# TRITERPENOIDS FROM EUPHORBIA MACULATA

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Key Word Index—Euphorbia maculata; Euphorbiaceae;  $\beta$ -amyrin acetate; taraxeryl acetate; lupenyl acetate;  $3\beta$ -acetoxy-30-norlupan-20-one;  $\alpha$ -amyrenonol; gult-5-en-3 $\beta$ -yl acetate; ursa-9 (11):12-dien-3 $\beta$ -ol; sitosterol.

Abstract—Two new triterpenoids were isolated together with  $\beta$ -amyrin acetate, taraxeryl acetate, lupenyl acetate,  $3\beta$ -acetoxy-30-norlupan-20-one,  $\alpha$ -amyrenonol and sitosterol from the whole herb of *Euphorbia maculata*. The structures of the new compounds were characterized as gult-5-en-3 $\beta$ -yl acetate and ursa-9(11):12-dien-3 $\beta$ -ol on the basis of chemical and spectral evidence.

### INTRODUCTION

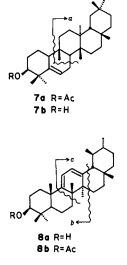
Euphorbia maculata L., an annual weed native to North America, is widely distributed in Japan. Its milky-white latex causes similar skin irritation to that of *E. supina*, belonging to the same genus [1]. Up to now, nothing has been reported about the biological activity or chemistry of *E. maculata*, though *E. supina* is used as a folk medicine for treatment of gastroenteric diseases such as diarrhoea and for healing purulent swellings [2, 3]. An examination of the less polar neutral extract of *E. maculata* led to the isolation of two new triterpene compounds along with five known triterpenoids and sitosterol. This paper describes the characterization of these compounds.

## **RESULTS AND DISCUSSION**

The neutral benzene extract of the dried whole herb afforded the known compounds,  $\beta$ -amyrin acetate (1), taraxeryl acetate (2), lupenyl acetate (3),  $3\beta$ -acetoxy-30norlupan-20-one (4),  $\alpha$ -amyrenonol (5) and sitosterol (6), and two new triterpenoids 7a and 8a. Compounds 1–3 and 6 were identified by direct comparison with authentic samples, 4 and 5 were confirmed by syntheses of the corresponding compounds, respectively (see Experimental). Compound 4 had been isolated previously from *Claoxylon polot* (Euphorbiaceae) [4] while 5 had been obtained from *Ilex buergeri* [5], *I. goshiensis* (Aquifoliaceae) [6] and *Canarium zeylanicum* (Burceraceae) [7].

Compound 7a,  $C_{32}H_{52}O_2$  (M<sup>+</sup> at m/z 468), had eight Me groups (<sup>1</sup>H NMR:  $\delta 0.85-1.16$ ), an axial acetoxy group geminal to a methine proton (IR: 1723 and 1238 cm<sup>-1</sup>; <sup>1</sup>H NMR:  $\delta 2.01$  (3H, s) and 4.69 (1H, t, J = 2.9 Hz)) and a trisubstituted double bond (<sup>1</sup>H NMR:  $\delta 5.56$  (1H, dt like); <sup>13</sup>C NMR:  $\delta 120.01$  (d) and 141.94 (s)). The EI-mass spectrum of 7a contained strong peaks due to retro-Diels-Alder cleavage of the B-ring at m/z 274 (fragment a) and 259 [a – Me] characteristic of a triterpene-5-ene skeleton [8]. These data suggested that 7a may be gult-5-en- $3\beta$ -yl acetate. This was confirmed in the following way. Alkaline hydrolysis of 7a furnished the alcohol 7b identical in all respects with an authentic sample.

Although gult-5-en-3 $\beta$ -ol as well as gult-5-en-3-one have isolated from natural sources including *Euphorbia* 



species [9-12], there appears to be no prior report of the acetate in nature.

