

**Synthesis of (1*S*,4*R*)-(-)-4-Hydroxy-2-cyclopentenyl Acetate by a Highly Enantioselective Enzyme-Catalyzed Transesterification in Organic Solvents<sup>1</sup>**

Fritz Theil,<sup>a</sup> Sibylle Ballschuh,<sup>a</sup> Hans Schick,<sup>\*a</sup> Monika Haupt,<sup>b</sup> Barbara Häfner,<sup>b</sup> Sigfrid Schwarz<sup>c</sup>

<sup>a</sup> Central Institute of Organic Chemistry of the Academy of Sciences of the GDR, Rudower Chaussee 5, DDR-1199 Berlin, German Democratic Republic

<sup>b</sup> Research Centre of Biotechnology, Alt-Stralau 62, DDR-1017 Berlin, German Democratic Republic

<sup>c</sup> VEB Jenapharm, Division of Research, Otto-Schott-Strasse 13, DDR-6900 Jena, German Democratic Republic

(1*S*,4*R*)-(-)-4-Hydroxy-2-cyclopentenyl acetate (**2**), a versatile intermediate in prostaglandin syntheses, was readily prepared by an efficient enzyme-catalyzed enantioselective monoacetylation of *cis*-2-cyclopenten-1,4-diol (**1**) with 2,2,2-trichloroethyl acetate in the organic solvent system triethylamine/tetrahydrofuran. The chemical yield reached nearly 50%. The enantiomeric excess of the crude product was 95%. It could be raised to more than 99% by a single recrystallization. Commercially available pancreatin, a crude enzyme preparation from porcine pancreas, was used as biocatalyst.

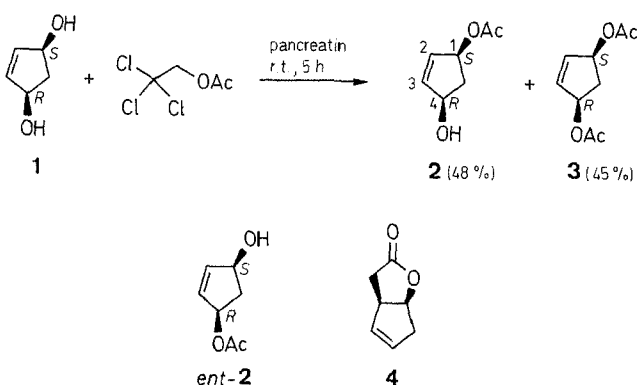
The enantiomerically pure (1*S*,4*R*)-(-)-4-hydroxy-2-cyclopentenyl acetate (**2**) belongs to a group of 1,4-disubstituted 2-cyclopentenes, which are attractive starting materials for the synthesis of prostaglandins<sup>2-5</sup> and other cyclopentanoid natural products.<sup>6</sup> Several approaches to prepare **2**<sup>3,7-10</sup> as well as *ent*-**2**<sup>11-13</sup> from the *meso*-compound **3** by an enantioselective microbial or enzymatic hydrolysis of the *R*- or *S*-acetoxy group have already been described. In principle, both compounds can be transformed via Claisen orthoester rearrangement into the enantiomerically pure lactone **4**, an important intermediate of many prostaglandin syntheses.<sup>14</sup> However, **2** is the preferred enantiomer, since it requires the lower number of reaction steps.<sup>13</sup>

Presently much attention has been focussed on the enzyme-catalyzed processes in organic solvents.<sup>15-19</sup> Following this direction, we have found a very efficient alternative to the above

mentioned procedures to convert the *meso*-diol **1**<sup>20,21</sup> into the monoacetate **2** by transesterification with 2,2,2-trichloroethyl acetate catalyzed by pancreatin in various organic solvents.

When the diol **1** is reacted in a mixture of dry tetrahydrofuran and triethylamine at ambient temperature for 5 hours with an excess of 2,2,2-trichloroethyl acetate in the presence of pancreatin, the starting material is completely converted into a mixture of the monoacetate **2** and the diacetate **3**.

