

The 1,4-linked disaccharide of hyaluronan:
synthesis of methyl
2-acetamido-2-deoxy- β -D-glucopyranosyl-
(1 \rightarrow 4)- β -D-glucopyranosiduronic acid

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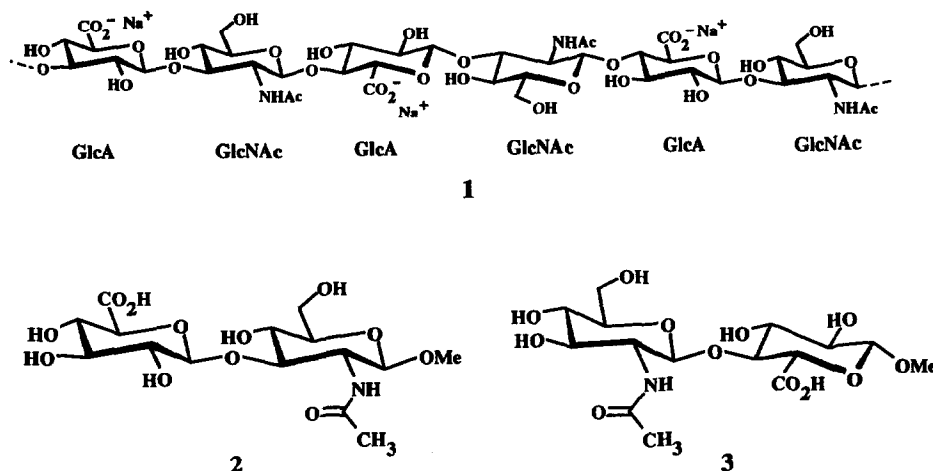
Hyaluronan (HA, **1**), a major component of the extracellular matrix of mammalian tissues [1], is a repeating polymer of 2-acetamido-2-deoxy-D-glucopyranose (GlcNAc) linked β ,1-4 to D-glucuronic acid (GlcA). The GlcA is bound β ,1-3 to the GlcNAc residue of the next subunit, hence the sequence GlcNAc(β 1-4)GlcA(β 1-3)GlcNAc. During the course of high-resolution NMR studies on the solution conformations of HA, it became necessary to fully characterize the methyl β -glycosides **2** and **3** of the two possible disaccharides derivable from this sequence. While the β ,1-3-linked dimer is available by chemical degradation [2] of polymeric HA, the β ,1-4-linked dimer **3** cannot be obtained in this manner. Several studies on the synthesis of HA fragments have been published [3], but the preparation of unprotected **3** has not been recorded. We now describe the successful synthesis of **3**.

Our initial strategy involved glycosylation of the free 4-hydroxyl group of methyl 2,3-di-O-benzyl-6-O-(4-methoxybenzyl)- β -D-glucopyranoside (**7**) with *N*-(3,4,6-tri-O-benzyl-2-deoxy-2-iodo- α -D-mannopyranosyl)benzenesulfonamide [4] (**4**). Glucose derivative **7** is available from methyl β -D-glucopyranoside in three steps in 57%

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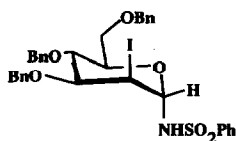


overall yield ¹ (1. *p*-MeOPhCH(OMe)₂, TsOH · H₂O, DMF, –MeOH, 50–70°C; 2. BnBr, NaH, RT; 3. CF₃CO₂H, NaBH₃CN, 3A MS, DMF, RT) [5]. Treatment of a mixture of 4 and 7 with 2.2 equiv of lithium tetramethylpiperide in THF containing AgOTf at –78°C, followed by warming to room temperature, afforded dimer 8 in 51% yield. Deprotection of the 6-OH (CAN, CH₃CN–H₂O, RT) [5], followed by Jones oxidation [6] and esterification (CH₂N₂, ether), produced 9 in 43% overall yield. Unfortunately, all attempts at removal of the *N*-sulfonyl and benzyl protecting groups to yield 3 resulted in incomplete deprotection and/or decomposition ².

