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ARTICLE TYPE

Enantioselective total Syntheses and determination of absolute configuration of marine toxins, oxazinins.

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The enantioselective total syntheses of natural marine toxins, oxazinin-1, -2, -4, -5, -6 and linear precursor preoxazinin-7 are described. The synthetic highlights include Sharpless asymmetric aminohydroxylation and dihydroxylation, Oxa-Michael reaction and intramolecular diastereoselective addition of an appropriate hydroxyl substituent to a 3-methyleneindolenine for the construction of ¹⁰ morpholine ring as key steps. The synthetic route also allowed the synthesis of the epi-preoxazinin and a structurally related secondary metabolite bursatellin isolated in 1980 from sea hare Bursatella leachii pleii and its epimer.

Introduction

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The Marine biotoxins pose serious threats to both human health ¹⁵ and fishery resources throughout the world¹. The problem has aggrevated in last two decades, earlier only few regions were affected in scattered locations, now in essence every costal region is affected, not only fish and shellfish but life of marine mammals, seabirds and other animal is also in danger due to the ²⁰ toxic algal species². In order to prevent or minimize such damage, lot of efforts are directed towards structure elucidation and synthetic confirmation of these toxins³. These marine biotoxins are found to be polyether compounds such as okadaic acid⁴, yessotoxin⁵ etc. During their investigation of toxic Adriatic ²⁵ mussels, Ciminiello *et al* have also isolated new type of toxins, which are completely different from polyether biotoxins isolated so for, but can raise further alarm for Sea life and public health.



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Figure 1. Structures of Oxazinin-1, -2, -3, -4, -5, -6, Preoxazinin-7 and Bursatellin

In 2001, Fattorusso and Ciminiello *et al* reported the isolation of Oxazinin-1 (1), -2 (2), and -3 (3)⁶ from the digestive glands of *Mytilus galioprovincialis* from northern Andriatic Sea. ⁵ Subsequentaly in 2006 cimminello *et al* reported isolation of another natural product Oxazinin-4 (4)⁷ from toxic mussels collected along the Emilia Romagna coasts, Italy. Again in 2007 same group reported isolation of three new natural products, namely, oxazinin-5 (5), -6 (6), and a linear precursor preoxazinin-

- ¹⁰ 7 (7)⁸. The structural elucidation of these oxazinins has been determined by extensive 2D NMR analysis and Molecular Mechanics and dynamics calculations. The absolute configuration of oxazinin-1 was established by application of the 9-AMA shift correlation method for β -chiral primary alcohols⁹. Possible ¹⁵ biogenetic pathway was proposed by Ciminiello *et al* leading to
- these novel class of cytotoxic marine natural products⁸. Oxazinin-1 (1) was shown to inhibit the proliferation of
- fibrosarcoma cell (WEHI-164) and J774 macrophages in vitro⁶. The originally assigned *cis*-stereochemistry at C-2 and C-6 centre ²⁰ in oxazinin-1 (1) and oxazinin-2 (2) was latter corrected as *trans*
- by Ciminiello *et al.* in 2006⁷. Because of their potential risk, these novel class of cytotoxic natural products could represent for human health and fish industries, comprehensive toxicological studies are needed. However their evaluation is hampered by the 25 limited availability of oxazinins from isolation, therefore efficient
- total synthesis of these natural products and their analogues are necessary.
- In this direction, Couladouros has reported total synthesis of a simple member of this family, oxazinin-3 (3) containing two ³⁰ stereocentres¹⁰. Subsequently in 2007, Baran group¹¹ also reported a 4 step synthesis of oxazinin-3 (3) using the direct coupling of indole to carbonyl compound under basic condition as key step. In 1980 Schmitz *et al* isolated a metabolite bursatellin¹² from the sea hare Bursatella leachii pleii. Initially ³⁵ Schmitz *et al* proposed structure **8**¹² for bursatellin, later in 1987 research group of G. Sodano¹³ collected two Mediterranean sea hares, *Bursatella leachii leachii and Bursatella leachii savignyana*, and isolated from them a compound that exhibited ¹H NMR spectral data nearly identical with that reported for ⁴⁰ bursatellin except for one additional low-field singlet observed at 8.20 ppm, this eventually lead to the revision of originally
- s.20 ppm, this eventually lead to the revision of originally proposed structure of bursatellin 8 to structure 9. Structurally bursatellin 9 is similar to preoxazinin-7 (7) except that the indole glyoxylic amide functionality in preoxazinin-7 (7) is replaced by 45 formamide group. In 1990 Sodano *et al*¹⁴ reported the
- ⁴⁵ formamide group. In 1990 Sodano *et al* reported the semisynthesis of bursatellin **9** from chloramphenicol, and later in 1991 Hiroshi *et al*¹⁵ reported the total synthesis of bursatellin **9** in 16 steps starting from L-tryptophan, both these syntheses are tedious and very low yielding.
- ⁵⁰ We recently initiated a research program directed towards the total synthesis of oxazinins. In preliminary communication¹⁶ we have reported the first total synthesis of oxazinin-5 (5), -6 (6), and their biogenetic precursor preoxazinin-7 (7). Prior to our report there were no reports of synthesis of oxazinins except for
- ss oxazinin-2 as described earlier. In this article we describe further studies in this area that include enantioselective total synthesis of oxazinin-1 (1) -2 (2), -4 (4) as well as epi-preoxazinin-7 (10) and *epi*-bursatellin 11.

60 Results and discussion.

Retrosynthetic analysis of oxazinins is depicted in scheme 1. It was envisaged that oxazinin-1 (1), -2 (2), and -4 (4) could be obtained from intermediate 12 by intramolecular regioselective addition of secondary hydroxyl group at -C-X position to 3-65 methyleneindolene moiety while oxazinin-5 (5) and -6 (6) could be prepared from same intermediate 12 by regioselective intramolecular addition of primary alcohol to 3methyleneindolene. Key intermediate 12 in turn could be generated in situ from chemoselective reduction of preoxazinin-7 70 (7) via 3-hydroxymethylindole intermediate. Preoxazinin-7 (7) could be easily obtained by coupling of 3-indoleglyoxylic acid chloride 14 and tyrosine derivative 13. To unambiguously establish the relative as well as absolute configuration of all oxazinin natural products, both (R, R) and (R, S) diastereomers of 75 unnatural amino acid derivative could be synthesized from ethyl cinnamate 15 by Sharpless asymmetric dihydroxylation¹⁷ and Sharpless asymmetric aminohydroxylation¹⁸ respectively.



80 Scheme 1. Retrosynthetic Analysis

To begin with the cinnamate **15** was prepared in two step from phydroxybenzaldehyde **16** by first protection of hydroxyl group as its benzyl ether followed by treatment of resultant aldehyde with PPh₃=CHCO₂Et. As discussed in retrosynthetic analysis ss cinnamate **15** was subjected to Sharpless asymmetric hydroxylation and Sharpless asymmetric aminohydroxylation, independently. Treatment of cinnamate **15** with AD mix-β gave diol **17** in 94% yield and 98% ee. Nosylation (Nscl, Et₃N, 92% Yield) of the diol **17** followed by treatment with sodium azide ⁹⁰ furnished hydroxyl azide **19** in 90% yield via nosylate **18**. SnCl₂ mediated reduction of the azido group within **19**, followed by in situ trapping of the resulting amine with $(Boc)_2O$ furnished amino derivative **20** in 80% overall yield. On the other hand Sharpless



Scheme 2. Synthesis of diastereomers of tyrosine derivative

- ⁵ asymmetric aminohydroxylation¹⁸ [NaOH, BocNH₂, ¹BuOCl, (DHQD)₂AQN, K₂OsO₂(OH)₄, nPrOH/H₂O (1:1)], of substrate 15 afforded directly boc protected amino alcohol 21 in 45% yield and 98% ee. Finally silylation of 20 and 21 (TBSCl, imidazole, 98% yield) gave the targeted (*R*, *R*) 22 and (*R*, *S*) 23 phenyl ¹⁰ alanine derivatives respectively. Hydrogenolysis of benzyl ether
- in 22 and 23 using Pd/C afforded phenols 24 and 25 respectively (*Scheme 2*).

Next we turned our attention towards the cyanoethylation of the phenolic hydroxyl group in compound 24 using oxa-Michael ¹⁵ reaction, our initial efforts using several standard bases failed to facilitate this reaction possibly due to retro-Michael fragmentation, as mentioned by the two independent groups of Sodano *et al*¹⁴ and Hiroshi *et al*¹⁵ in their synthesis of bursatellin 9, a metabolite isolated in 1980¹². Sodano *et al*.¹⁴ have also ²⁰ shown that bursatellin 9 on treatment with dil. aq. Na₂CO₃ undergoes retro-Michael fragmentation reaction to generate the corresponding phenol derivative. After trying several different conditions (**table 1**), finally to our delight treatment of aminophenol 24 with acrylonitrile using 20 mol% of triton-B

²⁵ under reflux condition generated the desired oxa-Michael product 26 in 92% yield.

After having fully functionalized amino ester derivative 26 in hand the stage was set for coupling reaction using

indoleglyoxylic chloride. Accordingly reduction of ester in 30 compound **26** using LiBH₄ followed by deprotection of boc group using TFA/CH₂Cl₂ afforded the amine 27. Coupling of amine 27

Table 1 : Optimization of oxa-Michael reaction



with indoleglyoxylic chloride [prepared in one step by treatment of indole with oxalyl chloride] followed by TBS-deprotection using TBAF afforded epi-preoxazinin (10). On the other hand amine 27 on refluxing with ethyl formate followed by TBS-40 deprotection generated epi-bursatellin (11) in 80% over all yield (*Scheme 3*).

After making epimers 10 and 11, we next turned our attention towards syntheses of preoxazinin-7 (7) and bursatellin 9. Oxa-Michael reaction of phenol 25 with acrylonitrile using 20 mol% 45 of triton-B under reflux condition as described earlier for diastereomer 24 generated the desired oxa-Michael product 13 in 92% yield. Reduction of ester 13 generated the primary alcohol 28 in 83% vield. Unlike the diastereomer 24, boc deprotection of compound 28 proved to be more complex and led to multiple 50 products and decomposition under various conditions such as TFA/CH₂Cl₂^{19a}, TFA-Triethylsilane^{19b}, 4M HCl in dioxane^{19c}. CAN^{19d}, TMSOTf^{19e}, and 10% H₂SO₄^{19f} and TMSI^{19g}. Interestingly treatment of 28 with stoichiometric amount of Yb(OTf)₃ under CH₂Cl₂ reflux conditions generated boc-55 deprotected amine 29 in excellent yield. Amine 29 on treatment with ethyl formate under reflux condition followed by TBS deprotection afforded the natural product bursatellin 9 in 80% vield (Scheme 4).

