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Design, synthesis and biological evaluation of new series of hexahydroquinoline and fused quinoline derivatives as potent inhibitors of wild-type EGFR and mutant EGFR (L858R and T790M)

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ABSTRACT: New series of hexahydroquinoline and fused quinoline derivatives were designed and synthesized. The thirty seven new compounds were screened for *in vitro* antitumor activity over HepG2, HCT-116 and MCF-7 cancer cell lines. Results indicated that compounds 2e, 2h, 5b, 5c, 6a, 7d and 9b have the strongest potency over the three cancer cells, and they were further screened for in vitro antitumor activity over NSCLC cell lines (A431 and H1975), as well as cytotoxicity against WI38 and WISH normal cell lines. Results revealed that 7d potently inhibited the growth of H1975 cells harboring EGFR^{T790M} mutation (IC₅₀ = 1.32 ± 0.2 µM) over A431 cells overexpressing EGFR^{WT} (IC₅₀ = 4.96 ± 0.3 µM). Moreover, the seven compounds displayed low cytotoxicity against the tested normal cells. The seven potent antitumor compounds were examined for their ability to inhibit the activity of EGFR^{WT}. The attained data manifested that 7d has remarkable EGFR^{WT} inhibitory activity (IC₅₀ = 0.083 ± 0.002 µM) compared to erlotinib (IC₅₀ = 0.067 ± 0.002 µM). Compound 7d was further studied for its enzymatic inhibitory activity against other eight human kinases, and it displayed outstanding inhibitory activity against EGFR^{L858R} and EGFR^{T790M} mutants (IC₅₀ = 0.053 ± 0.002 , 0.026 ± 0.001 μ M, respectively), as well as JAK3 (IC₅₀ = 0.069±0.003 μ M). Analysis of cell cycle evidenced that 7d induces cell cycle arrest in G2/M and pre-G1 phases in the tested cancer cells. In addition, cancer cell death induced by 7d was evidenced to take place *via* apoptosis supported by elevated Bax/Bcl-2 ratio in the tested cancer cells. Moreover, docking results confirmed the good binding interactions of 7d with EGFR^{WT}, EGFR^{L858R}, EGFR^{T790M} and JAK3, which came in agreement with the results of *in vitro* enzyme assay. Further, 7d is predicted to have good oral absorption, good drug-likeness properties and low toxicity risks in human.

Keywords: Hexahydroquinolines, Fused quinolines, Antitumor, EGFR inhibition, Cell cycle analysis, Apoptosis, Computational studies.

1. Introduction

Cancer is a hazardous disease affecting human in developed and developing countries [1]. Currently, treatment strategies include surgery [2], chemotherapy [3] and radiotherapy [4]. Despite the extensive research and rapid progress in cancer treatment, there is an urgent need for new therapies, and it is important to recognize new agents and new targets for cancer treatment. Receptor tyrosine kinases play a substantial role in signal transduction pathways that control cell division and differentiation. Epidermal growth factor receptor tyrosine kinase (EGFR-TK) belongs to a family of receptor tyrosine kinases that includes three other members (erbB2/HER-2, erbB3/HER-3 and erbB4/HER-4) [5,6].

EGFR-TK is an essential enzyme for intracellular signaling and cell transformation, and anomalous expression of this enzyme was reported in a variety of human malignancies, including non-small cell lung cancer (NSCLC) [7], hepatocellular [8], colorectal [9] and breast [10] cancers. As such, inhibition of EGFR-TK provides a rational basis for a class of targeted therapies, EGFR-TK inhibitors (EGFR-TKIs). EGFR-TKIs compete with ATP for ATP-binding site on EGFR. The ATP-binding site (Fig. 1) has the following characteristic features: Adenine binding region that contains two principal hydrogen bonds formed through interaction of N1 and N6 of the adenine ring with the corresponding amino acids. Lots of EGFR-TKIs use one of the two hydrogen bonds. Sugar pocket that represents a hydrophilic region. Hydrophobic binding regions and channels, though not used by ATP but play an important role in binding affinity and inhibitor selectivity. Phosphate binding region which is largely solvent exposed, and can be used for improving inhibitor selectivity [11].

In the last few years, the oncogenic role of EGFR has been more finely characterized due to the improved understanding of the mechanisms of receptor activation, the finding of somatic mutations of the receptor as well as mutations in the components of the signaling pathway, and in a great measure due to the excellent success of anti-EGFR therapies in clinic [12]. First-generation EGFR-TKIs such as gefitinib [13] and erlotinib [14] (Fig. 2) were proved to be effective in treatment of NSCLC; however, drug resistance rapidly develops. Hence, there is still an urgent need for effective clinical medicines that could overcome drug resistance in patients with advanced NSCLC harboring EGFR-mutations

Studies revealed that T790M 'gatekeeper' mutation in EGFR was the most common mechanism of resistance to the first-generation EGFR-TKIs [15,16]. Also, L858R mutation was reported as another mechanism of resistance to first-generation EGFR-TKIs [17]. To overcome resistance to first-generation EGFR-TKIs, the second-generation EGFR-TKIs (e.g., pelitinib

[18] and neratinib [19]) (Fig 2) were developed; however, acquired resistance to these agents invariably develops. Studies proved that T790M mutation in EGFR gene was also a common mechanism of resistance to the second-generation EGFR-TKIs [20]. Accordingly, third-generation EGFR-TKIs (e.g., osimertinib [21] and olmutinib [22]) (Fig. 2) were developed to overcome resistance to first- and second-generations EGFR-TKIs, and they enabled the irreversible binding to Cys797 residue in the ATP-binding site. In particular, osimertinib showed great selectivity toward the drug-resistant mutant EGFR^{T790M} over the wild-type EGFR (EGFR^{WT}), which made it an efficacious medical regimen for patients harboring EGFR^{T790M} mutation [20,23-25]. Because T790M [15,16] and L858R [17] are the frequent mutations that lead to resistance to EGFR-TKIs, we found it interesting to develop new drugs targeting EGFR^{L858R} and EGFR^{T790M} mutants to overcome resistance to first- and second-generations EGFR-TKIs in patients with advanced NSCLC harboring EGFR-mutations.

2. Rational and design

Quinoline is an important scaffold for a variety of anticancer agents that target a diverse range of receptor tyrosine kinases. The most widely studied of these receptors is EGFR-TK. Pelitinib (Figs. 2 and 3), a quinoline-3-carbonitrile derivative, is an anticancer agent with irreversible EGFR-TK inhibitory activity [18]. Furthermore, neratinib (Figs. 2 and 3) is an EGFR-TK inhibitor that could be used as an adjuvant therapy in patients with early stage breast cancer in which HER2 is overexpressed [19]. Moreover, quinoline-3-carbonitriles A [26] (Figs. 2 and 3) and **B** [26] (Fig. 2) displayed potent antitumor activity against human epidermoid (A431), breast (SKBR3) and colon (SW620) cancer cells, as well as potent EGFR-TK inhibitory activity. The quinoline-3-carboxamide derivative C [27] (Figs. 2 and 3) was found to elicit outstanding antitumor activity against MCF-7 cancer cells through targeting EGFR-TK. Further, the hexahydroquinoline-3-carboxylate D [28] (Figs. 2 and 3) was described as promising antitumor agent against human alveolar basal epithelial (A549) and small cell lung (H446) cancer cells through targeting EGFR-TK. All these quinoline derivatives were proved to act via competing with ATP for binding at the catalytic domain of EGFR-TK, and they share the essential pharmacophoric features required for EGFR-TK inhibition, including quinoline or hexahydroquinoline as a central heteroaromatic ring (occupies the adenine binding region), cyano, carboxamide or ester substitution for H-bond formation with the ATP-binding site, hydrophobic head (occupies the hydrophobic binding region I) and hydrophobic tail (occupies the hydrophobic binding region II). Taking these literature findings into consideration, and as an

attempt to prepare new antitumor EGFR-TKIs, the hexahydroquinoline scaffold was introduced as a central heteroaromatic ring similarly to the hexahydroquinoline EGFR-TKI **D**, and the cyano group or carboxylic (bioisostere of carboxamide) was retained at 3-postion. Further, the hydrophobic head at 4-position was replaced with various hydrophobic aromatic moieties (substituted with halogens as lipophilic electron-withdrawing groups); in addition, two methyl substituents were introduced at 7-position of hexahydroquinoline scaffold as a hydrophobic tail in order to prepare three new series of hexahydroquinolines **2a-j**, **3a-c** and **4a-h** that could target EGFR-TK (Fig. 3).

Furthermore, the bioactive pyrimidine E [29] (Figs. 4 and 5) possessed potent antitumor activity against the NSCLC cells (H3255, A431, HCC827, H1975 and PC-9) through targeting EGFR^{WT} and EGFR^{L858R}. Additionally, the pyrimidines F [30] (Figs. 4 and 5) and G [30] (Fig. 4) were established to display prominent antitumor activity against breast (MCF-7), colon (HCT-116) and human alveolar basal epithelial (A549) cancer cells through targeting EGFR-TK. Besides, several benzylideneamine derivatives H [31] (Figs. 4 and 5) were described as promising antitumor agents against human kidney epithelial (293T) cancer cells and EGFR-TKIs. Moreover, the phenylthiourea derivatives I [32] (Figs. 4 and 5) were proved to exhibit outstanding antitumor efficacy over MCF-7 cancer cells through targeting EGFR-TK. Furthermore, [1,2,4]triazole J [33] (Figs. 4 and 5) displayed interesting antitumor efficacy over HepG2, MCF-7 and HCT-116 cancer cells, as well as potent EGFR-TK inhibitory activity, whereas the [1,2,4]triazole derivatives K [34] (Figs. 4 and 5) were established as promising antitumor agents against A549 and small cell lung (H446) cancer cells via targeting EGFR-TK. Inspired by these findings and as an attempt to study the effect of replacement of the bicyclic quinoline scaffold in the small-molecule inhibitors with large tricyclic or tetracyclic scaffolds on the antitumor and EGFR-TK inhibitory activities, the concept of molecular hybridization was followed to prepare new series of fused quinoline derivatives. Molecular hybridization involves the combination of different pharmacophoric moieties with pertinent biological activities to obtain new hybrid compounds with improved activity, selectivity and safety [35,36]. Accordingly, new series of tetrahydropyrimidoquinolines **5a-f**, hexahydropyrimidoquinolines **6a,b**, and tetrahydro [1,2,4]triazolopyrimidoquinolines **9a,b** were designed through incorporation of pyrimidine and [1,2,4]triazole rings into the hexahydroquinoline scaffold (Fig. 5). Further, the manipulation strategy embraced the incorporation of benzylideneamine and phenylthiourea moieties into the hexahydropyrimidoquinoline scaffold (series 7a-d and 8a,b, respectively) (Fig. 5) in order to verify the importance of these moieties for the antitumor and EGFR-TK inhibitory activities.

