3 min (minor diastereomer).

(S)-3-(4-Methoxyphenyl)-5-((S)-2-nitro-1-phenylethyl)-5-(p-tolyl)thiazolidine-2,4-dione

(7r), White solid; Mp 108.6–110.1 °C; 90% ee; dr = 13:1; 41.1 mg (0.1 mmol), 89% yield; $[\alpha]_{D}^{22}$ +21.6 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.91 (d, *J* = 8.4, 2H), 7.60–7.12 (m, 7H), 6.90–6.67 (m, 2H), 6.56–6.24 (m, 2H), 4.99–4.76 (m, 2H), 4.44–4.41 (m, 1H), 3.75 (s, 3H), 2.40 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 173.0, 167.7, 160.1, 140.0, 132.5, 131.3,

130.2, 130.1, 129.4, 128.8, 128.3, 127.9, 124.9, 114.5, 75.7, 68.8, 55.5, 52.9, 21.0; HRMS (ESI) m/z 463.1335 (M+H⁺), Calcd for $C_{25}H_{23}N_2O_5S$ 463.1328. The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK ID-3 (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 70/30; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 29.1 min (minor, major diastereomer), 45.5 min (minor diastereomer), 57.9 min (major, major diastereomer), 82.4 min (minor diastereomer).

(S) - 5 - (4 - (tert - Butyl) phenyl) - 3 - (4 - methoxyphenyl) - 5 - ((S) - 2 - nitro - 1 - phenylethyl) thiazolidi

ne-2,4-dione (7s), White solid; Mp 108.6–110.2 °C; 87% ee; dr = 12:1; 43.8 mg (0.1 mmol), 87% yield; $[\alpha]_{D}^{22}$ +23.6 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.96 (d, *J* = 8.7, 2H), 7.61–7.37 (m, 7H), 6.87–6.68 (m, 2H), 6.51–6.21 (m, 2H), 5.05–4.74 (m, 2H), 4.46–4.43 (m, 1H), 3.75 (s, 3H), 1.37 (s, 9H); ¹³C NMR (75 MHz, CDCl₃) δ 173.0, 167.8, 160.2, 153.0, 132.5, 131.2, 130.1, 129.4, 128.8, 128.3, 127.7, 126.5, 124.7, 114.5, 75.7, 68.7, 55.5, 52.8, 34.7, 31.2; HRMS (ESI) m/z 505.1801 (M+H⁺), Calcd for C₂₈H₂₉N₂O₅S 505.1797. The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 8.3 min (minor, major diastereomer), 11.1 min (minor diastereomer), 13.7 min (major, major diastereomer), 41.3 min (minor diastereomer).

(*S*)-5-(Benzylthio)-3,5-diphenylthiazolidine-2,4-dione (9a), Colorless oil; 87% ee; 33.2 mg (0.1 mmol), 85% yield; $[\alpha]_{1D}^{22}$ -25.1 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.65 (d, *J* = 7.3, 2H), 7.53–7.25 (m, 6H), 7.18 (dd, *J* = 17.3, 8.8, 7H), 4.00 (d, *J* = 11.8, 1H), 3.66 (d, *J* = 11.8, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 172.0, 167.8, 135.6, 134.8, 132.8, 129.5, 129.4, 129.4, 129.4, 129.2, 128.8, 127.7, 127.5, 127.4, 66.3, 37.6; HRMS (ESI) m/z 392.0780 (M+H⁺), Calcd for C₂₂H₁₈NO₂S₂ 392.0779. The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 12.8 min (minor), 20.0 min (major).

