2-Deoxy-2-(substituted-methyl)analogs of β -Kdop

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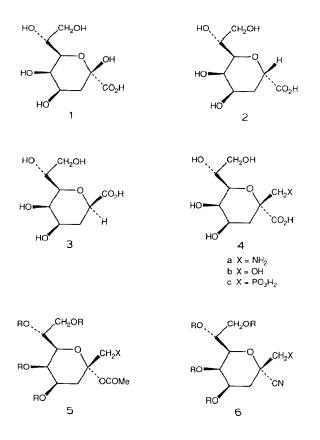
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ABSTRACT

Analogs of 3-deoxy-D-manno-octulosonic acid (Kdo) having the β -anomeric hydroxyl group replaced by a substituted-methyl group were prepared from Kdo. The substituted-methyl groups were derived from the carboxyl group of Kdo, whereas the carboxyl group in the analogs was introduced by the stereospecific reaction of the ketose azidomethyl acetate 12d with Me₃SiCN to give the nitrile 16. Hydrolysis of the nitrile group of 16 gave the azidocarboxylic acid 17a. Hydrogenation of 17a gave the amino acid 4a. Key steps in the preparation of the 2-hydroxymethyl and 2-phosphonomethyl analogs 4b and 4c, respectively, were the selective reduction of the azidomethyl derivative 17a to the aminomethyl derivative 17b, conversion of 17b by deamination to the β -lactone 19, and nucleophilic opening of the β -lactone ring of 19 with hydroxide and triethyl phosphite, respectively, to give 4b and 4c. An interesting contrast in the reaction with azide of the methyl and the (methylthio)methyl glycosides of the 1-O-alkylsulfonates 9b and 9f, respectively, was observed. Whereas the former reacted by direct displacement to give the unrearranged azide 9d, the reaction of the latter occurred with sulfur participation to give the rearranged azide 11.

INTRODUCTION

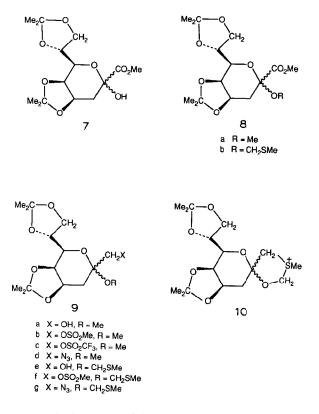
It was recently established in these laboratories¹ that, of the four interconverting tautomers of Kdo², only the β -pyranose 1 is bound to CMP-Kdo synthetase. Consistent with this observation was the discovery here³ that the 2,6-anhydro-3-deoxyaldonic acid analog 2 is a potent inhibitor of CMP-Kdo synthetase, whereas its 2-epimer 3 is virtually inactive. Although the analog 2 is locked to simulate the favorable β -pyranose form, it lacks the potential hydrogen-bond donor or acceptor present at C-2 in the β -pyranose tautomer (1) of Kdo. In the hope of preparing more potent inhibitors of CMP-Kdo synthetase based on the structure of 2, we planned the syntheses of 2-deoxy-2-(substituted-methyl) analogs 4a, 4b and 4c of β -Kdop (1). It was hoped that the substituted methyl groups of these analogs, with their potential hydrogen-bond donor or acceptor properties might interact with a binding site on CMP-Kdo synthetase normally occupied by HO-2 of 1, leading to stronger binding and consequently better competitive inhibitors. Our approach to the syntheses of 4a, 4b and 4c involved utilization of the Kdo carboxyl group as the precursor of the 2-(substituted-methyl) group. Subsequent application of the reaction of Me₃SiCN at the anomeric center of a suitable ketose derivative 5, thus derived, would introduce into the product 6 a nitrile group in a manner analogous to the reactions of aldose glycosyl acetates⁴. It was hoped that the favored stereochemistry of the reaction would lead to the glycosyl cyanide 6, which would be the



precursor of the carboxylic acids 4a, 4b and 4c. Alternative syntheses of C-glycosyl derivatives of Kdo utilizing enolate chemistry have been reported⁵.

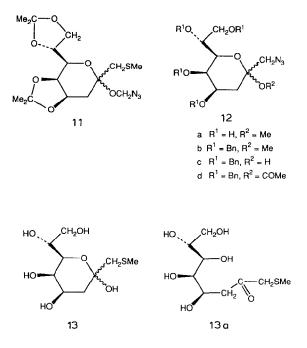
DISCUSSION

Treatment of Kdo with 2,2-dimethoxypropane in methanol, catalyzed by acidic resin gave the diisopropylidene acetal³ 7. The latter was converted by *O*-methylation into the methyl glycoside **8a**. Sodium borohydride reduction of **8a** gave the alcohol **9a** which was converted into the azide **9d** with sodium azide in DMF either via the methanesulfonate **9b** or the trifluoromethanesulfonate **9c**. Whereas the yield of the methyl glycoside **8a** by *O*-methylation of **7** was low, it was found that Albright– Goldman conditions⁶ applied to **7** gave the (methylthio)methyl glycoside **8b** in good yield. It was, therefore, hoped that the 2-*O*-(methylthio)methyl group of **8b** would provide a suitable protecting group for the C-2 of **7** required for preparation of the 2-(substituted-methyl) analogs. Sodium borohydride reduction of **8b** gave the desired alcohol **9e** in good yield, and the latter was readily converted into the methanesulfonate **9f**. In contrast to the vigorous conditions (125°, three days) required for conversion of the methanesulfonate **9b** of the methyl glycoside into the azide **9d** with sodium azide in



DMF, displacement of the methanesulfonate group of the (methylthio)methyl ether **9f** occurred under relatively mild conditions (100°, 20 h), under which the methanesulfonate **9b** of the methyl glycoside was quantitatively recovered. The difference in the ease of reaction with sodium azide in DMF of **9b** and **9f** was accounted for by the discovery that reaction of **9f** occurred with sulfur participation of the 2-*O*-(methylthio)methyl group *via* the five-membered cyclic sulfonium ion **10**. The latter **10** on reaction with azide ion gave rise to the rearranged product **11** rather than the product of direct displacement **9g**.