The designed new analogs were prepared and screened for antitumor and EGFR-TK inhibitory activities. However, inhibition of EGFR-TK activity is not sufficient to eradicate tumors, meaning that a single-target chemotherapeutic agent is not enough to treat the symptoms. Since single-targeted chemotherapeutics have limited antitumor activity, researchers have done their best to find out other better strategies for cancer remedy, for instance, the multi-target drug design strategy, which is based on the design and synthesis of several bioactive molecules that direct multiple targets to induce different alterations of cellular function [37-39]. Multitargeting chemotherapeutics have the potential to demonstrate additive or synergistic effects compared to the single-targeted ones; therefore, they attained an increasing interest in combating resistant cancer cells [40].

Structure-activity relationship study revealed that the 4-imino-3-(4-(dimethylamino)benzylidene)amino)hexahydropyrimidoquinoline derivative 7d has the most potent antitumor and EGFR^{WT} inhibitory activities, the promising potency of 7d might be attributed to presence of 4-(dimethylamino)benzylidene)amino moiety that offers an extra hydrophobic site, and an extra hydrogen bond acceptor site for binding interactions with the target. Additionally, compound 7d was evaluated for its ability to inhibit other eight enzymes from different classes of human tyrosine kinases (EGFR^{L858R}, EGFR^{T790M}, TYK2, JAK3, BTK, VEGFR-2, mTOR and HER-4) to ensure its multi-targeted activity, and hence good curative effect. Also, its capability to induce cell cycle arrest and cell apoptosis, as well as its effect on Bax/Bcl-2 ratio in HepG2, HCT-116 and MCF-7 cancer cells were assessed. Further, molecular docking studies utilizing the crystal structures of EGFR^{WT}, EGFR^{L858R}, EGFR^{T790M} and JAK3 were performed to investigate the possible binding patterns of the potent compound 7d with these target kinases.

3. Results and discussion

3.1. Chemistry

A general approach to synthesize the designed compounds is outlined in Schemes 1-3. The arylidenemalononitriles **1a-j** [41-48] were synthesized *via* reaction of araldehydes with malononitrile in ethanol [41,48]. In Scheme 1, 2-amino-4-aryl-7,7-dimethyl-5-oxo-5,6,7,8-tetrahydroquinoline-3-carbonitriles **2a-j** were prepared *via* reaction of arylidinemalononitriles **1a-j** with dimedone in refluxing ethanol in presence of ammonium acetate adopting the literature

procedure [49]. The structures of **2a-j** were confirmed on the basis of IR, ¹H NMR, ¹³C NMR, mass spectra and X-ray crystallography (Fig. 6).

Referring to Scheme 2, heating *ortho* aminonitriles **2a**, **2b** and **2f** with sulfuric acid 70% at 70 °C for 1-2 h afforded the corresponding *ortho* aminocarboxylic acid derivatives **3a-c**. Refluxing ortho aminonitriles 2a-d, 2f-h and 2j with excess triethyl orthoformate afforded the corresponding ethyl formimidates 4a-h following the literature procedure [49,50]. Compounds 4a-e and 4h were used as precursors for the synthesis of new pyrimidoquinoline derivatives. Adopting the reported method [49,50], compounds 4a-e and 4h were refluxed with ammonia solution 35% in ethanol to give the corresponding 4-aminopyrimidoquinoline derivatives 5a-f. Furthermore, the hexahydropyrimidoquinolines **6a**,**b** were synthesized *via* reaction of **4a**,**b** with hydrazine hydrate in ethanol at room temperature according to the reported procedure [51]. With respect to Scheme 3, the 3-amino-4-iminohexahydropyrimidoquinoline 6a was allowed to react with benzaldehyde derivatives in refluxing ethanol in presence of a catalytic amount of piperidine corresponding to produce the 3-(4-(un)substituted benzylidene)amino)hexahydropyrimidoquinolines 7a-d. The 3-(4-substituted phenyl)-1-(substituted hexahydropyrimidoquinolin-3-yl)thioureas 8a,b were synthesized via refluxing 3amino-4-iminohexahydropyrimidoquinoline 6a with the appropriate phenyl isothiocyanate in ethanol. Upon heating 3-amino-4-iminohexahydropyrimidoquinolines **6a,b** with triethyl orthoformate, the corresponding tetrahydrotriazolopyrimidoquinolines 9a,b were attained. The structures of the new compounds were ascertained by spectral and elemental analyses.

3.2. Biology

3.2.1. Cytotoxicity testing

3.2.1.1. In vitro antitumor testing

MTT assay was followed to assess the antitumor activity of the thirty seven new derivatives over hepatocellular (HepG2), colorectal (HCT-116) and breast (MCF-7) cancer cell lines [52-54], utilizing doxorubicin, erlotinib and osimertinib as reference drugs. Doxorubicin is one of the most effective chemotherapeutic agents against several types of human cancers, including HepG2 [55], HCT-116 [56] and MCF-7 [55] cancers. Two different mechanisms of action have been proposed for doxorubicin, (i) intercalation of DNA and inhibition of topoisomerase II leading to changes in chromatin structure, (ii) generation of free radicals and oxidative damage to biomolecules [57]. However, doxorubicin resistance is a serious problem

and several mechanisms such as reduction of drug uptake, activation of drug detoxification, increased drug efflux and DNA repair capacity, and deflecting apoptotic pathway [58,59] have been proposed.

Concentrations of compounds that induce 50% inhibition of cell viability (IC₅₀, μ M) were determined (Table 1). Results showed that **2e**, **2h**, **5b**, **5c**, **6a**, **7d** and **9b** are the most potent members against all selected cancer cells (IC₅₀ = 2.91-10.35 μ M) in comparison to the doxorubicin (IC₅₀ = 4.17-5.23 μ M), erlotinib (IC₅₀ = 4.16-11.21 μ M), and osimertinib (IC₅₀ = 8.44-9.72 μ M). Also, compounds **4d**, **4h** and **8b** were proved to have strong activity against HCT-116 cells (IC₅₀ = 3.85-9.82 μ M). Further, compounds **4g** and **7b** manifested strong activity against HepG2 cells (IC₅₀ = 10.17±0.9 and 9.50±0.8 μ M, respectively). Additionally, **4g** exhibited high potency over MCF-7 cells (IC₅₀ = 7.92 μ M). On the other hand, **2f** and **5d** demonstrated moderate efficacy over the three cancer cells (IC₅₀ = 13.64-20.61 μ M), whereas **8b** was proved to have moderate activity over HepG2 and MCF-7 cells (IC₅₀ = 14.49 and 12.57 μ M, respectively). Furthermore, compounds **4b**, **4g**, **7a** and **7b** were found to be moderately active over HCT-116 cells (IC₅₀ = 12.10-19.32 μ M). The rest of the tested compounds displayed moderate to weak activity over the three cancer cells.

3.2.1.1.1. Analysis of structure-activity relationship

Concerning the hexahydroquinolines 2a-j

With regard to analogs carrying substituted phenyl moiety at 4-position of hexahydroquinoline ring **2a-g**, it is evident that presence of 2,4-difluorophenyl moiety at 4-position of hexahydroquinoline ring led to promising antitumor potency over the three cancer cells (compound **2e**). Replacement of 2,4-difluorophenyl moiety with 2,6-difluorophenyl resulted in decreased activity over the three cancer cells (compound **2f** versus **2e**), and this might be due to the steric hindrance caused by the 2,6-difluoro substituents on the phenyl moiety in **2f**. Replacement of 2,6-difluorophenyl moiety in **2f** with 2-chloro-6-fluorophenyl resulted in decreased activity over the three cancer cells (compound **2d**), and this might be attributed to the increased lipophilicity of **2d** (LogP = 2.32) compared to **2f** (LogP = 1.80). Replacing 2-chloro-6-fluorophenyl moiety in **2d** with 2,6-dichlorophenyl resulted in decreased potency over the three cancer cells (compound **2c** versus **2d**), and this might be attributed to the increased lipophilicity of **2d** (LogP = 2.32). Replacing 2,6-dichlorophenyl moiety in **2c** with 2,3-dichlorophenyl resulted in decreased potency over all cancer cells (compound **2a** versus **2c**), while its replacement with 3,4-dichlorophenyl resulted in improved activity over HCT-116

and MCF-7 cells, and decreased activity over HepG2 (compound **2b** versus **2c**). Further, replacement of 2,4-difluorophenyl moiety in **2e** with biphenyl led to weak activity over the three cancer cells (compound **2g**). On the other hand, presence of [1,3]benzodioxol-4-yl moiety at 4-position of hexahydroquinoline resulted in high potency over the three cancer cells (compound **2h**). Replacement of [1,3]benzodioxol-4-yl moiety with naphthalen-2-yl led to weak activity over all cancer cells (compound **2i** versus **2h**). Contrariwise, replacement of naphthalen-2-yl moiety in **2i** with anthracen-9-yl resulted in improved potency over all cancer cells (compound **2j** versus **2i**).

Concerning the 2-aminohexahydroquinoline-3-carboxylic acid analogs 3a-c

Presence of 3,4-dichlorophenyl moiety at 4-position of hexahydroquinoline-3-carboxylic acid led to moderate activity over HepG2, and weak activity over HCT-116 and MCF-7 cells (compound **3b**). Replacement of 3,4-dichlorophenyl moiety in **3b** with 2,3-dichlorophenyl or 2,6-difluorophenyl resulted in weak or no activity over all cancer cells (compounds **3a** and **3c**, respectively).