The desired monoacetate **2** is isolated in 48% yield with an enantiomeric excess of 95% by flash chromatography on silica gel. The enantiomeric excess is increased to more than 99% by a single recrystallization from ether/*n*-hexane. The separated diacetate **3** can be hydrolyzed to the diol **1** and recycled.



In the absence of triethylamine the transesterification proceeds very slowly. Solvents like 1,4-dioxane or toluene are also suitable, but with respect to solubility problems and reaction rate, tetrahydrofuran proved to be the solvent of choice.

The described enzyme-catalyzed process was carried out in a multigram scale and represents a significant improvement for the synthesis of the enantiomerically pure prostaglandin intermediate **4**.

THF was dried with sodium wire. Et<sub>3</sub>N was distilled from and stored over KOH. Pancreatin, qualified as 6 × NF, is a mixture of crude porcine pancreatic enzymes with protease, amylase, and lipase activities. The product, purchased from Fa. Belger, Kleinmachnow, GDR, had a water content of 5.4% (Karl-Fischer-titration) and a lipase activity of 820 U/g (triolein as substrate). The <sup>1</sup>H-NMR spectrum was recorded at 100 MHz on a Tesla BS-567 spectrometer. Optical rotations were measured with the photoelectric polarimeter Polamat A (Carl Zeiss Jena) at 546 and 578 nm and extrapolated to 589 nm. Differential scanning microcalorimetry (DSC)<sup>22,23</sup> was performed on a DSC-1B (Perkin-Elmer). The enantiomeric excess (e.e.) was calculated on the basis of the optical rotation and of DSC measurements. Melting points were determined on a Boëtius micro melting point apparatus and are corrected. TLC was carried out on silica gel 60 F<sub>254</sub> (E. Merck) using EtOAc/*n*-hexane (2:1). For visualization, the plates were treated with 5% H<sub>2</sub>SO<sub>4</sub> in EtOH and heated to 150°C.

#### (1S,4R)-(-)-4-Hydroxy-2-cyclopentenyl Acetate (**2**):

Et<sub>3</sub>N (5 mL, 36 mmol), 2,2,2-trichloroethyl acetate (50 mL, 365 mmol), and pancreatin (25 g) are added to a solution of *cis*-2-cyclopentene-1,4-diol (**1**)<sup>20,21</sup> (5 g, 50 mmol) in THF (125 mL). After stirring at ambient temperature for 5 h, the suspension is filtered through celite. The filter cake is washed with EtOAc (3 × 20 mL). Then solvents and excess of 2,2,2-trichloroethyl acetate are distilled off under reduced pressure. The residue is purified by flash chromatography on silica gel (140 g, 25 × 4 cm, 0.063–0.04 mm) using *n*-hexane/EtOAc (2:1 and 1:1) as eluent. First diacetate **3** (4.1 g, 45%; colourless liquid; R<sub>f</sub> 0.75) is eluted, then monoacetate **2**. The TLC homogeneous product solidifies immediately after evaporation of the solvent affording colourless crystalline material; yield: 3.4 g (48%); mp 39–47°C; [α]<sub>D</sub><sup>20</sup> –62.7° (c = 1.0, CHCl<sub>3</sub>); e.e.: 95%. A single recrystallization from ether/*n*-hexane gives rise to enantiomerically pure **2**; R<sub>f</sub> 0.43; mp 46–48°C; [α]<sub>D</sub><sup>20</sup> –66.3°

(c = 1, CHCl<sub>3</sub>); e.e.: 99% (calculated from the optical rotation) and 99.7% (calculated from DSC measurements), respectively [Lit.<sup>9</sup> mp 49–50°C; [α]<sub>D</sub><sup>20</sup> –66° (c = 0.63, CHCl<sub>3</sub>), e.e.: 98%; Lit.<sup>12</sup> [α]<sub>D</sub><sup>20</sup> +66.3° for *ent*-**2** (c = 1.63, CHCl<sub>3</sub>), e.e.: 99%; Lit.<sup>11</sup> [α]<sub>D</sub><sup>22</sup> +75.0° for *ent*-**2** (c = 1.16, CHCl<sub>3</sub>), e.e. ~100%].