It was envisioned that amine **29** on coupling with indoleglyoxalic ⁶⁰ chloride **14** would generate the key intermediate **30**, which will eventually lead to synthesis of oxazinin 5 (**5**), -6 (**6**) and preoxazinin-7 (**7**). On the other hand, boc-deprotection of amino group in compound **13** followed by coupling with indoleglyoxylic acid and further functional group transformation would generate ⁶⁵ oxazinin-1 (**1**), -2 (**2**) and -4 (**4**). Accordingly coupling of amine **29** with indoleglyoxylic acid afforded ketoamide **30** in 70% yield. TBS-deprotection of compound **30** generated the natural product preoxazinin-7 (**7**) in excellent yield (*Scheme 5*). Reduction of ketone functionality in 3-indoleglyoxylic amide **30** using sodium borohydride generated the diastereomeric mixture of diols **31a,b** Which was then further treated with 10 mol% of PPTS in refluxing acetonitrile without further purification to 5 furnish morpholinones **32a** and **32b** as a 3:1 mixture of C-2 diastereomers. The two diastereomers **32a** and **32b** were



Scheme 3. Synthesis of epi-preoxazinin-7 and (+)-epi-bursatellin



10 Scheme 4. Synthesis of (-) bursatellin

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separated by preparative TLC. Finally TBS-deprotection of major **32a** and minor **32b** diastereomers provided natural products oxazinin-5 (5) and oxazinin-6 (6) respectively along with minor amount of cyanoethyl side chain cleaved compound **33a** and **33b** (Sakema 6). On the other hand, for the sumbases of overlaping 1

- 15 (Scheme 6). On the other hand, for the syntheses of oxazinin-1 (1), -2 (2) and -4 (4), boc group in compound 13 was deprotected using Yb(OTf)₃ under CH₂Cl₂ reflux conditions. The resultant amine was then coupled with indoleglyoxylic chloride using Et₃N to generate the intermediate 34 in excellent yield. TBS-
- ²⁰ deprodection of compound **34** using TBAF/THF afforded the alcohol **35** in 78% yield. To our surprise NaBH₄ reduction of indoleglyoxylic amide **35** not only reduced the ketone functionality but also the ester group to afford diastereomeric mixture of diols **36a,b**. This is one of the rare example of ester
- ²⁵ reduction using NaBH₄. The mixture of diols **36a,b** on treatment with 10 mol% PPTS in refluxing acetonitrile furnished the natural products oxazinin-1 (1) and -4 (4) along with small amount of oxazinin-5 (5) (3%) and oxazinin-6 (6) (2%) which were





Scheme 5. Synthesis of preoxazinin-7



Scheme 6. synthesis of Oxazinin-5 and -6



Scheme 7. Synthesis of Oxazinin-1 (1),-2 (2) and -4 (4)

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CD curves (for oxazinin-5,-6 and preoxazinin-7) identical with the natural oxazinin-1 (1), -2 (2), -4 (4), -5 (5), -6 (6) and preoxazinin-7 (7) reported in literature, thus confirming the structure of the natural products as well as their relative and s absolute stereochemistry.

Conclusions

In summary, we have completed the first enantioselective total syntheses of oxazinin-1, -2, -4, -5, -6 and their biogenetic linear precursor preoxazinin-7 along with structurally related metabolite ¹⁰ bursatelline and its epimer using, aminohydroxylation and oxa-Michael reaction as the key steps, in a longest linear sequence of 8 steps, from the cinnamate **15** with excellent overall yields. In addition, the absolute configuration of stereogenic center were firmly established by the total synthesis to be (2*S*, 5*R*, 6*R*) for ¹⁵ oxazinin-1 and -2, (2*R*, 5*R*, 6*R*) and for oxazinin-4, (2*R*, 5*R*, 7*R*) for oxazinin-5, (2*S*, 5*R*, 7*R*) for oxazinin-6 and (4*R*, 5*R*) for preoxazinin-7. The synthetic route developed here is general and efficient and could also be applied to the syntheses of other substituted morpholine related natural products.

20 Experimental section

General Information

All reactions were carried out under nitrogen or argon atmosphere with dry solvents under anhydrous conditions, unless otherwise mentioned. Anhydrous THF and diethyl ether were ²⁵ distilled from sodium-benzophenone and dichloromethane was distilled from calcium hydride. Yields refer to chromatographically pure material, unless otherwise stated. Reactions were monitored by thin-layer chromatography (TLC) carried out on 0.25 mm Merck silica gel plates (60F-254) using a LIV, light on a visualizing agant and on provided or

- ³⁰ UV light as a visualizing agent and an p-anisaldehyde or ninhydrin stain, and heat as developing agents. Merck silica gel (particle size 100-200 and 230-400 mesh) was used for flash column chromatography. Reagents were purchased at the highest commercial quality and used without further purification, unless ³⁵ otherwise stated. NMR spectra were recorded on either a Bruker
- ³⁵ otherwise stated. NMR spectra were recorded on either a Bruker Avance 200 (¹H: 200 MHz, ¹³C: 50MHz), Bruker Avance 400 (¹H: 400 MHz, ¹³C: 100MHz), Bruker Avance 500 (¹H: 500 MHz, ¹³C: 125 MHz), JEOL ECX 400 (¹H: 400 MHz, ¹³C: 100 MHz) JEOL ECX 500 (¹H: 500 MHz, ¹³C: 125 MHz). Mass
- ⁴⁰ spectrometric data were obtained using WATERS-Q-Tof Premier-ESI-MS. Optical rotations were measured on a Rudolph and JASCO using a 6.0 ml and 1.0 ml cell with a 100 and 50 mm path length are reported as $[\alpha]_D^{25}$ (c in g per ml solvent) at 25 °C. Enantiometic ratios (er) were determined by HPLC using
- ⁴⁵ grace denali RP-18 250 x 4.6 and kromasil 5- cellucoat 0.46cm x 25cm (detection at 230 nm). Diastereomeric ratios (dr) were determined by crude ¹H NMR. The following abbreviations were used to explain the multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, dd = doublet of doublet, dt = doublet of triplet, ⁵⁰ ddd = doublet of a doublet of a doublet, dm = doublet of a doublet of a doublet.
- multiplet, m = multiplet, br = broad.

Experimental procedures

(E)-ethyl 3-(4-(benzyloxy)phenyl)acrylate (15): To a solution of the aldehyde 16a (8.0 g, 37.7 mmol) in water (150 ml) was

⁵⁵ added PPh₃=CHCO₂Et salt (19.7 g, 56.5 mmol) and heated it to 90 °C for 2h. The reaction mixture was cooled to RT and extracted with CH₂Cl₂ (3 x 80 ml), washed with brine and dried over Na₂SO₄. Evaporation of the solvent and purification of the residue on a silica gel column using EtOAc-petroleum ether (1:9)
⁶⁰ as eluent furnished the ester **15** (*E/Z*-ratio : 95/5, 10.0 g, 94%), *Rf* = 0.30 (EtOAc-petroleum ether, 1:9). IR (neat): vmax/cm⁻¹ 3282, 1682 (C=O), 1632, 1603, 1585, 1515, 1439, 1371, 1279, 1189, 1033, 830; ¹H NMR (200 MHz, CDCl₃): δ 7.64 (d, *J* 16.0 Hz, 1H) 7.53-7.29 (m, 7H), 6.97 (dt, *J* 8.7 and 2.0 Hz, 2H), 6.31 (d, 65 *J* 16.0 Hz, 1H), 5.10 (s, 2H), 4.25 (q, *J* 14.3 and 7.1 Hz, 2H), 1.33 (t, *J* 7.1 Hz, 3H); ¹³C NMR (50 MHz, CDCl₃): δ 167.3, 160.5, 144.2, 136.5, 129.7, 128.6, 128.1, 127.4, 115.9, 115.2, 70.1, 60.3, 14.3; HRMS (ESI): *m/z* calcd for C₁₈H₁₉O₃ (M+H)

283.1334; found 283.1330.

(2R,3R)-ethyl 3-(4-benzyloxy)phenyl)-2,3-dihydroxypropanoate (17): To a solution of AD-mix β (86.6 g) in t-BuOH/H₂O (1:1, 800.0 ml) was added MeSO₂NH₂ (5.9 g, 61.9 mmol) at RT. The mixture was cooled to 0 °C, and ethyl cinnamate 15 (17.5 g, 75 61.9 mmol) was added dropwise. The resulting mixture was warmed to RT over a period of 16 h before it was quenched with Na₂SO₃ (200 ml, sat. aq. solution), extracted with EtOAc (3 \times 200 ml). The combined organic layers were washed with brine (200 ml), dried over Na₂SO₄. Evaporation of the solvent and 80 purification of the residue on silica gel column using ethyl acetate-petroleum ether (1:2) as eluent furnished the diol 17 (16.9 g, 86%); Rf = 0.40 (EtOAc-petroleum ether, 1:2). $[\alpha]_D^{25} = -10.6$ (c 4.7, CHCl₃); IR (neat): v_{max}/cm⁻¹ 3438, 3019, 1727, 1613, 1585, 1513, 1455, 1216, 1106, 1026, 747, 697; ¹H NMR (200 85 MHz, CDCl₃): δ 7.44-7.30 (m, 7H), 6.98 (d, J 8.7 Hz, 2H), 5.06 (s, 2H), 4.95 (dd, J 5.8 and 3.0 Hz, 1H), 4.33-4.29 (m, 1H), 4.25 (q, J 12.1 and 4.9 Hz, 2H), 3.17 (d, J 6.0 Hz, 1H), 2.75 (d, J 6.7 Hz, 1H), 1.25 (t, J 7.0 Hz, 3H); ¹³C NMR (50 MHz, CDCl₃): δ 172.7, 158.6, 136.9, 132.3, 128.6, 127.9, 127.6, 127.4, 114.7, 90 74.7, 74.2, 70.0, 62.1, 14.1; HRMS (ESI): m/z calcd for C₁₈H₂₀O₅Na (M+Na) 339.1208; found 339.1204.

(2*S*,3*R*)-ethyl-(4-(benzyloxy)phenyl)-3-hydroxy-2-(4nitrophe-

nylsulf onyloxy)propanoate (18): To a solution of diol 17 (11.2 95 g, 35.4 mmol) in CH₂Cl₂ (85 ml) were added NsCl (8.7g, 35.4 mmol) and Et₃N (7.4 ml, 53.1 mmol) at 0 °C. The reaction mixture was stirred for 6 h before it was quenched with HCl (20.0 ml, 5 % aq). Water was added to the reaction mixture, extracted with CH₂Cl₂ (3 x 100 ml), washed with brine and dried over 100 Na₂SO₄. Evaporation of the solvent and purification of the residue on a silica gel column using ethyl acetate-petroleum ether (1:4) furnished the nosylate **18** (16.3 g, 92%); Rf = 0.40 (EtOAcpetroleum ether 1:4). $[\alpha]_D^{25} = -40.0$ (c 0.9, CHCl₃); IR (neat): v_{max}/cm^{-1} 3437, 2926, 1760, 1610, 1532, 1455, 1378, 1350, 1188, 105 1094, 1026, 744, 617; ¹H NMR (200 MHz, CDCl₃): δ 8.23 (dt, J 9.1 and 4.3 Hz, 2H), 7.85 (dt, J 9.2 and 4.3 Hz, 2H), 7.45-7.32 (m, 5H), 7.13 (d, J 8.7 Hz, 2H), 6.80 (dt, J 8.8 and 4.8 Hz, 2H), 5.14 (d, J 4.2 Hz, 1H), 5.00 (s, 2H), 4.95 (d, J 4.0 Hz, 1H), 4.15 (q, J 14.3 and 7.2 Hz, 2H), 2.60 (s, 1H), 1.17 (t, J 7.2 Hz, 3H); ¹¹⁰ ¹³C NMR (50 MHz, CDCl₃): δ 166.4, 158.9, 150.5, 141.4, 136.4, 129.5, 129.1, 128.6, 128.1, 127.5, 127.4, 124.0, 114.7, 82.4, 73.0, 69.9, 62.5, 13.8; HRMS (ESI): *m/z* calcd for C₂₄H₂₃NO₉SNa

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70

(M+Na) 524.0991; found 524.1001.