Compound 8a,  $C_{30}H_{48}O$  (M<sup>+</sup> at m/z 424), was a triterpene dienol which had a hydroxyl group (IR: 3360 and  $1032 \text{ cm}^{-1}$ ) and a homoannular diene chromophore  $(UV: \lambda_{max} 281 \text{ nm}; \text{ IR}: 3030, 1638 \text{ and } 830 \text{ cm}^{-1})$ . Acetylation of 8a gave a monoacetate (8b). The <sup>1</sup>HNMR spectrum of 8b exhibited signals due to six tertiary methyl groups ( $\delta 0.86$ , 0.89, 0.91, 1.17 and 1.24), two secondary methyl groups ( $\delta 0.81$  (d, J = 6.1 Hz) and 0.93 (br s)), one acetoxy methyl singlet ( $\delta 2.06$ ) and a  $3\alpha$ -axial carbinolic methine proton [ $\delta 4.52$  (1H, dd, J = 10.5 and 5.7 Hz)]. Furthermore, signals due to two vicinal protons arranged in the centre of a cisoid diene system were observed as a pair of symmetrical doublets at  $\delta$  5.45 and 5.59 (each 1H, J = 5.8 Hz). In the EI-mass spectrum, two intense peaks arising from cleavage of both B-ring and D-ring were observed at m/z 255 (fragment c) and 271 (fragment b) for compound 8a and m/z 255 (fragment c) and 313 (fragment b) for compound 8b, respectively, indicating the presence of the cisoid diene at C-9 (11):12 of a pentacyclic triterpene carbon skeleton. All the above data suggested that 8a was ursa-9 (11):12-dien-3 $\beta$ -ol. This was confirmed by

direct comparison of **8b** with authentic ursa-9 (11):12dien- $3\beta$ -yl acetate.

Ito and his colleague [13] reported both the isolation of marsformosanone [ursa-9(11):12-dien-3-one] along with marsformoxide ( $11\alpha$ ,  $12\alpha$ -oxidotaraxer- $3\beta$ -yl acetate) from *Marsdenia formosana* (Asclepiadaceae) and the synthesis of ursa-9(11):12 dien- $3\beta$ -yl acetate by photochemical transformation of the latter compound [14]. To our knowledge, however, this is the first report of the isolation of compound **8a** in nature.

#### **EXPERIMENTAL**

Mps: uncorr; optical rotations: CHCl<sub>3</sub>, 1 dm cell; UV: EtOH; IR: KBr discs; <sup>1</sup>H NMR: 90 or 300 MHz, CDCl<sub>3</sub> with TMS as int. standard. <sup>13</sup>C NMR: 75.4 MHz, CDCl<sub>3</sub> with TMS as int. standard. MS: double focussing mass spectrometer (accelerating voltage of 3–6.5 kV; ionizing potential 70 eV). TLC: Merck silica gel HF<sub>254</sub>.

Plant material. E. maculata was collected in the field of Kitakatsuragi district, Nara Pref., Japan in September 1985. The material was identified by Mr M. Murata of the Department of Botany, Faculty of Science, Kyoto University, Kitashirakawaoiwake-cho, Sakyo-ku, Kyoto, Japan. A voucher specimen of the plant is on file at the Institute of Medicinal Chemistry, Osaka University of Pharmaceutical Sciences.

Extraction and isolation of constituents. The air-dried whole herb of the plant (13.3 kg) was extracted with  $C_6H_6$  (3 1×5). The combined  $C_6H_6$  soln was concd to *ca* 5 l, filtered, and then washed with 5% NaOH soln to remove acidic components. After evaporation of the solvent, the resulting dark-greenish residue (462 g) was subjected to silica gel CC (5 kg) using *n*-hexane, *n*hexane- $C_6H_6$  in different proportions and  $C_6H_6$  as eluants. Rechromatography of each residue collected from the fractions eluted with *n*-hexane- $C_6H_6$  (20:1, 10:1, 5:1 and 1:1) and  $C_6H_6$ yielded the following compounds in order of their polarity; 1 (2.240 g), 2 (120 mg) and 7a (58 mg) [from *n*-hexane- $C_6H_6$ (20:1)], 3 (123 mg) [from *n*-hexane- $C_6H_6$  (10:1)], 4 (72 mg) [from *n*-hexane- $C_6H_6$  (5:1)] 8a (12 mg) and 5 (85 mg) [from *n*hexane- $C_6H_6$  (1:1)] and 6 (9.235 g) [from  $C_6H_6$ ].