Considering the structures of the ethyl formimidates 4a-h

With respect to the analogs carrying substituted phenyl moiety at 4-position of hexahydroquinoline ring 4a-f, it is obvious that presence of 2-chloro-6-fluorophenyl moiety at 4position of hexahydroquinoline ring led to the highest activity over the three cancer cells (compound 4d). Replacing 2-chloro-6-fluorophenyl moiety in 4d with 2,6-dichlorophenyl or 2,6-difluorophenyl resulted in decreased activity over all tested cancer cells (compounds 4c and 4e, respectively). Replacing 2,6-dichlorophenyl moiety in 4c with 2,3-dichlorophenyl counterpart resulted in mild increase in activity over all cancer cells (compound 4a versus 4c), while its replacement with 3,4-dichlorophenyl resulted in apparent increase in activity over all selected cancer cells (compound 4b versus 4c), and we can conclude that the activity of the dichlorophenyl analogs 4a-c is influenced by the positions of the two chloro substituents, whereas the decreased activity in 4c compared to 4a and 4b might be due to the steric hinderance caused by the 2,6-dichloro substituents in 4c. On the other hand, the N-(4-((1,1'biphenyl)-4-yl)hexahydroquinolin-2-yl)formimidate **4f** showed the lowest activity amongst all tested N-(4-(substituted phenyl)hexahydroquinolin-2-yl)formimidates, and this might be attributed to the increased lipophilicity of 4f (LogP = 4.64) in comparison to 4a-e (LogP = 2.60-4.13). From another point of view, presence of the bicyclic [1,3]benzodioxol-4-yl moiety at 4position of hexahydroquinoline ring led to eminent potency over all selected cancer cells (compound 4g). Replacement of [1,3]benzodioxol-4-yl in 4g with anthracen-9-yl resulted in

mild increase in activity over HCT-116, and mild decrease in activity over HepG2 and MCF-7 cells (compound **4h** versus **4g**).

With respect to the hexahydropyrimidoquinolines 5a-f

Presence of 3,4-dichlorophenyl moiety at 5-position of hexahydroquinoline ring led to high potency over the three cancer cells (compound **5b**). Replacement of 3,4-dichlorophenyl moiety in **5b** with 2,3-dichlorophenyl resulted in weak activity over the three cancer cells (compound **5a**), while its replacement with 2,6-dichlorophenyl led to mild increase in activity over HCT-116, and mild decrease in activity over HepG2 and MCF-7 cancer cells (compound **5c** versus **5b**). Replacement of 2,6-dichlorophenyl moiety with 2,6-difluorophenyl resulted in decreased activity over all cancer cells (compound **5e** versus **5c**), and this might be attributed to the decreased lipophilicity of **5e** (LogP = 2.70) compared to **5c** (LogP = 3.73). Replacement of 3,4-dichlorophenyl moiety in **5b** with the polycyclic anthracen-9-yl resulted in decreased activity over the three cancer cells (compound **5f**), and this might be attributed to the bulkiness of the anthracene moiety.

Referring to the hexahydropyrimidoquinolines 6a,b

It is obvious that presence of 2,3-dichlorophenyl moiety at 5-position of hexahydropyrimidoquinoline nucleus led to high potency over the three cancer cells (compound **6a**), while its replacement with 3,4-dichlorophenyl led to weak activity over all cancer cells (compound **6b** versus **6a**).

Considering the structures of the hexahydropyrimidoquinoline derivatives 7a-d

Presence of unsubstituted benzylideneamino moietv at **3-position** of hexahydropyrimidoquinoline scaffold led to moderate activity over the three cancer cells (compound 7a). Replacement of unsubstituted benzylideneamino moiety in 7a with 4chlorobenzylideneamino led to increased activity over HepG2 and HCT-116 cells, and decreased activity over MCF-7 cells (compound 7b). Replacement of 4-chlorobenzylideneamino moiety in 7b with 4-bromobenzylideneamino led to apparent decrease in activity over the three cancer cells (compound 7c), and this might be attributed to the increased lipophilicity of 7c (LogP = 4.78) compared to 7b (LogP = 4.65). Contrariwise, replacement of 4-chlorobenzylidene moiety in 7b with 4-(dimethylamino)benzylidene led to compound 7d with close potency to doxorubicin and higher potency than erlotinib and osimertinib over the three cancer cells, and this confirms that presence of phenyl substituted with electron-donating group in 7d led to

higher potency than the unsubstituted phenyl (compound 7a) or phenyl substituted with electronwithdrawing group (compounds 7b,c).

Regarding the activity of 1-(substituted hexahydropyrimidoquinolin-3-yl)-3-(4-substituted phenyl)thiourea derivatives **8a,b**

Presence of 3-(4-chlorophenyl)thiourea moiety at 3-position of hexahydropyrimidoquinoline nucleus led to moderate activity over the three cancer cells (compound **8a**). Replacement of 3-(4-chlorophenyl)thiourea moiety in **8a** with 3-(4-methoxyphenyl)thiourea led to evident increase in activity over all cancer cells (compound **8b** versus **8a**), and this confirms that presence of phenyl substituted with electron-donating group improves the antitumor potency.

Referring to the tetrahydrotriazolopyrimidoquinolines 9a,b

Presence of 2,3-dichlorophenyl moiety at 12-position of tetrahydrotriazolopyrimidoquinoline nucleus led to moderate activity over HepG2 and HCT-116 cells, and weak activity over MCF-7 cells (compound 9a). Replacement of 2,3-dichlorophenyl moiety in 9a with 3,4-dichlorophenyl led to high potency over the three cancer cells (compound 9b versus 9a).

3.2.1.2. Cytotoxicity testing against NSCLC cell lines that overexpress $EGFR^{WT}$ (A431) and mutant $EGFR^{T790M}$ (H1975), as well as normal cell lines (W138 and WISH)

Cytotoxicity of compounds **2e**, **2h**, **5b**, **5c**, **6a**, **7d** and **9b** against NSCLC cell lines (A431, overexpressing EGFR^{WT}) and (H1975, harboring mutant EGFR^{T790M}), as well as lung fibroblast (WI38) and amnion epithelial (WISH) normal cell lines was assessed adopting MTT assay [52-54]. IC₅₀ values (μ M) were determined, and results are compared to doxorubicin, erlotinib and osimertinib (Table 2). Compound **7d** exhibited potent activity against H1975 cells (IC₅₀ = 1.32±0.2 μ M) with 3.8-fold selectivity over A431 cells (IC₅₀ = 4.96±0.3 μ M). The potency of **7d** over A431 cells was proved to be higher that of osimertinib (IC₅₀ = 5.63±0.3 μ M), whereas its potency over H1975 was half that of osimertinib (IC₅₀ = 0.69±0.1 μ M), and 6.45 times higher than that of erlotinib (IC₅₀ = 8.52±0.5 μ M). Similarly, compounds **2e**, **2h** and **5c** were proved to be more selective toward H1975 cells (IC₅₀ = 7.13±0.6, 3.89±0.4 and 4.46±0.4 μ M, respectively) compared to A431 cells (IC₅₀ = 9.82±1.0, 5.78±0.5 and 9.10±1.1 μ M, respectively). On contrary, compounds **5b**, **6a** and **9b** showed higher activity against A431 cells (T.89±0.6, 10.62±1.5 and 9.67±1.3 μ M, respectively) over H1975 cells (8.75±0.6, 12.31±1.7 and 10.11±1.3 μ M, respectively). Referring to results of cytotoxicity against normal cells (Table 2), the seven tested compounds showed lower cytotoxicity against W138 (IC₅₀ = 39.26-86.42 μ M) and WISH (IC₅₀ =

19.78-63.37 μ M) normal cells than doxorubicin (IC₅₀ = 6.72±0.5 and 3.18±0.2 μ M, respectively). In addition, the selectivity index (SI) values of **2e**, **2h**, **5b**, **5c**, **6a**, **7d** and **9b** toward HepG2, HCT-116, MCF-7, A431 and H1975 cancer cells versus WI38 and WISH normal cells were calculated (Table 3). The SI values more than 2 are considered as high selectivity [60,61]. Results indicated that the seven compounds have high SI values toward the five cancer cells versus the two normal cells (SI = 2.09-48.69), and they are expected to be safe antitumor agents. In addition, compound **7d** displayed the highest SI values toward all tested cancer cells (except HCT-116) with the highest SI values toward H1975 NSCLC cells harboring EGFR^{T790M} mutation (SI = 48.69 and 35.14) versus WI38 and WISH normal cells, respectively.

3.2.2. Mechanistic studies

3.2.2.1. Tyrosine kinase assay

The potent antitumor agents in the current study were subjected to further studies to investigate their potential mechanism of action. EGFRWT inhibitory activity of the potent antitumor compounds 2e, 2h, 5b, 5c, 6a, 7d and 9b was studied [62], and results are displayed as % inhibition of EGFR^{WT} (at 10 μ M) (Fig. 7) and IC₅₀ (μ M) (Table 4), and they are compared to erlotinib and osimertinib as reference drugs. Compound 7d demonstrated the highest EGFR^{WT} inhibitory activity (IC₅₀ = 0.083 ± 0.002 µM) close to that of erlotinib (IC₅₀ = 0.067 ± 0.002 µM) and about 6 times higher than that of osimertinib (IC₅₀ = 0.48 ± 0.009 µM), and this result comes in accordance with the antitumor screening results. In addition, compounds 2e, 2h and 5b displayed promising EGFR^{WT} inhibitory activity (IC₅₀ = 0.153 ± 0.003 , 0.129 ± 0.003 and 0.143±0.003 µM, respectively). Additionally, compounds 5c, 6a and 9b exhibited good EGFR^{WT} inhibitory activity (IC₅₀ = 0.216 ± 0.005 , 0.347 ± 0.007 and 0.459 ± 0.009 µM, respectively). Further, compound 7d was assessed for its inhibitory activity against EGFR^{L858R} and EGFR^{T790M} mutants [62]. Results are displayed as % inhibition (at 10 µM) (Fig. 8) and IC₅₀ (µM) (Table 5), and compared to erlotinib and osimertinib as reference drugs. Compound 7d displayed potent inhibitory activity against the drug-sensitive EGFR^{L858R} and the drug-resistant EGFR^{T790M} (IC₅₀ = 0.053 ± 0.002 and 0.026 ± 0.001 µM, respectively) compared to that of erlotinib (IC₅₀ = 0.045 ± 0.001 and 0.443 ± 0.002 µM, respectively) and osimertinib (IC₅₀ = 0.025 ± 0.001 and 0.022±0.001 µM, respectively). Actually, the activity of 7d was close to that of erlotinib over EGFR^{L858R} and 17 times higher than that of erlotinib over EGFR^{T790M}. Additionally, 7d displayed similar activity to that of osimertinib over EGFR^{T790M} and half the activity of osimertinib over EGFR^{L858R}, and we can conclude that **7d** has high selectivity toward mutant forms of EGFR over the wild-type form, and hence it might overcome resistance to EGFR-TKIs that develops due to L858R and T790M mutations of EGFR gene.

