NEW N^4 -HYDROXYCYTIDINE DERIVATIVES: SYNTHESIS AND ANTIVIRAL ACTIVITY

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Dedicated to Professor Antonín Holý on the occasion of his 70th birthday in recognition of his invaluable contributions to the area of medicinal chemistry.

Two series of N^4 -hydroxycytidine derivatives were synthesized and evaluated as potential antipox virus agents. Acylation of N^4 -hydroxycytidine (1) with an excess of acyl chloride or imidazolide yielded the corresponding N^4 -(acyloxy) derivatives. 5'-Phosphonates of 1 were prepared by the reaction of cytidine 5'-phosphonates with aqueous hydroxylamine hydrochloride (pH 6.0). Nucleoside 1 and its N^4 -(acyloxy) derivatives inhibited replication of pox viruses in cell cultures, N^4 -(pivaloyloxy)- and N^4 -(benzoyloxy)cytidines being the most potent. The synthesized 5'-phosphonates were more cytotoxic than the parent nucleoside. **Keywords**: Nucleosides; Nucleoside analogs; N^4 -Hydroxycytidine; Nucleoside phosphonates; Hydroxylamine; Pyrimidines; Antiviral agents; Pox viruses.

Nucleoside analogues show a high antiviral potential. Nucleoside-derived drugs are approved for therapy of AIDS and infections induced by herpex virus, varicella zoster virus, and hepatitis B virus. Of their shortcomings, one can note their cytotoxicity and insufficient bioavailability, which could be improved by proper modifications of their structures. N^4 -Hydroxy-cytidine (1) was shown to have a broad spectrum of antiviral activity. In particular, 1 inhibited in cell culture assays some clinically important infections, induced by viruses of severe acute respiratory syndrome, influenza A and B¹, measles¹, hepatitis C² and some others. In this paper we describe the synthesis of new derivatives of 1 and evaluation of their antiviral prop-

erties in Vero and LLC-MK2 cell cultures infected with viruses of the pox family.

We prepared a series of N^4 -(acyloxy) derivatives of **1** based on the data on the positive effect of hydrophobic substituents in the molecules of antivirals³. The synthesis of N^4 -(acyloxy) derivatives (**2a**-**2d**) was performed according to Scheme 1. Acylation of **1** in pyridine with a small excess of the acyl chloride led to benzoyl (**2a**) or pivaloyl (**2b**) derivatives of **1**. In the case of N^4 -[(adamantane-1-carbonyl)oxy]cytidine (**2c**) and N^4 -(triphenylacetoxy)cytidine (**2d**), imidazolides of adamantane-1-carboxylic or triphenylacetic acid were used as acylating agents. The products were isolated on a silica gel column in a gradient of concentrations of MeOH in CHCl₃ in 54–79% yields. We did not observe the formation of the corresponding *N*-acyl derivatives, which correlates with Mertes's and Smrt's data⁴ on the exclusive formation of *O*-derivatives when modified the N^4 -(hydroxyamino) group of N^4 -hydroxy-6-azacytidine.



(i) benzoyl chloride, Py, 4 °C, 20 h; (ii) pivaloyl chloride, Py, 4 °C, 20 h; (iii) 1-(adamantane-

-1-carbonyl)imidazolide, DMF, 20 °C, 20 h; (iv) 1-(triphenylacetyl)imidazolide, DMF, 20 °C, 20 h

Scheme 1

Another approach of designing new antivirals involves the introduction of 5'-phosphonate fragments into molecules of nucleoside analogues. For example, 5'-phosphonate of 3'-deoxy-3'-azidothymidine is used in med-

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ical practice as an anti-HIV drug (Nicavir®)⁵. Phosphonate derivatives of acyclic nucleosides have a broad spectrum of an antiviral activity and three of them are already marketed – HepseraTM (adenovir, PMEA), VireadTM (tenofovir, PMPA) and VistideTM (cidofovir, HPMPC)⁶.

5'-O-Phosphonate derivatives of cytidine 3a-3c were prepared according to the reported procedures⁷. The reaction of 3a-3c with aqueous hydroxylamine hydrochloride (pH 6) under the conditions described in ref.⁸ (Scheme 2) afforded derivatives 4a-4c, which were purified by reversedphase silica gel chromatography and isolated in 56–83% yields.



(i) NH₂OH·HCI/H₂O, pH 6.0, 36 °C, 24 h

SCHEME 2

The structures of the synthesized compounds were confirmed by UV, ¹H, ¹³C and ³¹P NMR, and mass spectra. It is noteworthy that substitution of the 4-amino group in the cytosine base by a hydroxylamine group resulted in a significant upfield shift of H-6, H-5 and H-1' resonances in the ¹H NMR spectra compared with the parent cytidine phosphonates.

