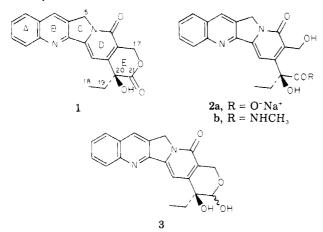
Prodrug Analogues of the Antitumor Alkaloid Camptothecin

John A. Adamovics¹ and C. Richard Hutchinson^{*2}

School of Pharmacy, University of Wisconsin, Madison, Wisconsin 53706. Received January 19, 1978

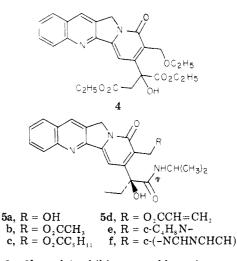
Two classes of (+)-camptothecin (1) derivatives are prepared by acylation of camptothecin-21-isopropylamide (5a). The 17-acetyl (5b) 17-hexanoyl (5c), and 17-acryloyl (5d) derivatives represent lipophilic substances with the potential for reconversion to 1 in vivo. A 17-pyrrolidinyl derivative 5e is prepared from 5b by a nucleophilic substitution reaction and represents a hydrophilic derivative of 1, which may not be able to be converted to 1 in vivo. It is reported also that 5b undergoes C-17 substitution by other nucleophiles (isopropylamine, imidazole, cyanide, and ethanethiol). The ester amide derivatives 5b-d are shown to have an antitumor activity in vivo (L1210, P388) less than 1 or its sodium salt 2a, and the amine amide derivative 5e is shown to be inactive in these assay systems. Both 5b and 5c exhibit apparent activity in vivo in the Lewis lung LL32 antitumor assay but no activity (9KB, L1210, P388) relative to 1. In one preliminary in vitro assay, 5c appeared to be nearly as active as 2a in causing the fragmentation of DNA and inhibiting thymidine's incorporation into DNA in intact HeLa cells. Consequently, the intact α -hydroxy- δ -lactone ring of 1 may not be absolutely necessary for antitumor activity.

Camptothecin (1), an alkaloid isolated from the rare



Chinese tree *Camptotheca acuminata* Decne in 1967 by Wall and co-workers,³ is a potent inhibitor of the growth of leukemia cells in vitro and shows good antitumor activity against murine L1210 and P388 leukemia in vivo.^{3,4} However, despite initial encouraging reports that 1 was active clincally against solid tumors of the gi tract,⁵ subsequent clinical evaluation of the drug led to discouraging results.⁶ Today it is used in the clinical treatment of cancer only in the People's Republic of China, with apparent success.⁷

Many analogues of 1 have been prepared in the course of studies to define structure-activity relationships (SAR)⁸ and to develop synthetic routes to this scarce compound.⁹ It was shown in one of the early SAR studies that the intact α -hydroxy- δ -lactone (ring E) must be present for full expression of the in vivo antitumor activity.^{8d,10} Thus, 20-deoxycamptothecin, 20-chlorocamptothecin, and camptothecin hemilactol (3) are essentially inactive in vivo.^{8d} The C-21 methylamide **2b** has ca. three-fifths the activity of 1 in the L1210 life-prolongation assay,^{8d} and the sodium salt 2a has ca. one-third the activity of 1 in the P388 in vivo assay.¹¹ All analogues of 1 consisting of only rings C, D, and E, or D and E, with or without additional aliphatic or aromatic functionality,^{8b} are inactive in vivo, despite some of them having moderate in vitro biological activity.^{8a} The only analogue of 1 that does not have the intact ring E, but which has good in vivo antitumor activity, is 4. This compound and four other analogues of 1 (in which the C-20 ethyl group was replaced by $-CH_2CH=CH_2$, $-CH_2C\equiv CH$, $-CH_2C_6H_5$, or $-CH_2COC_6H_5$) prepared by Sugasawa et al.¹² are the only analogues reported to date possessing in vivo antitumor activity equal to or better than 1.

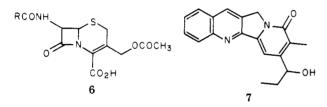


Since 2a, 2b, and 4 exhibit acceptable antitumor activity in vivo, we decided to prepare a series of camptothecin derivatives modeled after 2b, which would be expected to have enhanced lipophilicity or hydrophilicity in vivo relative to 1. Our rationale was based on the well-established relationship of drug lipophilicity and/or basicity to antitumor activity,¹³ e.g., the increased activity of MeCCNU over BCNU¹⁴ and the excellent in vivo antitumor activity of N-(trifluoroacetyl)adriamycin 14-valerate (AD 32).¹⁵ A second consideration was the potential for the reconversion of the derivatives to 1 in vivo. This would appear to be desirable for full expression of antitumor activity^{8c} and may be why 2a, 2b, and 4 exhibit good antitumor activity. (It has never been proven if the latter three drugs' antitumor activity is intrinsic or due to their conversion to 1 in vivo.) We describe herein the preparation of one series of such camptothecin derivatives, which exhibit moderate in vivo antitumor activity, as well as an unexpected, novel aspect of the chemistry of 1.