Compound **7d** was subjected to further studies against other six different human kinases containing SH2 domain (TYK2) or both SH2 and SH3 domains (JAK3 and BTK) and other human protein or lipid kinases (VEGFR-2, mTOR and HER-4) adopting the kinase profile assay [62], and using erlotinib as a reference drug. Results are expressed as % inhibition (at 10 μ M) (Fig. 8) and IC₅₀ (μ M) (Table 5). Compound **7d** exhibited potent enzymatic inhibitory activity against JAK3 (IC₅₀ = 0.069±0.003 μ M) which was proved to be 2 times higher than that of erlotinib (IC₅₀ = 0.146±0.003 μ M). In addition, it showed promising enzymatic inhibitory activity against the other tested human kinases (IC₅₀ = 0.089-0.298 μ M). Taking together, compound **7d** is considered to be a promising multi-kinase inhibitor (with the highest selectivity toward EGFR^{T790M} mutant) that might prevent drug resistance by interfering with intracellular signaling pathways or by inhibiting mutated binding sites or genes, and hence it could be used as a promising candidate compound for treatment of patients with advanced NSCLC harboring EGFR^{T790M} mutation.

3.2.2.1.1. Analysis of structure-activity relationship

EGFR^{WT} With respect to inhibitory activity. the 3-((4-(dimethylamino)benzylidene)amino)hexahydropyrimidoquinoline 7d demonstrated the highest EGFR^{WT} inhibitory activity (IC₅₀ = $0.083\pm0.002 \mu$ M), the potent inhibitory activity of 7d might be related to presence of 4-(dimethylamino)benzylidene moiety that carries an extra site for hydrophobic interaction as well as an extra hydrogen bond acceptor site. On the other hand, the 4-amino-5-(3,4-dichlorophenyl)tetrahydropyrimidoquinoline **5b** showed good inhibitory activity $(IC_{50} = 0.143 \pm 0.003 \mu M)$. Replacing 3,4-dichlorophenyl moiety in **5b** with 2,6-dichlorophenyl resulted in decreased inhibitory activity (compound 5c, $IC_{50} = 0.216 \pm 0.005 \mu M$), the decreased activity of 5c in comparison to 5b might be due to the steric hinderence caused by the 2,6dichloro substituents on the phenyl moiety in 5c. Replacing 4-aminotetrahydropyrimidoquinoline in 5b,c with 3-amino-4-iminohexahydropyrimidoquinoline resulted in obvious decrease in activity (compound **6a**, $IC_{50} = 0.347 \pm 0.007 \mu M$), and this might be attributed to the possibility of intramolecular hydrogen bonding interaction between 3-amino and 4-imino groups in 6a that hinders its bonding interaction with EGFR^{WT}. Further, replacing hexahydropyrimidoquinoline scaffold in 7d with hexahydroquinoline led to decreased activity (compounds 2e and 2h, IC_{50} =

0.153±0.003 and 0.129±0.003 μ M, respectively). Comparing the activity of compounds **2e** and **2h**, it is evident that presence of 3,4-difluorophenyl moiety at 4-position of hexahydroquinoline led to good EGFR^{WT} inhibitory activity (compound **2e**, IC₅₀ = 0.153±0.003 μ M). Replacing 3,4-difluorophenyl moiety in **2e** with [1,3]benzodioxol-4-yl led to increased potency (compound **2h**, IC₅₀ = 0.129±0.003 μ M), and this might be attributed to presence of two extra hydrogen bond acceptor sites in **2h**. On contrary, replacing the hexahydropyrimidoquinoline scaffold in **7d** with tetrahydrotriazolopyrimidoquinoline resulted in obvious decrease in activity (compound **9b**, IC₅₀ = 0.459±0.009 μ M), and this might be attributed to the decreased number of hydrogen bond acceptor and donor sites in **9b** as well as the absence of the extra hydrophobic site present in **7d**.

3.2.2.2. Analysis of cell cycle by flow cytometry assay

Several studies evidenced that EGFR-TKIs are capable of inducing cell cycle arrest and cellular apoptosis [63-66]. To further understand the antitumor mechanism of compound 7d, its effects on cell cycle progression in HepG2, HCT-116 and MCF-7 cells were examined (at the IC_{50} of the corresponding cell line) [67-69]. Analysis of cell cycle in HepG2 showed that 7d decreased % cells in G0/G1 phase (from 51.88% to 36.22%), and increased % cells in S phase (from 34.03% to 38.12%), G2/M phase (from 14.09% to 25.66%) and pre-G1 phase (from 1.44% to 17.85%) (Figs. 9 and 10). Analysis of cell cycle in HCT-116 illustrated that 7d decreased % cells in G0/G1 phase (from 48.52% to 34.05%) and S phase (from 35.61% to 27.44%), and increased % cells in G2/M phase (from 15.87% to 38.51%) and pre-G1 phase (from 1.19% to 22.49%) (Figs. 9 and 10). Regarding results of cell cycle analysis in MCF-7, 7d decreased % cells in G0/G1 phase (from 56.14% to 28.99%) and S phase (from 32.91% to 29.41%), and increased % cells in G2/M phase (from 10.95% to 41.6%) and pre-G1 phase (from 1.78% to 32.11%) (Figs. 9 and 10). From the attained results, we can conclude that 7d demonstrated considerable elevation in cell population of the three tested cancer cells in G2/M phase but at different intensities, and induced cellular apoptosis in pre-G1 phase (Figs. 9 and 10), and accordingly stopped the mitotic cycle.

3.2.2.3. Induction of cellular apoptosis

Cancer is one of the diseases where little cellular apoptosis develops leading to growth of malignant cells that will not die. Apoptosis is a well-known target in cancer therapeutics [70]. The mechanism of apoptosis is complex and involves different pathways [70]. Therefore, compound **7d** was examined for its capability to induce apoptosis in HepG2, HCT-116 and

MCF-7 cancer cells adopting the flow cytometry assay [67-69]. The flow cytometry assay results of cells pigmented with propidium iodide (PI) and annexin V-fluorescein isothiocyanate (annexin V-FITC) are shown in Fig. 11. The annexin V/PI double staining disclosed that after 24 h of exposure, **7d** (at the IC₅₀ of the corresponding cell line) induced early and late apoptosis in the three cancer cells compared to the untreated cells. Compound **7d** induced higher percentage of total apoptosis in MCF-7 cells compared to HepG2 and HCT-116 cells (Figs. 11 and 12). On contrary, it induced weak necrosis in the three treated cancer cells (Figs. 11 and 12) affirming that cell death induced by **7d** takes place mainly *via* apoptosis.

3.2.2.4. Analysis of levels of Bax and Bcl-2 proteins

Bcl-2 (B-cell lymphoma 2) family members are significant regulators of apoptosis that can be classified into three groups of proteins [71]. The first group protects against apoptosis (Bcl-2 itself) [71]. The second group is represented by Bax (Bcl-2 associated protein X) and Bak (Bcl-2 antagonist), which are the key activators of the apoptosis machinery in response to cellular stress stimuli [71]. The third group is comprised of different categories of proteins such as Bid (BH3 interacting death domain), Bim (Bcl-2 interacting mediator) and others [71]. Overexpression of the antiapoptotic protein Bcl-2 leads to antagonism of apoptosis and development of resistance of tumor cells to common chemotherapeutic agents. Thus, development of new drugs that can inhibit the antiapoptotic action of Bcl-2 is a very attractive strategy in the design of anticancer agents [72]. The effect of compound 7d on the expression levels of Bax and Bcl-2 proteins was determined after treatment of HepG2, HCT-116 and MCF-7 cancer cells with compound 7d (at 10 µM) [72] (Fig. 13). Compound 7d showed 5.71, 8.15 and 16.51-fold up-regulation in the level of the proapoptotic protein Bax in HepG2, HCT-116 and MCF-7 cells, respectively compared to the untreated cells, while it markedly down-regulated the level of the antiapoptotic protein Bcl-2 in HepG2, HCT-116 and MCF-7 cells up to 0.72, 0.53 and 0.31-fold compared to the untreated cells. These findings proved that induction of apoptosis by the active compound 7d was achieved through up-regulation of Bax and down-regulation of Bcl-2.