Confirmation of the structure of **11** was obtained from the nature of the products formed from **11** by mild acid-catalyzed hydrolysis. Whereas treatment of the methyl glycoside **9d** with 3:2 acetic acid-water selectively removed the isopropylidene groups to give the deprotected methyl glycoside **12a**, the same treatment of **11** gave a mixture of two products, both of which had lost not only the isopropylidene groups, but also the labile *O*-azidomethyl group. The more-polar product was characterized as one of the two possible pyranose anomers **13**, of the (methylthio)methyl ketoside by the magnitude of its coupling constant $J_{3,4}$ (12.6 Hz), which is characteristic of vicinal diaxial protons in six-membered rings. The ¹³C-n.m.r. spectrum of this product lacked a resonance near 86 p.p.m., which is characteristic of the C-5 carbon atoms of the furanose forms^{3a} of Kdo. ¹³C-N.m.r. studies showed that **13** was almost completely



converted into the less-polar product in acetic acid-water solution in about 20 h. ¹H-N.m.r. studies with 13 showed exchange in D_2O of the methylene protons of the (methylthio)methyl group as well as the C-3 methylene protons which may occur via the open form 13a.

The less-polar product 14 was characterized as a furanose structure by the resonance of its C-5 carbon at 86 p.p.m. (Table I), which is close to that observed^{2a} for the C-5 carbon resonances of both of the anomers of the furanose forms of Kdo. An anhydro form was suggested by the high-resolution f.a.b. mass spectrum. This was confirmed by n.m.r. studies (Table I), which established the formation of a C-2 to C-7 bridge from the absence of the hydroxyl groups at C-2 and C-7. The location of the hydroxyl groups in 14 was established both by the deuterium-induced β -isotope effects on the ¹³C resonances only on their hydroxyl-bearing carbons at C-4, C-6, and C-8, and by the vicinal coupling constants in Me₂SO between the C-4, C-6, and C-7 protons and the protons attached to their respective hydroxyl groups (Table I). Reduction of the azidomethyl glycoside 11 by LiAlH₄ gave rise to a product the n.m.r. spectrum and analysis of which were compatible with the ring-opened methylthio diol 15.

The azidomethyl methyl glycoside 12a was converted into the tetra-O-benzyl derivative 12b. Acid-catalyzed hydrolysis of 12b to give 12c, followed by acetylation, gave the glycosyl acetate 12d. Conversion of the glycosyl acetate 12d to the nitrile 16 was effected with Me₃SiCN in acetonitrile, catalyzed by boron trifluoride etherate. Base-catalyzed hydrolysis of 16 gave the carboxylic acid 17a. The conformation of the pyranose ring of 17a was established as 17a' from the coupling constant $J_{3a,4a} = 12.3$ Hz.

¹ H-N.m.r. data (Me_2SO-d_6) $\delta p.p.m. vs.$ $\delta_{Me_2SO} = 2.5$		Coupling constants (Me ₂ SO) (Hz)	¹ H-N.m.r. data (D_2O) $\delta p.p.m.$ vs. $\delta_{HOD} = 4.8$			$\Delta\delta$ ($H_2O/$ $D_2O)$
SMe	2.12		2.22	SMe	17.3	ь
H-1	2.74(2H)		2.95, 3.01	C-1	40.0	ь
H-2				C-2	109.3	-0.057
H-3a	1. 91		2.08	C-3	43.8	0.043
H-3b	2.32	$J_{3a,3b}$ 11.7	2.64	C-4	69.9	0.072
H-4	4.40	$J_{3a,4}$ 2.0, $J_{3b,4}$ 6.0	4.63	C-5	86.0	0.015
HO-4	4.95	J _{OH 4} 3.7		C-6	63.0	0.071
H-5	3.96	$J_{OH,4}^{J} 3.7$ $J_{4,5}^{b}$	4.27	C-7	76.2	0.020
H-6	3.37	J _{5.6} 3.8	3.69	C-8	62.0	0.082
HO-6	5.20	J _{OH,6} 3.9				
H-7	3.37	J ₆₇ 4.8	3.50			
H-8a	3.39	$J_{7.8a}^{0.7}$ 4.5	3.68			
H-8b	3.56	$J_{7,8b}$ 1.8, $J_{8a,8b}$ 10.0	3.79			
HO-8	4.49	$J_{OH,(8a,8b)}$ 4.7				

TABLE I^a

N.m.r. Data of Compound 14

^a Proton assignments were determined by selective homonuclear decoupling. Carbon ¹³C assignments were determined by a 2-D proton–carbon correlation. The ¹³C deuterium-induced isotope shifts were determined by obtaining a ¹³C-n.m.r. spectrum in both H₂O and D₂O solutions. The resonances of carbons bearing hydroxyl groups are shifted upfield when the OH proton is substituted with a deuteron. As the β -isotope shift is larger than other long-range isotope shifts, the carbons bearing hydroxyl groups are easily identified as C-4, C-6, and C-8. This is consistent with proton data obtained in Me₂SO. ^b Very small.

The configuration of 17a at C-2 could thus be established by heteronuclear decoupling by the method of Unger⁷, since the coupling constant $J_{3a,H-1} = 8.1$ Hz established the antiperiplanar relationship between the carboxyl group and the C-3-axial proton. This method was used to establish the C-2 configurations of related *C*-glycosyl analogs of Kdo.⁵

Catalytic hydrogenation of 17a gave the amino acid 4a.

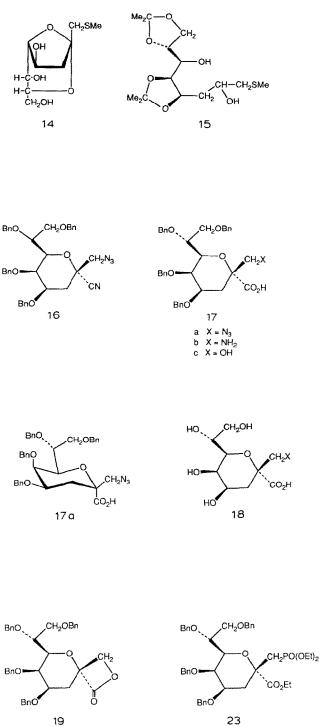
Treatment of the azido acid 17a with Zn in acetic acid gave a mixture of the amino acid 17b (49%) and the β -lactone 19 (40%). Deamination of the amino acid 17b with sodium nitrite in aqueous acetic acid gave the β -lactone 19.