β-Amyrin acetate (1). Colourless needles, mp 241–242.5° (MeOH–CHCl<sub>3</sub>),  $[\alpha]_D^{-3} + 68.9°$  (c 0.94) (lit. [15] mp 241–242°,  $[\alpha]_D + 81°$ ); IR ν<sub>max</sub> 1722 and 1240 (OAc), 1635 and 812 cm<sup>-1</sup> (-C=C-H); <sup>1</sup>H NMR (90 MHz): δ0.84 (3H, s, H-28), 0.88 (12H, s, H-23, 24, 29, 30), 0.98 (6H, s, H-25, 26), 1.14 (3H, s, H-27), 2.07 (3H, s, OAc), 4.54 (1H, dd, J = 11, 6 Hz, H-3α), 5.21 (1H, t, J = 3.5 Hz, H-12); EIMS m/z (rel. int.): 468 [M]<sup>+</sup> (3.5), 453 (2), 408 [M – HOAc]<sup>+</sup> (4), 218 (100), 203 (47). Compound 1 was identified by direct comparison with the sample prepared by acetylation (Ac<sub>2</sub>O–C<sub>5</sub>H<sub>5</sub>N, room temp.) of authentic β-amyrin (mmp, TLC, MS and <sup>1</sup>H NMR).

Taraxeryl acetate (2). Colourless needles, mp 299-302° (EtOAc),  $[\alpha]_D^{23} + 16.9°$  (c 0.52) (lit. [16] mp 304-305°,  $[\alpha]_D^{20} + 9°$ ); IR  $\nu_{max}$  1720 and 1243 (OAc), 1638 and 815 cm<sup>-1</sup> (-C=C-H); <sup>1</sup>H NMR (300 MHz):  $\delta 0.82$  (3H, s), 0.86 (3H, s), 0.88 (3H, s). 0.9 (3H, s), 0.91 (3H, s), 0.95 (6H, s), 1.07 (3H, s), 2.04 (3H, s, OAc), 4.46 (1H, dd, J = 11, 6.5 Hz, H-3\alpha), 5.53 (1H, dd, J = 8.3, 3.7 Hz, H-15); EIMS m/z (rel. int.): 468 [M] <sup>+</sup> (12), 453 (6), 408 [M - HOAc] <sup>+</sup> (5), 344 (31), 329 (23), 284 (25), 269 (30), 204 (100). Compound **2** was identified by direct comparison with the sample prepared by acetylation (Ac<sub>2</sub>O-C<sub>5</sub>H<sub>5</sub>N, room temp.) of taraxerol isolated from *Taraxacum officinale* [17] (mmp, TLC, IR, MS and <sup>1</sup>H NMR).

*Lupenyl acetate* (3). Colourless needles, mp  $215-216^{\circ}$  (MeOH-CHCl<sub>3</sub>),  $[\alpha]_{D}^{23} + 35.5^{\circ}$  (c 0.35) (lit. [18] mp  $221-222^{\circ}$ .

 $[\alpha]_D + 39^\circ$ ); IR  $\nu_{max}$  1727 and 1239 (OAc), 3060, 1637 and 865 cm<sup>-1</sup> (-C=CH<sub>2</sub>); <sup>1</sup>H NMR (90 MHz):  $\delta$ 0.79 (3H, s, H-28), 0.85 (9H, s, H-23, 24, 25), 0.95 (3H, s, H-27), 1.04 (3H, s, H-26), 1.70 (3H, s, H-30), 2.04 (3H, s, OAc), 4.47 (1H, dd, J = 11, 6 Hz, H-3\alpha), 4.56 and 4.67 (each 1H, dd, J = 2.5 Hz, H-29); EIMS m/z (rel. int.): 468 [M]<sup>+</sup> (92), 453 (21), 408 [M - HOAc]<sup>+</sup> (21), 249 (33), 218 (64), 204 (58), (100). Compound **3** was identified by direct comparison with an authentic sample of lupenyl acetate (mmp, TLC, IR, MS and <sup>1</sup>H NMR).