(2S,3R)-ethyl-2-azido-3-(4-(benzyloxy)phenyl)-3-hydroxypropanoate (19): To a solution of nosylate 18 (15.0 g, 29.8 mmol) in 5 DMF (80.0 ml) was added sodium azide (11.6 g, 179.7 mmol) at RT. The resulting reaction mixture was heated at 50 °C for 38h. After cooled to RT, the reaction mixture was concentrated in vacuo, extracted with ethyl acetate (3 x 100 ml), washed with brine and dried over Na₂SO₄. Evaporation of the solvent and 10 purification of the residue on a silica gel column using ethyl acetate-petroleum ether (2:3) furnished azide 19 (9.2 g, 90%), Rf = 0.40 (EtOAc-petroleum ether 2:3). $[\alpha]_D^{25} = +63.8$ (c 6.0, CHCl₃); IR (neat): v_{max}/cm⁻¹ 3443, 2112, 1574, 1397, 1115, 618, 536; ¹H NMR (200 MHz, CDCl₃): δ 7.43-7.30 (m, 8H), 7.00 (dt, 15 J 8.8 and 5.1 Hz, 2H), 4.98 (dd, J 7.0 and 4.4 Hz, 1H), 4.26 (q, J 14.3 and 7.0 Hz, 2H), 4.10 (d, J 7.0 Hz, 1H), 2.78 (d, J 4.4 Hz, 1H), 1.58 (s, 1H), 1.28 (t, J 7.1 Hz, 3H); ¹³C NMR (50 MHz, CDCl3): 8 170.0, 159.0, 136.7, 131.3, 128.6, 128.0, 127.9, 127.4, 114.9, 73.6, 69.9, 66.8, 62.1, 14.0; HRMS (ESI): m/z calcd for 20 C18H19N3O4Na (M+Na) 364.1273; found 364.1282.

(2S,3R)-ethyl 3-(4-(benzyloxy)phenyl)-2-(tert-butoxycarbonyl-

- amino)-3-proxypropanoate (20): To a solution of azide 19 (4.0 g, 11.7 mmol) in dioxane (60.0 ml) was added a solution of 25 SnCl₂•2H₂O (13.2 g, 58.5 mmol) in H₂O/dioxane (4:1, 100.0 ml) at 0 °C. The resulting mixture was warmed to RT over a period of 5 h before it was cooled to 0 °C. Solid NaHCO₃ was slowly added until pH 8-9 was reached. Boc₂O (3.8 g, 17.6 mmol) was added at 0 °C and the reaction mixture was warmed to RT over a 30 period of 15 h before it was acidified by slow addition of HCl (2 N) until pH 5-6 was reached. The reaction mixture was filtered through a mixture of silica gel and celite, and the filtrate was extracted with EtOAc (4×30 ml). The combined organic layers were dried over Na2SO4 Evaporation of the solvent and 35 purification of the residue on silica gel column using ethyl acetate-petroleum ether (1:3) as eluent furnished the Boccarbamate 20 (3.1 g, 80%). Rf = 0.40 (EtOAc-petroleum ether, 1:3). $\left[\alpha\right]_{D}^{25} = -50.2$ (c 9.6, CHCl₃); IR (neat): v_{max}/cm^{-1} 3436, 2979, 2932, 1715, 1612, 1586, 1511, 1455, 1369, 1243, 1170, 40 1114, 1025, 864, 752; ¹H NMR (200 MHz, CDCl₃): δ 7.43-7.30 (m, 5H), 7.20 (d, J 8.6 Hz, 2H), 6.94 (dt, J 8.7 and 2.8 Hz, 2H), 5.32 (br d, J 7.5 Hz, 1H), 5.12 (br s, 1H), 5.04 (s, 2H), 4.65 (dd, J
- 7.6 and 3.9 Hz, 1H), 4.13 (q, J 14.3 and 7.2 Hz, 2H), 4.04 (br s, 1H), 1.43 (s, 9H), 1.19 (t, J 7.2 Hz, 3H); ¹³C NMR (100 MHz, 45 CDCl₃): δ 169.8, 158.5, 156.3, 136.8, 131.6, 128.5, 127.9, 127.4, 127.3, 114.5, 80.5, 74.6, 69.9, 61.6, 59.7, 28.2, 14.0 HRMS
- (ESI): m/z calcd for $C_{23}H_{30}NO_6$ (M+H) 416.2073; found 416.2065.
- 50 (2S,3R)-ethyl 3-(4-(benzyloxy)phenyl)-2-((tert-butoxycarbonyl)amino)-3-(tert-butyldimethylsilyloxy)propanoate (22): To a solution of the alcohol 20 (2.0 g, 4.8 mmol) in DMF (15.0 ml) were added TBSCl (1.0 g, 6.2 mmol), imidazole (0.5 g, 7.2 mmol) and stirred magnetically at RT for 12 h. Water was added
- 55 to the reaction mixture, extracted with ethyl acetate (3 x 20 ml), washed with brine and dried over Na₂SO₄. Evaporation of the solvent and purification of the residue on a silica gel column using ethyl acetate-petroleum ether (1:9) furnished the TBS ether

22 (2.3 g, 92%) as colorless oil, Rf = 0.40 (EtOAc-petroleum 60 ether 1:9). $[\alpha]_D^{25} = -57.9$ (c 11.8, CHCl₃); IR (neat): v_{max}/cm^{-1} 3446, 2931, 2858, 1718, 1611, 1511, 1367, 1250, 1170, 1094, 1050, 939, 837; ¹H NMR (200 MHz, CDCl₃): δ 7.36-7.18 (m, 7H), 6.85 (d, J 8.7 Hz, 2H), 5.22 (d, J 8.3 Hz, 1H), 4.94 (s, 3H), 4.43 (dd, J 8.3 and 4.2 Hz, 1H), 3.98 (q, J 14.2 and 7.1 Hz, 2H), 65 1.35 (s, 9H), 1.05 (t, J 7.1 Hz, 3H), 0.84 (s, 9H), 0.00 (s, 3H), -0.19 (s, 3H); ¹³C NMR (50 MHz, CDCl₃): δ 169.7, 158.1, 154.6, 136.8, 132.7, 128.3, 127.7, 127.2, 114.0, 79.3, 74.9, 69.7, 60.8, 60.6, 28.1, 25.5, 17.9, 13.8, -5.0, -5.5; HRMS (ESI): m/z calcd for C₂₉H₄₄NO₆ (M+H) 530.2938; found 530.2944.

(2R, 3R)-ethyl 2-(tert-butoxycarbonylamino)-3-(tertbutyldimethylsilyloxy)-3-(4-hydroxyphenyl)propanoate (24): To a solution of the ester 22 (2.0 g, 3.8 mmol) in methanol was added 5% Pd/C (40.0 mg) and shaken in hydrogenation 75 atmosphere at RT for 12 h. The reaction mixture was filtered on celite using ethyl acetate. Evaporation of the solvent and purification of the residue on a silica gel column using ethyl acetate-petroleum ether (1:4) as eluent furnished hydroxy ester 24 (1.6 g, 97%) as a colorless oil, Rf = 0.30 (EtOAc-petroleum ether ⁸⁰ 1:4). $[\alpha]_D^{25} = -50.3$ (*c* 9.7, CHCl₃); IR (neat): v_{max}/cm^{-1} 3434, 3020, 2859, 1694, 1616, 1516, 1369, 1169, 1084, 834; ¹H NMR (200 MHz, CDCl₃): δ 7.49 (br s, 1H), 7.11 (d, J 8.6 Hz, 2H), 6.73 (d, J 8.5 Hz, 2H), 5.26 (d, J 8.6 Hz, 1H), 4.91 (d, J 4.2 Hz, 1H), 4.44 (dd, J 8.5 and 4.4 Hz, 1H), 4.05 (q, J 14.2 and 7.0 Hz, 2H), 85 1.37 (s, 9H), 1.12 (t, J 7.0 Hz, 3H), 0.84 (s, 9H), 0.00 (s, 3H), -0.19 (s, 3H); ¹³C NMR (50 MHz, CDCl₃): δ 170.4, 156.0, 155.1, 131.3, 127.4, 114.9, 80.1, 75.1, 61.2, 60.3, 28.2, 25.5, 17.9, 13.8, -4.9, -5.5; HRMS (ESI): m/z calcd for C₂₂H₃₈NO₆Si (M+H) 440.2468; found 440.2475.

(2R,3R)-ethyl 2-(tert-butoxycarbonylamino)-3-(tertbutyldimethylsilyloxy)-3-(4-(2-

cyanoethoxy)phenyl)propanoate (26): To a solution of the phenol 24 (1.0 g, 2.3 mmol) in acrylonitrile (10.0 ml) was added 95 catalytic amount of triton B and refluxed for 8 h. Excess acrylonitrile was removed under reduced pressure and the reaction mixture was diluted with water, extracted with CH₂Cl₂, washed with brine and dried over Na₂SO₄. Evaporation of the solvent and purification of the residue on silica gel column using 100 ethyl acetate-petroleum ether (1:4) as eluent furnished the cyanoester 26 (1.0 g, 92%). Rf = 0.40 (EtOAc-petroleum ether, 1:4). $\left[\alpha\right]_{D}^{25} = -53.3$ (c 10.7, CHCl₃); IR (neat): v_{max}/cm^{-1} 3444, 3020, 2931, 2240, 1712, 1612, 1510, 1250, 1170, 1115, 1080, 1056, 930, 757, 669; ¹H NMR (200 MHz, CDCl₃): δ 7.23 (d, J 105 8.6 Hz, 2H), 6.80 (d, J 8.7 Hz, 2H), 5.20 (d, J 8.3 Hz, 2H), 4.95 (d, J 3.8 Hz, 1H), 4.40 (dd, J 8.3 and 4.0 Hz, 1H), 4.13 (t, J 6.3 Hz, 2H), 4.03 (q, J 14.4 and 7.3 Hz), 2.77 (t, J 6.3 Hz, 2H), 1.36 (s, 9H), 1.10 (t, J 7.2 Hz, 3H), 0.84 (s, 9H), 0.00 (s, 3H), -0.19 (s, 3H); ¹³C NMR (50 MHz, CDCl₃): δ 169.6, 157.0, 154.7, 133.9, 110 127.5, 117.1, 113.9, 79.6, 75.0, 62.5, 61.0, 60.7, 28.2, 25.5, 18.5, 18.5, 17.9, 13.9, -4.9, -5.4; HRMS (ESI): m/z calcd for C₂₅H₄₁N₂O₆Si (M+H) 493.2734; found 493.2735.