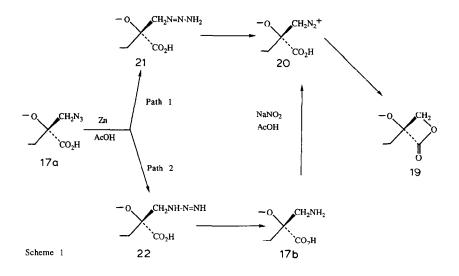
Formation of the β -lactone by nitrous acid deamination of 17b, which must proceed via the diazonium ion 20 (Scheme 1), suggests that partitioning of the azido acid 17a between the amino acid 17b and the lactone 19 on treatment with Zn and acetic acid occurs via the isomeric azoamino compounds 21 and 22 (Scheme 1). The former leads to the lactone 19 via the diazonium ion 20, while the latter leads to the amino acid 17b.

Base-catalysed hydrolysis of the lactone 19 smoothly gave rise to the hydroxy acid 17c which was O-debenzylated by catalytic hydrogenation to give 4b.

Opening of the lactone ring of 19 with triethyl phosphite gave the triethyl ester 23, which was deprotected by base-catalysed hydrolysis and catalytic hydrogenation to give

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the phosphonic acid 4c. The corresponding reaction of β -propiolactone has been reported⁸.

All of the new analogs **4a**, **4b** and **4c** showed only weak activities as CMP-Kdo synthetase inhibitors compared with 2,6-anhydro-3-deoxy-D-glycero-D-galacto-octonic acid ("2-deoxy-a-carboxy Kdo", **2**). The concentrations $[I_{s0} (\mu M]$ of compounds **2**, **4a**, **4b**, and **4c** which caused 50% inhibition of CMP-Kdo synthetase were 10.5, 11 000, 1100, and 9200, respectively.

EXPERIMENTAL

General methods. — N.m.r. spectra were determined at 300 MHz with a General Electric GN 300 nuclear magnetic resonance spectrometer, high-resolution mass spectra with a Kratos MS50 mass spectrometer, and i.r. spectra with a Perkin–Elmer model 283B or a Nicolet 60 SX FT spectrometer. All compounds had i.r. absorptions characteristic of the chromophores present. Optical rotations were determined with a Perkin–Elmer model 241 digital polarimeter. Gravity chromatography was performed with Merck, Darmstadt 70–230 mesh silica gel, and flash chromatography was carried out with Merck, Darmstadt 230–400 mesh silica gel. Extractions with CHCl₃ were carried out by shaking the mixtures or solutions with mixtures of CHCl₃ and 5% aq. NaHCO₃. The CHCl₃ solutions were separated and dried (MgSO₄), the CHCl₃ was evaporated under diminished pressure, and the residues were dried under high vacuum.

Methyl 3-deoxy-4,5;7,8-di-O-isopropylidene-D-manno-2-octulopyranosonate (7). — A mixture of 10 g (36.6 mmol) of Kdo ammonium salt, 2,2-dimethoxypropane (200 mL), dry MeOH (200 mL), and MeOH-washed and dried AG 50Wx8 (H^+) resin (20 g) was stirred for 3 d at room temperature. The mixture was filtered, the filtrate was evaporated and the residue flash chromatographed (80:20:1.5:1 PhMe-EtOAc-MeOH–Et₃N) to yield 6.4 g (53%) of 7 as a white solid. An analytical sample was prepared by gravity chromatography with the same solvent system; m.p. $132-134^{\circ}$, $[a]_{p}^{20} + 28.2^{\circ}$ (c 1, CHCl₃); ¹H-n.m.r. (CDCl₃): δ 1.92 (dd, 1 H, $J_{3,4}$ 4.9, $J_{3,3'}$ 14.4 Hz, H-3), 2.51 (dd, 1 H, $J_{3'4}$ 6.6 Hz, H-3'), and 3.82 (s, 3 H, CO₂Me).

Anal. Calc. for C₁₅H₂₄O₈: C, 54.20; H, 7.28. Found: C, 53.57; H, 7.44.

Methyl (methyl 3-deoxy-4,5;7,8-di-O-*isopropylidene*-D-manno-2-octulopyranosid) onate (**8a**). — Methyl iodide (1.87 mL, 30 mmol) was added to a stirred mixture of 3.32 g (10 mmol) of **7**, 4.63 g (20 mmol) of Ag₂O, and 0.11 g (0.3 mmol) of Bu₄NI in 80 mL of dry DMF. After stirring for 24 h, the mixture was filtered and the insoluble salts washed with DMF. The combined filtrates were evaporated under diminished pressure. Chloroform was added to the residue and the solution was filtered through a Celite mat. The filtrate was washed with H₂O, dried (MgSO₄) and evaporated. Flash chromatography with 1:10 EtOAc-CHCl₃ gave 1.3 g (38%) of **8a** as a solid; m.p. 114–117°; ¹H-n.m.r. (CDCl₃): δ 1.32, 1.38, 1.43, 1.45 (4 s, 12 H, CMe₂), 1.87 (dd, 1 H, J_{3,4} 3, J_{3,3'} 15 Hz, H-3), 2.78 (dd, 1 H, J_{3'4} 3.6 Hz, H-3'), 3.23 (s, 3 H, OMe), and 3.78 (s, 3 H, CO₂Me). Anal. Calc. for C₁₆H₂₆O₈: C, 55.48; H, 7.57. Found: C, 54.60; H, 7.50.

Methyl 3-deoxy-4,5;7,8-di-O-isopropylidene-D-manno-2-octulopyranoside (9a). — To a stirred solution of 776 mg (2.34 mmol) of the ester 8a in a 18 mL of MeOH, cooled in an ice-water bath, was added a freshly prepared solution of 614 mg (16.2 mmol) of NaBH₄ in 6 mL of water. Stirring was continued with cooling for 1 h and then at room temperature overnight. Extraction with CHCl₃ gave a syrup which on chromatography using 1:1 EtOAc-CHCl₃ gave 527 mg (74%) of 9a as a syrup; ¹H-n.m.r. (CDCl₃): δ 1.33, 1.36, 1.42, 1.44 (4s, 12 H, CMe₂), 1.68 (dd, 1 H, J_{3,4} 3.9, J_{3,3}, 15.9 Hz, H-3), 2.47 (dd, 1 H, J_{3,4} 3.6 Hz, H-3'), and 3.26 (s, 3 H, OMe).