 $3\beta$ -Acetoxy-30-norlupan-20-one (4). Colourless needles, mp 276–278° (MeOH–CHCl<sub>3</sub>),  $[\alpha]_D^{23} + 3.5°$  (c 0.33) (lit. [4] mp 262–263°,  $[\alpha]_D^{20}$  + 9.0°); IR  $\nu_{max}$ : 1722 and 1248 (OAc), 1706 cm<sup>-1</sup> (C=O); <sup>1</sup>H NMR (90 MHz): δ0.77 (3H, s, H-28), 0.86 (9H, s, H-23, 24, 25), 0.97 (3H, s, H-27), 1.03 (3H, s, H-26), 2.06 (3H, s, OAc), 2.16 (3H, s, H-29), 4.48 (1H, dd, J = 11, 6 Hz, H-3 $\alpha$ ); EIMS m/z (rel. int.): 470 [M]<sup>+</sup> (2), 410 [M - HOAc]<sup>+</sup> (23), 395 (11), 367 [M -HOAc-MeCO]<sup>+</sup> (19), 231 (11), 203 (36), 189 (100). Compound 4 (20 mg) was hydrolysed with 0.05 M KOH-EtOH to give  $3\beta$ -hydroxy-30-norlupan-20-one (18 mg), mp 238-240°,  $[\alpha]_{D}^{23} - 13.8^{\circ}$  (c 0.42) (lit. [19] mp 237–239°,  $[\alpha]_{D} - 10^{\circ}$ ); IR  $v_{max}$ : 3458 and 1037 (OH), 1687 cm<sup>-1</sup> (C=O); <sup>1</sup>H NMR (90 MHz): 0.77 (6H, s, H-23, 28), 0.84 (3H, s, H-24), 0.97 (6H, s, H-25, 27), 1.02 (3H, s, H-26), 2.15 (3H, s, H-29), 3.20 (1H, dd, J = 11, 6.5 Hz, H-3 $\alpha$ ); EIMS m/z (rel. int.): 428 [M]<sup>+</sup> (9), 413 (3), 410 (33), 395 (14), 367  $[M - H_2O - MeCO]^+$  (15), 207 (51), 189 (100).

Compound 4 was identified by direct comparison with synthetic  $3\beta$ -acetoxy-30-norlupan-20-one (mmp, TLC, IR, MS and <sup>1</sup>H NMR).

Synthesis of  $3\beta$ -acetoxy-30-norlupan-20-one. To a soln of lupenyl acetate (27 mg) and OsO<sub>4</sub> (1 mg) in dioxane (7 ml) containing H<sub>2</sub>O (1.5 ml) NaIO<sub>4</sub> (36 mg) was gradually added under stirring and the mixture was allowed to stand at room temp. for 72 hr. The reaction mixture was treated with H<sub>2</sub>S and the precipitate of OsS was filtered off. Dilution of the filtrate with H<sub>2</sub>O (30 ml) gave a ppt. which was extracted with CHCl<sub>3</sub>. The usual work-up, followed by purification with prep. TLC (Merck silica gel PF<sub>254</sub>, C<sub>6</sub>H<sub>6</sub>-CHCl<sub>3</sub>-EtOAc 2:2:1) afforded 3 $\beta$ -acetoxy-30-norlupan-20-one (25 mg), mp 276-278° (MeOH-CHCl<sub>3</sub>), identical in all respects with compound 4.