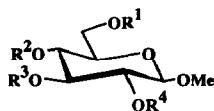
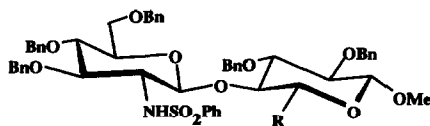
Since deprotection of 9 proved problematic, we sought a glycosyl donor with a more cooperative *N*-protecting group. After some exploration, we settled on the widely used glycosyl halide, 3,4,6-tri-*O*-acetyl-2-deoxy-2-phthalimido-*D*-glucopyranosyl bromide [8] (10). An advantage of 10 was its ability to couple with a glycosyl acceptor (13) having C-6 already oxidized, thereby reducing the number of transformations required on the coupled product. Glucose derivative 13 is available from methyl β-*D*-glucopyranoside in five steps in 31% overall yield (1. *p*-MeOPhCH(OMe)₂, TsOH · H₂O, DMF, –MeOH, 50–70°C; 2. BnBr, NaH, RT; 3. Me₃SiCl, NaBH₃CN, 3A MS, CH₃CN, RT [5]; 4. CrO₃ · 2 pyr, Ac₂O, ^tBuOH, CH₂Cl₂–DMF, RT [9]; 5. CAN, CH₃CN–H₂O, RT [5]). The reaction of 13 and 10

¹ Abbreviations used: CAN, ceric ammonium nitrate; MS, molecular sieves; pyr, pyridine; RT, room temperature; MPM, *p*-methoxyphenylmethyl; AgOTf, silver trifluoromethanesulfonate. In NMR assignments for 3 and its precursors G = glucopyranosyl unit or subunit; U = glucopyranosyluronic acid subunit; N = 2-acetamido-2-deoxy-β-*D*-glucopyranosyl subunit.

² Danishefsky and co-workers [7] have recently employed 2-(trimethylsilyl)ethanesulfonamide in their 'sulfonamidoglycosylation' procedure, in place of benzenesulfonamide. The eventual removal of the 2-(trimethylsilyl)ethanesulfonamide group is accomplished under conditions similar to those used for the removal of trimethylsilylethoxymethyl (SEM) protecting groups.



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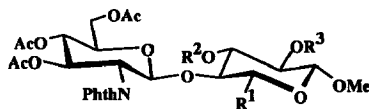
5: $R^1, R^2 = \text{CHPh-}p\text{-(OMe)}$, $R^3 = R^4 = \text{H}$ 6: $R^1, R^2 = \text{CHPh-}p\text{-(OMe)}$, $R^3 = R^4 = \text{Bn}$ 7: $R^1 = \text{MPM}$, $R^2 = \text{H}$, $R^3 = R^4 = \text{Bn}$ 8: $R = \text{CH}_2\text{OMPM}$ 9: $R = \text{CO}_2\text{Me}$

with AgOTf in the presence of *sym*-collidine [10] and 3A MS in CH_2Cl_2 at $-30 \rightarrow 25^\circ\text{C}$ provided the protected disaccharide glycoside **14** in 94% yield. Removal of the benzyl protecting groups (30% Pd-C , H_2 , $\text{EtOAc-H}_2\text{O}$) provided **15** in 75% yield. Unfortunately cleavage of the *tert*-butyl ester using formic acid [11] produced, in addition to **16**, an unidentified side product which we were unable to remove.

The reaction of **7** and **10** with AgOTf in the presence of *sym*-collidine and 3A MS in CH_2Cl_2 [10] at $-30 \rightarrow 25^\circ\text{C}$ provided the protected disaccharide glycoside **17**, which could not be purified. However, removal of the 6-OH protecting group (CAN , $\text{CH}_3\text{CN-H}_2\text{O}$, RT) cleanly produced **18** in 70% yield from **7**. Jones