(1R,2S)-1-(tert-butyldimethylsilyloxy)-1-(4-(2tert-butyl 115 cyanoethoxy)phenyl)-3-hydroxypropan-2-ylcarbamate (26a): To a magnetically stirred solution of the cyanoester 26 (2.0 g, 4.1

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mmol) in ether/methanol (20:10 ml) was added LiBH₄ (0.45 g, 20.5 mmol) and stirred for 4 h at RT. Solvent was evaporated under reduced pressure, water (15.0 ml) was added to the residue and extracted with ether (3 x 20 ml). The combined extract was 5 washed with brine (15.0 ml) and dried over Na₂SO₄. Evaporation of the solvent and purification of the residue on a silica gel column using ethyl acetate-petroleum ether (1:2) as eluent furnished the alcohol 26a (1.5 g, 83%), Rf = 0.30 (EtOAcpetroleum ether, 1:2). $[\alpha]_D^{25} = -17.9$ (*c* 4.3, CHCl₃); IR (neat): 10 v_{max}/cm⁻¹ 3445, 2930, 2260, 1701, 1510, 1392, 1367, 1246, 1170, 1086, 868, 557; ¹H NMR (200 MHz, CDCl₃): δ 7.25 (d, J 8.5 Hz, 2H), 6.81 (d, J 8.6 Hz, 2H), 5.39 (d, J 7.8 Hz, 1H), 5.06 (br s, 1H), 4.12 (t, J 6.3 Hz, 2H), 3.75 (dd, J 11.5 and 2.5 Hz, 1H), 3.48-3.34 (m, 2H), 2.76 (t, J 6.3, Hz, 2H), 1.38 (s, 9H), 0.84 (s, 9H), 15 0.00 (s, 3H), -0.18 (s, 3H); ¹³C NMR (50 MHz, CDCl₃): δ 156.9, 155.5, 134.3, 127.1, 117.1, 114.3, 79.4, 76.8, 62.5, 60.9, 56.5, 28.3, 25.7, 18.5, 18.0, -5.0, -5.5; HRMS (ESI): m/z calcd for $C_{23}H_{39}N_2O_5Si$ (M+H) 451.2628; found 451.2628.

20 N-((1R,2R)-1-(4-(2-cyanoethoxy)phenyl)-3-hydroxy-1-((trim-

ethylsilyl) oxy) propan-2-yl)formamide (27a) : To a solution of boc-amine 26a (90.0 mg, 0.20 mmol) in CH₂Cl₂ (4.0 ml) was added TFA (0.19 ml, 2.00 mmol) at 0 °C and stirred for 8 h at RT. It was quenched with NaHCO₃ and concentrated on vacuo. 25 The residue was dissolved in ethyl formate (6.0 ml) and refluxed for 6 h. Evaporation of the solvent and purification of the residue on a silica gel column using MeOH-CH₂Cl₂ (1:19) furnished TBS-protected bursatellin 27a (50.0 mg, 70%, 2 steps) as colorless oil, Rf = 0.30 (MeOH-CH₂Cl₂ 1:19). $[\alpha]_D^{25} = +13.0$ (c ³⁰ 1.1, CHCl₃); IR (neat): vmax/cm⁻¹ 3357, 2928, 2856, 2280,1746, 1670, 1512, 1381, 1245, 1080, 1040, 837; ¹H NMR (200 MHz, CDCl₃, peaks due to major rotamer): δ 8.21 (s, 1H), 7.26 (d, J 8.6 Hz, 2H), 6.84 (d, J 8.7 Hz, 2H), 6.44 (d, J 7.6 Hz, 1H), 5.05 (d, J 2.7 Hz, 1H), 4.14 (t, J 6.3 Hz, 2H), 3.93-3.82 (m, 2H), 3.45-3.36 35 (m, 1H), 2.78 (t, J 6.3 Hz, 2H), 0.85 (s, 9H), 0.00 (s, 3H), -0.20 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 161.0, 157.3, 134.0, 128.0, 127.3, 117.1, 114.5, 62.6, 60.7, 54.6, 25.8, 18.6, 17.9, -4.9, -5.3; HRMS (ESI): m/z calcd for C₁₉H₃₀N₂O₄SiNa (M+Na) 401.1873; found 401.1867;

40 Synthesis of epi-bursatellin (11): To a magnetically stirred solution of the TBS protected bursatellin 27a (30.0 mg, 0.08 mmol) in THF (2.0 ml) was added 1 M solution of TBAF in THF (0.08 ml, 0.08 mmol) and stirred for 1 h at 0 °C. Solvent was 45 evaporated under reduced pressure, water (3.0 ml) was added to the residue and extracted with CH₂Cl₂ (3 x 10 ml). The combined extract was washed with brine (6.0 ml) and dried over Na₂SO₄. Evaporation of the solvent and purification of the residue on a silica gel column using MeOH-CH₂Cl₂ (1:9) as eluent furnished $_{50}$ the (+)-bursatellin 11 (16.8 mg, 80%), Rf = 0.40 (MeOH:CH₂Cl₂, 1:9). $\left[\alpha\right]_{D}^{25} = +7.1$ (c 0.4, MeOH); IR (neat): vmax/cm⁻¹ 3362, 2977, 2243, 1660, 1450, 1377, 1174, 1099; ¹H NMR (400 MHz, CDCl₃): δ 8.27 (s, 1H), 7.37 (d, J 8.3 Hz, 2H), 6.94 (d, J 8.5 Hz, 2H), 6.42 (br d, J 6.8 Hz, 1 H), 5.06 (d, J 3.3 Hz, 1H), 4.22 (t, J 55 6.5 Hz, 2H), 4.14 (dd, J 7.5 and 3.8 Hz,1H), 3.90 (dd, J 10.8 and 2.8 Hz, 1H), 3.66 (d, J 11.0 Hz, 1H), 3.06 (br s, 1H) 2.85 (t, J 6.3 Hz, 2H), 2.57 (br s, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 161.3, 18.7;

60 N-((1R,2R)-1-((tert-butyldimethylsilyl)oxy)-1-(4-(2-cyanoethoxy)phenyl)-3-hydroxypropan-2-yl)-2-(1H-indol-3-yl)-2-oxoacetamide (27b): To a solution of indole (1.0 g, 8.6 mmol) in anhydrous diethyl ether (15.0 ml) at 0 °C, freshly distilled oxalyl 65 chloride (0.8 ml, 9.5 mmol) was added dropwise. The reaction mixture was stirred at 0 °C for 0.5 h and then allowed to warm to RT. The resulting yellow crystals of indoleglyoxylic chloride 14 were collected by filtration, washed with cold anhydrous diethyl ether and dried under vacuum (1.6 g, 15.7 mmol). The compound 70 14 was used without further purification in next step. To a solution of boc-amine 26a (100 mg, 0.2 mmol) in CH₂Cl₂ (5.0 ml) was added TFA (0.15 ml, 2.0 mmol) and stirred for 2 h. aq. NaHCO₃ was added to the reaction mixture and extracted with CH₂Cl₂ (3 x 10 ml). The organic layer was washed with brine and 75 dried over Na₂SO₄. After evaporation of the solvent, the residue was dissolved in CH2Cl2, cooled to 0 °C. To the cold solution was added Et₃N (17.0 mg, 0.16 mmol) followed by 3-indoleglyoxalyl chloride 14 (40.0 mg, 0.14 mmol) and stirred magnetically at RT for 12 h. Water (10.0 ml) was added to the reaction mixture and ⁸⁰ extracted with CH₂Cl₂ (3 x 10 ml). Combined organic layers were washed with brine and dried over anhydrous Na₂SO₄. Evaporation of the solvent and purification of the residue on a silica gel column using MeOH:CH₂Cl₂ (1:19) furnished the TBSprotected preoxazinin **27b** (73 mg, 70%). $[\alpha]_D^{25} = +24.3$ (c 1.4, ⁸⁵ CHCl₃); IR (neat): vmax/cm⁻¹ 3436, 3010, 2930, 2400, 2280, 1611, 1511, 1438, 1214, 1124, 1045, 929, 756, 618; ¹H NMR (200 MHz, CDCl₃): § 9.27 (s, 1H), 8.88 (d, J 3.3 Hz, 1H), 8.41-8.45 (m, 1H), 8.13 (d, J 8.6 Hz, 1H), 7.45-7.29 (m, 5H), 6.86 (d, J 8.7 Hz, 2H), 5.05 (d, J 4.0 Hz, 1H), 4.16 (t, J 6.3 Hz, 2H), 4.07-90 3.94 (m, 2H), 3.71-3.60 (m, 1H), 3.04 (d, J 7.2 Hz, 1H), 2.82 (t, J 6.2 Hz, 2H), 1.75 (s, 1H), 0.92 (s, 9H), 0.07 (s, 3H), -0.15 (s, 3H); ¹³C NMR (50 MHz, CDCl₃): δ 180.4, 162.5, 157.2, 138.1, 135.7, 134.2, 127.5, 126.5, 124.2, 123.3, 122.4, 117.4, 144.4, 113.2, 111.7, 75.8, 62.5, 61.0, 56.2, 25.7, 18.6, 18.0, -4.9, -5.3; HRMS 95 (ESI): m/z calcd for C₂₈H₃₆N₃O₅Si (M+H) 522.2424; found 522.2424.

Synthesis of epi-preoxazinin-7 (10): To the solution of 27b (60.0 mg, 0.15 mmol) in THF (4.0 ml) was added TBAF (1.0 M 100 in THF, 0.15 ml, 0.15 mmol). The resulting mixture was stirred for 1 h at 0 °C, before it was concentrated in vacuo. Water (5.0 ml) was added to the residue and extracted with ether (3 x 20 ml). The combined extract was washed with brine (13.0 ml) and dried over Na₂SO₄. Evaporation of the solvent and purification of the ¹⁰⁵ residue on a silica gel column using ethyl acetate-petroleum ether (9:1) as eluent furnished the epi-preoxazinin-7 10 (27.0 mg, 80 %). Rf = 0.30 (EtOAc-petroleum ether, 9:1). $[\alpha]_D^{25} = +40.4$ (c 0.25, MeOH); IR (neat): vmax/cm⁻¹ 3434, 3340, 3162, 2977, 2270, 1662, 1622, 1450, 1377, 1113, 1089, 929, 757; ¹H NMR 110 (400 MHz, CD₃CN): δ 10.19 (br s, 1H), 8.78 (d, J 3.3 Hz, 1H), 8.27 (dd, J 5.8 and 2.0 Hz, 1H), 7.71 (d, J 9.5 Hz, 1H), 7.54 (dd, J 7.5 and 4.0 Hz, 1H), 7.36-7.25 (m, 4H), 6.87 (dt, J 8.7 and 2.0 Hz, 2H), 4.82 (dd, J 6.4 and 5.0 Hz, 1H), 4.19-4.07 (m, 3H), 3.92 (d, J 4.8 Hz, 1H), 3.84-3.60 (m, 2H), 3.23 (t, J 5.8 Hz, 1H), 2.81 ¹¹⁵ (t, J 6.1 Hz, 2H); ¹³C NMR (100 MHz, CD₃CN): δ 182.4, 163.7, 158.7, 140.1, 137.5, 136.3, 129.3, 127.8, 125.1, 124.2, 122.9,

157.5, 133.9, 127.2, 117.1, 114.8, 75.4, 62.7, 61.4, 54.2, 29.7,

119.6, 115.5, 113.6, 74.3, 64.3, 62.2, 57.5, 19.5; HRMS (ESI): *m/z* calcd for C₂₂H₂₁N₃O₅Na (M+Na) 430.1379; found 430.1370.