<sup>1</sup>H-NMR (CDCl<sub>3</sub>/TMS): δ = 1.58 (dt, 1H, J = 15, 4 Hz, H<sub>a</sub> of CH<sub>2</sub>); 1.86 (s, 1H, OH, exchangeable with D<sub>2</sub>O); 1.99 (s, 3H, CH<sub>3</sub>); 2.74 (dt, 1H, J = 15, 8 Hz, H<sub>b</sub> of CH<sub>2</sub>); 4.65 (m, 1H, CHOH); 5.42 (m, 1H, HCOAc); 5.98 (dd, 2H, J = 15, 6 Hz, CH=CH).

Received: 4 January 1988; revised: 2 March 1988

- (1) Enzymes in Organic Synthesis; Part 1.
- (2) Tanaka, T., Kurozumi, S., Toru, T., Miura, S., Kobayashi, M., Ishimoto, S. *Tetrahedron* **1976**, *32*, 1713.
- (3) Takano, S., Tanigawa, K., Ogasawara, K. *J. Chem. Soc. Chem. Commun.* **1976**, 189.
- (4) Nara, M., Terashima, S., Yamada, S. *Tetrahedron* **1980**, *36*, 3161.
- (5) Noyori, R., Suzuki, M. *Angew. Chem.* **1984**, *96*, 854; *Angew. Chem. Int. Ed. Engl.* **1984**, *23*, 847.
- (6) Harre, M., Raddatz, P., Walenta, R., Winterfeldt, E. *Angew. Chem.* **1982**, *94*, 496; *Angew. Chem. Int. Ed. Engl.* **1982**, *21*, 480.
- (7) Miura, S., Kurozumi, S., Toru, T., Tanaka, T., Kobayashi, M., Matsubara, S., Ishimoto, S. *Tetrahedron* **1976**, *32*, 1893.
- (8) Laumen, K., Schneider, M. *Tetrahedron Lett.* **1984**, *25*, 5875.
- (9) Laumen, K., Reimerdes, E.H., Schneider, M., Görisch, H. *Tetrahedron Lett.* **1985**, *26*, 407.
- (10) Wang, Y.-I., Chen, C.-S., Girdaukas, G., Sih, C.J. *J. Am. Chem. Soc.* **1984**, *106*, 3695.
- (11) Sugai, T., Mori, K. *Synthesis* **1988**, 19.
- (12) Deardorff, D.R., Matthews, A.J., McMeekin, D.S., Craney, C.L. *Tetrahedron Lett.* **1985**, *26*, 5615; *Tetrahedron Lett.* **1986**, *27*, 1255.
- (13) Laumen, K., Schneider, M. *P. J. Chem. Soc. Chem. Commun.* **1986**, 1298.
- (14) Tömösközi, I., Gruber, L., Kovács, G., Székely, I., Simonidesz, V. *Tetrahedron Lett.* **1976**, 4639.
- (15) Klibanov, A.M. *Chem. Technol.* **1986**, *16*, 354, and references cited therein.
- (16) Cambou, B., Klibanov, A.M. *J. Am. Chem. Soc.* **1984**, *106*, 2687.
- (17) Therisod, M., Klibanov, A.M. *J. Am. Chem. Soc.* **1986**, *108*, 5638.
- (18) Ramos Tombo, G.M., Schär, H.-P., Fernandez i Busquets, X., Ghisalba, O. *Tetrahedron Lett.* **1986**, *27*, 5707.
- (19) Hemmerle, H., Gais, H.-J. *Tetrahedron Lett.* **1987**, *28*, 3471.
- (20) Schenck, G.O., Dunlap, D.E. *Angew. Chem.* **1956**, *68*, 248.
- (21) Kaneko, C., Sugimoto, A., Tanaka, S. *Synthesis* **1974**, 876.
- (22) Wilen, S.H., Collet, A., Jacques, J. *Tetrahedron* **1977**, *33*, 2725.
- (23) Schurig, V. *Kontakte (Darmstadt)*, **1985**, 54.