Methyl 1-azido-1,3-dideoxy-4,5;7,8-di-O-isopropylidene-D-manno-2-octulopyranoside (9d). — To a stirred solution of the alcohol (9a) [prepared by NaBH₄ reduction of 1.38 g (3.98 mmol) of ester 8a, as just described] in 22 mL of pyridine, cooled in an ice-water bath, was added 1.10 mL (6.53 mmol) of $(CF_3SO_2)_2O$. Stirring was continued with cooling for 1 h. Extraction with CHCl₃ gave the trifluoromethanesulfonate 9c as a syrup. The latter was immediately dissolved in 34 mL of Me₂SO and treated with 1.05 g (16.2 mmol) of NaN₃. The resulting solution was heated for 3 h at 75°. Extraction with CHCl₃ gave a syrup which on chromatography with 10:1 PhMe₃-EtOAc gave 0.685 g (50%) of 9d; $[a]_{D}^{25} + 41°$ (c 1, MeOH); ¹H-n.m.r. (CDCl₃): δ 1.33, 1.36, 1.41, 1.43 (4s, 12 H, CMe₂), 1.66 (ddd, 1 H, J_{3,3}, 16.5, J_{3,4} 3.9 Hz, H-3), 2.41 (dd, 1 H, H-3'), 3.27 (s, 3 H, OMe), 3.38 (d, 1 H, J_{AB} 13.5 Hz, CH_AH_BN₃), and 3.65 (dd, 1 H, J_{BH-3} 1.5 Hz, CH_AH_BN₃).

Anal. Calc. for $C_{15}H_{25}N_3O_6$: C, 52.46; H, 7.34; N, 12.24. Found: C, 52.78; H, 7.56; N, 12.19.

Methyl 3-deoxy-4,5;7,8-di-O-isopropylidene-1-O-methylsulfonyl-D-manno-2-octulopyranoside (**9b**). — To a stirred solution of 514 mg (1.6 mmol) of the alcohol **9a** in 7 mL of pyridine, cooled in an ice-water bath was added 0.30 mL (3.8 mmol) of MeSO₂Cl. Stirring was continued with cooling for 1 h and then for 1.5 h at room temperature. Extraction with CHCl₃ gave the methanesulfonate **9b** as a syrup; ¹H-n.m.r. (CDCl₃): δ 1.34, 1.38, 1.42, 1.44 (4s, 12 H, CMe₂), 1.66 (ddd, 1 H, $J_{3,4}$ 3.6, $J_{3,3'}$ 16.5 $J_{1,3}$ 1.5 Hz, H-3), 2.46 (dd, 1 H, $J_{3',4}$ 3.6 Hz, H-3'), 3.08 (s, 3 H, OSO₂Me), and 3.32 (s, 3 H, OMe). Attempted reaction of the methanesulfonate **9b** with NaN₃ in DMF at 100°. — A. A stirred solution of the methanesulfonate **9b**, prepared as just described, 427 mg (6.57 mmol) of NaN₃, and 15 mL of Me₂SO was heated for 18 h at 100°. Extraction with CHCl₃ gave 629 mg of recovered **9b**.

B. A stirred solution of 581 mg (1.46 mmol) of the methanesulfonate **9b**, 849 mg (13 mmol) of NaN₃, and 15 mL of Me₂SO was heated at 125° for 3 d. Extraction with CHCl₃ gave 439 mg of syrup which on chromatography using 20:1 CHCl₃-EtOAc gave 277 mg (55%) of **9d**, identical with that prepared as already described.

Methyl [(methylthio)methyl 3-deoxy-4,5;7,8-di-O-isopropylidene-D-manno-2octulopyranosid]onate (**8b**). — Dry Me₂SO (47 mL) was added to a stirred solution of 5.4 g (16 mmol) of 7 in 47 mL of Ac₂O. The solution was stirred for 22 h at room temperature. To the solution was added 500 mL of EtOAc and the resulting solution was washed thoroughly with H₂O, 10% aqueous NaCl, and was evaporated under diminished pressure. The residue was flash chromatographed (1:8 EtOAc–PhMe) to give 5.8 g (91% **8b** as a syrup; ¹H-n.m.r. (CDCl₃): δ 1.32, 1.38, 1.39, 1.43 (4s, 12 H, CMe₂), 1.88 (dd, 1 H, J_{3,4}2.7, J_{3,3} 15.5 Hz, H-3), 2.92 (dd, 1 H, J_{3',4} 3.8 Hz, H-3'), 2.17 (s, 3 H, SMe), and 3.77 (s, 3 H, CO₂Me).

(Methylthio)methyl 3-deoxy-4,5;7,8-di-O-isopropylidene-D-manno-2-octulopyranoside (9e). — To a stirred solution of 3.4 g (8.7 mmol) of the methyl ester **8b** in 60 mL of MeOH was added a freshly prepared solution of 2.74 g (72.4 mmol) of NaBH₄ in 30 mL of water, and stirring was continued overnight. Extraction with CHCl₃ gave the alcohol **9e** as a syrup; ¹H-n.m.r. (CDCl₃): δ 1.33, 1.37, 1.40, 1.41 (4s, 12 H, CMe₂), 1.58 (dd, 1 H, $J_{3,4}$ 3.6, $J_{3,3'}$ 16.5 Hz, H-3), 2.61 (dd, 1 H, $J_{3',4}$ 3.0 Hz, H-3'), 2.28 (s, 3 H, SMe), 2.96 (t, 1 H, $J_{1,OH}$ 7.2 Hz, disappears on D₂O exchange, OH), 3.67 (d, 2 H, singlet after D₂O exchange H-1), 4.56 (d, 1 H, J_{AB} = 12 Hz, H_A of OCH_AH_BSMe), and 4.85 (d, 1 H, H_B).