(2S,3R)-ethyl 3-(4-(benzyloxy)phenyl)-2-((tert-butoxycarbony-5 I)amino)-3-hydroxypropanoate (21): A round bottom flask was charged with sodium hydroxide (30.5 ml of 1 N solution) and diluted with water (45.0 ml) in a dark fume hood. Part of this alkaline solution (5.0 ml) was transferred into a vial to dissolve K₂[OsO₂(OH)₄] (147 mg, 0.4 mmol). With vigorous stirring, n-10 propanol (40.0 ml) and tert-butylcarbamate (3.6 g, 31.0 mmol) were added to the flask, followed by dropwise addition of freshly prepared tert-butyl hypochlorite (3.5 ml, 3.05 mmol). After five minutes, n-propanol solution (35.0 ml) of (DHQD)₂AQN (343 mg, 0.4 mmol) and ethyl cinnamate 15 (2.82 g, 1.0 mmol), and 15 the aqueous $K_2[OsO_2(OH)_4]$ solution were added. After 2 h, the solution was quenched with 5.0 g sodium bisulfite, added water and extracted with ethyl acetate (3 x 50 ml), washed with brine and dried over Na2SO4. Evaporation of the solvent and purification of the residue on a flash silica gel column using ethyl 20 acetate-petroleum ether (1:4) furnished the hydroxy ester 21 (2.14 g, 45%, 98% ee), Rf = 0.30 (EtOAc-petroleum ether 1:4). $[\alpha]_{D}^{25} =$ -12.8 (c 4.0, CHCl₃); IR (neat): vmax/cm⁻¹ 3421, 2930, 1716, 1690, 1512, 1392, 1244, 1162, 1025; ¹H NMR (200 MHz, CDCl₃): δ 7.50-7.20 (m, 7H), 6.94 (td, J 8.7 and 2.0 Hz, 2H), 25 5.35 (d, J 8.8 Hz, 1H), 5.12 (t, J 3.4 Hz, 1H), 5.04 (s, 2H), 4.45 (br d, J 7.4 Hz, 1H), 4.18 (q, J 14.3 and 7.2 Hz, 2H), 2.9 (br s, 1H), 1.35 (s, 9H), 1.22 (t, J 7.1 Hz, 3H); ¹³C NMR (50 MHz, CDCl₃): δ 171.0, 158.6, 155.8, 136.9, 132.4, 128.6, 128.0, 127.5, 127.4, 114.7, 80.1, 73.7, 70.0, 61.7, 59.7, 28.2, 14.1; HRMS $_{30}$ (ESI): m/z calcd for $C_{23}H_{30}NO_6$ (M+H) 416.2073; found 416.2086.

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(2*S*,3*R*)-ethyl 3-(4-(benzyloxy)phenyl)-2-((*tert*-butoxycarbonyl)amino)-3-((*tert*-butyldimethylsilyl)oxy)propanoate (23):

- To a solution of the alcohol **21** (4.0 g, 9.6 mmol) in DMF (25.0 ml) were added TBSCl (1.9 g, 12.5 mmol), imidazole (0.9 g, 14.4 mmol) and stirred magnetically at RT for 12 h. Water was added to the reaction mixture, extracted with ethyl acetate (3 x 20 ml), washed with brine and dried over Na₂SO₄. Evaporation of the ⁴⁰ solvent and purification of the residue on a silica gel column using ethyl acetate-petroleum ether (1:9) furnished the TBS ether **23** (4.9 g, 98%) as colorless oil, Rf = 0.40 (EtOAc-petroleum ether 1:9). $[\alpha]_D^{25} = -16.5$ (*c* 5.4, CHCl₃); IR (neat): v_{max}/cm^{-1} 2930, 1720, 1643, 1510, 1367, 1250, 830, 778; ¹H NMR (200 ⁴⁵ MHz, CDCl₃): δ 7.50-7.20 (m, 7H), 6.94 (dt, *J* 8.7 and 2.0 Hz, 2H), 5.30-5.10 (m, 2H), 5.05 (s, 2H), 4.38 (dd, *J* 7.5 and 2.5 Hz,
- 2H), 5.30-5.10 (m, 2H), 5.05 (s, 2H), 4.38 (dd, J /.5 and 2.5 Hz, 1H), 4.30-4.10 (m, 2H), 1.35 (s, 9H), 1.31 (t, J 7.1 Hz, 3H), 0.89 (s, 9H), 0.00 (s, 3H), -0.15 (s, 3H); ¹³C NMR (50 MHz, CDCl₃): δ 170.0, 158.3, 154.9, 137.1, 133.0, 128.6, 127.9, 127.5, 114.3, 50 79.6, 75.2, 69.9, 61.0, 60.9, 53.5, 28.4, 25.8, 18.2, 14.1, -4.8, -5.3; UDN(C) (50)
- HRMS (ESI): m/z calcd for C₂₉H₄₄NO₆Si (M+H) 530.2938; found 530.2941.
- (2*S*,3*R*)-ethyl 2-(*tert*-butoxycarbonylamino)-3-(*tert*butyldimethylsilyloxy)-3-(4-hydroxyphenyl)propanoate (25): To a solution of the ester 23 (2.0 g, 3.8 mmol) in methanol was added 5% Pd/C (40.0 mg) and shaken in hydrogenation atmosphere at RT for 12 h. The reaction mixture was filtered on celite using ethyl acetate. Evaporation of the solvent and

⁶⁰ purification of the residue on a silica gel column using ethyl acetate-petroleum ether (1:4) as eluent furnished hydroxy ester 25 (1.6 g, 97%) as a colorless oil, *Rf* = 0.30 (EtOAc-petroleum ether 1:4). [α]_D²⁵ = -17.8 (*c* 2.3, CHCl₃); IR (neat): v_{max}/cm⁻¹ 3434, 3020, 2859, 1694, 1616, 1516, 1369, 1169, 1084, 834; ¹H NMR
⁶⁵ (400 MHz, CDCl₃): δ 7.22 (d, *J* 8.5 Hz, 2H), 6.97 (s, 1H), 6.79 (d, *J* 8.5 Hz, 2H), 5.50 (br d, *J* 9.3 Hz, 1H), 5.22 (s, 1H), 4.30-4.05 (m, 3H), 1.36 (s, 9H), 1.27 (t, *J* 7.0 Hz, 3H), 0.89 (s, 9H), 0.01 (s, 3H), -0.15 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 172, 157.9, 156.9, 133.7, 129.1, 116.0, 80.4, 75.6, 62.6, 62.3, 28.9, 70 26.4, 19.0, 14.8, -4.0, -4.8; HRMS (ESI): *m/z* calcd for C₂₂H₃₈NO₆Si (M+H) 440.2468; found 440.2467.

(2*S*,3*R*)-ethyl 2-(*tert*-butoxycarbonylamino)-3-(*tert*-butyldimethylsilyloxy)-3-(4-(2-

- ⁷⁵ **cyanoethoxy)phenyl)propanoate (13)** : To a solution of the phenol **25** (1.0 g, 2.3 mmol) in acrylonitrile (10.0 ml) was added catalytic amount of triton B and refluxed for 8 h. Excess acrylonitrile was removed under reduced pressure and the reaction mixture was diluted with water, extracted with CH₂Cl₂,
- ⁸⁰ washed with brine and dried over Na₂SO₄. Evaporation of the solvent and purification of the residue on silica gel column using ethyl acetate-petroleum ether (1:4) as eluent furnished the ester **13** (1.0 g, 92%). *Rf* = 0.40 (EtOAc-petroleum ether, 1:4). $[\alpha]_D^{25} = -15.6$ (*c* 2.0, CHCl₃); IR (neat): v_{max}/cm⁻¹ 3447, 3020, 2931, 2240,
- ⁸⁵ 1712, 1612, 1510, 1215, 1171, 1086, 757, 669; ¹H NMR (200 MHz, CDCl₃): δ 7.30 (d, *J* 8.3 Hz, 2H), 6.87 (d, *J* 8.4 Hz, 2H), 5.35-5.00 (m, 2H), 4.40-4.05 (m, 5H), 2.82 (t, *J* 6.3 Hz, 2H), 1.35 (s, 9H), 1.32 (t, *J* 7.2 Hz, 3H), 0.89 (s, 9H), 0.00 (s, 3H), -0.15 (s, 3H); ¹³C NMR (50 MHz, CDCl₃): δ 170.5, 157.1., 155.3, 133.9, 90 127.5, 117.1, 114.1, 79.5, 74.2, 62.5, 61.3, 60.5, 28.1, 25.5, 18.4,
- 17.9, 13.9, -4.8, -5.7; HRMS (ESI): m/z calcd for $C_{25}H_{41}N_2O_6Si$ (M+H) 493.2734; found 493.2727.
- tert-butyl (1R,2R)-1-(tert-butyldimethylsilyloxy)-1-(4-(2-95 cyanoethoxy)phenyl)-3-hydroxypropan-2-ylcarbamate (28): To a magnetically stirred solution of the cyano ester 13 (2.0 g, 4.1 mmol) in ether/methanol (20:10 ml) was added LiBH₄ (0.5 g, 20.5 mmol) and stirred for 4 h at RT. Solvent was evaporated under reduced pressure, water (15.0 ml) was added to the residue 100 and extracted with ether (3 x 20 ml). The combined extract was washed with brine (15.0 ml) and dried over Na₂SO₄. Evaporation of the solvent and purification of the residue on a silica gel column using ethyl acetate-petroleum ether (1:2) as eluent furnished the alcohol 28 (1.5 g, 83%), Rf = 0.30 (EtOAc-¹⁰⁵ petroleum ether, 1:2). $[\alpha]_D^{25} = -22.2$ (*c* 4.2, CHCl₃); IR (neat): v_{max}/cm⁻¹ 3442, 3020, 2931, 2255, 1700, 1510, 1392, 1367, 1216, 1172, 1086, 838; ¹H NMR (200 MHz, CDCl₃): δ 7.25 (d, J 8.6 Hz, 2H), 6.86 (d, J 8.6 Hz, 2H), 5.0-4.8 (m, 2H), 4.18 (t, J 6.3 Hz, 2H), 3.75-3.50 (m, 3H), 2.83 (t, J 6.3, Hz, 2H), 1.37 (s, 9H), 0.90 110 (s, 9H), 0.06 (s, 3H), -0.16 (s, 3H); ¹³C NMR (50 MHz, CDCl₃): δ 156.9, 156.1, 135.0, 127.6, 117.2, 114.2, 79.5, 72.9, 62.7, 62.5, 58.5, 28.2, 25.7, 18.5, 18.0, -4.7, -5.3; HRMS (ESI): m/z calcd for C₂₃H₃₉N₂O₅Si (M+H) 451.2628; found 451.2620.