 $\alpha$ -Amyrenonol (5). Colourless needles, mp 204-205.5° (MeOH-CHCl<sub>3</sub>),  $[\alpha]_{D}^{23}$  + 114.9° (c 0.21) (lit. [6] mp 206-208°,  $[\alpha]_{\rm D} + 96^{\circ}$ ; UV  $\lambda_{\rm max}$  250.5 nm ( $\epsilon$  7,600) [-C=C-C=O]; IR  $v_{max}$  3430 and 1038 (OH), 1658 (-C=C-C=O), 1620 and 838 cm<sup>-1</sup> (-C=C-H); <sup>1</sup>H NMR (300 MHz):  $\delta$ 0.81 (3H, d, J = 6 Hz), 0.81 (3H, s), 0.95 (3H, br s), 1.00 (3H, s), 1.14 (3H, s), 1.17 (6H, s), 1.30 (3H, s), 3.23  $(1H, dd, J = 11.5, 5.5 Hz, H-3\alpha)$ , 5.54  $(1H, dd, J = 11.5, 5.5 Hz, H-3\alpha)$ , 5.54  $(1H, dd, J = 11.5, 5.5 Hz, H-3\alpha)$ , 5.54  $(1H, dd, J = 11.5, 5.5 Hz, H-3\alpha)$ , 5.54  $(1H, dd, J = 11.5, 5.5 Hz, H-3\alpha)$ , 5.54  $(1H, dd, J = 11.5, 5.5 Hz, H-3\alpha)$ , 5.54  $(1H, dd, J = 11.5, 5.5 Hz, H-3\alpha)$ , 5.54  $(1H, dd, J = 11.5, 5.5 Hz, H-3\alpha)$ , 5.54  $(1H, dd, J = 11.5, 5.5 Hz, H-3\alpha)$ , 5.54  $(1H, dd, J = 11.5, 5.5 Hz, H-3\alpha)$ , 5.54  $(1H, dd, J = 11.5, 5.5 Hz, H-3\alpha)$ , 5.54 (1H, dd, J = 11.5, 5.5 Hz)s, H-12); EIMS m/z (rel. int.): 440 [M]<sup>+</sup> (16), 425 (6), 422 (4), 273 (100), 232 (84). Acetylation of 5 (22 mg) with  $Ac_2O$  and  $C_5H_5N$ (1:1, 2 ml) gave an acetate (21 mg), mp 286-288° (MeOH-CHCl<sub>3</sub>),  $[\alpha]_D^{23} + 116^\circ$  (c 0.21) (lit. [20] mp 286-288°,  $[\alpha]_{D} + 101^{\circ}$ ; UV $\lambda_{max}$  251 nm ( $\varepsilon$  10000); IR  $v_{max}$ : 1715 and 1245 (OAc),  $1642 \text{ cm}^{-1}$  (-C=C-C=O); <sup>1</sup>H NMR (300 MHz):  $\delta 0.81$  (3H, d, J = 6.1 Hz), 0.82 (3H, s), 0.88 (3H, s), 0.88 (3H, s), 0.94 (3H, br s), 1.17 (3H, s), 1.19 (3H, s), 1.29 (3H, s), 2.05 (3H, s, OAc), 4.55 (1H, dd, J = 11.5, 5.5 Hz, H-3 $\alpha$ ), 5.54 (1H, s, H-12); EIMS m/z (rel. int.):  $482 [M]^+ (12), 422 [M - HOAc]^+ (11), 407 [M - HOAc - Me]^+$ (10), 273 (100), 232 (58). This acetate was identified by direct comparison with a synthetic sample of a-amyrenonyl acetate (3\beta-acetoxyurs-12-en-11-one) (mmp, TLC, IR, MS and <sup>1</sup>HNMR).

Synthesis of  $\alpha$ -amyrenonyl acetate.  $\alpha$ -Amyrin (25 mg) was acetylated (Ac<sub>2</sub>O-C<sub>5</sub>H<sub>5</sub>N 1:1, 4 ml) as usual to give  $\alpha$ -amyrin acetate (25 mg), mp 225-227°, M<sup>+</sup> at m/z 468 (C<sub>32</sub>H<sub>52</sub>O<sub>2</sub>). A mixture of  $\alpha$ -amyrin acetate (20 mg) in HOAc (15 ml) and CrO<sub>3</sub> (20 mg) in 90% HOAc (3.5 ml) was heated on a steam bath for 2 hr. The reaction mixture was worked up as usual yielding a residue, which on purification by prep. TLC ( $C_6H_6$ -EtOAc 5:1) afforded  $\alpha$ -amyrenonyl acetate, mp 286-288°,  $[\alpha]_D^{23} + 117^\circ$  (c 0.35), identical in all respects with the acetate of compound 5.

Sitosterol (6). Colourless needles, mp 137-139° (MeOH-CHCl<sub>3</sub>) (lit. [21] mp 138-139°). Compound 6 was identified by direct comparison with an authentic sitosterol (TLC, IR and MS).

Gult-5-en-3 $\beta$ -yl acetate (7a). Colourless needles, mp 190–191.5° (MeOH–CHCl<sub>3</sub>), [ $\alpha$ ]<sub>D</sub><sup>3</sup> + 76.8° (c 0.50) (lit. [5] mp 192–194°, [ $\alpha$ ]<sub>D</sub> + 79°); IR v<sub>max</sub>: 1723 and 1238 (Ac), 1639 and 814 (-C<sub>1</sub>=C-H), 1380 and 1360 cm<sup>-1</sup> (gem dimethyl); <sup>1</sup>H NMR (300 MHz):  $\delta$ 0.85, 0.95, 0.99, 1.00, 1.05, 1.07, 1.10, 1.16 (each 3H, s, 8 × tert, Me), 2.01 (3H, s, OAc), 4.69 (1H, t, J = 2.9 Hz, H-3 $\alpha$ ), 5.56 (1H, dt like, J = 5.5, 1.8 Hz, H-6); <sup>13</sup>C NMR:  $\delta$ 16.41 (q), 18.41 (q), 18.91 (t), 19.58 (q), 21.22 (q), 23.49 (t), 25.04 (q), 25.49 (t), 28.28 (s), 29.11 (q), 30.14 (s), 30.40 (t), 32.03 (C × 2, q and t), 32.36 (q), 33.09 (t), 34.48 (q), 34.62 (t), 34.78 (s), 35.10 (t), 36.02 (t), 37.88 (s), 38.98 (t), 39.09 (s), 39.31 (s), 43.11 (d), 47.38 (d), 49.58 (d), 78.60 (d), 120.01 (d), 141.94 (s), 170.85 (s); EIMS m/z (rel. int.): 468 [M]<sup>+</sup> (15), 408 [M - HOAc]<sup>+</sup> (9), 393 [M - Me - HOAc]<sup>+</sup> (8), 274 [fragment a] (88), 259 [a - Me] (100), 245 (8), 205 (6).