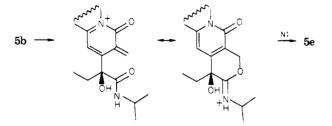
Synthesis of Camptothecin Derivatives. (+)-Camptothecin formed a comparatively stable amide (5a) on reaction with excess isopropylamine at reflux temperature overnight. This amide was unstable in solution or to silica gel chromatography (\rightarrow 1) but could be acylated at C-17 to give the ester amides 5b-d in excellent yield using standard methodology. The ester amides 5b and 5c are soluble in organic solvents of medium (CHCl₃, THF) to low (C₆H₆) polarity and thus represent lipophilic derivatives of 1. They also are rapidly transformed back to 1 in dilute aqueous-alcoholic acid or base at 25 °C, implying that they could be recoverted to 1 in vivo fairly easily. This is a very tenuous assumption, since in vivo the requisite hydrolysis may well be achieved only enzymatically, which might not occur readily if appropriately unspecific amidases and esterases are not available.

It was anticipated that the acrylate amide 5d would undergo Michael addition by secondary amines to give basic ester amide derivatives of 1. These should have been water soluble (as their salts) or at physiological pH should have shown enhanced binding to DNA, the presumed primary site of the biological action of 1.8ª Surprisingly, reaction of 5d with neat pyrrolidine at 25 °C gave 5e (73%) by C-17 substitution rather than the expected 1.4-addition product. That 5e had been formed was clearly evident from its positive response to Draggendorf's reagent (which 1 does not exhibit), its ability to be extracted into dilute aqueous acid, and its NMR and mass spectral characteristics. This amine amide derivative was not transformed to 1 in acid but was readily transformed in dilute aqueous bicarbonate at 25 °C. Consequently, it seems unlikely that 5e would be reconverted to 1 in vivo as readily as 5b-d.

By subsequent experimentation, it was revealed that the acetate amide 5b also underwent substitution by pyrrolidine to give 5e, as well as by several other nucleophiles (isopropylamine, imidazole, cyanide, and sulfide). These latter substitution products of 5b were formed in a moderate to good yield and were only partially characterized due to their instability to chromatography and recrystallization (Experimental Section). Nevertheless, it was clear that 5b has an unusually electrophilic carbon at C-17, whose ease of nucleophilic substitution is precedented by the chemistry of cephalosporin antibiotics (6) and



the synthesis of mappicine (7) from 1. In the case of 6, many derivatives of the antibiotic have been made by substitution at the carbon bearing the acetoxy group despite the fact that the enamine center ($N^1C^2C^3$), which could assist in displacement of the acetate group, is deactivated by electron-withdrawing substituents.¹⁶ Mappicine (7) is prepared via sodium borohydride reduction of an intermediate 17-formyl-20-keto degradation product of 1, which underscores the electrophilicity of the C-17 carbon atom.¹⁷ It may be that substitution at C-17 of **5b** is facilitated by the following mechanistic rationale:



Biological Activity. The antitumor activity of 5b-f was compared to that of 1 and 2a at equimolar dosage levels in vivo (mice) in the P388 and L1210 assays; 5b and 5c were also assayed in the melanocytic melanoma B-16 and Lewis lung carcinoma test systems but without reference to 1 or 2a as a control. These testing results are shown in Table I as percentage increase in life span vs. dosage, along with the comparative in vitro cytotoxic activity of 1 and 5b-f. It is immediately clear that none of these camptothecin derivatives have an antitumor

activity equal to or greater than 1 or 2a. The potency of the two most active compounds, 5b and 5d, is not greater than 85% of that of 1 at the optimal 9.5 μ mol/kg dosage level. The pyrrolidinamide 5e is essentially inactive in vivo but seems to have significant in vitro activity in the L12110 cell cytotoxicity assay, whereas the imidazolamide 5f is inactive in both assay systems. Both 5b and 5c appear to have moderate activity in the Lewis lung carcinoma assay but not against melanocytic melanoma B-16 in which 1 exhibits high activity (2a, somewhat lower activity).¹¹ However, we caution that these particular data may not be significant and *must* be confirmed by retesting with appropriate controls (of 1 or 2a), since both 1 and 2a are known to give erratic results in the Lewis lung assay.¹⁸