(Methylthio) methyl 3-deoxy-4,5;7,8-di-O-isopropylidene-1-O-methylsulfonyl-D-manno-2-octulopyranoside (9f). — To a stirred solution of the alcohol 9e, prepared as just described, in 45 mL of pyridine, cooled in an ice bath, was added 1.8 mL (23.3 mmol) of MeSO₂Cl. Stirring was continued for 1 h with cooling and then for 1 h at room temperature. Extraction with CHCl₃ gave the methanesulfonate 9f as a syrup; ¹H-n.m.r. (CDCl₃): δ 1.32, 1.38, 1.42, 1.44 (4s, 12 H, CMe₂), 1.87 (dd, 1 H, $J_{3,4}$ 3.0, $J_{3,3}$ 16.2 Hz, H-3), 2.78 (dd, 1 H, $J_{3,4}$ 4.2 Hz, H-3'), 2.23 (s, 3 H, SMe), and 3.78 (s, 3 H, OSO₂Me).

Azidomethyl 3-deoxy-4,5;7,8-O-di-isopropylidene-1-S-methyl-1-thio-D-manno-2octulopyranoside (11). — A stirred solution of the methanesulfonate 9f, prepared as just described, 2.4 g (36.9 mmol) of NaN₃ and 84 mL of Me₂SO was heated for 20 h at 100°. Extraction with CHCl₃ gave 3.68 g of 11 (100% based on 8b) as a syrup; ¹H-n.m.r. (CDCl₃): δ 1.31, 1.35, 1.40, 1.43 (4s, 12 H, CMe₂), 1.77 (ddd, 1 H, $J_{3,4}$ 3.6, $J_{3,3}$ · 16.5, $J_{1,3}$ 1.5 Hz, H-3), 2.70 (dd, 1 H, $J_{3',4}$ 4.2 Hz, H-3'), 2.18 (s, 3 H, SMe, 2.86 (d, 1 H, J_{AB} 13.8 Hz, H_A of CH_AH_BSMe), 3.02 (bd, 1 H, H_B), 4.73 (d, 1 H, $J_{A'B'}$ 9.0 Hz, $H_{A'}$ of OCH_{A'}H_{B'}N₃), and 4.77 (d, 1 H, H_{B'}).

Methyl 1-azido-1,3-dideoxy-D-manno-2-octulopyranoside (12a). — A solution of 277 mg (0.8 mmol) of the azide 9d in 15 mL of 3:2 AcOH-water was kept for 19 h at

room temperature. Solvent was evaporated under diminished pressure. The residue was dissolved in 10 mL of water and the water was removed by lyophilization leaving 216 mg (100%) of **12a**; ¹H-n.m.r. (D₂O): δ 1.81 (t, 1 H, $J_{3a,4} = J_{3a,3e} = 13.2$ Hz, H-3a), 1.97 (dd, 1 H, $J_{3e,4}$ 5.4 Hz, H-3e), 3.27 (s, 3 H, OMe), 3.31 (d, 1 H, J_{AB} 13.5 Hz, $CH_AH_BN_3$), and 3.66 (d, 1 H, $CH_AH_BN_3$).

3-Deoxy-1-S-methyl-1-thio-D-manno-2-octulopyranose (13) and 2,7-anhydro-3deoxy-1-S-methyl-1-thio-a-D-manno-2-octulofuranose (14). — A solution of 457 mg (1.2 mmol) of the azide 11 in 25 mL of 3:2 AcOH-water was kept at room temperature overnight. The solvent was evaporated under diminished pressure. The residue was dissolved in water and the water was removed by lyophilization. Chromatography using 10:1 EtOAc- MeOH gave 143 mg (52%) of 14 in the early fractions; $[a]_{D}^{19} + 38.4^{\circ}$ (c 1, MeOH); ¹H-n.m.r. (see Table I); exact mass: calc. for C₉H₁₇O₅S (MH)⁺ 237.0797, found 237.0797; and 62.4 mg (22.5%) of 13 in the later fractions; $[a]_{D}^{21} + 43.5^{\circ}$ (c 1, MeOH); ¹H-n.m.r. (D₂O): δ 1.82 (t, 1 H, J_{3a,3e}, J_{3a,4} 12.6 Hz, H-3a), 1.97 (dd, 1 H, J_{3e,4} 5.4, J_{3a,3e} 12.6 Hz, H-3e), 2.18 (s, 3 H, SMe), 2.86 (d, 1 H, J_{AB} 14.1 Hz, CH_AH_BSMe), and 2.76 (d, 1 H, CH_AH_BSMe); exact mass calc. for C₉H₁₉O₆S (MH)⁺ 255.0902, found 255.0904.

3-Deoxy-4,5;7,8-di-O-isopropylidene-1-S-methyl-1-thio-D-glycero-D-galacto-(or D-talo)-octitol (15). — To a stirred slurry of 73.5 mg (1.94 mmol) of LiAlH₄ in 6 mL of ether was added dropwise a solution of 305 mg (0.78 mmol) of the azide 11 in 10 mL of ether. Stirring was continued for 5 h followed by addition of 3 mL of MeOH. Extraction with ether gave 195 mg of syrup which on chromatography with 1:1 CHCl₃–EtOAc gave 138 mg (52%) of 15 as a syrup; ¹H-n.m.r. (CDCl₃): δ 1.34, 1.38, 1.39, 1.52 (4s, 12 H, CMe₂), and 2.16 (s, 3 H, SMe).

Anal. Calc. for $C_{15}H_{27}O_6S$: C, 53.71; H, 8.11; S, 9.56. Found: C, 53.84; H, 8.32; S, 9.11.

Methyl 1-azido-4,5,7,8-tetra-O-benzyl-1,3-dideoxy-D-manno-2-octulopyranoside (12b). — To a stirred solution of 1.19 g (4.52 mmol) of the azide 12a in 79 mL of DMF, under nitrogen, was added 1.78 g (44.3 mmol) of 60% NaH. Stirring was continued for 0.5 h. The resulting stirred mixture was cooled in an ice-water bath and 0.143 g (0.387 mmol) of Bu₄NI was added followed by 4.3 mL of PhCH₂Br. After the addition was complete, stirring was continued for 1 h at room temperature and then overnight at room temperature. Extraction with CHCl₃ gave 5.2 g of a dark-brown syrup. Flash chromatography using 50:1 PhMe–EtOAc followed by gravity chromatography using 50:1 PhMe–EtOAc gave 1.49 g (52.8%) of 12b; $[a]_D^{25} + 18.7^\circ$ (c 1, CHCl₃); ¹H-n.m.r. (CDCl₃): δ 2.08 (dd, 1 H, $J_{3a,3e}$ 12, $J_{3e,4}$ 5 Hz, H-3e), 2.18 (t, 1 H, $J_{3a,4}$ 12 Hz, H-3a), 3.18 (s, 3 H, OMe), 3.20 (d, 1 H, J_{AB} 13.5 Hz, $CH_AH_BN_3$), 3.35 (d, 1 H, $CH_AH_BN_3$).