(*S*)-5-(Benzylthio)-3-(4-chlorophenyl)-5-phenylthiazolidine-2,4-dione (9b), Colorless oil; 88% ee; 37.4 mg (0.1 mmol), 88% yield; $[\alpha]_{D}^{22}$ –20.8 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃)

δ 7.63 (d, J = 7.4, 2H), 7.44–7.24 (m, 5H), 7.23–6.99 (m, 7H), 3.96 (d, J = 12.0, 1H), 3.65 (d, J = 12.0, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 171.7, 167.6, 135.4, 135.3, 134.8, 131.1, 129.7, 129.5, 129.4, 129.2, 128.8, 128.7, 127.8, 127.5, 66.3, 37.7; HRMS (ESI) m/z 426.0390 (M+H⁺), Calcd for C₂₂H₁₇ClNO₂S₂ 426.0389. The ee was determined by HPLC analysis. CHIRALPAK IF (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 8.0 min (minor), 10.4 min (major).

(*S*)-5-(Benzylthio)-5-phenyl-3-(*p*-tolyl)thiazolidine-2,4-dione (9c), Colorless oil; 90% ee; 36.8 mg (0.1 mmol), 91% yield; $[\alpha]_{D}^{22}$ –21.9 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.66 (d, *J* = 7.4, 2H), 7.46–7.27 (m, 5H), 7.20–7.15 (m, 5H), 7.05 (d, *J* = 7.9, 2H), 4.00 (d, *J* = 11.8, 1H), 3.66 (d, *J* = 11.8, 1H), 2.31 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 172.1, 167.9, 139.7, 135.7, 134.9, 130.2, 130.1, 129.4, 129.3, 129.2, 128.7, 127.7, 127.5, 127.1, 66.3, 37.6, 21.3; HRMS (ESI) m/z 406.0934 (M+H⁺), Calcd for C₂₃H₂₀NO₂S₂ 406.0935. The ee was determined by HPLC analysis. CHIRALPAK IF (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 10.2 min (minor), 12.6 min (major).

(*S*)-5-(Benzylthio)-3-(4-methoxyphenyl)-5-phenylthiazolidine-2,4-dione (9d), Colorless oil; 91% ee; 41.3 mg (0.1 mmol), 98% yield; $[\alpha]_{D}^{22}$ –24.2 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.65 (d, *J* = 7.4, 2H), 7.42–7.23 (m, 3H), 7.22–6.99 (m, 7H), 6.91 (d, *J* = 8.3, 2H), 4.00 (d, *J* = 11.8, 1H), 3.70 (d, *J* = 17.5, 3H), 3.66 (d, *J* = 11.8, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 172.2, 168.1, 160.2, 135.7, 134.9, 129.4, 129.4, 129.2, 128.8, 128.6, 127.7, 127.5, 125.4, 114.8, 66.2, 55.6, 37.6; HRMS (ESI) m/z 422.0884 (M+H⁺), Calcd for C₂₃H₂₀NO₃S₂ 422.0885. The ee was determined by HPLC analysis. CHIRALPAK IF (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 11.6 min (minor), 16.6 min (major).

(S)-5-(Benzylthio)-5-(4-(tert-butyl)phenyl)-3-(4-methoxyphenyl)thiazolidine-2,4-dione

(9e), Colorless oil; 87% ee; 39.6 mg (0.1 mmol), 83% yield; $[\alpha]_{D}^{22}$ -20.1 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.67 (d, *J* = 7.7, 2H), 7.44 (d, *J* = 7.8, 2H), 7.26 (s, 5H), 7.19 (d, *J* = 7.9, 2H), 7.02 (d, *J* = 8.1, 2H), 4.10 (d, *J* = 11.9, 1H), 3.85 (s, 3H), 3.80 (s, 1H), 1.35 (s, 9H);

¹³C NMR (75 MHz, CDCl₃) δ 172.3, 168.2, 160.2, 152.5, 135.1, 132.6, 129.4, 128.7, 128.6, 127.6, 127.2, 126.1, 125.5, 114.8, 66.2, 55.6, 37.6, 34.7, 31.2; HRMS (ESI) m/z 478.1517 (M+H⁺), Calcd for C₂₇H₂₈NO₃S₂ 478.1511. The ee was determined by HPLC analysis. CHIRALPAK IF (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 9.3 min (minor), 11.5 min (major).