4. Computational studies

4.1. Binding mode of compound 7d to EGFRWT, EGFRL858R, EGFRT790M and JAK3

In order to provide a deep insight into the binding mode of compound **7d** (potent EGFR^{WT}, EGFR^{L858R}, EGFR^{T790M} and JAK3 inhibitor) to EGFR^{WT}, EGFR^{L858R}, EGFR^{T790M} and JAK3, a

study based on the crystal structures of EGFRWT (PDB code: 3W2S) [73], EGFRL858R (PDB code: 4LQM) [74], EGFR^{T790M} (PDB code: 2JIV) [75] and JAK3 (PDB code: 5TTV) [76] was accomplished applying "molecular operating environment (MOE) version 2014.0901" Chemical Computing Group Inc. software [77]. Results of docking of 7d into the ATP-binding pocket of EGFR^{WT} illustrated that it interacts with EGFR^{WT} through two hydrogen bonding interactions, one formed between NH of pyrimidoquinoline and Asp800, and the second one formed between oxygen atom of C=O and Ser720, as well as one arene-H interaction between the phenyl of benzylidene moiety and Leu844 (Fig. 14). Results of docking of 7d into EGFR^{L858R} revealed that it forms one hydrogen bonding interaction between the Cl at 3-position of phenyl ring and Met793, as well as two arene-H interactions, one formed between the phenyl of benzylidene moiety and Leu718, and the second one formed between the 5-phenyl moiety and Val726 (Fig. 15). Docking results of 7d into EGFR^{T790M} showed that it forms two hydrogen bonding interactions through N1 and N3 of pyrimidoquinoline with Met793, as well as one arene-H interaction between the phenyl of benzylidene moiety and Gly796 (Fig. 16). Further, docking results of 7d into the binding site of JAK3 indicated that it binds to JAK3 through two hydrogen bonding interactions, one formed between the C=NH group and Arg911, and the second one formed between N1 and Lys830, as well as one arene-H interaction between the 5-phenyl moiety and Arg953 (Fig. 17). Summary of residues involved in arene-H interaction, number of hydrogen bonds, residues involved in hydrogen bonding interaction and docking interaction energy (kcal/mol) of compound 7d with EGFR^{WT}, EGFR^{L858R}, EGFR^{T790M} and JAK3 is presented in Table 6. These results came in agreement with the results of in vitro enzyme assay, and gave a preliminary conception of the binding mode of the new analogs.

4.2. In silico studies

In silico studies have a significant rule in discovery of new drugs, since they are of great importance in predicting the physicochemical properties, pharmacokinetics, and toxicity of drugs of biological interest [78]. For the new compounds, Lipinski's rule [79] and Veber's parameters [80] were analyzed using Molinspiration software [81]. Further, ADME properties were predicted using PreADMET software [82]. Likewise, toxicity risks in human and drug-likeness were prophesied applying Osiris property explorer software [83].

4.2.1. Molinspiration calculations

Lipinski's rule helps to predict whether a biologically active compound is likely to have the chemical and physical properties that ensure good oral absorption or not [79]. Likewise, topological polar surface area (TPSA) and number of rotatable bonds (Nrotb) affect the extent of drug absorption [80]. Molinspiration software [81] was applied for analysis of TPSA, Nrotb and Lipinski's rule parameters of the new compounds. Results (Table 7) illustrated that all of the new compounds agreed with the adequate criteria of TPSA, Nrotb and Lipinski's rule with zero or one violation of Lipinski's rule, hence they are predicted to have good oral absorption.

4.2.2. ADME prediction

PreADMET software [82] was utilized to analyze the ADME characteristics of the new compounds. Blood-brain barrier (BBB) penetration, plasma protein binding (%PPB) and human intestinal absorption (%HIA) were predicted, and results are listed in Table 8. It was reported that drugs with BBB (C_{brain}/C_{blood}) values more than 1 are predicted to be CNS-active candidates, whereas those with BBB (C_{brain}/C_{blood}) values less than 1 are foreseen to be CNS-inactive candidates [84]. Compounds **2a**, **2c**, **5a**, **5c**, **6a**,**b**, **7a-d**, **8a**,**b** and **9a** are foreseen to be CNS-active candidates (BBB penetration = 1.05-6.24). On contrary, the remaining compounds are foreseen to be CNS-inactive candidates (BBB penetration = 0.05-0.89). Additionally, all of the analyzed compounds are predicted to display excellent plasma protein binding (%PPB = 62.55-100.00%), and excellent human intestinal absorption (%HIA = 88.79-97.22%).

4.2.3. Prediction of toxicity risks in human and drug-likeness

The new derivatives were analyzed for prediction of various toxicity risks in human (tumorigenicity, irritancy and mutagenicity) and drug-likeness using Osiris software [83]. Results (Table 9) manifested that all of the analyzed compounds are expected to be non-tumorigenic (except 2j, 4h and 5f), non-irritating (except 2j, 4h and 5f) and non-mutagenic (except 2j, 4h and 5f).

Drug-likeness rules are used for studying the drug-like properties of a molecule. A positive value indicates that the compound has fragments which are frequently present in commercial drugs [85]. Results (Table 9) showed that **3a,b, 5a-c, 6a,b, 7a-d, 8a,b** and **9a,b** have acceptable drug-likeness values.

5. Conclusion

Results of antitumor assay assured that compounds 2e, 2h, 5b, 5c, 6a, 7d and 9b are the most potent members over HepG2, HCT-116 and MCF-7 cancer cells ($IC_{50} = 2.91-10.13 \mu M$). Moreover, compound 7d exhibited enhanced inhibitory potency over H1975 NSCLC cells harboring mutated EGFR^{T790M} (IC₅₀ = $1.32\pm0.2 \mu$ M) over A431 cells overexpressing EGFR^{WT} $(IC_{50} = 4.96 \pm 0.3 \mu M)$, and hence 7d is considered to be a promising and selective candidate for effective treatment of patients with EGFR^{T790M}-mutated NSCLC cell lines. In addition, the seven active antitumor compounds displayed low cytotoxicity over WI38 and WISH normal cells (IC₅₀ = 19.78-86.42 μ M), and they might be utilized as potent and safe antitumor agents. Results of EGFR^{WT} inhibitory activity revealed that 7d has the strongest inhibitory activity (IC₅₀ = 0.083±0.002 µM). Further, compound 7d showed potent inhibitory activity against EGFR^{L858R} and EGFR^{T790M} mutants (IC₅₀ = 0.053 ± 0.002 and 0.026 ± 0.001 µM, respectively), and hence it might overcome resistance to EGFR-TKIs. Additionally, 7d exhibited outstanding inhibitory activity against JAK3 (IC₅₀ = $0.069\pm0.003 \mu$ M) which was proved to be 2-fold higher than that of erlotinib (IC₅₀ = $0.146\pm0.005 \mu$ M). Analysis of cell cycle demonstrated that **7d** induces cell cycle arrest in G2/M and pre-G1 phases in HepG2, HCT-116 and MCF-7 cancer cells. Furthermore, death of cancer cells induced by 7d was affirmed to occur mainly via apoptosis supported by elevation of Bax/Bcl-2 ratio. Docking results affirmed the precise fit of 7d into the binding site of EGFR^{WT}, EGFR^{L858R}, EGFR^{T790M} and JAK3 enzymes. Moreover, results of Molinspiration and Osiris calculations proved that 7d is predicted to have good oral absorption, good drug-likeness properties and low toxicity risks in human. An overview on the attained results emphasized that 7d is a new potent multi-kinase inhibitor with promising antitumor activity against H1975 NSCLC cells harboring EGFR^{T790M} mutation. These advantages together with the low cytotoxicity of 7d against normal cells indicate that 7d is considered to be an auspicious lead compound for future evolution of new more potent and more selective antitumor agents against resistant NSCLC cells harboring EGFR mutations.

6. Experimental

6.1. Chemistry

Stuart melting point (SMP30) apparatus was utilized for determining the melting points (°C). IR spectra were recorded in KBr disc on Unicam SP 1000 IR spectrometer (v in cm⁻¹). ¹H

and ¹³C NMR spectra were recorded on Bruker Avance III HD FT-high resolution (400 MHz) and (500 MHz). Chemical shifts are expressed in δ ppm with reference to tetramethylsilane. Mass spectra were recorded on Triple Quad Tandem spectrometer (ESI) at 4.0 and 3.5 kV for positive and negative ion modes, respectively, and on direct probe controller inlet part of thermo scientific ISQ single quadrupole GC-MS spectrometer (ISQ LT) utilizing X-calibur software, Regional Center of Mycology and Biotechnology (RCMB), Al-Azhar University, Nasr city, Cairo, Egypt. Elemental analyses (% C, H, N) were achieved, and they were in harmony with the predicted structures within ±0.4% of the calculated values. Reaction times were monitored by TLC plates (Silica gel 60 F254), and spots were visualized by UV (366 nm). Hexane/ethyl acetate (9:1) was used for elution. The arylidenemalononitriles **1a-j** were synthesized adopting the previously reported procedure [41,48].

6.1.1. Synthesis of 2-amino-4-aryl-7,7-dimethyl-5-oxo-1,4,5,6,7,8-hexahydroquinoline-3carbonitriles 2a-j

A mixture of dimedone (0.7 g, 0.005 mol), ammonium acetate (1.93 g, 0.025 mol) and arylidenemalononitrile 1a-j (0.005 mol) in absolute ethanol (10 mL) was refluxed for 5-7 h. The precipitate formed was filtered, dried and crystallized from ethanol to yield the hexahydroquinoline derivatives 2a-j.

6.1.1.1. 2-Amino-4-(2,3-dichlorophenyl)-7,7-dimethyl-5-oxo-1,4,5,6,7,8-hexahydroquinoline-3-carbonitrile (2a)

Pale yellow powder, yield 85%, m.p. 253-255 °C. IR: 3445, 3323 (NH₂); 3204 (NH); 2191 (C=N); 1663 (C=O). ¹H NMR (500 MHz, DMSO- d_6): δ 0.97 (s, 3H, CH₃), 1.03 (s, 3H, CH₃), 2.07 (d, J = 16.5 Hz, 1H, CH), 2.24 (d, J = 16.0 Hz, 1H, CH), 2.50-2.52 (m, 2H, CH₂), 4.77 (s, 1H, C₄-H), 7.12 (s, 2H, NH₂, D₂O-exchangeable), 7.15 (d, J = 8.0 Hz, 1H, Ar-H), 7.29 (t, J = 7.5 Hz, 1H, Ar-H), 7.45-7.47 (dd, J = 1.5, 1.5 Hz, 1H, Ar-H). ¹³C NMR: δ 26.9, 28.3, 31.8, 33.7, 49.9, 56.3, 111.6, 119.1, 128.3, 128.5, 128.8, 130.2, 131.8, 144.4, 158.7, 163.3, 195.7. ESI-MS, m/z: 362.10 [M+H]⁺, 360.30 [M-H]⁺. Anal. Calcd. (Found) for C₁₈H₁₇Cl₂N₃O (361.07): C, 59.68 (59.35); H, 4.73 (4.44); N, 11.60 (11.92).