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11: $R^1 = \text{CH}_2\text{OH}$, $R^2 = \text{MPM}$, $R^3 = R^4 = \text{Bn}$ 12: $R^1 = \text{CO}_2^t\text{Bu}$, $R^2 = \text{MPM}$, $R^3 = R^4 = \text{Bn}$ 13: $R^1 = \text{CO}_2^t\text{Bu}$, $R^2 = \text{H}$, $R^3 = R^4 = \text{Bn}$ 14: $R^1 = \text{CO}_2^t\text{Bu}$, $R^2 = R^3 = \text{Bn}$ 15: $R^1 = \text{CO}_2^t\text{Bu}$, $R^2 = R^3 = \text{H}$ 16: $R^1 = \text{CO}_2\text{H}$, $R^2 = R^3 = \text{H}$ 17: $R^1 = \text{CH}_2\text{OMPM}$, $R^2 = R^3 = \text{Bn}$ 18: $R^1 = \text{CH}_2\text{OH}$, $R^2 = R^3 = \text{Bn}$ 19: $R^1 = \text{CO}_2\text{Me}$, $R^2 = R^3 = \text{Bn}$ 20: $R^1 = \text{CO}_2\text{Me}$, $R^2 = R^3 = \text{H}$

oxidation [6], followed by esterification (CH_2N_2 , ether), provided **19** in 58% yield. Finally, a four-step sequence accomplished the removal of all protecting groups and the acylation of the free amine function (1. H_2 , 30% Pd-C, EtOAc- H_2O ; 2. aq NaOH, MeOH, 0°C ; 3. $\text{N}_2\text{H}_4 \cdot \text{H}_2\text{O}$, EtOH, reflux; 4. Ac_2O , MeOH) [8,11] to provide **3** in 52% overall yield from **19**.

1. Experimental

Spectroscopic measurements. — ^1H and ^{13}C NMR spectra were recorded in CDCl_3 on a Varian UNITY 500 MHz spectrometer at resonance frequencies of 499.84 and 125.67 MHz, respectively, at RT. Spectroscopic data for **3** were obtained at 37°C on a sample dissolved in D_2O , with HOD (δ 4.76 ppm) serving as the internal ^1H reference and the acetamido methyl carbon (δ 23.3 ppm) serving as the internal ^{13}C reference. Optical rotations were determined on a Perkin-Elmer model 141 polarimeter. The mass of compound **3** was determined by negative-ion high resolution FABMS at the University of Illinois School of Chemical Sciences Mass Spectrometry Laboratory.

Methyl 4,6-O-(4-methoxybenzylidene)- β -D-glucopyranoside (5).—A solution of methyl β -D-glucopyranoside (20.0 g, 103 mmol), 4-methoxybenzaldehyde dimethyl acetal [5] (28.6 g, 157 mmol), and *p*-toluenesulfonic acid monohydrate (0.18 g, 0.95 mmol) in anhyd DMF (103 mL) was heated at 50°C on a rotary evaporator under water aspirator pressure (~ 22 mmHg) for 1 h. The temperature was then increased to 70°C and the mixture was concentrated in volume to ca. 40 mL. This remaining solution was poured into a stirred slurry of ice (50 g), satd aq NaHCO_3 (100 mL), and diethyl ether (100 mL). The white precipitate that formed was filtered, washed with hexanes (3×100 mL), H_2O (2×100 mL), and dried in vacuo over P_2O_5 to yield 27.8 g (87%) of a white solid which was identified as **5**; mp 176 – 177°C (from EtOAc); MS: *m/z* calcd for $\text{C}_{15}\text{H}_{20}\text{O}_7$, 312.1209; found, 312.1222; ^1H NMR: 2.71 (s, 1 H, OH), 2.84 (s, 1 H, OH), 3.37 (ddd, 1 H, *J* 10, 9.5, 5.0 Hz, G5), 3.40–3.54 (m, 2 H, G2, G3), 3.50 (s, 3 H, OMe), 3.67–3.76 (m, 2 H, G6, G6'), 3.73 (s, 3 H, ArOMe), 4.25 (d, 1 H, *J* 7.5 Hz, G1), 4.27 (dd, 1 H, *J* 10.5, 5.0 Hz, G4), 5.42 (s, 1 H, ArCH), 6.82–6.84 (m, 2 H, Ar), 7.34–7.36 (m, 2 H, Ar); ^{13}C NMR: δ 55.24, 57.40, 66.28, 68.55, 73.10, 74.40, 80.46, 101.75, 104.08, 113.65, 127.56, 129.40, 160.19.