Stability of compounds **2a**, **2b** was evaluated in PBS and normal human blood serum at 37 °C according to the reported procedures⁹. Compounds **2a**, **2b** were stable in PBS to give only traces of **1** after 10 days. Half-life periods of compounds **2a**, **2b** in human blood serum were shown to be 40 ± 5 and 45 ± 5 min, respectively, to give only **1**.

Antiviral activity of the synthesized compounds was studied in Vero and LLC-MK2 kidney cell cultures infected with various pox viruses: vaccinia (VV), monkeypox (MPV) and cowpox (CPV) viruses. Antiviral efficiency was determined using a Neutral Red uptake assay according to ref.¹⁰ The data were graphed as shown in Fig. 1 for **2a** and summarized in the Table I. Compound **1** effectively suppressed replication of pox viruses (VV, MPV CPV) and was moderately toxic. Antiviral activities of **1** were higher than those of cidofovir, used as reference preparation in the antipox virus study-ing¹⁰, although the latter demonstrated a considerably lower toxicity. As a

result, selectivity indexes (SI) of **1** were comparable with those of cidofovir (3.6–15.3 versus 1.2–9.5 for different viruses and cell cultures). Antiviral properties of compounds **2a** and **2b** were higher than those of parent **1** due to lower cytotoxicity and higher activity in both cell cultures. In LLC-MK-2 cells SI values of compounds **2a** and **2b** achieved 50–100, whereas SI values of **1** were 5–15 for different viruses. The similar results were obtained in Vero cells (IS 10–67 for compounds **2a** and **2b** versus 3.6–5.6 for parent **1**). The introduction of more bulky substituents into the N^4 -position of the heterocyclic base (compounds **2c** and **2d**) dramatically increased cytotoxicity and did not effect antiviral activity.

As a rule, nucleoside phosphonates exhibit a lower activity in cell assays if compared with parent nucleosides, but due to a more potent reduction of cytotoxicity, the SI of these compounds are higher^{5,11}. Cytotoxicities of N^4 -hydroxycytidine 5'-phosphonate derivatives **4** in Vero cell culture were higher then that of the parent nucleoside (CC₅₀ 5.4–38 µM) and SI values of compounds **4a** and **4b** were low (3 and 4.4 correspondingly) Compound **4c** was inactive in these experiments. A similar result was obtained for 4'-thio-5-ethyl-2'-deoxyuridine, whose phosphonates were neither more active as anti herpes simplex virus agents nor less toxic⁹.



Fig. 1

Representative data from a single experiment for **2a** against various pox viruses on LLC-MK2 cells. Data are shown for a single replicate set of assays. IC₅₀ values (μ g/ml) for each strain are shown below the graph, as the R^2 fit of the points to the curve

TABLE I

Antiviral activity and cytotoxicity of N^4 -alkyloxycytidines **2a–2d** and 5'-phosphonate derivatives of N^4 -hydroxycytidine **4a–4c** in Vero and LLC-MK2 cell cultures

Compd	Cell culture	СС ₅₀ µм	VV		MPV		CPV	
			IC ₅₀ , µм	IS	IC ₅₀ , µм	IS	IC ₅₀ , µм	IS
1	Vero	50	8.9	5.6	10.1	5.0	14.0	3.6
	MK-2	77	15.1	5.1	5.0	15.3	13.9	5.76
2a	Vero	165	4.1	40.2	7.9	20.7	19.7	8.4
	MK-2	275	2.7	102	2.3	118	4.8	57.5
2b	Vero	175	2.6	67	8.4	21	18.0	9.7
	MK-2	480	4.7	103	2.6	186	6.7	77
2c	Vero	31	6.7	4.7	4.6	6.7	5.0	6.2
	MK-2	102	7.3	13.1	11.4	8.9	31.6	3.2
2d	Vero	23	1.3	17.7	2.9	8.2	6.4	3.6
	MK-2	47	1.4	34	4.0	11.7	22.5	2.1
4a	Vero	6.6	1.5	4.4	n.d.	n.d.	n.d.	n.d.
4b	Vero	5.4	1.8	3.0	n.d.	n.d.	n.d.	n.d.
4 c	Vero	38	38	1	n.d.	n.d.	n.d.	n.d.
Cido- fovir	Vero	360	44.1	8.1	37.6	9.5	47.7	7.5
	MK-2	360	83.5	4.3	112.2	3.0	298.7	1.2

VV, vaccinia virus; MPV, monkeypox virus; CPV, cowpox virus.