We cannot assess at this time if any of our camptothecin derivatives have intrinsic antitumor activity in vivo. From our observations during the preparation of 5b-f, it was clear that these substances are not highly stable to chromatography on silica gel or recrystallization. Trace amounts of 1 would reappear during such manipulations, even though crystalline samples could be obtained with sharp melting points. Consequently, it is very likely that a small amount of 1 was formed from these derivatives in vivo. Reversion of 5e and 5f to 1 in vivo seems least likely based on their chemical properties, which could account for their inactivity in vivo.

On the other hand, 5c had an activity identical to 1 and **5b** ca. one-half that of 1 in their effects on Hela cell DNA sedmentation in vitro; 5c was as active as 2a in inhibiting the incorporation of radioactive thymidine into acid-insoluble material in HeLa cells.¹⁹ Although these results are preliminary, subject to retesting, they indicate that either 5b and 5c may have intrinsic antitumor activity. To the extent that the in vitro activity of camptothecin analogues is meaningful vis-a-vis their antitumor effects in vivo,^{8a,c} this latter conclusion is consistent with the conclusion made by Sugasawa et al.¹² regarding the antitumor activity of 4, whose conversion to 1 in vivo would not occur as readily as 5b-d. Recently, analogous observations have been reported about AD-32, i.e., that it may have an intrinsic antitumor activity in vivo without transformation to adriamycin.^{15b}

We acknowledge that it remains to be proven clearly if the intact E ring of 1 is absolutely essential for antitumor activity.

Experimental Section

All solvents and liquid reagents were redistilled prior to use. Evaporation in vacuo was done at ≤ 40 °C on a rotary evaporator. Thin-layer (TLC) and preparative-layer (PLC) chromatography was done using EM silica gel purchased from Brinkmann Inc., New York. IR spectra were determined with a Perkin-Elmer 257 infrared spectrophotometer. ¹H and ¹³C NMR spectra were recorded on a Bruker HX-90E nuclear magnetic resonance spectrometer at 90 or 22.63 MHz, respectively. Mass spectra were run on an AEI-MS-9 mass spectrometer interfaced to a Nova 2 data system or on a Finnegan quadrupole 1015 GC-mass spectrometer interfaced to a Finnegan M6000 data system. Optical rotations were taken on a Perkin-Elmer 241 polarimeter. Elemental analyses were done by Galbraith Laboratories, Knoxville, Tenn. Melting points were determined on a Koefler hot stage and are uncorrected.

Isopropylamide of Camptothecin (5a). (+)-Camptothecin (116 mg, 0.33 mmol, NSC 94600, supplied by the NCI, lot no. 69-12-16-01) was mixed with excess dry isopropylamine (20-25 mL), and the resulting slurry was refluxed for 18 h with magnetic stirring under N₂ to suppress the C-5 oxidation of $1.^{20}$ The final reaction mixture was a clear, light-amber solution, TLC analysis of which (CHCl₃-MeOH, 95:5) showed that complete conversion to 5a had been achieved (R_f 0.55 relative to 1). Removal of excess isopropylamine in vacuo gave 5a as a light-amber solid glass