Anal. Calc. for $C_{37}H_{41}N_3O_6$: C, 71.25; H, 6.63; N, 6.74. Found: C, 71.58; H, 6.80; N, 6.10.

1-Azido-4,5,7,8-tetra-O-benzyl-1,3-dideoxy-D-manno-2-octulopyranose (12c). — A stirred solution of 1.49 g (2.39 mmol) of the methyl glycoside 12b in 70 mL of AcOH was heated to 80° and 25 mL of $2M H_2SO_4$ was added over a period of 12 min. After the addition was complete, heating was continued for 1.5 h. The resulting solution was cooled in an ice-water bath and then shaken with a mixture of 200 mL of CHCl₃ and 500

mL of ice-water. The CHCl₃ solution was separated and washed with 500 mL of 5% NaHCO₃ (considerable foaming), and then with 500 mL of water. The aqueous solutions were washed with CHCl₃. The CHCl₃ solutions were combined and dried (MgSO₄). Evaporation of the CHCl₃ under diminished pressure and chromatography of the residue with 20:1 CHCl₃-EtOAc gave 576 mg (40%) of 12c as a syrup; ¹H-n.m.r. (CDCl₃): δ 1.89 (dd, 1 H, $J_{3e,4}$ 6, $J_{3e,3a}$ 12 Hz, H-3*e*), 2.03 (t, 1 H, $J_{3a,4}$ 12 Hz, H-3*a*), 2.93 (d, 1 H, J_{AB} 12.9 Hz, CH_AH_BN₃), and 3.33 (d, 1 H, CH_AH_BN₃).

I-O-Acetyl-1-azido-4,5,7,8-tetra-O-benzyl-1,3-dideoxy-D-manno-2-octulopyranose (12d). — To a stirred solution of 3.31 g (5.4 mmol) of the azide 12c in 65 mL of pyridine, cooled in an ice-water bath, was added dropwise over a period of 10 min, 26 mL of Ac₂O followed by 0.1024 g of 4-dimethylaminopyridine. Stirring was continued with cooling for 1 h and then overnight at room temperature. The resulting solution was poured into 500 mL of water and the resulting suspension was cooled in an ice-water bath. Solid NaHCO₃ was added portionwise with stirring. Extraction with CHCl₃ gave an orange syrup which on chromatography using 20:1 PhMe-EtOAc gave 2.71 g (77%) of 12d; ¹H-n.m.r. (CDCl₃): δ 1.92 (s, 3 H, OCOMe), 2.16 (dd, 1 H, $J_{3a,4} = J_{3a,3e} = 12.3$ Hz, H-3a), 2.51 (t, 1 H, $J_{3e,4}$ 4.8, $J_{3a,3e}$ 12.3 Hz, H-3a), and CH₂N₃ (not resolved).

1-Azido-4,5,7,8-tetra-O-*benzyl-1,3-dideoxy-a*-D-manno-2-octulopyranosyl cyanide (16). — To a stirred solution of 3.85 g (5.9 mmol) of 12d in 28 mL of dry MeNO₂ was added 6.0 mL (45 mmol) of Me₃SiCN, followed by a few drops of BF₃-Et₂O. The solution was stirred at room temperature for 1 h. Extraction with CHCl₃ and flash chromatography with 40:1 PhMe-EtOAc yielded 3.45 g (94%) of 16 as a light syrup; $[a]_{D}^{22} + 39^{\circ}$ (c 1, CHCl₃); ¹H-n.m.r. (CDCl₃): δ 2.03 (dd, 1 H, $J_{3e,4}$ 3.3, $J_{3e,3a}$ 12.1 Hz, H-3e), 2.28 (t, 1 H, $J_{3a,4}$ 12.1 Hz, H-3a), 3.32 (d, 1 H, J_{AB} 12.9 Hz, $CH_AH_BN_3$), and 3.50 (d, 1 H, $CH_AH_BN_3$).

Anal. Calc. for C₃₇H₃₈N₄O₅: C, 71.82; H, 6.19; N, 9.05. Found: C, 71.54; H, 6.03; N, 9.38.

2,6-Anhydro-2-C-(azidomethyl)-4,5,7,8-tetra-O-benzyldeoxy-D-glycero-D-talooctonic acid (17a). — A solution prepared from 3.45 g (5.57 mmol) of 16, 18 g (0.45 mol) of NaOH, 45 mL of water and 180 mL of EtOH was stirred and heated under nitrogen for 4 days at 70°. The resulting solution was cooled to room temperature and shaken with excess M HC1. Extraction with CHCl₃ gave 3.29 g of a syrup which on flash chromatography using 20:2:0.1 PhMe–EtOAc–AcOH gave 2.89 g (81%) of 17a as a syrup; $[a]_{D}^{22}$ + 10.2° (c 1, CHCl₃); ¹H-n.m.r. (CDCl₃): δ 2.15 (t, 1 H, $J_{3a,3e} = J_{3a,4} = 12.3$ Hz, H-3a), 2.61 (dd, 1 H, $J_{3e,4}$ 4.2 Hz, H-3e), 3.30 (d, 1 H, J_{AB} 12.8 Hz, $CH_AH_BN_3$), and 3.49 (d, 1 H, $CH_AH_BN_3$).

Anal. Calc. for C₃₇H₃₉N₃O₇: C, 69.68; H, 6.16; N, 6.59. Found: C, 69.84; H, 6.84; N, 6.55.