6.1.1.2. 2-Amino-4-(3,4-dichlorophenyl)-7,7-dimethyl-5-oxo-1,4,5,6,7,8-hexahydroquinoline-3-carbonitrile (2b) [86]

White crystals, yield 85%, m.p. 243-245 °C [86]. IR: 3531, 3473 (NH₂); 3327 (NH); 2187 (C=N); 1687 (C=O). ¹H NMR (500 MHz, DMSO- d_6): δ 0.99 (s, 3H, CH₃), 1.04 (s, 3H, CH₃), 2.06 (d, J = 16.0 Hz, 1H, CH), 2.28 (d, J = 16.0 Hz, 1H, CH), 2.36 (d, J = 20.0 Hz, 1H, CH),

2.54 (d, J = 20.0 Hz, 1H, CH), 5.20 (s, 1H, C₄-H), 7.14 (s, 2H, NH₂, D₂O-exchangeable), 7.25 (t, J = 8.0 Hz, 1H, Ar-H), 7.34-7.36 (dd, J = 0.8, 0.8 Hz, 1H, Ar-H), 7.42-7.45 (dd, J = 1.2, 1.2 Hz, 1H, Ar-H). ¹³C NMR: δ 24.1, 27.2, 35.9, 45.8, 53.7, 53.9, 106.5, 113.5, 123.9, 124.0, 125.3, 130.2, 130.7, 132.0, 153.9, 158.5, 191.4. ESI-MS, *m/z*: 362.10 [M+H]⁺, 360.30 [M-H]⁺. Anal. Calcd. (Found) for C₁₈H₁₇Cl₂N₃O (361.07): C, 59.68 (59.92); H, 4.73 (4.96); N, 11.60 (11.22).

6.1.1.3. 2-Amino-4-(2,6-dichlorophenyl)-7,7-dimethyl-5-oxo-1,4,5,6,7,8-hexahydroquinoline-3-carbonitrile (2c) [87]

Pale yellow powder, yield 85%, m.p. 240-242 °C [87]. IR: 3473, 3323 (NH₂); 3175 (NH); 2189 (C=N); 1684 (C=O). ¹H NMR (500 MHz, DMSO- d_6): δ 0.98 (s, 3H, CH₃), 1.04 (s, 3H, CH₃), 2.06-2.07 (m, 1H, CH), 2.25-2.34 (m, 1H, CH), 2.34-2.38 (m, 1H, CH), 2.52 (m, 1H, CH), 5.19 (s, 1H, C₄-H), 7.11 (s, 2H, NH₂, D₂O-exchangeable), 7.23 (t, *J* = 7.5 Hz, 1H, Ar-H), 7.33-7.35 (dd, *J* = 1.5, 1.5 Hz, 1H, Ar-H), 7.43-7.45 (dd, *J* = 1.5, 1.0 Hz, 1H, Ar-H). ¹³C NMR: δ 26.9, 31.6, 32.6, 41.3, 49.9, 56.0, 110.0, 119.0, 128.5, 129.0, 134.1, 135.8, 159.4, 163.8, 195.7. ESI-MS, *m*/*z*: 362.10 [M+H]⁺, 360.40 [M-H]⁺. Anal. Calcd. (Found) for C₁₈H₁₇Cl₂N₃O (361.07): C, 59.68 (59.31); H, 4.73 (4.65); N, 11.60 (11.42).

6.1.1.4. 2-Amino-4-(2-chloro-6-fluorophenyl)-7,7-dimethyl-5-oxo-1,4,5,6,7,8hexahydroquinoline-3-carbonitrile (2d)

White powder, yield 72%, m.p. 197-199 °C. IR: 3415, 3331 (NH₂); 3212 (NH); 2197 (C=N); 1683 (C=O). ¹H NMR (500 MHz, DMSO- d_6): δ 0.93 (s, 3H, CH₃), 1.04 (s, 3H, CH₃), 2.06 (d, J = 16.0 Hz, 1H, CH), 2.26 (d, J = 16.5 Hz, 1H, CH), 2.35 (d, J = 18.0 Hz, 1H, CH), 2.55 (d, J = 18.0 Hz, 1H, CH), 4.87 (s, 1H, C₄-H), 7.12 (s, 2H, NH₂, D₂O-exchangeable), 7.13-7.25 (m, 3H, Ar-H). ¹³C NMR: δ 26.3, 28.6, 29.5, 31.7, 49.8, 56.0, 114.7, 115.1, 119.2, 125.2, 125.5 (d, J = 3.5 Hz), 129.2 (d, J = 9.2 Hz), 133.6 (d, J = 4.1 Hz), 158.2, 159.3, 163.5, 195.7. ESI-MS, *m/z*: 346.10 [M+H]⁺. Anal. Calcd. (Found) for C₁₈H₁₇ClFN₃O (345.10): C, 62.52 (62.34); H, 4.96 (4.69); N, 12.15 (12.35).

6.1.1.5. 2-Amino-4-(2,4-difluorophenyl)-7,7-dimethyl-5-oxo-1,4,5,6,7,8-hexahydroquinoline-3-carbonitrile (2e)

Pale green powder, yield 70%, m.p. 187-189 °C. IR: 3424, 3333 (NH₂); 3227 (NH); 2179 (C=N); 1685 (C=O). ¹H NMR (500 MHz, DMSO- d_6): δ 0.91 (s, 3H, CH₃), 1.03 (s, 3H, CH₃), 2.06 (d, J = 16.0 Hz, 1H, CH), 2.28 (d, J = 16.0 Hz, 1H, CH), 2.34 (d, J = 17.5 Hz, 1H, CH), 2.54-2.58 (dd, J = 1.5, 1.0 Hz, 1H, CH), 4.62 (s, 1H, C₄-H), 7.01 (t, J = 8.5 Hz, 2H, Ar-H), 7.12 (s, 2H, NH₂, D₂O-exchangeable), 7.26-7.32 (m, 1H, Ar-H). ¹³C NMR: δ 25.0, 26.0, 28.7, 31.8, 49.8, 54.3, 110.2, 111.7 (d, J = 25.0 Hz), 119.1 (t, J = 15.5 Hz), 119.5, 129.1 (t, J = 9.5 Hz), 159.3, 159.7 (d, J = 9.5 Hz), 161.7 (d, J = 10.0 Hz), 163.5, 195.7. EI-MS, m/z: 329.30 (2.19, M⁺), 83.10 (100.00). Anal. Calcd. (Found) for C₁₈H₁₇F₂N₃O (329.35): C, 65.64 (65.35); H, 5.20 (5.46); N, 12.76 (12.43).

6.1.1.6. 2-Amino-4-(2,6-difluorophenyl)-7,7-dimethyl-5-oxo-1,4,5,6,7,8-hexahydroquinoline-3-carbonitrile (2f)

White crystals, yield 70%, m.p. 255-257 °C. IR: 3408, 3333 (NH₂); 3217 (NH); 2199 (C=N); 1685 (C=O). ¹H NMR (500 MHz, DMSO-*d*₆): δ 0.91 (s, 3H, CH₃), 1.03 (s, 3H, CH₃), 2.06 (d, *J* = 16.5 Hz, 1H, CH), 2.26-2.36 (m, 2H, CH₂), 2.57 (d, *J* = 17.5 Hz, 1H, CH), 4.62 (s, 1H, C₄-H), 7.01 (t, *J* = 16.0 Hz, 2H, Ar-H), 7.13 (s, 2H, NH₂, D₂O-exchangeable), 7.25-7.31 (m, 1H, Ar-H). ¹³C NMR: δ 25.0, 26.0, 28.7, 31.8, 49.8, 54.4, 110.2, 111.7 (d, *J* = 25.0 Hz), 119.1 (t, *J* = 15.5 Hz), 119.5, 129.1 (t, *J* = 10.8 Hz), 159.3, 159.6 (d, *J* = 7.4 Hz), 161.6 (d, *J* = 7.1 Hz), 163.5, 195.7. ESI-MS, *m*/*z*: 328.40 [M-H]⁺. Anal. Calcd. (Found) for C₁₈H₁₇F₂N₃O (329.13): C, 65.64 (65.34); H, 5.20 (5.41); N, 12.76 (12.43).

6.1.1.7. 2-Amino-4-((1,1'-biphenyl)-4-yl)-7,7-dimethyl-5-oxo-1,4,5,6,7,8-hexahydroquinoline-3carbonitrile (2g)

White crystals, yield 90%, m.p. 242-244 °C. IR: 3384, 3318 (NH₂); 3208 (NH); 2193 (C=N); 1682 (C=O). ¹H NMR (500 MHz, DMSO-*d*₆): δ 0.97 (s, 3H, CH₃), 1.04 (s, 3H, CH₃), 2.12 (d, *J* = 16.0 Hz, 1H, CH), 2.26 (d, *J* = 15.5 Hz, 1H, CH), 2.53 (s, 2H, CH₂), 4.21 (s, 1H, C₄-H), 7.05 (s, 2H, NH₂, D₂O-exchangeable), 7.22 (d, *J* = 8.5 Hz, 2H, Ar-H), 7.33 (t, *J* = 7.5 Hz, 1H, Ar-H), 7.44 (t, *J* = 7.5 Hz, 2H, Ar-H), 7.58 (d, *J* = 8.5 Hz, 2H, Ar-H), 7.62 (d, *J* = 7.5 Hz, 2H, Ar-H). ¹³C NMR: δ 26.9, 28.4, 31.9, 35.3, 50.0, 58.1, 112.6, 119.8, 126.6, 126.7, 127.3, 127.8, 128.9, 138.5, 139.9, 144.0, 158.5, 162.5, 195.8. ESI-MS, *m/z*: 367.70 [M-H]⁺. Anal. Calcd. (Found) for C₂₄H₂₃N₃O (369.18): C, 78.02 (78.32); H, 6.27 (6.40); N, 11.37 (11.64).