$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$					ED ₅₀ , μmol/mL (relative activity)			
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	assay system: in vitro ^d	1	2a	5b	5c	5d	5e	5f
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	9KB	$< 2.8 imes 10^{-6}$ (100)	q	$7.8 imes 10^{-4}$ (2.8)	8.1×10^{-5} (29)	6.9×10^{-5} (25)	2.2×10^{-5} (7.8)	Ą
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	P388	8.6×10^{-6} (100)	q	7.4×10^{-3} (0.08)	1.2×10^{-3} (0.01)	3.9×10^{-4} (0.45)	1.1×10^{-4} (0.13)	> 200
c 1 2a 5b 5c 5d 5e 5d 5e -11.2 ^d $81 (100; 29)$ $67 (11.1; 29)$ $97 (12.8; 28.5)$ $155 (14.3; 28.3)$ $155 (6.5; 14.1)$ $125 (62.5; 136)^{d}$ $2e$ -11.2 ^d $81 (100; 29)$ $67 (11.1; 29)$ $97 (12.8; 28.5)$ $155 (14.3; 28.3)$ $155 (6.5; 14.1)$ $125 (62.5; 136)^{d}$ $111 (5.1; 11)$ $111 (5.1; 12)$ $111 (5.1; 12)$ $111 (5.1; 12)$ $111 (5.1; 12)$	L1210	2.8×10^{-6} (100)	b	p q	$5.5 imes 10^{-4}$ (2.0)	7.6×10^{-4} (2.7)	6.5×10^{-5} (23)	> 200
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$					Γ/C, % ^e (mg/kg; μmol/	kg)		
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	in vivo ^c	1	2a	5b	Š	5d	Se	5f
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$, 10.3-11.2 ^d	$\begin{array}{c} 81 \ (10.0; \ 29) \\ 204 \ (3.3; \ 9.5) \\ 147 \ (1.1; \ 3.2) \\ 137 \ (0.4; \ 1.1) \end{array}$	$\begin{array}{c} 67 \ (11.1; \ 29) \\ 85 \ (3.7; \ 9.5) \\ 168 \ (1.2; \ 3.1) \\ 139 \ (0.4; \ 1.0) \end{array}$	97 (12.8; 28.5) 173 (4.3; 9.6) 126 (1.4; 3.1) 126 (0.5: 1.1)	$155 (14.3; 28.3) \\139 (4.8; 9.5) \\107 (1.6; 3.2) \\107 (0.5; 1.0)$	$155 (6.5; 14.1) \\165 (2.5; 5.4)' \\i$	$\begin{array}{c} 125 \ (62.5; 136)^{g} \\ 126 \ (18.9; 41) \\ 111 \ (5.1; 11) \end{array}$	$\begin{array}{c} 92 \ (45.0; 100) \\ 89 \ (23.0; 50) \\ 105 \ (6.4; 14) \end{array}$
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	0, 8.8 ^d	$ \begin{array}{c} 85 (h) \\ 168 \\ 131 \\ 114 \\ 114 \\ \end{array} $	74(h) 114 147 116	105(h) 147 116 110		$\frac{125}{i}(6.5; 14.1)$	113 (h) 101 109	q
$ b \qquad b \qquad 156 (48.6; 107) \qquad 134 (44.4; 58) \qquad b \qquad b \\ 166 (15.7; 35) \qquad 147 (14.6; 29) \\ 147 (4.0; 9) \qquad 163 (4.0; 8) \\ \end{array} $	20.2^{d}	9	p	$\frac{114}{105} (43.5; 97) \\ 105 (14.8; 33) \\ 100 (4.9; 11) \\ 100 (4.9; 11) \\ 100 \\ 10$	$egin{array}{c} 109 & (46; 91) \ 109 & (14.6; 29) \ 110 & (4.0; 8) \end{array}$	q	q	Ą
	$s lung, 20.4^d$	р	b	$\begin{array}{c} 156 \ (48.0; 107) \\ 166 \ (15.7; 35) \\ 147 \ (4.0; 9) \end{array}$	$\begin{array}{c} 134 \ (44.4; 88) \\ 147 \ (14.6; 29) \\ 163 \ (4.0; 8) \end{array}$	q	q	q
	oride salt. ^{<i>h</i>} Sam	e dosages used as in	the P388 assay. ^t	hydrochloride salt. h Same dosages used as in the P388 assay. t Insufficient drug for assays at !ower level.	ssays at lower level.		hydrochloride salt. h Same dosages used as in the P388 assay. i Insufficient drug for assays at lower level.	

Table I. Cytotoxicity and Antitumor Activity of Camptothecin and its Analogues

(100%): IR (KBr) ν 3300, 1648, 1580–1510 cm⁻¹; ¹H NMR (CDCl₃) δ 1.10 [d + t, H-18 + (CH₃)₂⁻], 2.28 (m, H-19), 4.08 [m, (CH₃)₂CHN], 4.85 (s, H-17), 5.03 (s, H-5), 6.73 (d, J = 8 Hz, NH), 7.51–8.11 (6 arom H).

