

Pyrrolizidine Alkaloids: Evidence for *N*-(4-Aminobutyl)-1,4-diaminobutane (Homospermidine) as an Intermediate in Retronecine Biosynthesis

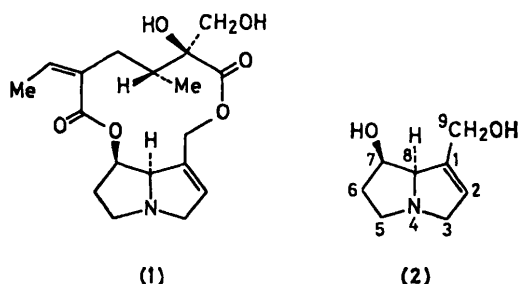
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Summary [1-amino-¹⁵N; 1-¹³C]Putrescine (**3**) is incorporated into retronecine (**2**) in *Senecio isatideus* plants with a labelling pattern consistent with the formation of a symmetrical C₄-N-C₄ intermediate; the intermediate is shown to be homospermidine (**4**) by ¹⁴C-labelling experiments.

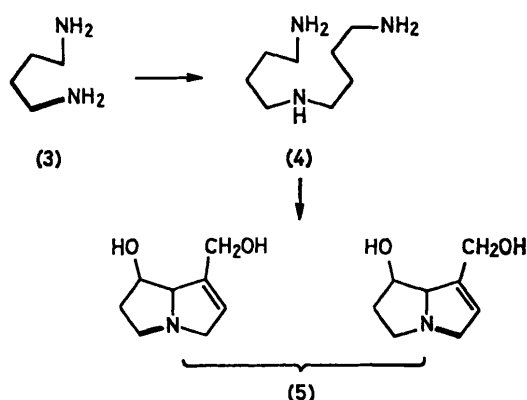
PUTRESCINE [as (**3**)] is the most efficient precursor so far found for retronecine (**2**),¹ the base portion of many pyrrolizidine alkaloids. Retronecine is formed by alkaline hydrolysis of retrorsine (**1**), the major alkaloid present in *Senecio isatideus* plants. ¹³C-Labelled putrescines have recently been used to establish the complete labelling pattern in retronecine (**2**).² The results indicate that two putrescine molecules combine to form retronecine with

nearly equal labelling in both halves of the molecule. This suggests, but does not prove, that a later C_4-N-C_4 symmetrical intermediate is involved in retronecine biosynthesis.³ We believed that the use of [^{13}C - ^{15}N]-labelled putrescine (3) would provide this proof. Biosynthesis of



retronecine (5) from (3) via a symmetrical intermediate [such as (4)] should produce two ^{13}C - ^{15}N couplings in the $\{^1H\}$ - ^{13}C n.m.r. spectrum of retronecine (Scheme).

The *N*-benzyloxycarbonyl derivative of 1-amino-3-bromopropane was treated with $K^{13}C^{15}N$ (B.O.C. Prochem Ltd., containing 90.6% ^{13}C and 99.4% ^{15}N) to give the corresponding nitrile [$\{^1H\}$ - ^{13}C n.m.r. spectrum ($CDCl_3$)



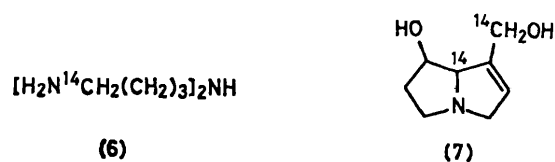
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δ 119.0 p.p.m. (d, J 17 Hz)], which was catalytically hydrogenated to give [1-amino- ^{15}N ; 1- ^{13}C]putrescine (3) isolated and recrystallised as its dihydrochloride (28% overall yield) [$\{^1H\}$ - ^{13}C n.m.r. spectrum (D_2O) δ 39.6 p.p.m. (d, J 5.1 Hz)].

Introduction of the [^{13}C - ^{15}N]-labelled precursor (3), together with [1,4- ^{14}C]putrescine dihydrochloride (5 μ Ci) into two three-month old *Senecio isatideus* plants was carried out as described previously.¹ Retrorsine (1) was extracted and recrystallised to constant specific activity (2.2% specific incorporation). Alkaline hydrolysis of retrorsine gave retronecine (2) isolated and recrystallised as its hydrochloride with the same specific activity.

Comparison of the 25 and 90 MHz $\{^1H\}$ - ^{14}C n.m.r.

spectra of labelled retronecine hydrochloride² taken in D_2O , with unlabelled material run under the same conditions, showed enrichment factors of 0.4% for the signals at δ 55.4 (C-5) and 80.6 p.p.m. (C-8), and 0.5% for those at δ 62.6 (C-3) and 59.2 p.p.m. (C-9). This corresponds to a total enrichment factor of 1.8%, and a specific ^{13}C incorporation of ca. 2%. In addition, the resolution-enhanced spectra showed the presence of doublets at δ 55.4 (J 4.5 Hz) and 62.6 p.p.m. (J 5 Hz) with enrichment factors of 0.2–0.25%. The presence of ^{13}C - ^{15}N species in retronecine hydrochloride was confirmed by observation of the 36.5 MHz $\{^1H\}$ - ^{15}N n.m.r. spectrum taken in D_2O , which showed ^{13}C - ^{15}N satellites (J ca. 5 Hz) in addition to the natural-abundance signal at δ 311.2 p.p.m. upfield from external nitromethane. The fact that C-3 and C-5 of retronecine are both enriched approximately equally with ^{13}C - ^{15}N species [as in (5)] provides strong evidence for the involvement of a symmetrical, C_4-N-C_4 , intermediate in retronecine biosynthesis.



A reasonable possibility for this symmetrical intermediate is *N*-(4-aminobutyl)-1,4-diaminobutane (homospersmidine) (4), a known plant constituent.⁴ Accordingly ^{14}C -labelled homospersmidine (6) was synthesized as follows. The *N*-benzyloxycarbonyl derivative of 4-aminobutanoic acid was condensed with 3-bromopropylamine. Treatment of the protected bromoamide with $K^{14}CN$, followed by hydrogenation and reduction with borane in tetrahydrofuran, yielded ^{14}C -labelled homospersmidine (6), isolated and recrystallised as its trihydrochloride (20% yield from the bromoamide). The feeding experiment was carried out as usual.¹ Retrorsine (1) was isolated, diluted with inactive alkaloid, and recrystallised to constant specific activity (0.5% total incorporation, 0.3% specific incorporation). Retronecine (7), derived from retrorsine by base hydrolysis, had the same specific activity. Treatment of retronecine with OsO_4 - HIO_4 gave formaldehyde (from C-9),⁵ isolated as its dimedone derivative, containing $44 \pm 4\%$ of the retronecine activity. Modified Kuhn-Roth oxidation of retronecine gave β -alanine [from C-(5 + 6 + 7)], isolated as its *N*-2,4-dinitrophenyl derivative, with $2 \pm 2\%$ of the retronecine activity. These results are consistent with the intact incorporation of homospersmidine (6) into retronecine (7). Moreover, homospersmidine was isolated in a radioactive form after feeding DL-[5- ^{14}C]ornithine to an *S. isatideus* plant. After 24 h, the plant was macerated in 0.4 M aqueous trichloroacetic acid containing inactive homospersmidine trihydrochloride (35 mg). The *N*'-substituted *N*-phenylthiourea derivative of homospersmidine was prepared and purified as described for other polyamines.⁶ Recrystallisation to constant specific activity gave a radioactive derivative containing 0.5% of the original activity fed, thus indicating that homospersmidine is a normal intermediate in retronecine biosynthesis.

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