 CC_{50} is the minimal concentration of the compound, at which 50% cell death occurs. IC_{50} is the concentration of compounds at which proliferation of virus is inhibited by 50%. IS = CC_{50}/IC_{50} ; n.d., not determined.

To summarize, modification of N^4 -hydroxycytidine **1** by acylation of the N^4 -position may have some hopeful prospects, whereas its 5'-phosphonylation seems to be a dead lock.

EXPERIMENTAL

UV spectra (λ , nm; ϵ) were measured on a UV-2401 spectrophotometer (Shimadzu, Japan). ¹H NMR spectra were recorded on an AMX III-400 (Bruker, U.S.A.) with a working frequency of 400 MHz for ¹H NMR (Me₄Si and sodium 3-(trimethylsilyl)propane-1-sulfonate (DSS) were internal standards for CDCl₃ or DMSO-*d*₆ and D₂O solutions, respectively), 101 MHz for ¹³C NMR and 162 MHz for ³¹P NMR. ³¹P NMR spectra were recorded with ¹H-decoupling and 85% H₃PO₄ as an external standard. Chemical shifts (δ -scale) are given in ppm; cou-

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pling constants (*J*) in Hz. Mass spectra were registered on a Compact MALDI 4 spectrometer (Kratos Analytical, U.S.A.). Melting points were determined on a Kofler block and are uncorrected.

Starting Materials and Solvents

Adamantane-1-carboxylic acid, Et_3N , benzoyl chloride and pivaloyl chloride were purchased from Fluka; pyridine, dioxane, DMF, triphenylacetic acid, 1,1'-carbonyldiimidazole were from Aldrich; Silica gel 60 (40–63 µm) and LiChroprep RP-18 (25–40 µm) were from Merck, hydroxylamine hydrochloride was from Reakhim (Russia). Cytidine 5'-(ethoxycarbonyl)phosphonate (**3a**), cytidine 5'-phosphonate (**3b**) and 5'-O-(phosphonomethyl)cytidine (**3c**) were prepared according to published procedures⁷. Cidofovir was a kind gift of Dr Rosenberg (Institute of Organic Chemistry and Biochemistry, Academy of Sciences of the Czech Republic).

 N^4 -(Benzoyloxy)cytidine (2a) and N^4 -(Pivaloyloxy)cytidine (2b)

Benzoyl chloride or pivaloyl chloride (0.34 mmol) was added to a precooled (-20 °C) solution of **1** (80 mg, 0.31 mmol) in dry pyridine (2 ml). The mixture was kept at 4 °C for 20 h, MeOH (1 ml) was added and the solvents were evaporated. The residue was dissolved in chloroform (0.5 ml), applied onto a silica gel column (2×22 cm), and eluted with a gradient of methanol in chloroform (0 \rightarrow 20%, 750 ml). The target fractions were evaporated, the residue was crystallized from MeOH to give **2a** (76 mg, 72%) and **2b** (89 mg, 79%).

 N^4 -(Benzoyloxy)cytidine (2a): For C₁₆H₁₇N₃O₇ MS: m/z 362.3 (M⁺). M.p. 208–209 °C. UV (MeOH): λ_{max} 276 (7200), 231 (7300). ¹H NMR (CD₃OD): 7.54 m, 6 H (C₆H₅, H-6); 5.91 d, 1 H, $J_{1',2'}$ = 4.7 (H-1'); 5.82 d, 1 H, $J_{5,6}$ = 8.1 (H-5); 4.18 m, 2 H (H-2', H-3'); 3.99 t, 1 H, $J_{4',3'} \sim J_{4',5'}$ = 2.8 (H-4'); 3.82 dd, 2 H, J_{gem} = 12.7, $J_{5'a,4'}$ = 3.9 (H-5'a); 3.76 dd, 2 H J_{gem} = 12.7, $J_{5'b,4'}$ = 3.7 (H-5'). ¹³C NMR (CD₃OD): 61.09 (C-5'); 69.99 (C-2'); 72.93 (C-3'); 84.78 (C-4'); 87.31 (C-1'); 96.32 (C-5); 128.34 (CH-m-Bz); 129.86 (CH-i-Bz); 130.01 (CH-*o*-Bz); 133.03 (CH-*p*-Bz); 133.30 (C-6); 134.35 (C-2); 149.07 (C-4); 179.07 (C=O).