6.1.1.8. 2-Amino-4-([1,3]benzodioxol-4-yl)-7,7-dimethyl-5-oxo-1,4,5,6,7,8-hexahydroquinoline-3-carbonitrile (2h)

Greyish green crystals, yield 65%, m.p. 231-233 °C. IR: 3392, 3330 (NH₂); 3216 (NH); 2190 (C=N); 1686 (C=O). ¹H NMR (500 MHz, DMSO- d_6): δ 0.94 (s, 3H, CH₃), 1.03 (s, 3H, CH₃), 2.06 (d, J = 15.5 Hz, 1H, CH), 2.26 (d, J = 15.5 Hz, 1H, CH), 2.39 (d, J = 17.0 Hz, 1H, CH), 2.50-2.58 (m, 1H, CH), 4.24 (s, 1H, C₄-H), 5.83 (s, 1H, Dioxole-H), 6.02 (s, 1H, Dioxole-H), 6.58-6.62 (m, 1H, Ar-H), 6.70-6.74 (m, 2H, Ar-H), 6.98 (s, 2H, NH₂, D₂O-exchangeable). ¹³C NMR: δ 26.3, 28.7, 31.8, 32.1, 50.0, 57.6, 100.7, 107.1, 110.9, 119.7, 121.3, 121.5, 126.2, 144.5, 147.1, 150.2, 163.0, 195.7. EI-MS, *m/z*: 338.18 (100.00, M⁺+1), 337.25 (63.10, M⁺). Anal. Calcd. (Found) for C₁₉H₁₉N₃O₃ (337.38): C, 67.64 (67.41); H, 5.68 (5.48); N, 12.46 (12.22).

6.1.1.9. 2-Amino-7,7-dimethyl-4-(naphthalen-2-yl)-5-oxo-1,4,5,6,7,8-hexahydroquinoline-3carbonitrile (2i) [88]

White powder, yield 75%, m.p. 255-257 °C [88]. IR: 3348, 3312 (NH₂); 3172 (NH); 2190 (C=N); 1684 (C=O). ¹H NMR (500 MHz, DMSO- d_6): δ 0.95 (s, 3H, CH₃), 1.03 (s, 3H, CH₃), 2.07 (d, J = 16.5 Hz, 1H, CH), 2.25 (d, J = 16.0 Hz, 1H, CH), 2.54 (d, J = 4.5 Hz, 2H, CH₂), 4.35 (s, 1H, C₄-H), 7.07 (s, 2H, NH₂, D₂O-exchangeable), 7.26-7.28 (dd, J = 1.5, 2.0 Hz, 1H, Ar-H), 7.44-7.49 (m, 2H, Ar-H), 7.66 (s, 1H, Ar-H), 7.83-7.85 (m, 2H, Ar-H), 7.88 (d, J = 7.5 Hz, 1H, Ar-H). ¹³C NMR: δ 26.7, 28.4, 31.8, 35.9, 50.0, 58.1, 112.5, 119.8, 125.5, 125.6, 125.7, 126.2, 127.5, 127.7, 128.1, 132.0, 132.8, 142.1, 158.5, 162.6, 195.8. ESI-MS, *m/z*: 344.30 [M+H]⁺, 341.90 [M-H]⁺. Anal. Calcd. (Found) for C₂₂H₂₁N₃O (343.17): C, 76.94 (76.72); H, 6.16 (6.34); N, 12.24 (12.52).

6.1.1.10. 2-Amino-4-(anthracen-9-yl)-7,7-dimethyl-5-oxo-1,4,5,6,7,8-hexahydroquinoline-3carbonitrile (2j)

Dark orange crystals, yield 56%, m.p. 227-228 °C. ¹H NMR (500 MHz, DMSO-*d₆*): δ 0.86 (s, 3H, CH₃), 1.02 (s, 3H, CH₃), 1.88 (d, *J* = 16.0 Hz, 1H, CH), 2.13 (d, *J* = 16.0 Hz, 1H, CH), 2.53 (d, *J* = 17.5 Hz, 1H, CH), 2.62 (d, *J* = 18.0 Hz, 1H, CH), 5.98 (s, 1H, C₄-H), 7.05 (s, 2H, NH₂, D₂O-exchangeable), 7.41-7.46 (m, 2H, Ar-H), 7.57-7.70 (m, 2H, Ar-H), 8.06-8.09 (m, 3H, Ar-H), 8.52 (s, 1H, Ar-H), 8.65 (d, *J* = 9.0 Hz, 1H, Ar-H). ¹³C NMR: δ 27.5, 28.5, 29.4, 31.5, 50.0, 58.3, 113.9, 119.3, 124.5, 126.0, 126.8, 128.8, 129.8, 130.8, 131.5, 134.6, 158.1, 162.0, 195.7. EI-MS, *m/z*: 395.19 (18.65, M⁺+2), 394.27 (42.81, M⁺+1), 393.38 (49.91, M⁺), 83.20 (100.00). Anal. Calcd. (Found) for C₂₆H₂₃N₃O (393.49): C, 79.36 (79.66); H, 5.89 (5.79); N, 10.68 (10.29).

6.1.2. Synthesis of 2-amino-7,7-dimethyl-5-oxo-4-(substituted phenyl)-1,4,5,6,7,8hexahydroquinoline-3-carboxylic acids 3a-c

The appropriate *ortho* aminonitrile **2a**, **2b** and **2f** (0.002 mol) was heated with sulfuric acid 70% (6 mL) at 70 °C for 1-2 h. The reaction mixture was cooled and neutralized with 10% NaOH. The precipitated solid was filtered and crystallized from ethanol/water (1:1) to afford the corresponding *ortho* aminocarboxylic acids **3a-c**.

6.1.2.1. 2-Amino-4-(2,3-dichlorophenyl)-7,7-dimethyl-5-oxo-1,4,5,6,7,8-hexahydroquinoline-3carboxylic acid (3a)

White powder, yield 60%, m.p. 192-194 °C. IR: 3452 (CO<u>OH</u>); 3216, 3123 (NH₂, NH); 1692 (<u>CO</u>OH); 1640 (C=O). ¹H NMR (400 MHz, DMSO-*d*₆): δ 0.99 (s, 3H, CH₃), 1.07 (s, 3H, CH₃),

2.16-2.27 (dd, J = 16.0, 16.0 Hz, 2H, CH₂), 2.44 (d, J = 29.6 Hz, 2H, CH₂), 2.95 (s, 1H, NH, D₂O-exchangeable), 4.17 (s, 1H, C₄-H), 7.12 (d, J = 7.2 Hz, 1H, Ar-H), 7.37 (s, 1H, Ar-H), 7.57 (d, J = 7.6 Hz, 1H, Ar-H), 10.21 (s, 1H, COOH, D₂O-exchangeable). ¹³C NMR: δ 28.9, 32.9, 33.4, 38.2, 50.4, 94.0, 112.0, 127.3, 129.2, 129.6, 131.3, 131.5, 144.7, 153.8, 170.4, 180.7, 195.0. EI-MS, *m/z*: 381.31 (35.12, M⁺), 353.28 (100.00). Anal. Calcd. (Found) for C₁₈H₁₈Cl₂N₂O₃ (381.25): C, 56.71 (56.49); H, 4.76 (4.44); N, 7.35 (7.72).

6.1.2.2. 2-Amino-4-(3,4-dichlorophenyl)-7,7-dimethyl-5-oxo-1,4,5,6,7,8-hexahydroquinoline-3carboxylic acid (3b)

White powder, yield 65%, m.p. 207-209 °C. IR: 3449 (CO<u>OH</u>); 3218, 3124 (NH₂, NH); 1693 (<u>CO</u>OH); 1641 (C=O). ¹H NMR (500 MHz, DMSO- d_6): δ 0.96 (s, 3H, CH₃), 1.04 (s, 3H, CH₃), 2.17-2.24 (m, 2H, CH₂), 2.41-2.49 (m, 2H, CH₂), 2.96 (s, 1H, NH, D₂O-exchangeable), 4.14 (s, 1H, C₄-H), 7.09 (d, *J* = 7.5 Hz, 1H, Ar-H), 7.34 (s, 1H, Ar-H), 7.55 (d, *J* = 8.0 Hz, 1H, Ar-H), 10.20 (s, 1H, COOH, D₂O-exchangeable). ¹³C NMR: δ 28.5, 32.4, 32.9, 37.7, 49.9, 78.0, 111.5, 126.8, 128.7, 128.8, 130.8, 131.0, 144.2, 153.3, 170.0, 173.1, 194.5. Anal. Calcd. (Found) for C₁₈H₁₈Cl₂N₂O₃ (381.25): C, 56.71 (56.43); H, 4.76 (4.39); N, 7.35 (7.55).

6.1.2.3. 2-Amino-4-(2,6-difluorophenyl)-7,7-dimethyl-5-oxo-1,4,5,6,7,8-hexahydroquinoline-3carboxylic acid (3c)

Pale green powder, yield 62%, m.p. 245-246 °C. IR: 3453 (CO<u>OH</u>); 3220 (NH); 3160, 3126 (NH₂); 1710 (<u>CO</u>OH); 1630 (C=O). ¹H NMR (500 MHz, DMSO-*d*₆): δ 0.88 (s, 3H, CH₃), 1.01 (s, 3H, CH₃), 2.01 (d, *J* = 16.0 Hz, 1H, CH), 2.16 (d, *J* = 16.0 Hz, 1H, CH), 2.25-2.30 (m, 2H, CH₂), 3.08 (s, 1H, NH, D₂O-exchangeable), 4.48 (s, 1H, C₄-H), 6.98 (s, 2H, Ar-H), 7.25 (s, 1H, Ar-H), 10.19 (s, 1H, COOH, D₂O-exchangeable). ¹³C NMR: δ 26.5, 28.7, 32.2, 35.6, 50.1, 78.0, 108.5, 111.6 (d, *J* = 25.0 Hz), 119.7 (t, *J* = 17.9 Hz), 128.6 (t, *J* = 9.5 Hz), 153.0, 159.6 (d, *J* = 8.4 Hz), 161.5 (d, *J* = 9.5 Hz), 169.3, 194.3. EI-MS, *m/z*: 348.23 (28.79, M⁺), 231.30 (100.00). Anal. Calcd. (Found) for C₁₈H₁₈F₂N₂O₃ (348.35): C, 62.06 (62.41); H, 5.21 (5.50); N, 8.04 (8.42).

6.1.3. Synthesis of ethyl *N*-(4-aryl-3-cyano-7,7-dimethyl-5-oxo-1,4,5,6,7,8-hexahydroquinolin-2-yl)formimidates 4a-h

A solution of *ortho* aminonitrile **2a-d**, **2f-h** and **2j** (0.001 mol) in triethyl orthoformate (10 mL) was refluxed for 7-10 h. Excess triethyl orthoformate was evaporated under reduced pressure and the solid attained was crystallized from ethanol to produce the ethyl formimidates **4a-h**.