Methyl 2,3-di-O-benzyl-4,6-O-(4-methoxybenzylidene)- β -D-glucopyranoside (6).—Sodium hydride (60%, 13.8 g, 346 mmol) was washed with anhyd hexanes (3×30 mL) and then dispersed in anhyd DMF (420 mL). A solution of **5** (26.9 g, 86 mmol) in anhyd DMF (78 + 38 + 12 mL) was added dropwise to the NaH slurry at RT. After 5 min BnBr (25.0 mL, 210 mmol) was added dropwise over a 15-min period. The reaction was quenched with MeOH (74 mL) after 3 h, diluted with EtOAc (1000 mL), and washed with H_2O (3×390 mL). The combined aq phases were extracted with ether (800 mL). The combined organic phases were dried (MgSO_4) and concentrated in vacuo to give 43.3 g of an off-white residue, which was

recrystallized from EtOH–acetone to yield 34.1 g (2 crops, 80%) of a white solid identified as **6**; mp 151°C (from EtOH–acetone); $[\alpha]_D -39.0^\circ$ (c 1.19, CHCl_3); MS: m/z calcd for $\text{C}_{29}\text{H}_{32}\text{O}_7$, 492.2148; found, 492.2155; ^1H NMR: δ 3.41 (td, 1 H, J 10.0, 5.0 Hz, G5), 3.45 (dd, 1 H, J 8.3, 7.5 Hz, G2), 3.59 (s, 1 H, OMe), 3.67 (dd, 1 H, J 9.5, 9.0 Hz, G3), 3.77 (ABX, 2 H, J_{AB} 13.0, J_{AX} 10, J_{BX} 10 Hz, G6, G6'), 3.82 (s, 3 H, ArOMe), 4.35 (dd, 1 H, J 10.5, 5.0 Hz, G4), 4.42 (d, 1 H, J 7.5 Hz, G1), 4.82 (ABq, 2 H, J 11.0 Hz, CH_2Ph), 4.85 (ABq, 2 H, J 11.0 Hz, CH_2Ph), 5.54 (s, 1 H, ArCH), 6.90–6.92 (m, 2 H, Ar), 7.24–7.26 (m, 12 H, Ar); ^{13}C NMR: δ 57.18, 57.33, 65.89, 68.65, 74.96, 75.15, 80.74, 81.38, 82.11, 101.02, 105.12, 113.50, 127.24, 127.50, 127.57, 127.92, 127.95, 128.19, 128.24, 129.77, 138.37, 138.47, 159.93. Anal. Calcd for $\text{C}_{29}\text{H}_{32}\text{O}_7$ (492.571): C, 70.70; H, 6.55. Found: C, 70.57; H, 6.67.