N^4 -[(Adamantane-1-carbonyl)oxy]cytidine (2c) and N^4 -(Triphenylacetoxy)cytidine (2d)

1,1'-Carbonyldiimidazole (81 mg, 0.5 mmol) was added to a solution of adamantine-1-carboxylic or triphenylacetic acid (0.33 mmol) in DMF (5 ml) and the mixture was kept at 20 °C for 1 h, then the mixture was added to a precooled (-20 °C) solution of 1 (78 mg, 0.3 mmol) in DMF (3 ml). After 20 h at 4 °C, the solvent was evaporated and the residue was partitioned between chloroform (20 ml) and water (5 ml). The organic layer was washed with water (5 ml), dried over anhydrous Na₂SO₄, and evaporated. The residue was dissolved in chloroform (0.5 ml), applied onto a silica gel column (2.5 × 9 cm), and eluted with a gradient of methanol in chloroform (0 \rightarrow 7%, 300 ml). The target fractions were evaporated in vacuum to give 68 mg of **2c** (54%) or 94 mg of **2d** (59%) as a white foam.
$$\begin{split} &N^4 - [(Adamantane-1-carbonyl)oxy]cytidine~(\textbf{2c}):~\text{For}~C_{20}\text{H}_{27}\text{N}_3\text{O}_7~(421.2)~\text{MS:}~m/z~420~(\text{M}^+).\\ &UV~(\text{MeOH}):~\lambda_{\text{max}}~276~(7200),~230~(7250).~^1\text{H}~\text{NMR:}~(\text{CDCl}_3):~9.75~\text{s},~1~\text{H}~(3\text{-NH});~7.05~\text{d},~1~\text{H},\\ &J_{6,5} = 8.1~(\text{H-6});~5.82~\text{d},~1~\text{H},~J_{1',2'} = 5.6~(\text{H-1'});~5.8~\text{d},~1~\text{H}~(\text{H-5});~4.27~\text{t},~1~\text{H},~J_{2',1'}\sim~^3J_{2',3'} = 5.6~(\text{H-2'});~4.17~\text{t},~1~\text{H},~J_{3',2'}\sim~^3J_{3',4'} = 5.6~(\text{H-3'});~4.04~\text{m},~1~\text{H}~(\text{H-4'});~3.7~\text{dd},~2~\text{H},~J_{\text{gem}} = 12.7,~^3J_{5'a,4'} = 3.1~(\text{H-5'a});~3.61~\text{dd},~2~\text{H},~J_{5'b,4} = 4.4~(\text{H-5'b});~1.94~\text{m},~15~\text{H}~(\text{Ada}). \end{split}$$

 λ_{max}^{3} (Triphenylacetoxy)cytidine (2d): For $C_{29}H_{27}N_3O_7$ (529.6) MS: m/z 528 (M⁺). UV (MeOH): λ_{max} 270 (7400), 228 (7300). ¹H NMR: (CDCl₃): 9.45 s, 1 H (3-NH); 7.46 d, 1 H, $J_{6,5}$ = 8.06 (H-6); 7.2 m, 15 H (3 C_6H_5); 6.81 d, 1 H (H-5); 5.86 d, 1 H, $J_{1',2'}$ = 5.0 (H-1'); 4.27 t, 1 H, $J_{2',1'} \sim J_{2',3'}$ = 5.2 (H-2'); 4.16 t, 1 H, $J_{3',2'} \sim J_{3',4'}$ = 5.1 (H-3'); 4.12 m, 1 H (H-4'); 3.82 dd, 2 H, J_{gem} = 12.7, $J_{5'a,4'}$ = 3.8 (H-5'a); 3.76 dd, 2 H, $J_{5'b,4'}$ = 4.2 (H-5'b).

 N^4 -Hydroxycytidine 5'-(Ethoxycarbonyl)phosphonate (**4a**), N^4 -Hydroxycytidine 5'-Phosphonate (**4b**) and N^4 -Hydroxy-5'-O-(phosphonomethyl)cytidine (**4c**)

A solution of hydroxylamine hydrochloride (174 mg, 2.5 mmol) in water (3 ml, pH 6.0) was added to the solution of corresponding cytidine derivative **3a–3c** (0.5 mmol) in water (3 ml). The mixture was kept at 36 °C for 24 h, then concentrated to 3 ml. The residue was applied onto a LiChroprep RP-18 column (2 × 17 cm) and eluted in a linear gradient of MeOH (0 \rightarrow 5%, 600 ml) in aqueous 0.01 M NH₄HCO₃. The target fractions were evaporated, co-evaporated with water (2 × 5 ml), the residue was dissolved in water (5 ml) and freeze-dried to yield 148 mg (72%) of **4a**, 121 mg (71%) of **4b** or 153 mg (83%) of **4c** as ammonium salts.