¹¹⁵ N-((1R,2R)-1-(tert-butyldimethylsilyloxy)-1-(4-(2cyanoethoxy)phenyl)-3-hydroxypropan-2-yl)formamide (28a): To a solution of boc-amine 28 (80.0 mg, 0.2 mmol) in CH₂Cl₂

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110

(8.0 ml) was added Yb(OTf)₃ (110.0 mg, 0.2 mmol) and refluxed for 8 h. It was guenched with NaHCO₃ and concentrated on vacuo. The residue was dissolved in ethyl formate (6.0 ml) and refluxed for 6 h. Evaporation of the solvent and purification of 5 the residue on a silica gel column using MeOH-CH₂Cl₂ (1:19) furnished TBS-protected bursatellin 28a (47.6 mg, 71%, 2 steps) as colorless oil, Rf = 0.30 (MeOH-CH₂Cl₂1:19). $[\alpha]_D^{25} = -15.1$ (c 4.4, CHCl₃); IR (neat): v_{max}/cm⁻¹ 3358, 2928, 2856, 2280, 1746, 1672, 1512, 1389, 1245, 1080, 1047, 837; ¹H NMR (500 MHz, 10 CDCl₃, peaks due to major rotamer): δ 8.14 (s, 1H), 7.25 (d, J 15.0 Hz, 2H), 6.86 (d, J 8.8 Hz, 1H), 6.10 (d, J 7.3 Hz, 1H), 4.97 (d, J 3.3 Hz, 1H), 4.19 (t, J 6.5 Hz, 2H), 4.07-4.01 (m, 1H), 3.68 (d, J 5.8 Hz, 2H), 2.84 (t, J 6.0 Hz, 2H), 0.91 (s, 9H), 0.08 (s, 3H), -0.14 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 165.0, 161.6, 15 157.3, 134.7, 127.8, 127.4, 117.2, 114.5, 114.5, 73.5, 72.3, 63.0, 62.6, 60.8, 57.2, 25.8, 25.8, 18.7, 18.2, -4.6, -5.2; HRMS (ESI): m/z calcd for $C_{19}H_{30}N_2O_4SiNa$ (M+Na) 401.1873; found 401.1869.

- 20 Synthesis of (-)-bursatellin (9): To a magnetically stirred solution of the TBS protected bursatellin 28a (20.0 mg, 0.05 mmol) in THF (2.0 ml) was added 1 M solution of TBAF in THF (0.05 ml, 0.05 mmol) and stirred for 1 h at 0 °C. Solvent was evaporated under reduced pressure, water (3.0 ml) was added to 25 the residue and extracted with CH₂Cl₂ (3 x 8 ml). The combined extract was washed with brine (6.0 ml) and dried over Na₂SO₄. Evaporation of the solvent and purification of the residue on a silica gel column using MeOH-CH₂Cl₂ (1:9) as eluent furnished the (-)-bursatellin 9 (11.2 mg, 80%), Rf = 0.40 (MeOH:CH₂Cl₂ ³⁰ 1:9). $[\alpha]_{\rm D}^{25} = -8.6$ (*c* 0.6, MeOH); IR (neat): $v_{\rm max}/{\rm cm}^{-1}$ 3360, 2977, 2240, 1660, 1450, 1377, 1174, 1089; ¹H NMR (400 MHz, CD₃OD): δ 7.90 (s, 1H), 7.23 (d, J 8.5 Hz, 2H), 6.82 (d, J 8.5 Hz, 2H), 4.80 (d, J 3.6 Hz, 1H), 4.07 (t, J 6.0 Hz, 2H), 4.01-3.96 (m, 1H), 3.54 (dd, J 10.8 and 6.2 Hz, 1H), 3.38 (dd, J 10.8 and 6.0 ³⁵ Hz, 1H), 2.80 (t, J 6.0 Hz, 2H); ¹³C NMR (100 MHz, CD₃OD): δ 164.0, 159.0, 136.7, 128.6, 119.0, 115.4, 71.9, 64.2, 62.5, 57.1, 19.0;
- 40 N-((1R,2R)-1-((tert-butyldimethylsilyl)oxy)-1-(4-(2-cyanoethoxy)phenyl)-3-hydroxypro-pan-2-yl)-2-(1H-indol-3-yl)-2-oxoacetamide (30): To a solution of boc-amine 28 (100 mg, 0.2 mmol) in CH₂Cl₂ (10.0 ml) was added Yb(OTf)₃ (138 mg, 0.2 mmol) and refluxed for 8 h. Aq. NaHCO₃ was added to the 45 reaction mixture and extracted with CH₂Cl₂ (3 x 10 ml). The organic layer was washed with brine and dried over Na_2SO_4 . After evaporation of the solvent, the residue was dissolved in CH₂Cl₂, cooled to 0 °C. To the cold solution was added Et₃N (17.0 mg, 0.16 mmol) followed by 3-indoleglyoxalyl chloride 14 50 (40.0 mg, 0.14 mmol) and stirred magnetically at RT for 12 h. Water (10.0 ml) was added to the reaction mixture and extracted with CH₂Cl₂ (3 x 10 ml). Combined organic layers were washed
- with brine and dried over anhydrous Na₂SO₄. Evaporation of the solvent and purification of the residue on a silica gel column 55 using MeOH:CH₂Cl₂ (1:19) furnished the TBS-protected
- preoxazinin-7 **30** (73.0 mg, 70%). $[\alpha]_D^{25} = -24.0$ (*c* 0.9, CHCl₃); IR (neat): v_{max}/cm^{-1} 3392, 3020, 2930, 2400, 2280, 1635, 1511, 1423, 1215, 1045, 929, 758, 669; ¹H NMR (400 MHz, CDCl₃): δ

9.58 (s, 1H), 8.71 (dd, J 5.1 and 3.4 Hz, 1H), 8.39 (d, J 7.5 Hz, 60 1H), 7.92 (d, J 9.0 Hz, 1H), 7.40-7.20 (m, 5H), 6.73 (d, J 8.7 Hz, 2H), 5.00 (d, J 3.1 Hz, 1H), 4.15-4.05 (m, 1H), 4.02 (t, J 6.2 Hz, 2H), 3.85-3.70 (m, 2H), 2.72 (t, J 6.2 Hz, 2H), 0.92 (s, 9H), 0.07 (s, 3H), -0.15 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 180.3, 163.0, 157.0, 138.5, 135.8, 134.7, 127.4, 126.5, 124.1, 123.3, 65 122.2, 117.4, 114.2, 112.9, 111.9, 72.73, 62.73, 62.36, 58.1, 25.8, 18.6, 18.1, -4.61, -5.25; HRMS (ESI): m/z calcd for C₂₈H₃₆N₃O₅S₁ (M+H) 522.2424; found 522.2424.

Synthesis of preoxazinin-7 (7): To the solution of 30 (80.0 mg, 70 0.2 mmol) in THF (5.0 ml) was added TBAF (1.0 M in THF, 0.2 ml, 0.2 mmol). The resulting mixture was stirred for 1 h at 0 °C, before it was concentrated in vacuo. Water (5.0 ml) was added to the residue and extracted with ether (3 x 20 ml). The combined extract was washed with brine (15.0 ml) and dried over Na₂SO₄. 75 Evaporation of the solvent and purification of the residue on a silica gel column using ethyl acetate-petroleum ether (9:1) as eluent furnished the preoxazinin-7 7 (48.7 mg, 78 %). Rf = 0.30(EtOAc-petroleum ether, 9:1). $[\alpha]_D^{25} = -11.0$ (*c* 0.6, MeOH); IR (neat): v_{max}/cm⁻¹ 3480, 3340, 3162, 2977, 2270, 1660, 1622, 1450, ⁸⁰ 1377, 1174, 1089, 929; ¹H NMR (400 MHz, CD₃CN): δ 10.12 (br s, 1H), 8.81 (s, 1H), 8.31 (d, J 9.1 Hz, 1H), 7.79 (br s, 1H), 7.54 (d, J 9.1 Hz, 1H), 7.30 (dd, J 9.1 and 7.3 Hz, 2H), 7.29 (dd, J 9.1 and 7.3 Hz, 2H), 6.88 (d, J 8.9 Hz, 2H), 5.00 (t, J 3.6 Hz, 1H), 4.14 (t, J 6.2 Hz, 2H), 4.10-4.00 (m, 1H), 3.86 (d, J 4.0 Hz, 85 1H), 3.70 (dd, J 11.4 and 5.6 Hz, 1H), 3.63 (dd, J 11.4 and 5.6 Hz, 2H), 2.81 (t, J 6.2 Hz, 2H); ¹³C NMR (100 MHz, CD₃CN): δ 181.8, 163.6, 158.4, 139.9, 137.2, 136.5, 128.5, 127.6, 124.9, 124.0, 122.7, 119.3, 115.3, 113.5, 113.3, 72.5, 64.0, 63.1, 57.8, 19.2, HRMS (ESI): m/z calcd for $C_{22}H_{21}N_3O_5Na$ (M+Na) 90 430.1379; found 430.1382.

3-(4-((S)-((3S,6R)-6-(1H-indol-3-yl)-5-oxomorpholin-3-yl)(tertbutyldimethylsilyloxy)methyl)phenoxy)propanenitrile (32a) & 3-(4-((S)-((3S,6S)-6-(1H-indol-3-yl)-5-oxomorpholin-3-yl)(tert-95 butyldimethylsilyloxy)methyl)phenoxy)propanenitrile (32b): To a cold solution (0 °C) of 30 (104.0 mg, 0.2 mmol) in MeOH was added NaBH₄ (40.0 mg, 1 mmol) and stirred vigorously at the same temperature for 30 min. Excess MeOH was removed from reaction mixture under reduced pressure. Added water (10.0 100 ml), extracted with ethyl acetate (3 x 8 ml), washed with brine (10.0 ml) and dried over Na₂SO₄ and concentrated. To the crude alcohol was added acetonitrile (50.0 ml), catalytic (10.0 mg) pyridine para-toluene sulfonate (PPTS) and refluxed for 1 h. The reaction mixture was quenched with aq. NaHCO₃ (20.0 ml), 105 extracted with ethyl acetate (3 x 8 ml), washed with brine and dried over Na₂SO₄. Evaporation of the solvent and purification of the residue using preparative thin layer chromatography furnished the diastereomers 32a (23.0 mg) and 32b (58.0 mg) (yield 78% for two steps).