2-C-(Aminomethyl-2,6-anhydro-4,5,7,8-tetra-O-benzyl-3-deoxy-D-glycero-Dtalo-octonic acid (17b) and 2,6-anhydro-4,5,7,8-tetra-O-benzyl-3-deoxy-2-C-(hydroxymethyl)-D-glycero-D-talo-octonic acid lactone (19) by Zn-AcOH reduction of 17a. — To a stirred solution of 171 mg (0.27 mmol) of the azido acid 17a in 8.6 mL of AcOH was added 1.06 g of Zn dust. Stirring was continued for 2 h. The Zn was removed by filtration. Acetic acid was evaporated under diminished pressure leaving a syrup which on chromatography using 20:2:0.1 CHCl₃–MeOH–HCl gave in the early fractions 60 mg (40%) of the β -lactone **19**; $[a]_{D}^{21}$ + 0.47° (*c* 1, MeOH); ¹H-n.m.r. (CDCl₃): δ 2.26 (dd, 1 H, $J_{3e,4}$ 4.0, $J_{3e,3a}$ 12.6 Hz, H-3*e*), and 2.48 (t, 1 H, $J_{3a,4}$ 12.6 Hz, H-3*a*).

Anal. Calc. for C₃₂H₃₈O₇: C, 74.72; H, 6.44. Found: C, 74.46; H, 6.35.

In the later fractions there was obtained 106 mg(49%) of the hydrochloride of the amino acid **17b**; ¹H-n.m.r. (CDCl₃): broad peaks, no definition.

2,6-Anhydro-4,5,7,8-tetra-O-benzyl-3-deoxy-2-C-(hydroxymethyl)-D-glycero-Dtalo-octonic acid lactone (19). — A. By deamination of 17b. — To a stirred solution of 102 mg (0.16 mmol) of the hydrochloride of the amino acid 17b, 2.0 mL of AcOH and 2.0 mL of water was added 82 mg of NaNO₂. Stirring was continued for 1 h. Extraction with CHCl₃ gave 89 mg of a syrup. Chromatography of 69 mg of the latter using 40:1 PhMe–EtOAc gave 29 mg (31%) of the β -lactone 19, identical with that prepared as just described.

B. Preparative. — To a stirred solution of 1.36 g (2.1 mmol) of the azido acid 17a in 70 mL of AcOH was added 8.4 g of Zn dust. Stirring was continued for 2 h and the Zn was removed by filtration through a Celite mat. The AcOH was evaporated under diminished pressure leaving a mixture of the amino acid 17b and the lactone 19. To a stirred solution of the latter mixture in 40 mL of AcOH and 20 mL of water was added 0.810 g of NaNO₂. Stirring was continued for 1 h. The resulting suspension was shaken with a mixture of 500 mL of water and 200 mL of CHCl₃. The CHCl₃ solution was separated and washed with 500 mL of water. The aqueous solutions were washed in series with four 100-mL portions of CHCl₃. The CHCl₃ solutions were combined and dried (MgSO₄). The solvent was evaporated under diminished pressure, leaving a syrup which on flash chromatography gave 0.668 g (52.7%) of the β -lactone 19 identical with that prepared as just described.

2,6-Anhydro-4,5,7,8-tetra-O-benzyl-3-deoxy-2-C-(hydroxymethyl)- β -D-manno-2-D-glycero-D-talo-octonic acid (17c). — A suspension of 458 mg (0.77 mmol) of the lactone 19 in 30 mL of tert-BuOH was stirred for 15 min, and 15 mL of 3M NaOH was added. Stirring was continued for 26 h. The resulting suspension was shaken with a mixture of 250 mL of cold 0.5M HCl and 100 mL of CHCl₃. The CHCl₃ solution was separated and washed with 250 mL of 10% NaCl. The aqueous solutions were washed with CHCl₃ and the CHCl₃ solutions were combined and dried (MgSO₄). Evaporation of solvent left 464 mg of white glass which on flash chromatography with 10:10:0.1 PhMe-EtOAc-AcOH gave 418 mg (89%) of 17c as a rigid white glass; $[a]_{p}^{23} - 2.5^{\circ}$ (c 1, CHCl₃); ¹H-n.m.r. (CDCl₃): δ 2.11 (t, 1 H, $J_{3a,3e} = J_{3a,4} = 12.6$ Hz, H-3a), and 2.59 (dd, 1 H, $J_{3e,4}$ 4.5 Hz, H-3e).

Anal. Calc. for C₃₇H₄₀O₈: C, 72.53; H, 6.58. Found: C, 72.34; H, 6.71.

Ethyl 2,6-anhydro-4,5,7,8-tetra-O-benzyl-2-C-(diethylphosphonomethyl)-3-deoxy-D-glycero-D-talo-octonate (23). — A stirred solution of 518 mg (0.87 mmol) of 19 in 45 mL of $(EtO)_3P$ was heated for 4 h at 100°. The major portion of the $(EtO)_3P$ was evaporated under high vacuum and residual $(EtO)_3P$ was removed by co-distillation with PhMe under diminished pressure. Chromatography of the residue using 2:1 PhMe-EtOAc gave 329 mg (50%) of **23** as a syrup; $[a]_{p}^{20} + 0.16^{\circ}$ (c 1, MeOH); ¹H-n.m.r. (CDCl₃): δ 1.1-1.3 (m, 6 H, OCH₂Me), 2.2-2.4 [m, 3 H, H-3a and CH₂P(OEt)₂], and 2.58 (dd, 1 H, J_{1e4} 5.1, J_{1e3a} 12 Hz, H-3e).

Anal. Calc. for C₄₃H₅₃O₁₀P: C, 67.88; H, 6.78. Found: C, 67.26; H, 6.78.

2-C-(Aminomethyl)-2,6-anhydro-3-deoxy-D-glycero-D-talo-octonic acid hydrochloride (4a). — The azido acid 17a (393 mg, 0.53 mmol) was catalytically hydrogenated in 100 mL of 0.2M HCl in MeOH with 390 mg of Pd black under 3 atm. of hydrogen. After removal of the major portion of the catalyst by filtration, solvent was evaporated under diminished pressure. Residual HCl was removed by co-distillation with EtOH under diminished pressure. The residue was dissolved in water and filtered through a Millipore type EH membrane. Lyophilization gave 238 mg of a glass. Gel filtration in MeOH through Sephadex LH20 gave 77 mg (100%) of the hydrochloride of 4a as a glass; $[a]_{p}^{20} + 51^{\circ}$ (c 1, MeOH); ¹H-n.m.r. (D₂O): δ 1.72 (t, 1 H, $J_{3a,4} = J_{3a,3e} = 12.6$ Hz, H-3a), 2.30 (dd, 1 H, $J_{3e,4}$ 4.5 Hz, H-3e), 3.18 (d, 1 H, J_{AB} 12.6 Hz, $CH_AH_BNH_2$), and 3.25 (d, 1 H, $CH_AH_BNH_2$); exact mass calc. for $C_9H_{18}NO_7$ (MH)⁺ 252.1083, found 252.1080.