Methyl 2,3-di-O-benzyl-4,6-O-(4-methoxybenzyl)- β -D-glucopyranoside (7).—A solution of $\text{CF}_3\text{CO}_2\text{H}$ (15.6 mL, 203 mmol) in anhyd DMF (120 mL) over 3A MS at 0°C was added to a slurry of **6** (10.0 g, 20.3 mmol), NaBH_3CN (6.72 g, 102 mmol), and crushed 3A MS (10.0 g) in anhyd DMF (160 mL) at RT. After 17 h the mixture was filtered through Celite into iced satd aq NaHCO_3 (280 mL). The aq phase was extracted with CH_2Cl_2 (5×160 mL). The combined organic phases were washed with satd aq NaHCO_3 (280 mL), water (280 mL), and satd aq NaCl (280 mL), dried (MgSO_4), and concentrated in vacuo to give a white solid. This was purified by flash column chromatography [12] (7 : 1 toluene–EtOAc) to yield 8.45 g (84%) of a colorless oil, which slowly solidified to a white solid and was identified as **7**: $[\alpha]_D -18.1^\circ$ (c 1.10, CHCl_3); ^1H NMR: δ 3.33–3.38 (m, 1 H, G5), 3.34 (dd, 1 H, J 9.0, 7.5 Hz, G2), 3.39 (dd, 1 H, J 9.0, 9.0 Hz, G3), 3.50 (s, 3 H, OMe), 3.50–3.54 (m, 1 H, G4), 3.65 (ABX, 2 H, J_{AB} 10.4, J_{AX} 5.4, J_{BX} 4.6 Hz, G6, G6'), 3.74 (s, 3 H, ArOMe), 4.26 (d, 1 H, J 7.5 Hz, G1), 4.46 (ABq, 2 H, J 11.8 Hz, CH_2Ph), 4.75 (ABq, 2 H, J 11.3 Hz, CH_2Ph), 4.77 (ABq, 2 H, J 11.5 Hz, CH_2Ph), 6.79–6.83 (m, 2 H, Ar), 7.18–7.31 (m, 12 H, Ar); ^{13}C NMR: δ 55.14, 56.98, 69.90, 71.62, 73.21, 73.92, 74.55, 75.13, 81.67, 83.91, 104.64, 113.73, 127.54, 127.67, 127.83, 127.98, 128.25, 128.39, 129.26, 129.90, 138.42, 138.59, 159.19. Anal. Calcd for $\text{C}_{29}\text{H}_{34}\text{O}_7$ (494.584): C, 70.43; H, 6.93. Found: C, 70.57; H, 7.03.

Methyl O-(2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 4)- β -D-glucopyranosiduronic acid (3).—A slurry of **7** (1.04 g, 2.1 mmol), AgOTf (1.88 g, 7.3 mmol), crushed 3A MS, and 2,4,6-collidine (0.83 mL, 6.3 mmol) in anhyd CH_2Cl_2 (20 mL) was stirred at RT for 1 h then cooled to -30°C . A slurry of **10** (3.75 g, 7.6 mmol) and crushed 3A MS in anhyd CH_2Cl_2 (10 mL) was stirred at RT for 1 h, then added dropwise to the above slurry at -30°C . After 0.5 h the mixture was allowed to warm to RT and stirred a further 4 h. The mixture was filtered through Celite, the solids were washed with CH_2Cl_2 , and the combined filtrate was washed successively with satd aq NaHCO_3 (30 mL), M aq $\text{Na}_2\text{S}_2\text{O}_3$ (3×25 mL), water (25 mL), 10% aq citric acid (25 mL), satd aq NaHCO_3 (25 mL), H_2O (25 mL), and satd aq NaCl (25 mL), dried (Na_2SO_4), and concentrated in vacuo to give a bright-yellow foam. The foam was purified by flash column chromatography (5 : 1 toluene–EtOAc) to yield 1.85 g of a sticky off-white foam that was identified as impure methyl O-(3,4,6-tri-O-acetyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)-(1 \rightarrow 4)-2,3-di-O-benzyl-6-O-(4-methoxybenzyl)- β -D-glucopyranoside (**17**).