TBS-protected oxazinin-5 (32a): $[\alpha]_D^{25} = +19.1$ (*c* 2.0 CHCl₃); IR (neat): v_{max}/cm^{-1} 3396, 3269, 2928, 2856, 1668(C=O), 1612, 1512, 1462, 1238, 1105, 858, 839, 780, 744, 618; ¹H NMR (CDCl₃, 400 MHz): δ 8.35 (br s, 1H), 7.68 (d, J 7.8 Hz, 1H), 115 7.35-7.00 (m, 6H), 6.89 (d, J 8.6 Hz, 1H), 6.55 (br s, 1H), 5.51 (s, 1H), 4.63 (d, J 8.3 Hz, 1H), 4.18 (t, J 6.3 Hz, 2H), 3.70-3.55 (m, 2H), 3.41 (dd, J 11.5 and 5.0 Hz, 1H), 2.83 (t, J 6.3 Hz, 2H), 1.64 (br s, 1H), 0.91 (s, 9H), 0.10 (s, 3H), -0.19 (s, 3H); 13 C NMR (CDCl₃, 100 MHz): δ 169.4, 157.8, 136.4, 133.7, 128.2, 126.1, 124.6, 122.4, 120.0, 119.5, 117.1, 114.7, 111.3, 76.2, 73.8, 62.6, s 61.5, 58.6, 25.8, 18.6, 18.1, -4.5, -5.0; HRMS (ESI): *m/z* calcd for C₂₈H₃₆N₃O₄Si (M+H) 506.2475; found 506.2486.

Synthesis of TBS-protected oxazinin-6 (32b): $[α]_D^{25} = -28.4$ (*c* 4.5, CHCl₃); IR (neat): v_{max} /cm⁻¹ 3398, 2927, 2856, 1668 (C=O), 10 1611, 1511, 1459, 1248, 1097, 859, 838, 779, 743, 617; ¹H NMR (CDCl₃, 500 MHz): δ 8.65 (br s, 1H), 7.71 (d, *J* 7.6 Hz, 1H), 7.35-7.00 (m, 6H), 6.81 (d, *J* 8.6 Hz, 2H), 6.62 (s, 1H), 5.51 (s, 1H), 4.61 (d, *J* 8.5 Hz, 1H), 4.12 (t, *J* 6.1 Hz, 2H), 3.70-3.50 (m, 2H), 3.50-3.40 (m, 1H), 2.79 (t, *J* 6.1 Hz, 2H), 0.90 (s, 9H), 0.09 15 (s, 3H), -0.21 (s, 3H). ¹³C NMR (CDCl₃, 125 MHz): δ 169.7, 157.7, 136.5, 133.5, 128.2, 126.2, 124.9, 122.3, 119.9, 119.4, 117.1, 114.6, 111.5, 110.9, 76.1, 73.8, 62.5, 61.1, 58.6, 25.8, 18.6, 18.1, -4.5, -5.0; HRMS (ESI): *m/z* calcd for C₂₈H₃₆N₃O₄Si(M+H) 506.2475; found 506.2463.

Oxazinin-5 (5) and (2*R*,5*S*)-5-((*S*)-hydroxy(4-hydroxyphenyl)methyl)-2-(1H-indol-3-yl)morpholin-3-one

(33a): To an ice cold (0 °C) solution of the compound 32a (22.0 mg, 0.043 mmol) in anhydrous THF (2.0 ml) was added dropwise 25 1 M solution of TBAF in THF (0.08 ml, 0.08 mmol) and stirred magnetically for 1 h. The reaction mixture was quenched with aq. NaHCO₃ (5.0 ml), extracted with ethyl acetate (5 ml x 3), washed with brine and dried over Na2SO4. Evaporation of the solvent and purification of the residue using preparative TLC furnished 30 oxazinin-5 5 (11.0 mg, 65%) and the diol 33a (2.5 mg, 15%). $[\alpha]_D^{25} = +6.9$ (c 0.4, MeOH); IR (neat): v_{max}/cm^{-1} 3360, 3302, 2924, 2853, 2257, 1660, 1609, 1513, 1456, 1306, 1240, 1177, 1106, 1049, 838, 749, 618; ¹H NMR (CDCl₃, 400 MHz); δ 9,29 (br s, 1H), 7.58 (d, J 8.1 Hz, 1H), 7.40 (d, J 8.3 Hz, 1H), 7.34 (d, 35 J 8.5 Hz, 2H), 7.27 (d, J 2.7 Hz, 1H), 7.15 (dd, J 8.1 and 7.1 Hz, 1H), 7.05 (dd, J 8.1 and 7.1 Hz, 1H), 6.96 (d, J 8.8 Hz, 1H), 5.29 (s, 1H), 4.58 (dd, J 8.1 and 4.2 Hz, 1H), 4.19 (t, J 6.1 Hz, 2H), 3.86 (d, J 4.1 Hz, 1H), 3.85-3.70 (m, 1H), 3.59 (dd, J 12.0 and 3.9 Hz, 1H), 3.50 (dd, J 12.0 and 7.6 Hz, 1H), 2.85 (t, J 6.1 Hz, ⁴⁰ 2H); ¹³C NMR (CDCl₃, 100 MHz): δ 169.2, 158.0, 136.5, 134.1, 128.1, 126.1, 125.3, 121.8, 119.4, 118.2, 114.6, 112.0, 111.5, 73.9, 73.6, 63.1, 63.0, 57.5, 18.1; HRMS (ESI): m/z calcd for C₂₂H₂₁N₃O₄Na (M+Na) 414.1430; found 414.1418.

45 (2R,5S)-5-((S)-hydroxy(4-hydroxyphenyl)methyl)-2-(1H-

- indol-3-yl)morpholin-3-one (33a): $[a]_D^{25} = +5.3$ (c 0.6, MeOH); IR (neat): v_{max} /cm⁻¹ 3362, 3169, 2923, 2853, 1653, 1612, 1463, 1186, 1108, 967, 838, 745, 618; ¹H NMR (CDCl₃, 400 MHz): δ 9.30 (br s, 1H), 7.58 (d, *J* 8.0 Hz, 1H), 7.39 (d, *J* 8.0 Hz, 1H),
- ⁵⁰ 7.25 (s, 1H), 7.20 (d, *J* 7.3 Hz, 2H), 7.18-7.00 (m, 3H), 6.85-6.75 (m, 2H), 5.28 (s, 1H), 4.49 (dd, *J* 8.1 and 3.9 Hz, 1H), 3.85-3.75 (m, 2H), 3.56 (dd, *J* 12.0 and 3.9 Hz, 1H), 3.47 (dd, *J* 11.7 and 7.8 Hz, 1H); ¹³C NMR (CDCl₃, 100 MHz): δ 169.3, 156.9, 136.4, 132.2, 128.0 (2C), 126.2, 125.3, 121.8, 119.4 (2C), 115.2 (2C), ⁵⁵ 111.9, 74.1, 73.6, 63.2, 57.6; HRMS (ESI): *m/z* calcd for C₁₉H₁₈N₂O₄Na (M+Na) 361.1164; found 361.1181.

Oxazinin-6 (6) and (2*S*,5*S*)-5-((*S*)-hydroxy(4-

hydroxyphenyl)methyl)-2-(1H-indol-3-yl)morpholin-3-one

60 (33b): To an ice cold (0 °C) solution of the compound 32b (20.0 mg, 0.04 mmol) in anhydrous THF (2.0 ml) was added dropwise 1 M solution of TBAF in THF (0.08 ml, 0.08 mmol) and stirred magnetically for 1 h. The reaction mixture was quenched with aq. NaHCO₃ (5.0 ml), extracted with ethyl acetate (5 ml x 3), washed 65 with brine and dried over Na₂SO₄. Evaporation of the solvent and purification of the residue using preparative TLC furnished oxazinin-6 6 (9.3 mg, 60%) and the diol 33b (3.7 mg, 24%). $\left[\alpha\right]_{D}^{25} = +9.0$ (c 0.5, MeOH); IR (neat): v_{max}/cm^{-1} 3360 (OH), 3163, 2924, 2857, 2256 (CN), 1661 (C=O), 1618, 1513, 1456, ⁷⁰ 1306, 1241, 1177, 1107, 1051, 838, 749, 618; ¹H NMR (CD₃CN, 400 MHz): δ 9.37 (br s, 1H), 7.62 (d, J 7.8 Hz, 1H), 7.43 (d, J 8.0 Hz, 1H), 7.35-7.26 (m, 2H), 7.25-7.20 (m, 1H), 7.18-7.14 (m, 1H), 7.09-7.05 (m, 1H), 6.92 (d, J 8.3 Hz, 2H), 6.86 (br s, 1H), 5.38 (s, 1H), 4.71 (dd, J 7.8 and 3.9 Hz, 1H), 4.17 (t, J 6.1 Hz, 75 2H), 3.91 (d, J 3.9 Hz, 1H), 3.68-3.52 (m, 3H), 2.84 (t, J 6.1 Hz, 2H); ¹³C NMR (CD₃CN, 100 MHz): δ 170.1, 158.8, 137.3, 135.4, 129.3, 127.3, 126.3, 122.7, 120.6, 120.2, 119.2, 115.6, 112.7, 112.5, 75.3, 74.5, 64.2, 62.7, 58.2, 19.0.

80 (2S,5S)-5-((S)-hydroxy(4-hydroxyphenyl)methyl)-2-(1H-

indol-3-yl)morpholin-3-one (33b): $[\alpha]_D^{25} = +7.0$ (*c* 0.6, MeOH); IR (neat): v_{max}/cm^{-1} 3360 (OH), 2923, 2853, 1652 (C=O), 1613, 1457, 1182, 1108, 966, 838, 745, 618; ¹H NMR (CDCl₃, 400 MHz): δ 9.38 (br s, 1H), 7.63 (d, *J* 7.6 Hz, 1H), 7.43 (d, *J* 8.1 Hz, 85 1H), 7.30-7.00 (m, 6H), 6.87 (br s, 1H), 6.85-9.70 (m, 2H), 5.38 (s, 1H), 4.70-4.60 (m, 1H),3.83 (br s, 1H), 3.65-3.50 (m, 3H).¹³C NMR (CDCl₃, 100 MHz): δ 170.2, 157.8, 137.5, 133.4, 129.2, 127.5, 126.4, 122.9, 120.5, 120.4, 116.2, 112.7, 112.6, 75.6, 74.5, 62.8, 58.6; HRMS (ESI): *m/z* calcd for C₁₉H₁₈N₂O₄Na (M+Na) 90 361.1164; found 361.1158.