(S)-5-(Benzylthio)-5-(4-chlorophenyl)-3-(4-methoxyphenyl)thiazolidine-2,4-dione (9f), Colorless oil; 88% ee; 40.0 mg (0.1 mmol), 88% yield; $[\alpha]_{D}^{22}$ –31.3 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.60 (d, *J* = 8.2, 2H), 7.29 (d, *J* = 8.3, 2H), 7.28–6.94 (m, 7H), 6.92 (d, *J* = 8.5, 2H), 3.99 (d, *J* = 12.0, 1H), 3.76 (s, 3H), 3.69 (d, *J* = 12.0, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 171.9, 167.6, 160.2, 135.4, 134.7, 134.2, 129.3, 129.2, 129.0, 128.8, 128.5, 127.7, 125.3, 114.8, 65.6, 55.6, 37.7; HRMS (ESI) m/z 456.0492 (M+H⁺), Calcd for C₂₃H₁₉ClNO₃S₂ 456.0495. The ee was determined by HPLC analysis. CHIRALPAK IF (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 15.0 min (minor), 16.8 min (major).

(S)-5-(Benzylthio)-5-(3-fluorophenyl)-3-(4-methoxyphenyl)thiazolidine-2,4-dione (9g), Colorless oil; 89% ee; 41.7 mg (0.1 mmol), 95% yield; $[\alpha]_D^{22}$ -29.1 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.41 (s, 2H), 7.29 (d, *J* = 7.7, 1H), 7.21–7.03 (m, 7H), 7.03–6.85 (m, 3H), 3.99 (d, *J* = 11.9, 1H), 3.75 (s, 3H), 3.69 (d, *J* = 11.9, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 171.8, 167.6, 162.8 (d, 1JC-F = 246.4 Hz), 160.2, 138.1 (d, 1JC-F = 7.3 Hz), 134.7, 130.7 (d, 1JC-F = 8.2 Hz), 129.3, 128.8, 128.5, 127.7, 125.2, 123.3 (d, 1JC-F = 3.0 Hz), 114.6 (d, 1JC-F = 21.0 Hz), 115.0 (d, 1JC-F = 24.2 Hz), 114.8, 65.4, 55.6, 37.7; HRMS (ESI) m/z 440.0791 (M+H⁺), Calcd for C₂₃H₁₉FNO₃S₂ 440.0790. The ee was determined by HPLC analysis. CHIRALPAK IF (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 10.4 min (minor), 12.8 min (major).

(S)-5-((4-Chlorobenzyl)thio)-3-(4-methoxyphenyl)-5-phenylthiazolidine-2,4-dione (9h), Colorless oil; 88% ee; 43.6 mg (0.1 mmol), 96% yield; $[\alpha]_{D}^{22}$ -30.2 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.62 (d, *J* = 7.0, 2H), 7.31 (d, *J* = 6.8, 3H), 7.19–7.00 (m, 6H), 6.92 (d, *J*

= 8.3, 2H), 3.94 (d, J = 12.3, 1H), 3.75 (s, 3H), 3.65 (d, J = 12.2, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 172.2, 167.8, 160.2, 135.6, 133.6, 133.5, 130.7, 129.3, 129.2, 128.9, 128.5, 127.5, 125.3, 114.8, 66.0, 55.6, 37.0; HRMS (ESI) m/z 456.0490 (M+H⁺), Calcd for C₂₃H₁₉CINO₃S₂ 456.0495. The ee was determined by HPLC analysis. CHIRALPAK IF (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 13.1 min (minor), 25.8 min (major).