CAN (2.30 g, 4.2 mmol) was added to a solution of crude **17** (1.85 g, ~0.56 mmol) in 9:1 CH₃CN–H₂O (32 mL). After 3.5 h the mixture was concentrated in vacuo and the residue was dissolved in EtOAc (125 mL). The solution was washed with satd aq NaHCO₃ (40 mL), H₂O (40 mL), and satd aq NaCl (40 mL), dried (MgSO₄), and concentrated in vacuo to give an off-white foam. This was purified by flash column chromatography (3:2 toluene–EtOAc) to yield 0.96 g (58% from **7**) of white foam identified as methyl *O*-(3,4,6-tri-*O*-acetyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)-(1 → 4)-2,3-di-*O*-benzyl-β-D-glucopyranoside: $[\alpha]_D +37.8^\circ$ (*c* 0.50, CHCl₃); ¹H NMR: δ 1.82 (s, 3 H, Ac), 1.96 (s, 3 H, Ac), 1.98 (s, 3 H, Ac), 3.20 (dtd, 1 H, *J* 9.5, 2.0, 1.5 Hz, G5), 3.32 (dd, 1 H, *J* 8.8, 7.8 Hz, G2), 3.34–3.38 (m, 1 H, N5), 3.46 (s, 3 H, OMe), 3.47–3.50 (m, 1 H, G6), 3.56 (br d, 1 H, *J* 12.5 Hz, G6'), 3.65, (dd, 1 H, *J* 9.0, 9.0 Hz, G3), 3.75 (dd, 1 H, *J* 12.3, 2.3 Hz, N6), 3.83, (dd, 1 H, *J* 9.0, 8.5 Hz, G4), 4.02 (dd, 1 H, *J* 12.3, 3.8 Hz, N6'), 4.24 (dd, 2 H, *J* 10.3, 8.8 Hz, N2), 4.24 (d, 1 H, *J* 7.5 Hz, G1), 4.68 (ABq, 2 H, *J* 11.0 Hz, CH₂Ph), 4.97 (ABq, 2 H, *J* 12.0 Hz, CH₂Ph), 5.12 (dd, 1 H, *J* 10.0, 9.5 Hz, N4), 5.69 (dd, 1 H, *J* 10.8, 9.3 Hz, N3), 5.81 (d, 1 H, *J* 8.5 Hz, N1), 7.17–7.37 (m, 10 H, Ar), 7.75–7.89 (AA'BB', 4 H, Phth); ¹³C NMR: δ 20.30, 20.50, 20.61, 55.37, 57.05, 60.82, 61.33, 68.31, 70.76, 71.73, 74.04, 74.12, 74.58, 75.69, 82.10, 82.88, 98.09, 104.53, 123.40, 125.23, 126.10, 127.02, 127.54, 127.94, 128.16, 128.20, 128.27, 128.96, 131.54, 134.13, 138.14, 139.13, 169.34, 170.03, 170.57.

A solution of chromium trioxide (0.517 g, 5.17 mmol) in 3 M aq H₂SO₄ (0.27 mL concd H₂SO₄–1.40 mL H₂O) was added to a solution of **18** (0.43 g, 0.54 mmol) in 3:2 acetone–CH₂Cl₂ (7.0 mL) at 0°C. After 15 min the mixture was allowed to warm to RT. The reaction was quenched with EtOH after 4 h and the mixture was filtered. The filtrate was concentrated in vacuo to remove volatiles and the remaining aq phase was extracted with CHCl₃ (4 × 10 mL). The combined organic phases were washed with H₂O (10 mL) and satd aq NaCl (10 mL), dried (Na₂SO₄), and concentrated in vacuo to give a brown foam, which was treated with diazomethane generated from Diazald® [13]. The resulting brown solid was purified by flash column chromatography (5:1 toluene–EtOAc) to yield 0.26 g (58%) of a colorless oil identified as methyl *O*-(3,4,6-tri-*O*-acetyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)-(1 → 4)-(methyl 2,3-di-*O*-benzyl-β-D-glucopyranosid)uronate (**19**): $[\alpha]_D -1.9^\circ$ (*c* 0.12, CHCl₃); ¹H NMR: δ 1.86 (s, 3 H, Ac), 2.00 (s, 6 H, Ac), 3.42 (dd, 1 H, *J* 9.0, 7.5 Hz, U2), 3.46 (s, 3 H, OMe), 3.63 (dd, 1 H, *J* 9.0, 9.0 Hz, U3), 3.64 (s, 3 H, CO₂Me), 3.61–3.65 (m, 1 H, N5), 3.71 (d, 1 H, *J* 10.0 Hz, U5), 3.91 (dd, 1 H, *J* 12.0, 2.0 Hz, N6), 4.11–4.16 (m, 2 H, N6', U4), 4.24 (dd, 1 H, *J* 11.0, 8.5 Hz, N2), 4.27 (d, 1 H, *J* 7.5 Hz, U1), 4.70 (ABq, 2 H, *J* 11.3 Hz, CH₂Ph), 4.95 (ABq, 2 H, *J* 11.8 Hz, CH₂Ph), 5.14 (dd, 1 H, *J* 10.3, 9.3 Hz, N4), 5.58 (d, 1 H, *J* 8.0 Hz, N1), 5.78 (dd, 1 H, *J* 10.8, 9.3 Hz, N3), 7.17–7.40 (m, 10 H, Ar), 7.75–7.79 (m, 2 H, Phth), 7.89 (br s, 2 H, Phth); ¹³C NMR: δ 20.36, 20.55, 20.62, 52.54, 55.06, 57.18, 61.54, 68.53, 70.53, 71.63, 74.06, 74.54, 74.60, 77.49, 81.30, 82.07, 97.54, 104.75, 123.38, 125.26, 126.69, 127.20, 127.56, 127.93, 128.19, 128.22, 129.00, 134.05, 134.08, 134.12, 134.16, 138.13, 138.93, 168.24, 169.43, 170.71, 170.61.