*N*⁴-Hydroxycytidine 5'-(ethoxycarbonyl)phosphonate (**4a**): For C₁₂H₁₈N₃O₁₀P (395.3) MS: *m/z* 394 (M⁺). UV: (H₂O, pH 2) λ_{max} 282 (10400), 221 (8990); (H₂O, pH 7) λ_{max} 271 (8100), 235 (8400). ¹H NMR (D₂O): 6.88 d, 1 H, *J*_{6,5} = 8.4 (H-6); 5.64 d, 1 H, *J*_{1',2'} = 6.5 (H-1'); 5.51 d, 1 H (H-5); 4.04-4.00 m, 2 H (H-2', H-3'); 3.97-3.95 m, 2 H (CH₂CH₃); 3.93 m, 1 H (H-4'); 3.91-3.87 m, 2 H (H-5'); 1.18 t, 3 H, *J*_{CH2.CH3} = 7.1 (CH₂CH₃). ³¹P NMR (D₂O): -4.51 s.

 N^4 -Hydroxycytidine 5'-phosphonate (**4b**): For C₉H₁₄N₃O₈P (323.2) MS: m/z 322 (M⁺). UV: (H₂O, pH 2) $\lambda_{\rm max}$ 282 (10300), 222 (8950); (H₂O, pH 7) $\lambda_{\rm max}$ 269 (8150), 235 (8600). ¹H NMR (D₂O): 6.85 d, 1 H, J_{5,6} = 8.1 (H-6); 6.49 d, 1 H, J_{H,P} = 641 (H-P); 5.52 d, 1 H (H-5); 5.64 d, 1 H, J_{1',2'} = 6.5 (H-1'); 4.05–3.99 m, 2 H (H-2', H-3'); 3.92 m, 1 H (H-4'); 3.83–3.73 m, 2 H (H-5'). ³¹P NMR (D₂O): 7.12 s. ¹³C NMR (D₂O): 60.48 d, J_{C,P} = 2.28 (C-5'); 67.46 (C-2'); 69.85 (C-3'); 80.28 d, J_{C,P} = 7.63 (C-4'); 85.69 (C-1'); 96.27 (C-5); 128.62 (C-6); 144.11 (C-2); 148.64 (C-4).

 N^4 -Hydroxy-5'-O-(phosphonomethyl)cytidine (4c): For $\rm C_{10}H_{16}N_3O_9P$ (353.2) MS: m/z 352 (M⁺). UV: (H₂O, pH 2) $\lambda_{\rm max}$ 281 (10500), 221 (9050); (H₂O, pH 7) $\lambda_{\rm max}$ 271 (8200), 235 (8500). ¹H NMR (D₂O): 7.40 d, 1 H, $J_{6,5}$ = 5.3 (H-6); 5.72 m, 2 H (H-5, H-1'); 4.17 t, 1 H, $J_{2',1'}$ = 5.3, $J_{2',3'}$ = 5.3 (H-2'); 4.10 t, 1 H, $J_{3',2'}$ = 4.4, $J_{3',4'}$ = 5.0 (H-3'); 4.00 d, 1 H, $J_{4',3'}$ = 3.1 (H-4'); 3.75 dd, 1 H, $J_{\rm gem}$ = 11.2, $J_{5'a,4'}$ = 2.5 (H-5'H); 3.65 dd, 1 H, $J_{5'b,4'}$ = 3.4 (H-5'b); 3.55 d and 3.57 d, 2 H, $J_{\rm Ha,P}$ = $J_{\rm Hb,P}$ = 9.0 (PCH₂O). ³¹P NMR (D₂O): 15.94 s. ¹³C NMR (D₂O): 68.72 d, $J_{\rm C,P}$ = 156 (C-P); 70.46 (C-2'); 72.37 (C-3'); 73.82 d, $J_{\rm C,P}$ = 9.8 (C-5'); 83.41 (C-4'); 86.89 (C-1'); 98.33 (C-5); 130.28 (C-6); 143.46 (C-2); 149.45 (C-4).

The *in vitro* cytotoxicity and antiviral effects of synthesized compounds in Vero, and LLC-MK2 kidney cell cultures infected with viruses of pox virus family (vaccinia, monkeypox and cowpox viruses) were determined using a neutral red uptake assay as described previously¹⁰. The data were plotted and analyzed by using the software program (Molecular Devices, Menlo Park, U.S.A.).

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