(*S*)-3-(4-Methoxyphenyl)-5-((4-methylbenzyl)thio)-5-phenylthiazolidine-2,4-dione (9i), Colorless oil; 87% ee; 42.2 mg (0.1 mmol), 99% yield; $[\alpha]_{D}^{22}$ –10.0 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.77 (d, *J* = 7.0, 2H), 7.44 (d, *J* = 7.9, 3H), 7.14 (dt, *J* = 16.1, 8.1, 6H), 7.02 (d, *J* = 7.5, 2H), 4.08 (d, *J* = 11.6, 1H), 3.85 (s, 3H), 3.75 (d, *J* = 11.9, 1H), 2.33 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 170.3, 166.2, 158.2, 135.5, 133.9, 129.7, 127.5, 127.4, 127.2, 126.7, 125.6, 123.5, 112.8, 64.4, 53.7, 35.4, 19.2; HRMS (ESI) m/z 436.1042 (M+H⁺), Calcd for C₂₄H₂₂NO₃S₂ 436.1041. The ee was determined by HPLC analysis. CHIRALPAK IF (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 13.7 min (minor), 29.3 min (major).

(S)-3-(4-Methoxyphenyl)-5-((3-methylbenzyl)thio)-5-phenylthiazolidine-2,4-dione (9j),

Colorless oil; 89% ee; 42.6 mg (0.1 mmol), 98% yield; $[\alpha]_D^{22}$ –19.4 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.77 (d, *J* = 7.0, 2H), 7.52 – 7.34 (m, 3H), 7.20 (d, *J* = 8.8, 3H), 7.03 (d, *J* = 10.0, 5H), 4.08 (d, *J* = 11.7, 1H), 3.86 (s, 3H), 3.74 (d, *J* = 11.8, 1H), 2.31 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 170.3, 166.2, 158.2, 136.5, 133.8, 132.8, 128.2, 127.4, 127.2, 126.7, 126.7, 126.5, 125.6, 124.5, 123.5, 112.8, 64.4, 53.7, 35.6, 19.4; HRMS (ESI) m/z 436.1037 (M+H⁺), Calcd for C₂₄H₂₂NO₃S₂ 436.1041. The ee was determined by HPLC analysis. CHIRALPAK IF (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 13.9 min (minor), 17.4 min (major).

(*S*)-3-(4-Methoxyphenyl)-5-phenyl-5-(phenylthio)thiazolidine-2,4-dione (9k), Colorless oil; 12% ee; 32.5 mg (0.1 mmol), 80% yield; $[\alpha]_{D}^{22}$ -5.1 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.86 (d, *J* = 7.2, 2H), 7.70 (d, *J* = 7.5, 2H), 7.60–7.33 (m, 6H), 6.88 (d, *J* = 8.5, 2H),

6.66 (d, J = 8.4, 2H), 3.80 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 171.6, 167.9, 160.0, 137.7, 135.5, 131.1, 129.5, 129.3, 129.1, 128.4, 127.7, 125.0, 114.6, 72.2, 55.5; HRMS (ESI) m/z 408.0729 (M+H⁺), Calcd for C₂₂H₁₈NO₃S₂ 408.0728. The ee was determined by HPLC analysis. CHIRALPAK IF (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 13.3 min (major), 16.3 min (minor).

(*R*)-5-(Benzylsulfonyl)-3-(4-methoxyphenyl)-5-phenylthiazolidine-2,4-dione (10), White solid; Mp 133.1–134.5 °C; 91% ee; 183.5 mg (0.5 mmol), 81% yield; $[\alpha]_{D}^{22}$ –21.1 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.77 (s, 2H), 7.45 (s, 3H), 7.29–7.02 (m, 8H), 6.98 (d, J = 8.2, 2H), 4.66 (d, J = 12.9, 1H), 3.92 (d, J = 12.9, 1H), 3.80 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 168.4, 167.4, 160.6, 131.5, 130.7, 130.6, 129.5, 129.2, 128.8, 128.7, 128.3, 125.0, 124.8, 115.0, 83.8, 55.6, 55.2; HRMS (ESI) m/z 454.0705 (M+H⁺), Calcd for C₂₃H₂₀NO₅S₂ 454.0703; The ee was determined by HPLC analysis. CHIRALPAK IF (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 22.6 min (minor), 30.1 min (major).

ASSOCIATED CONTENT

Supporting Information.

General information, optimization of the reaction conditions of sulfenylation, determination of the absolute configuration by X-ray crystallography, copies of HPLC and NMR Spectra. This material is available free of charge via the Internet at <u>http://pubs.acs.org</u>.