2,6-Anhydro-3-deoxy-2-C-(hydroxymethyl)-D-glycero-D-talo-octonic acid (4b). — The hydroxy acid 17c (391 mg, 0.64 mmol) was catalytically hydrogenated in 150 mL of 0.2M HCl with 390 mg of Pd black under 3 atm of hydrogen. Catalyst was removed by filtration and solvent was evaporated under diminished pressure. Residual hydrochloric acid was removed by co-distillation with MeOH under diminished pressure. The residue was dissolved in water and filtered through a Millipore type EH membrane. Lyophilization gave (100%) of 4b as a glass; $[a]_{p}^{23} + 58.6^{\circ} (c 1, MeOH)$; ¹H-n.m.r. (D₂O): δ 1.71 (t, 1 H, $J_{3a,3e} = J_{3a,4} = 15$ Hz, H-3a), and 2.20 (dd, 1 H, $J_{3e,4}$ 4.5 Hz, H-3e); exact mass: calc. for C₉H₁₇O₈ (MH)⁺ 253.0923, found 253.0925.

2,6-Anhydro-3-deoxy-2-C-(phosphonomethyl)-D-glycero-D-talo-octonic acid (4c). — A stirred solution of 329 mg (0.43 mmol) 23 in 22 mL of a solution prepared from 2 g of NaOH, 5 mL of water and 20 mL of EtOH was heated overnight at 70° under nitrogen and then allowed to cool. The mixture was shaken with a mixture of 100 mL of CHCl₃ and 200 mL of MHCl. The CHCl₃ solution was separated and washed with 200 mL of water. The aqueous solutions were washed with CHCl₃. The CHCl₃ solutions were combined and dried (MgSO₄). Evaporation of the CHCl₃ left a residue which was hydrogenated with 270 mg of Pd black in 50 mL of 0.2m HCl in MeOH under 3 atm of hydrogen. Conventional isolation of the product gave 164 mg of glass. The latter was dissolved in 10 mL of 2m NaOH and the resulting solution was stirred and heated for 20 h at 70° under N₂. The solution was cooled, diluted with 25 mL of water and acidified by addition of 10 g of Dowex HCR-S (H⁺) resin. After removal of the resin by filtration, lyophilization gave 134 mg (98%) of 4c as a syrup; $[a]_{D}^{20} + 46^{\circ}$ (c 1, MeOH); ¹H-n.m.r. (CD₃OD): δ 2.13 (t, 1 H, $J_{3a,3e} = J_{3a,4} = 12.6$ Hz, H-3a), and 2.2–2.4 (m, 3 H, H-3e and CH₂PO₃H₂); exact mass: calc. for C₉H₁₈O₁₀P (MH)⁺ 317.0637, found 317.0637.

CMP-Kdo synthetase assay. — CMP-Kdo synthetase was isolated from *Escherichia coli* by the method of Goldman and Kohlbrenner⁹. The reaction was monitored with a coupled assay performed at 30° in semi-micro cuvettes containing 50mm Hepes (pH 7.6), mM Kdo, 0.5mM MgCl₂, mM DTT, 1.8 mg of glycogen, 7.8 units of inorganic pyrophosphatase, 10 units of phosphorylase a, 13 units of phosphoglucomutase, 15 units of D-glucose 6-phosphate dehydrogenase, 0.36 mg of NADP, and CMP-Kdo synthetase in a final volume of 1 mL. After a 6-min. pre-incubation period, the reaction was initiated by the addition of 10 μ L of diluted CMP-Kdo synthetase. The change in absorption at 340 nm was measured with a Gilford Response spectrophotometer which was programmed to calculate reaction rates. The reaction was linear between 2 and 5 min after CMP-Kdo synthetase addition. The apparent K_m of Kdo was 0.32mM.

REFERENCES

- 1 W. E. Kohlbrenner and S. W. Fesik, J. Biol. Chem., 260 (1985) 14695-14699.
- 2(a) R. Cherniak, R. G. Jones, and D. S. Gupta, Carbohydr. Res., 75 (1979) 39-49; (b) F. M. Unger, Adv. Carbohydr. Chem. Biochem., 38 (1981) 323-388.
- 3(a) P. Lartey, D. Riley, R. Hallas, W. Rosenbrook, Jr., D. Norbeck, D. Grampovnik, W. Kohlbrenner, N. Wideburg, and A. G. Pernet, *Abstr. Pap. Am. Chem. Soc. Mtg.*, 193 (1987) MEDI-68; (b) P. Lartey, D. Norbeck, J. Tadanier, C. Maring, and C.-M. Lee, *ibid.*, 193 (1987) MEDI-69; (c) A. Claesson, K. Luthman, K. Gustafsson, and G. Bondesson, *Biochem. Biophys. Res. Commun.*, 143 (1987) 1063–1068.
- 4(a) F. G. De las Heras and P. Fernandez-Resa, J. Chem. Soc. Perkin Trans. 1 (1982) 903-907; (b) M. T. G. Lopez, F. G. De las Heras, and A. S. Felix, J. Carbohydr. Chem., 6 (1987) 273-279.
- 5(a) D. W. Norbeck, J. B. Kramer, and P. A. Lartey, J. Org. Chem., 52 (1987) 2174–2179; (b) K. Luthman, M. Orbe, M. T. Wagland, and A. Claesson, *ibid.*, 52 (1987) 3777–3784.
- 6 J. D. Albright and L. Goldman, J. Am. Chem. Soc., 89 (1967) 2416-2423.
- 7 F. M. Unger, D. Stix, and G. Schultz, Carbohydr. Res., 80 (1980) 191-195.
- 8 R. L. McConnell and H. W. Coover, J. Am. Chem. Soc., 78 (1956) 4453-4455.
- 9 R. Goldman and W. Kohlbrenner, J. Bacteriol., 163 (1985) 256-261.