17-Acetylcamptothecin-21-isopropylamide (5b). A sample of crude 5a (from 0.33 mmol of 1) was dissolved in CHCl₃-CH₂Cl₂ (1:1, 2-3 mL) and mixed with 1 mL each of Ac₂O and dry pyridine. This reaction mixture was stirred magnetically for ca. 3.5 h at room temperature and then the solvents and excess reagents were removed in vacuo (oil pump) by a MeOH-C₆H₅CH₃ azeotrope. (Longer acetylation times resulted in the formation of a C-20 acetate byproduct.) The resulting solid residue was purified by PLC in CHCl₃-MeOH (100:6) to give 5b (150 mg, 100%): yellow needles, mp 204-206 °C (isopropyl alcohol); IR (CHCl₃) ν 3410, 3300, 1735, 1660, 1600, 1585, 1520, 1460, 1240 cm⁻¹; ¹H NMR (CDCl₃) δ 1.15 (t, J = 6.5 Hz, H-18), 1.21 [d, J = 6.1 Hz, (CH₃)₂CH], 2.06 (s, OCOCH₃), 2.42 (m, H-19), 3.47 (br d, $J \approx 3$ Hz, OH), 4.07 [m, (CH₃)₂CHN], 5.13 (s, H-5), 5.48 (q, $J_{AB} = 11.8$ Hz, H-17), 6.76 (d, J = 8.3 Hz, NH), 7.45-8.11 (6 arom H); MS, m/e (rel intensity), 390 (M⁺ - 59) (5), 373 (15), 303 (35); [α]²⁴_D +87° (c 0.06, CHCl₃).

17-Hexanoylcamptothecin-21-isopropylamide (5c). A sample of crude 5a (from 0.33 mmol of 1) was dissolved in CHCl₃-CH₂Cl₂ by magnetic stirring. Hexanoyl chloride (0.1 mL, 0.6 mmol) and ethyldiisopropylamine (0.1 mL, 0.6 mmol) were added to this solution, and the reaction mixture was stirred for 4 h at room temperature. The solvents and excess reagents were removed in vacuo (oil pump), and the resulting residue was purified by PLC in CHCl₃-EtOH (100:6), twice developed, to give 5c (122 mg, 71%): pale-yellow needles, mp 187-187.5 °C (EtOH-H₂O); IR (CHCl₃) ν 3405, 3300, 1730, 1661, 1600, 1585, 1520, 1460, 1240, 1165 cm⁻¹; ¹H NMR (CDCl₃) δ 1.10 (t, H-18), 1.20 (d + m), 2.35 (m, H-19), 3.48 (d, OH), 5.48 (q, $J_{AB} = 12.7$ Hz, H-17), 6.82 (d, J = 8.0, NH); MS, m/e (rel intensity), 360 (M⁺ - 145) (15), 303 (5), 77 (100); $[\alpha]^{24}_{D} + 81^{\circ}$ (c 0.06, CHCl₃). Anal. (C₂₉H₃₅N₃O₅) C, H, N.

TLC analysis of crystalline **5c** in the above solvent system showed six trace contaminants having pale-blue fluorescence under short-wave UV light: R_f 0.18, 0.48, 0.67, 1.27, 1.43, 1.70 relative to **5c**. These also appeared after elution of **5c** (clearly a single band) from PLC adsorbents with EtOH. Similar impurities appeared in crystalline or PLC pure **5b**. These may be forming during TLC analysis on silica gel, since they were absent or appeared in lesser amounts in TLC analysis on Al₂O₃ (same solvents), on cellulose (toluene-EtOAc, 7:2), or on silica gel (acetone or EtOAc).

17-Acryloylcamptothecin-21-isopropylamide (5d). A sample of crude 5a (122 mg, 0.3 mmol) was acylated with acryloyl chloride (0.8 mmol), by the same procedures as used for the preparation of 5c, to give 5d (57 mg, 41%): mp 188–194.5 °C (isopropyl alcohol); IR (CHCl₃) ν 3500, 3408, 1725, 1660, 1655, 1585, 1405 cm⁻¹; ¹H NMR (CDCl₃) δ 1.16 (d + t, H-18 + 6 H), 2.47 (m, H-19), 3.99 (m, NCH), 5.07 (q, $J_{AB} = 19.5$ Hz, H-5), 5.58 (q, $J_{AB} = 12.0$ Hz, H-17), 5.71–6.53 (m, J = 2, 10, and 18 Hz, CH=CH₂), 6.87 (d, J = 7.8 Hz, NH), 7.35–8.01 (6 arom H); MS, m/e (rel intensity), 443 (M⁺ - 18) (5), 402 (5), 390, 373 [M⁺ - (O₂CCH=CH₂ + OH)] (20), 360 (25), 304 (65), 219 (85), 44 (100). Anal. (C₂₆H₂₇O₅N₃·*i*-PrOH) C, H, N.