AUTHOR INFORMATION

Corresponding Author

*E-mail: chmjzy@henu.edu.cn

[§]L.J. and L.B. made equal contributions to this work

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

We are grateful for the grants from NSFC (No. 21072044, 21672052) and Henan province

(14IRTSTHN006, 152300410057, 162300410002).

REFERENCES

- (1) (a) Lesyk, R. B.; Zimenkovsky, B. S. Curr. Org. Chem. 2004, 8, 1547. (b) Murugan, R.; Anbazhagan, S.; Narayanan, S. S. Eur. J. Med. Chem. 2009, 44, 3272. (c) Tomašič, L. P. Curr. Med. Chem. 2009, 16, 1596. (d) Mohler, D. L.; Shen, G.; Dotse, A. K. Bioorg. Med. Chem. Lett. 2000, 10, 2239. (e) Giles, R. G.; Lewis, N. J.; Quick, J. K.; Sasse, M. J.; Urguhart, M. W. J.; Youssef, L. Tetrahedron 2000, 56, 4531. (f) Kaminskyy, D.; Zimenkovsky, B.; Lesyk, R. Eur. J. Med. Chem. 2009, 44, 3627.
- (2) Saltiel, A. R.; Olefsky, J. M. Diabetes 1996, 45, 1661.
- (3) (a) Mandal, M.; Tang, H.; Xiao, L.; Su, J.; Li, G.; Yang, S.-W.; Pan, W.; Tang, H.; Dejesus, R.; Hicks, J.; Lombardo, M.; Chu, H.; Hagmann, W.; Pasternak, A.; Gu, X.; Jiang, J.; Dong, S.; Ding, F.-X.; London, C.; Biswas, D.; Young, K.; Hunter, D. N.; Zhao, Z.; Yang, D.; *PCT Int. Appl.* 2015, WO 2015112441 A1 20150730. (b) York, B. M. Jr. *U.S.* 1991, US 5070100 A 19911203. (c) Iwataki, I.; Rudchenko, V. F.; Kurz, T.; Semer, C. R.; Kucharek, T. A.; Geffken, D. *U.S.* 2002, US 6476055 B1 20021105. (d) Mihm, G.; Hauel, N.; Ries, U.; Meel, J. van; Wienen, W.; Entzeroth, M. Ger. *Offen*, 1995, DE 4408497 A1 19950921. (e) Desai, R. C.; Han, W.; Metzger, E. J.; Bergman, J. P.; Gratale, D. F.; MacNaul, K. L.; Berger, J. P.; Doebber, T. W.; Leung, K.; Moller, D. E.; Heck, J. V.; Sahoo, S. P. *Bioorg. Med. Chem. Lett.* 2013, *13*, 2795. (f) Falk, K.; Roetzschke, O. *PCT Int. Appl.* 2006, WO 2006029891 A2 20060323. (g) Pedras, M. S. C.; Hossain, M. Org. *Biomol. Chem.* 2006, *4*, 2581. (h) Pedras, M. S. C.; Hossain, *Bioorg. Med. Chem.* 2007, *15*, 5981.
- (4) (a) Wheeler, H. L. Am. Chem. J. 1901, 345. (b) Wheeler, H. L.; Johnson, T. B. Am. Chem. J. 1902, 680.