(2S,3R)-ethyl 2-(2-(1H-indol-3-yl)-2-oxoacetamido)-3-(tertbutyldimethylsilyloxy)-3-(4-(2-cyanoethoxy)phenyl)propaneate (34): To a solution of boc-amine 13 (100 mg, 0.2 mmol) in 95 CH₂Cl₂ (10.0 ml) was added Yb(OTf)₃ (138 mg, 0.2 mmol) and refluxed for 8 h. aq. NaHCO3 was added to the reaction mixture and extracted with CH₂Cl₂ (3 x 10 ml). The organic layer was washed with brine and dried over Na₂SO₄. After evaporation of the solvent, the residue was dissolved in CH₂Cl₂, cooled to 0 °C. 100 To the cold solution was added Et₃N (17.0 mg, 0.16 mmol) followed by 3-indoleglyoxalyl chloride 14 (40.0 mg, 0.14 mmol) and stirred magnetically at RT for 12 h. Water (10.0 ml) was added to the reaction mixture and extracted with CH₂Cl₂ (3 x 10 ml). Combined organic layers were washed with brine and dried 105 over anhydrous Na₂SO₄. Evaporation of the solvent and purification of the residue on a silica gel column using (EtOAcpetroleum ether, 1:1) furnished the TBS-protected compound 34 (114 mg, 80%). $[\alpha]_D^{25} = -50.0$ (*c* 1.5, CHCl₃); IR (neat): v_{max}/cm^{-1} 3393, 3313, 3060, 2954, 2929, 2856, 2255, 1738, 1686, 1511, 110 1472, 1340, 1052, 939, 780, 651; ¹H NMR (500 MHz, CDCl₃): δ 9.43 (s, 1H), 8.54 (d, J 3.4 Hz, 1H), 8.11 (d, J 8.1 Hz, 1H), 7.34 (d, J 7.9 Hz, 1H), 7.31-7.24 (m, 4H), 6.74 (d, J 8.8 Hz, 2H), 5.39

(d, *J* 2.0 Hz, 1H), 4.69 (dd, *J* 9.8 and 2.1 Hz, 1H), 4.31-4.17 (m, 2 H), 4.07-4.00 (m, 2H), 2.73 (t, *J* 6.1 Hz, 2H), 0.91 (s, 9H), 0.02 ¹¹⁵ (s, 3H), -0.15 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 179.9, 169.5, 162.3, 157.2, 138.2, 135.8, 133.7, 127.5, 126.4, 124.0, 123.2, 122.2, 117.5; 114.2, 112.9, 11.8, 74.3, 62.4, 61.9, 59.4, 29.7, 25.7, 18.6, 18.0, 14.1, -4.6, -5.5; HRMS (ESI): m/z calcd for $C_{30}H_{37}N_3O_6S_iNa$ (M+Na) 586.2349; found 586.2345.

5 (2S,3R)-ethyl 2-(2-(1H-indol-3-vl)-2-oxoacetamido)-3-(4-(2cyanoethoxy)phenyl)-3-hydroxypropanoate (35): To the solution of 34 (80.0 mg, 0.14 mmol) in THF (5.0 ml) was added TBAF (1.0 M in THF, 0.20 ml, 0.2 mmol). The resulting mixture was stirred for 1 h at 0 °C, before it was concentrated in vacuo. 10 Water (5.0 ml) was added to the residue and extracted with ether (3 x 20 ml). The combined extract was washed with brine (15.0 ml) and dried over Na₂SO₄. Evaporation of the solvent and purification of the residue on a silica gel column using ethyl acetate-petroleum ether (7:3) as eluent furnished the 35 (49.7 mg, 15 78 %). Rf = 0.40 (EtOAc-petroleum ether, 7:3). $[\alpha]_D^{25} = -59.0$ (c 0.25, MeOH); IR (neat): v_{max}/cm⁻¹ 3377, 2926, 2854, 2255, 1736, 1680, 1632, 1458, 1371, 1157, 1050, 751; ¹H NMR (500 MHz, CD₃CN): δ 10.14 (s, 1H), 8.68 (d, J 3.4 Hz, 1H), 8.27-8.25 (m, 1H), 7.95 (d, J 8.8 Hz, 1H), 7.52-7.49 (m, 1H), 7.32-7.26 (m, 20 4H), 6.85-6.83 (m, 2H), 5.27 (s, 1H), 4.67 (dd, J 9.1 and 3.4 Hz, 1H), 4.20-4.08 (m, 5H), 2.78 (t, J 6.0 Hz, 2H), 1.21 (t, J 7.1 Hz, 3H); ¹³C NMR (125 MHz, CD₃CN): δ 181.6, 171.0, 163.7, 159.0, 140.2, 137.5, 135.1, 128.8, 127.8, 125.3, 124.4, 123.1, 119.6, 115.6, 113.7, 73.6, 64.3, 62.8, 59.9, 19.5, 14.8; HRMS (ESI): m/z 25 calcd for C₂₄H₂₃N₃O₆Na (M+Na) 472.1485; found 472.1489;

Synthesis of oxazinin 1 and 4: To a cold solution (0 °C) of 35 (110 mg, 0.25 mmol) in MeOH was added NaBH₄ (80.0 mg, 2 mmol) and stirred vigorously at the same temperature for 30 min. ³⁰ Excess MeOH was removed from reaction mixture under reduced pressure. Added water (10.0 ml), extracted with ethyl acetate (3 x 8 ml), washed with brine (10.0 ml) and dried over Na₂SO₄ and concentrated. To the crude alcohol was added acetonitrile (50.0 ml), catalytic (15.0 mg) pyridine para-toluene sulfonate (PPTS) ³⁵ and refluxed for 1 h. The reaction mixture was quenched with aq. NaHCO₃ (20.0 ml), extracted with ethyl acetate (3 x 8 ml), washed with brine and dried over Na₂SO₄. Evaporation of the solvent and purification of the residue using preparative thin layer chromatography furnished the diastereomers 1 (23 mg) and 4 (57 mg) (yield 60% for two steps).

Oxazinin-1 (1): $[\alpha]_D^{25} = +28.6$ (*c* 0.28 MeOH); IR (neat): v_{max}/cm^{-1} 3421, 2923, 2852, 2253(CN), 1660(C=O), 1614, 1540, 1514, 1458, 1384, 1177, 1050, 830, 839, 780, 746; ¹H NMR ⁴⁵ (CD₃CN, 500 MHz): δ 9.39 (br s, 1H), 7.59 (d, *J* 7.9 Hz, 1H), 7.43 (d, *J* 7.9 Hz, 1H), 7.30 (d, *J* 2.1 Hz, 1H) 7.21 (d, *J* 8.8 Hz, 2H), 7.14 (t, *J* 7.3, 1H), 7.00 (t, *J* 7.3 Hz, 1H), 6.88 (d, *J* 8.8 Hz, 2H), 6.69 (s, 1H), 5.59 (s, 1H), 4.62 (d, *J* 8.3 Hz, 1H), 4.15 (t, *J* 6.1 Hz, 2H), 3.74-3.71 (m, 1H), 3.41 (dd, *J* 11.3 and 3.0 Hz, 1H), 50 3.28 (dd *J* 11.3 and 5.8 Hz, 1H), 2.82 (t, *J* 6.1 Hz, 2H); ¹³C NMR (CD₃CN, 125 MHz): δ 170.2, 159.0, 137.4, 131.8, 130.0, 127.5, 126.1, 122.8, 120.4, 120.0, 119.1, 115.4, 112.4, 112.3, 72.9, 71.0,

(M+Na) 414.1430 found 414.1439. ⁵⁵ **Oxazinin-4 (4):** $[\alpha]_D^{25} = +34.0$ (*c* 1.0 MeOH); IR (neat): v_{max}/cm^{-1} 3410, 2921, 2850, 2253(CN), 1736, 1661(C=O), 1613, 1513, 1462, 1238, 1353, 1177, 1045, 831, 746; ¹H NMR (CD₃CN, 500)

63.9, 61.9, 59.9, 19.0; HRMS (ESI): *m/z* calcd for C₂₂H₂₁N₃O₄Na

- MHz): δ 9.28 (br s, 1H), 7.68 (d, *J* 7.2 Hz, 1H), 7.41 (d *J* 8.0 Hz, 60 2 H), 7.36 (d, *J* 8.5 Hz, 2H), 7.32 (d, *J* 2.6 Hz, 1H), 7.15 (t, *J* 7.1 Hz, 1H), 7.07 (t, *J* 8.0 Hz, 1H), 6.93 (, *J* 6.3 Hz, 2H), 6.68 (s, 1H), 5.43 (s, 1H), 4.74 (d, *J* 9.7 Hz, 1H), 4.16 (t, *J* 6.3 Hz, 2H), 3.89-3.86 (m, 1H), 3.43 (dd, *J* 11.4 and 2.6 Hz, 1H), 3.31 (dd, *J* 10.8 and 6.1 Hz, 1H), 3.10 (br s, 1H), 2.82 (t, *J* 6.0 Hz, 2H); ¹³C 65 NMR (CD₃CN, 125 MHz): δ 170.8, 158.3, 137.6, 132.1, 129.9, 127.1, 126.4, 122.9, 120.6, 120.5, 119.3, 115.7, 113.4, 112.6, 78.0, 75.7, 64.1, 62.0, 60.8, 19.2; HRMS (ESI): *m/z* calcd for $C_{22}H_{21}N_3O_4Na$ (M+Na) 414.1430; found 414.1439
- 70 Oxazinin-2 (2): To the solution of 1 (35.0 mg, 0.1 mmol) in THF (2.0 ml) was added 1N NaOH solution (1.0 N in H₂O, 0.4 ml, 0.4 mmol). The resulting mixture was stirred for 1 h at RT, before it was guenched with 1N HCl. Water (5.0 ml) was added to the residue and extracted with ethyl acetate (3 x 20 ml). The 75 combined extract was washed with brine (15.0 ml) and dried over Na₂SO₄. Evaporation of the solvent and purification of the residue on a silica gel column using ethyl acetate-petroleum ether (9:1) as eluent furnished the **2** (22 mg, 80 %). $[\alpha]_D^{25} = +6.0$ (*c* 0.1, MeOH); IR (neat): v_{max}/cm⁻¹ 3318, 2923, 1651, 1613, 1515, ⁸⁰ 1456, 1377, 1172, 1086, 1057, 976, 754; ¹H NMR (CD₃CN, 500 MHz): δ 9.37 (br s, 1H), 7.59 (d, J 7.7 Hz, 1H), 7.43 (d J 8.3 Hz, 2 H), 7.29 (d, J 2.6 Hz, 1H), 7.16 (t, J 7.1 Hz, 1H), 7.09 (d, J 8.5 Hz, 2H), 7.00 (, J 6.8 Hz, 1H), 6.74(d, J 8.5 Hz, 2H), 6.63 (s, 1H), 5.57 (s, 1H) 4.57 (d, J 9.2 Hz, 1H), 3.73-3.69 (m, 1H), 3.44 (dd, J ⁸⁵ 11.4 and 2.6 Hz, 1H), 3.26 (br s, 1H), 3.03 (br s, 1H); ¹³C NMR (125 MHz, CD₃CN): δ 170.2, 158.0, 137.4, 130.0, 129.9, 127.5, 126.1, 122.8, 120.3, 120.1, 116.0, 112.4, 112.4, 72.9, 71.2, 62.1, 59.9; HRMS (ESI): *m/z* calcd for C₁₉H₁₉N₂O₄ (M+H) 339.1345; found 339.1343.

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Notes and references

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20

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