17-Pyrrolidinylcamptothecin-21-isopropylamide (5e). Acetate amide **5b** (90 mg, 0.2 mmol) was dissolved in pyrrolidine (ca. 1 mL), and the mixture was stirred magnetically for 30 min at room temperature. The excess pyrrolidine was removed in vacuo, and the residue was purified by PLC in CHCl₃-EtOH (90:10). Several blue fluorescing bands (short-wave UV light) were seen; the major band (R_f 0.4) that reacted with Draggendorf's reagent was eluted from the adsorbent with EtOH to give **5e** (70 mg, 76%) as a pale-yellow solid: mp 203-209 °C dec (EtOH-H₂O); IR (film) ν 3400, 1650, 1590 cm⁻¹; ¹H NMR (CDCl₃) δ 1.25 [m, (CH₃)₂ + H-18], 1.92 (m, β -CH₂ of pyrrolidine), 2.38 (m, H-19), 2.98 (m, α -CH₂ of pyrrolidine), 4.18 [m + q, $J_{AB} = 11$ Hz, (CH₃)₂CH- + H-17], 5.17 (s, H-5), 7.33 (d, J = 8 Hz, NH), 7.48-8.28 (6 arom H); ¹³C NMR (CDCl₃) δ 8.1 (C-18), 22.8 and 23.6 [(CH₃)₂ + β -CH₂ of pyrrolidine], 32.8 (C-19), 41.5 [NCH-(CH₃)₂], 48.8 (C-17), 50.4 (C-5), 52.5 (α -CH₂ of pyrolidine), 79.8 (C-20), 102.1 (C-14), 125.9-131.1 (C-6 to -12), 143.8 (C-3), 148.8 (C-15), 151.6 (C-13), 157.5 (C-2), 161.7 (C-16a), 173.8 (C-21) (these assignments were made by reference to the ¹³C NMR spectrum of compounds 1²¹ and **5b** and SFOR decoupling experiments); UV (MeOH) 370, 293 (sh), 254 nm; UV (MeOH + 0.1 N HCI) 365, 254 nm; MS, m/e (rel intensity), 460.2449 ($C_{28}H_{32}N_4O_3$, calcd: 460.2473) (10), 391 (15), 303 (100); $[\alpha]^{24}_{D} + 108^{\circ}$ (c 0.01, CHCl₃). Substitution Products of 5b. The following compounds were

prepared only once and partially characterized.

17-Isopropylaminocamptothecin-21-isopropylamide. A small sample of 5b was stirred in neat isopropylamine for 20 min at room temperature. One major product was formed, which reacted positively with Draggendorf's reagent: IR (film) ν 3400, 1650, 1590 cm⁻¹; ¹H NMR (CDCl₃) δ 1.03 (m, 15 H), 2.27 (m, H-19), 3.35 (m, 1 H), 4.03 (m + q, 3 H), 5.13 (s, H-5), 6.02 (d, J = 10 Hz, NH); MS, m/e (rel intensity), 448 (M⁺·) (5), 390 (M⁺· - 58) (50), 303 (85).

17-Imidazolylcamptothecin-21-isopropylamide (5f). A sample of 5b (18 mg, 0.04 mmol) and imidazole (10 mg) were dissolved in Me₂SO (1 mL), and the reaction was stirred magnetically for 30 h at 50 °C. The reaction solution was poured into H_2O (15 mL) and extracted with CHCl₃ (3 × 5 mL), and the combined CHCl₃ extracts were dried (Na₂SO₄) and evaporated in vacuo. The resulting residue was purified by PLC in CHCl₃-MeOH (90:10) to give a single, Draggendorf-positive substance (18 mg, 100%): mp 189–194 °C; IR (film) ν 3200, 1650, 1600 cm⁻¹; ¹H NMR (Me₂SO-d₆) δ 0.92 (6, 3 H), 1.01, 1.09 (dd, 6 H), 2.18 (m, H-19), 3.9 (m, 1 H), 5.25 (br s, H-5), 5.39 (br s, H-17), 6.39-8.65 (9 arom H + NH); MS, *m/e* (rel intensity), 457.2124 (C₂₆H₂₇N₅O₃, calcd: 457.2114] (7), 390 (9), 389 (9), 371 (21), 360 (20), 303 (100), 68 (58).

17-Cyanocamptothecin-21-isopropylamide. A solution of **5b** (34 mg, 0.075 mmol) and KCN (100 mg, 1.53 mmol) in Me₂SO (1 mL) was stirred for 5 min at room temperature. The Me₂SO was removed in vacuo (oil pump), and the resulting residue was purified by PLC in CHCl₃-MeOH (95:5) to give a single product (27 mg, 90%): IR (film) ν 3250, 1670, 1600 cm⁻¹: ¹H NMR (CDCl₃) δ 1.3 (m, 9 H), 2.4 (m, H-19), 3.02 (q, $J_{AB} = 18$ Hz, 2 H), 3.94 (m, 1 H), 5.30 (br s, H-5), 7.30–8.42 (6 arom H); MS, m/e (rel intensity), 390 [M⁺ - CN] (6), 373 [M⁺ - (CN + OH)] (2), 360 (21), 303 (10), 44 (100).