- (5) Hamersma, J. A. M.; Speckamp, W. N. Tetrahedron 1982, 38, 3255.
- (6) (a) Han, Z.; Chen, W.; Dong, S.; Yang, C.; Liu, H.; Pan, Y.; Yan, L.; Jiang, Z. Org. Lett. 2012, 14, 4670. (b) Qiao, B.; Liu, X.; Duan, S.; Yan, L.; Jiang, Z. Org. Lett. 2014, 16, 672.
 (c) Huang, L.; Li, J.; Zhao, Y.; Ye, X.; Liu, Y.; Yan, L.; Tan, C.-H.; Liu, H.; Jiang, Z. J. Org. Chem. 2015, 80, 8933. (d) Zhu, B.; Qiu, S.; Li, J.; Coote, M. L.; Lee, R.; Jiang, Z. Chem. Sci. 2016, 7, 6060.
- (7) (a) Zhang, W.; Tan, D.; Lee, R.; Tong, G.; Chen, W.; Qi, B.; Huang, K.-W.; Tan, C.-H.; Jiang, Z. Angew. Chem. Int. Ed. 2012, 51, 10069. (b) Zhu, B.; Zhang, W.; Lee, R.; Han, Z.; Yang, W.; Tan, D.; Huang, K.-W.; Jiang, Z. Angew. Chem. Int. Ed. 2013, 52, 6666. (c) Qiao, B.; An, Y.; Liu, Q.; Yang, W.; Liu, H.; Shen, J.; Yan, L.; Jiang, Z. Org. Lett. 2013, 15, 2358. (d) Bai, X.; Jing, Z.; Liu, Q.; Ye, X.; Zhang, G; Zhao, X.; Jiang, Z. J. Org. Chem. 2015, 80, 12686. (e) Zhu, B.; Lee, R.; Li, J.; Ye, X.; Hong, S.-N.; Qiu, S.; Coote, M. L.; Jiang, Z. Angew. Chem. Int. Ed. 2016, 55, 1299. (f) Jiao, L.; Zhao, X.; Liu, H.; Ye, X.; Li, Y.; Jiang, Z. Org. Chem. Front. 2016, 3, 470.
- (8) (a) Taylor, E. C.; Berchtold, G. A.; Goeckner, N. A.; Stroehmann, F. G. J. Org. Chem. 1961, 26, 2715. (b) Taylor, E. E. Jr.; Wolinsky, J.; Lee, H.-H. J. Am. Chem. Soc. 1964, 76, 1866. (c) Potts, K. T.; Chen, S. J.; Kane, J.; Marshall, J. L.; J. Org. Chem. 1977, 42, 1633. (d) Majcen Le Maréchal, A.; Robert, A.; Leban, I. Tetrahedron 1990, 46, 453.
- (9) See the supplemental materials.
- (10) CCDC1430350 (7f) and CCDC1489694 (11) contain the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif..
- (11) For selected examples on asymmetric sulfenylation: (a) Marigo, M.; Wabnitz, T. C.; Fielenbach, D.; Jørgensen, K. A. Angew. Chem. Int. Ed. 2005, 44, 794. (b) Franzén, J.; Marigo, M.; Fielenbach, D.; Wabnitz, T. C.; Kjærsgaard, A.; Jørgensen, K. A. J. Am. Chem. Soc. 2005, 127, 18296. (c) Liao, K.; Zhou, F.; Yu, J.-S.; Gao, W.-M.; Zhou, J. Chem. Commun. 2015, 51, 16255. For selected reviews, see: (d) Zhao, X.; Shen, J.; Jiang, Z. Mini-Rev. Org. Chem. 2014, 11, 424. (e) Chauhan, P.; Mahajan, S.; Enders, D. Chem.

Rev. 2014, *114*, 8807.

- (12) Salaski, E. J.; Salaski, E. J.; Ayral-Kaloustian, S.; Epstein, J. W. PCT Int. Appl. 2003, WO 2003018011 A1 20030306.
- (13) For the selected reviews, see: (a) Chai, Z.; Zhao, G.; *Catal. Sci. Technol.* 2012, *2*, 29. (b)
 Serdyuk, O. V.; Heckel, C. M.; Tsogoeva, S. B. *Org. Biomol. Chem.* 2013, *11*, 7051. (c)
 Zhao, X.; Zhu, B.; Jiang, Z. *Synlett* 2015, *26*, 2216.
- (14) Vallejos, J. C.; Legrand, O.; Christidis, Y. Bull. Soc. Chim. Fr. 1997, 134, 101.

catalyst <mark>G</mark>

catalyst <mark>K</mark>

ЮМе