Alkaline hydrolysis of **7a**. Compound **7a** (22 mg) was hydrolysed with 0.05 M KOH in EtOH (13 ml) on a steam bath for 2 hr. After usual work-up, the crude product was chromatographed on silica gel to give the alcohol **7b** (20 mg) as colourless needles, mp 210–213° (MeOH–CHCl<sub>3</sub>),  $[\alpha]_D^{23} + 63.3°$  (c 0.71) (lit. [5] mp 210–211.5°,  $[\alpha]_D + 64°$ ); IR  $v_{max}$ : 3428 and 1037 cm<sup>-1</sup> (OH); <sup>1</sup>H NMR (90 MHz):  $\delta 0.85$ , 0.96, 0.99, 1.01, 1.05, 1.10, 1.14, 1.17 (each 3H, s, 8 × tert Me), 3.47 (1H, t, J = 3Hz, H-3\alpha), 5.63 (1H, m, w/2 = 4 Hz, H-6); EIMS m/z (rel. int.): 426 [M]<sup>+</sup> (4), 408 [M - H<sub>2</sub>O]<sup>+</sup> (22), 274 [fragment a] (97), 259 [a - Me] (100), 245 (17), 205 (51). **7b** was identified by direct comparison with an authentic sample of gult-5-en-3 $\beta$ -ol (mmp, TLC, IR, MS and <sup>1</sup>H NMR).

*Ursa*-9 (11): 12-*dien*-3 $\beta$ -ol (8a). Colourless needles, mp 150–151.5° (MeOH – CHCl<sub>3</sub>),  $[\alpha]_D^{23}$  + 362.6° (*c* 0.31) (lit. [20] mp 157–158°,  $[\alpha]_D^{20.5}$  + 360°); UV  $\lambda_{max}$  281 nm (*e* 9900) [homo-annular diene]; IR  $v_{max}$ : 3360 and 1032 (OH), 3030, 1638 and 830 cm<sup>-1</sup> (diene); <sup>1</sup>H NMR (300 MHz):  $\delta$ 0.82, 0.86, 0.89, 1.03, 1.18, 1.21 (each 3H, *s*, 6 × tert Me), 0.81 (3H, *d*, *J* = 6 Hz) and 0.93 (3H, *br s*) [2 × sec Me], 3.24 (1H, *dd*, *J* = 11, 5.5 Hz, H-3 $\alpha$ ), 5.45 and 5.60 (each 1H, *d*, *J* = 5.8 Hz, H-11, 12); EIMS *m/z* (rel. int.): 424 [M]<sup>+</sup> (100), 406 [M – H<sub>2</sub>O]<sup>+</sup> (5), 271 [fragment *b*] (34), 255 [fragment c] (94).

Acetylation of compound **8a**. A soln of **8a** (7 mg) in  $C_5H_5N$  (0.5 ml) was acetylated with  $Ac_2O$  (0.5 ml) at room temp. overnight. After usual work-up, the crude product was chromatographed over silica gel to give an acetate (**8b**) as colourless prisms, mp 165–166° (MeOH–CHCl<sub>3</sub>,  $[\alpha]_D^{23} + 341.2°$  (c 0.18) (lit. [13] mp 165–167°,  $[\alpha]_D^{25} + 336°$ ); UV  $\lambda_{max}$  281 nm ( $\epsilon$  10000) (homoannular diene); IR  $v_{max}$ : 1736 and 1243 (OAc), 3030, 1638 and 830 cm<sup>-1</sup> (diene); <sup>1</sup>H NMR (300 MHz):  $\delta$ 0.86 (3H, s), 0.89 (6H, s), 0.91 (3H, s), 1.17 (3H, s), 1.24 (3H, s), [6 × tert Me], 0.81 (3H, d, J = 6.1 Hz) and 0.93 (3H, br s) [2 × sec Me], 2.06 (3H, s, OAc), 4.52 (1H, dd, J = 10.5, 5.7 Hz, H-3a), 5.45 and 5.59 (each 1H, d, J = 5.8 Hz, H-11, 12); <sup>13</sup>C NMR:  $\delta$ 16.75 (q, C-29), 17.42\* (q, C-24), 17.55\* (q, C-25), 18.20 (t, C-6), 21.33 (q,