A slurry of **19** (0.209 g, 0.255 mmol) and 30% Pd–C in 10:1 EtOAc–H₂O (11 mL) was subjected to H₂ (51 psig) in a Parr apparatus. After 4 days the mixture

was filtered through Celite and concentrated in vacuo to give a yellow oil. The oil was purified by flash column chromatography (2:1 EtOAc–toluene) to yield 0.125 g (77%) of a white foam that was identified as **20**. Iced N aq NaOH (5.8 mL) was added to a solution of **20** (0.125 g, 0.195 mmol) in MeOH (23 mL) at 0°C. After 2.5 h the mixture was neutralized to pH 7 with glacial AcOH (0.6 mL) and concentrated in vacuo to give a white solid. The white solid was dissolved in anhyd EtOH (18 mL), the solution was degassed with N₂, hydrazine monohydrate (3.1 mL, 64 mmol) was added, and the mixture was heated to 85°C. After 18.5 h the solution was concentrated in vacuo while removing the residual H₂O as a toluene–EtOH azeotrope to give a white solid. The solid was dissolved in anhyd MeOH (6.0 mL), the solution was cooled to 0°C, and Ac₂O (1.5 mL) was added. After 2 h the mixture was concentrated in vacuo and azeotroped with toluene to give a white solid, which was dissolved in H₂O, passed through a column of AG 50W-X4, and concentrated in vacuo to give a yellow solid. This solid was purified by flash column chromatography (1:1 CHCl₃–MeOH) to yield 0.0503 g (62%) of a white solid which was identified as **3**. The TLC and the ¹H and ¹³C NMR spectra of **3** all showed it to be a single compound, > 99% pure based upon the signal-to-noise ratios of the NMR spectra. ¹H NMR (D₂O): 2.04 (s, 3 H, Ac), 3.32 (dd, 1 H, *J* 9.5, 8.0 Hz, U2), 3.45–3.49 (m, 2 H, N4, N5), 3.52 (dd, 1 H, *J* 10.5, 9.0 Hz, N3), 3.54 (s, 3 H, OMe), 3.58 (dd, 1 H, *J* 9.5, 8.5 Hz, U3), 3.69 (m, 1 H, N2) 3.70 (m, 1 H, U5), 3.73 (m, 1 H, U4), 3.76–3.77 (m, 1 H, N6'), 3.90–3.93 (m, 1 H, N6), 4.37 (d, 1 H, *J* 7.8 Hz, U1), 4.54 (d, 1 H, *J* 8.5 Hz, N1); ¹³C NMR (D₂O): δ 23.3 (CH₃C = O), 55.96 (N2), 56.99 (OMe), 60.28 (N6), 69.46 (N4/N5), 72.43 (U2), 73.58 (U3,N3), 75.58 (N5/N4), 76.35 (U5), 79.70 (U4), 100.43 (N1), 103.86 (U1), 173.99 (C = O), 174.56 (C = O). MS: *m/z* calcd. for C₁₅H₂₄NO₁₂, 410.1299; found, 410.1294.

Acknowledgments

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