17-Ethylmercaptocamptothecin-21-isopropylamide. A sample of 5b (20 mg, 0.044 mmol) was dissolved in Me₂SO (1 mL) containing EtSH (0.1 mL) and solid K₂CO₃ (5 mg). The reaction mixture was stirred magnetically for 1 h at room temperature; then the Me₂SO and excess EtSH were evaporated in vacuo (oil pump). The resulting solid residue was extracted with CHCl₃-MeOH (4:1), and the soluble material was purified by PLC in CHCl₃-MeOH (95:5) to yield one major product: IR (film) ν 3280, 1650, 1600 cm⁻¹; ¹H NMR (CDCl₃) δ 1.2 (m, 12 H), 2.40 (m, 2 H), 2.80 (m, 2 H), 4.06 [m, (CH₃)₂CH), 4.08 (q, J_{AB} = 13.5 Hz, 2 H), 5.07 (br s, H-5), 6.85 (d, J = 8 Hz, NH), 7.5–8.4 (6 arom H); MS, m/e (rel intensity), 451 [M⁺·] (4), 390 (25), 372 [M⁺· – (EtSH + OH)] (35), 360 (50), 303 (100).

Acknowledgment. We thank Mr. Gary Girdaukas, School of Pharmacy, and Mr. Mel Micke, University of Wisconsin Biochemistry Department, for low-resolution mass spectral determinations; Mr. Jim Blackbourn, School of Pharmacy, for NMR spectral determinations; and the National Institutes of Health (CA 17127) for partial research support. We are particularly indebted to Mr. Brent Hasse, University of Wisconsin School of Pharmacy, and Professor S. Horwitz, Albert Einstein College of Medicine, New York City, for in vitro assays.

References and Notes

- Postdoctoral fellow of the National Cancer Institute (CA 05876), 1977.
- (2) Career Development Awardee of the National Cancer Institute (CA 00253), 1976–1981.
- M. E. Wall, M. C. Wani, C. E. Cook, K. H. Palmer, A. T. McPhail, and G. A. Sim, J. Am. Chem. Soc., 88, 3888 (1967). It is also found in two other plants: T. R. Govindachari and N. Viswanathan, Indian J. Chem., 10, 453 (1972); S. Tafur, J. D. Nelson, D. C. Delong, and G. H. Svoboda, Lloydia, 39, 261 (1976).
- (4) B. J. Abbott, Cancer Treat. Rep., 60, 1007 (1976).

- (5) J. A. Gottlieb, A. M. Gaurino, J. B. Call, V. T. Oliverio, and J. B. Block, Cancer Chemother. Rep., 54, 461 (1970).
- (6) J. A. Gottlieb and J. K. Luce, *Cancer Chemother. Rep.*, 56, 103 (1972); C. G. Moertel, A. J. Schutt, R. J. Reitmeier, and R. G. Hahn, *ibid.*, 56, 95 (1972); F. M. Muggia, P. J. Creaven, H. H. Hansen, M. H. Cohen, and O. S. Selawry, *ibid.*, 56, 515 (1972).
- (7) G. R. Pettit, China Q., 59, 789 (1976).
- (a) S. B. Horwitz, Antibiotics, 3, 48-57 (1975); (b) J. A. Bristol, D. L. Comins, R. W. Davenport, M. J. Kane, R. E. Lyle, J. R. Maloney, D. E. Portlock, and S. B. Horwitz, J. Med. Chem., 18, 535 (1975); (c) M. E. Wall and M. C. Wani, Annu. Rev. Pharmacol. Toxicol., 17, 117 (1977); (d) J. L. Hartwell and B. J. Abbott, Adv. Pharmacol. Chemother., 7, 137-138 (1969).
- (9) Reviews: (a) A. G. Schultz, Chem. Rev., 73, 385 (1973); (b)
 M. Shamma and V. St. Georgiev, J. Pharm. Sci., 63, 163 (1974).
- (10) M. E. Wall, Biochem. Physiol. Alkaloid, Int. Symp. 4th, 1969, 77–87 (1972).
- (11) M. E. Wall, personal communication, June 2, 1978.
- (12) T. Sugasawa, T. Toyoda, N. Uchida, and K. Yamaguchi, J. Med. Chem., 19, 675 (1976).
- (13) For example, cf. G. P. Wheeler, B. J. Bowden, J. A. Grimsley,

and H. H. Lloyd, *Cancer Res.*, **34**, 194 (1974); W. C. J. Ross in *Handb. Exp. Pharmakol.*, **38**, 33 (1975); P. J. Cox and P. B. Farmer, *Cancer Treat. Rev.*, **4**, 47, 119 (1977).