<u>Me</u>-COO-), 21.52 (q, C-30), 22.16 (q, C-26), 24.28 (t, C-2), 25.46 (q, C-27), 26.10 (t, C-16), 28.16 (q, C-23), 28.23 (t, C-15), 28.72 (q, C-28), 31.22<sup>†</sup> (t, C-21), 31.97<sup>†</sup> (t, C-7), 33.69 (s, C-17), 37.00 (t, C-1), 37.88 (s, C-10), 38.55 (s, C-4), 39.00<sup>‡</sup> (d, C-19), 39.47<sup>‡</sup> (d, C-20), 40.69 (s, C-8), 41.36 (t, C-22), 43.10 (s, C-14), 51.23 (d, C-5), 57.33 (d, C-18), 80.62 (d, C-3), 115.51 (d, C-11), 123.00 (d, C-12), 141.37 (s, C-13), 154.18 (s, C-9), 171.02 (s, MeCOO) [\*<sup>†</sup><sup>‡</sup> values may be interchangeable]; EIMS m/z (rel. int.): 466 [M]<sup>+</sup> (100), 406 [M - HOAc]<sup>+</sup> (5), 313 [fragment b] (9), 255 [fragment c] (56). Compound **8b** was identified by direct comparison with the authentic sample of ursa-9 (11):12-dien-3 $\beta$ -yl acetate (mmp, TLC, IR, MS and <sup>1</sup>H NMR).

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#### REFERENCES

- 1. Matsunaga, S. and Morita, R. (1983) Phytochemistry 22, 605.
- Nanba, T. and Mikage, M. (1983) Poisonous Plants, p. 74. Hoikusha, Tokyo.
- Okuda, T. (1986) Encyclopedia of Natural Medicine, p. 312. Hirokawa, Tokyo.
- 4. Hui, W. H., Li, M. M. and Lee, Y. C. (1977) Phytochemistry 16, 607.
- Yagishita, K. and Nishimura, M. (1961) Agric. Biol. Chem. 25, 517.
- Yagishita, K. and Nishimura, M. (1961) Agric. Biol. Chem. 25, 844.
- 7. Bandaranayake, W. M. (1980) Phytochemistry 19, 255.
- 8. Budzikiewicz, H., Brauman, J. I. and Djerassi, C. (1965) Tetrahedron 21, 1855.
- 9. Fisher, F. G. and Seiler, N. (1961) Ann. 664, 162.
- Hui, W. H., Ko, P. D. S., Lee, Y. C. and Li, M. M. (1975) Phytochemistry 14, 1063.
- Gaind, K. N., Singla, A. K., Boar, R. B. and Copsey, D. B. (1976) Phytochemistry 15, 1999.
- Ohmoto, T., Uzawa, S. and Imazeki, N. (1972) Shoyakugaku Zasshi 26, 47.
- 13. Ito, K. and Lai, J. (1979) Chem. Pharm. Bull. 27, 210.
- 14. Ito, K. and Lai, J. (1979) Chem. Pharm. Bull. 27, 2248.
- 15. Ageta, H. and Arai, Y. (1983) Phytochemistry 22, 1801.
- 16. Hui, W. H. and Li, M. M. (1976) Phytochelistry 15, 563.
- 17. Burrows, S. and Simpson, J. C. E. (1983) J. Chem. Soc. 2042.
- 18. Kircher, H. W. (1980) Phytochemistry 19, 2701.
- Thompson, M. J. and Bowers, W. S. (1965) Phytochemistry 7, 845.
- Ewen, E. S., Spring, F. S. and Vickerstaff, T. (1939) J. Chem. Soc. 1303.
- 21. Fieser, L. and Fieser, M. (1959) Steroids, p. 351. Reinhold, New York.