- (14) S. K. Carter, F. M. Schabel, Jr., L. E., Broder, and T. P. Johnston, Adv. Cancer. Res., 16, 273 (1972).
- (15) (a) M. Isreal, E. J. Modest, and E. Frei, Cancer Res., 35, 1365 (1975); (b) cf. later studies by these investigators in Cancer. Treat. Rep., 62, 111, 119 (1978).
- (16) C. F. Murphy and J. A. Webber, in "Cephalosporins and Penicillins", E. H. Flynn, Ed., Academic Press, New York, 1972, Chapter 4. This may not be of mechanistic significance, since the sulfur atom of 6 also is felt to play an important role in the displacement reaction.
- (17) T. R. Govindachari, K. R. Ravindranath, and N. Viswanathan, J. Chem. Soc., Perkin Trans. 1, 1215 (1974).
- (18) Dr. Matt Suffness, National Cancer Institute, personal communication, June 1, 1978.
- (19) Professor S. Horwitz, Albert Einstein College of Medicine, New York, preliminary unpublished results.
- (20) K. Krohn, H.-W. Ohlendorf, and E. Winterfeldt. Chem. Ber., 109, 1389 (1976).
- (21) C. R. Hutchinson, A. H. Heckendorf, P. E. Daddona, E. Hagman, and E. Wenkert, J. Am. Chem. Soc., 96, 5609 (1974).

Synthesis and Antitumor Activity of Water-Soluble (2-Chloroethyl)nitrosoureas

Tetsuo Suami,* Kin-ichi Tadano,

Department of Applied Chemistry, Faculty of Engineering, Keio University, Hiyoshi, Yokohama 223, Japan

and William T. Bradner

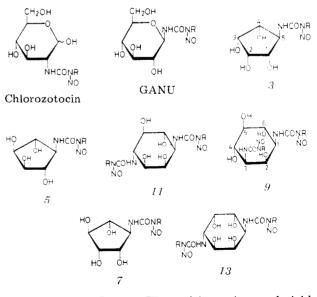
Bristol Laboratories, Syracuse, New York 13201. Received January 19, 1978

Six water-soluble (2-chloroethyl)nitrosoureido derivatives of cyclopentanetetrols and cyclohexanetetrols have been prepared. Their antitumor activities were determined against leukemia L1210 in mice.

Steptozotocin is a naturally occurring antitumor antibiotic¹⁻⁴ and has a unique structure: 2-(3-methyl-3-nitrosoureido)-2-deoxy-D-glucopyranose.^{5,6} In this antibiotic, the methylnitrosoureido group seems to be an essential functional group for biological activity, and the carbohydrate moiety might play an important role as a carrier of the functional group. This has been verified by the fact that the introduction of the functional group into a polyhydroxycyclohexane system did not result in loss of activity against leukemia L1210.⁷ Also, O-methylation of an anomeric hydroxyl group of streptozotocin⁸ and its analogues⁹ was not detrimental for activity.

Montgomery and his co-workers showed in their systematic studies on N-nitrosoureas^{10,11} that a replacement of the methyl group in this functional group with a 2-chloroethyl group markedly enhanced the activity of the N-nitrosoureas against leukemia L1210. Accordingly, the clinically useful BCNU,¹⁰ CCNU,¹¹ and MeCCNU¹¹ were synthesized. More recently, 2-[3-(2-chloroethyl)-3-nitrosoureido]-2-deoxy-D-glucopyranose (chlorozotocin)¹² exhibited strong antileukemic activity,¹³⁻¹⁵ and its positional isomer, 1-(2-chloroethyl)-3-(β -D-glucopyranosyl)-1-nitrosourea (GANU)¹⁶ showed marked activity against leukemia L1210.¹⁷

Now we report a synthesis of six water-soluble (2chloroethyl)nitrosoureido derivatives of cyclopentanetetrols and cyclohexanetetrols and their antileukemic activities



against leukemia L1210. The melting points and yields of compounds 2-13 are listed in Tables I and II.

Chemistry. A synthesis of 5-[3-(2-chloroethyl)-3nitrosoureido]-1,2,3,4-cyclopentanetetrols was exemplified in the case of its (1,2,3,4/5) stereoisomer.¹⁸ When tetra-*O*-acetyl-(1,2,3,4/5)-5-acetamido-1,2,3,4-cyclo-