6.1.3.1. Ethyl N-(4-(2,3-dichlorophenyl)-3-cyano-7,7-dimethyl-5-oxo-1,4,5,6,7,8hexahydroquinolin-2-yl)formimidate (4a)

Pink crystals, yield 60%, m.p. 204-206 °C. ¹H NMR (500 MHz, DMSO- d_6): δ 0.99 (s, 3H, CH₃), 1.04 (s, 3H, CH₃), 1.28 (t, J = 7.5 Hz, 3H, OCH₂CH₃), 2.12 (d, J = 16.5 Hz, 1H, CH), 2.25 (d, J = 16.0 Hz, 1H, CH), 2.58 (s, 2H, CH₂), 4.27-4.30 (q, J = 7.5 Hz, 2H, OCH₂CH₃), 5.00 (s, 1H, C₄-H), 7.28 (d, J = 7.5 Hz, 1H, Ar-H), 7.34 (t, J = 7.5 Hz, 1H, Ar-H), 7.53-7.54 (dd, J = 1.5, 1.5 Hz, 1H, Ar-H), 8.56 (s, 1H, N=CH). ¹³C NMR: δ 14.1, 27.1, 28.2, 31.9, 33.3, 49.9, 64.2, 80.4, 110.7, 116.6, 128.6, 129.5, 129.6, 130.5, 132.1, 156.6, 162.3, 163.3, 163.8, 195.7. EI-MS, *m/z*: 422.22 (42.81, M⁺+4), 420.21 (18.88, M⁺+2), 418.25 (29.43, M⁺), 83.13 (100.00). Anal. Calcd. (Found) for C₂₁H₂₁Cl₂N₃O₂ (418.32): C, 60.30 (60.54); H, 5.06 (5.39); N, 10.05 (10.44).

6.1.3.2. *Ethyl N-(4-(3,4-dichlorophenyl)-3-cyano-7,7-dimethyl-5-oxo-1,4,5,6,7,8hexahydroquinolin-2-yl)formimidate* (4b)

Pink crystals, yield 63%, m.p. 205-207 °C. ¹H NMR (500 MHz, DMSO- d_6): δ 0.98 (s, 3H, CH₃), 1.04 (s, 3H, CH₃), 1.28 (t, J = 7.0 Hz, 3H, OCH₂CH₃), 2.11 (d, J = 16.0 Hz, 1H, CH), 2.25 (d, J= 16.5 Hz, 1H, CH), 2.56 (s, 2H, CH₂), 4.28-4.29 (q, J = 7.0 Hz, 2H, OCH₂CH₃), 4.89 (s, 1H, C₄-H), 7.33 (d, J = 8.0 Hz, 1H, Ar-H), 7.40 (d, J = 7.5 Hz, 1H, Ar-H), 7.59 (s, 1H, Ar-H), 8.55 (s, 1H, N=CH). Anal. Calcd. (Found) for C₂₁H₂₁Cl₂N₃O₂ (418.32): C, 60.30 (60.19); H, 5.06 (5.36); N, 10.05 (10.31).

6.1.3.3. *Ethyl N-(4-(2,6-dichlorophenyl)-3-cyano-7,7-dimethyl-5-oxo-1,4,5,6,7,8hexahydroquinolin-2-yl)formimidate* (4c)

White crystals, yield 72%, m.p. 215-216 °C. ¹H NMR (500 MHz, DMSO-*d*₆): δ 0.99 (s, 3H, CH₃), 1.04 (s, 3H, CH₃), 1.28 (t, *J* = 7.0 Hz, 3H, OCH₂CH₃), 2.09 (d, *J* = 15.5 Hz, 1H, CH), 2.24-2.28 (dd, *J* = 5.0, 5.5 Hz, 1H, CH), 2.45 (d, *J* = 17.5 Hz, 1H, CH), 2.50 (d, *J* = 18.0 Hz, 1H, CH), 4.28-4.32 (q, *J* = 7.0 Hz, 2H, OCH₂CH₃), 5.41 (s, 1H, C₄-H), 7.28-7.33 (m, 1H, Ar-H), 7.38-7.45 (m, 1H, Ar-H), 7.49-7.52 (m, 1H, Ar-H), 8.57 (s, 1H, N=CH). ¹³C NMR: δ 14.3, 27.6, 28.7, 32.4, 37.2, 50.4, 64.7, 80.8, 111.2, 117.1, 128.9, 129.9, 131.0, 132.5, 157.1, 162.8, 164.3, 196.3. Anal. Calcd. (Found) for C₂₁H₂₁Cl₂N₃O₂ (418.32): C, 60.30 (60.50); H, 5.06 (5.41); N, 10.05 (10.32).

6.1.3.4. *Ethyl N-(4-(2-chloro-6-fluorophenyl)-3-cyano-7,7-dimethyl-5-oxo-1,4,5,6,7,8hexahydroquinolin-2-yl)formimidate* (4d)

Pink crystals, yield 63%, m.p. 196-197 °C. IR: 3448 (NH); 2209 (C=N); 1668 (C=O). ¹H NMR (500 MHz, DMSO- d_6): δ 0.95 (s, 3H, CH₃), 1.04 (s, 3H, CH₃), 1.29 (t, J = 7.0 Hz, 3H, OCH₂CH₃), 2.08-2.12 (d, J = 16.5 Hz, 1H, CH), 2.28 (d, J = 15.5 Hz, 1H, CH), 2.43-2.46 (m,

1H, CH), 2.59 (d, J = 17.5 Hz, 1H, CH), 4.28-4.32 (q, J = 7.0 Hz, 2H, OCH₂CH₃), 5.08 (s, 1H, C₄-H), 7.18-7.36 (m, 3H, Ar-H), 8.58 (s, 1H, N=CH). ¹³C NMR: δ 13.8, 26.3, 28.5, 31.1, 31.8, 49.8, 56.0, 64.2, 78.2, 108.2, 109.8, 114.8 (d, 100.0 Hz), 116.7, 126.1 (d, 120.0 Hz), 130.0 (d, 10.7 Hz), 133.9, 156.2-157.8 (m), 162.2, 163.9 (d, 11.0 Hz), 195.7. EI-MS, *m/z*: 404.32 (1.89, M⁺+2), 401.69 (5.32, M⁺), 55.13 (100.00). Anal. Calcd. (Found) for C₂₁H₂₁ClFN₃O₂ (401.87): C, 62.76 (62.42); H, 5.27 (5.36); N, 10.46 (10.30).

6.1.3.5. Ethyl N-(3-cyano-4-(2,6-difluorophenyl)-7,7-dimethyl-5-oxo-1,4,5,6,7,8hexahydroquinolin-2-yl)formimidate (4e)

White powder, yield 59%, m.p. 192-193 °C. IR: 3392 (NH); 2190 (C=N); 1668 (C=O). ¹H NMR (500 MHz, DMSO-*d*₆): δ 0.93 (s, 3H, CH₃), 1.05 (s, 3H, CH₃), 1.29 (t, *J* = 7.5 Hz, 3H, OCH₂CH₃), 2.10 (d, *J* = 16.5 Hz, 1H, CH), 2.29 (d, *J* = 16.5 Hz, 1H, CH), 2.46 (d, *J* = 18.0 Hz, 1H, CH), 2.60 (d, *J* = 18.0 Hz, 1H, CH), 4.28-4.32 (q, *J* = 7.5 Hz, 2H, OCH₂CH₃), 4.85 (s, 1H, C₄-H), 7.07 (t, *J* = 9.0 Hz, 2H, Ar-H), 7.33-7.39 (m, 1H, Ar-H), 8.56 (s, 1H, N=CH). ¹³C NMR: δ 13.8, 26.2, 28.5, 30.7, 31.9, 49.8, 64.3, 78.8, 109.3, 111.9 (d, *J* = 23.9 Hz), 116.9, 117.2 (t, *J* = 16.8 Hz), 130.0 (t, *J* = 10.8 Hz), 157.1, 159.7, 161.7, 162.2, 163.9, 195.7. EI-MS, *m/z*: 387.10 (1.78, M⁺+2), 386.29 (7.45, M⁺+1), 385.25 (32.80, M⁺), 383.26 (100.00). Anal. Calcd. (Found) for C₂₁H₂₁F₂N₃O₂ (385.41): C, 65.44 (65.12); H, 5.49 (5.26); N, 10.90 (10.61).

6.1.3.6. *Ethyl N-(4-((1,1'-biphenyl)-4-yl)-3-cyano-7,7-dimethyl-5-oxo-1,4,5,6,7,8hexahydroquinolin-2-yl)formimidate* (4*f*)

Yellow crystals, yield 65%, m.p. 180-182 °C. ¹H NMR (500 MHz, DMSO- d_6): δ 0.99 (s, 3H, CH₃), 1.05 (s, 3H, CH₃), 1.29 (t, J = 6.5 Hz, 3H, OCH₂CH₃), 2.16 (d, J = 16.0 Hz, 1H, CH), 2.27 (d, J = 16.5 Hz, 1H, CH), 2.60 (s, 2H, CH₂), 4.28-4.32 (q, J = 7.0 Hz, 2H, OCH₂CH₃), 4.46 (s, 1H, C₄-H), 7.31-7.36 (m, 3H, Ar-H), 7.45 (t, J = 8.0 Hz, 2H, Ar-H), 7.61-7.64 (m, 4H, Ar-H), 8.56 (s, 1H, N=CH). ¹³C NMR: δ 13.9, 27.1, 28.2, 31.8, 31.9, 50.0, 64.1, 82.3, 111.5, 117.3, 126.7, 126.9, 127.5, 128.4, 129.0, 139.1, 139.7, 142.0, 156.1, 162.0, 163.1, 195.8. EI-MS, *m*/*z*: 428.39 (4.07, M⁺+2), 427.32 (21.08, M⁺+1), 426.32 (100.00, M⁺). Anal. Calcd. (Found) for C₂₇H₂₇N₃O₂ (425.53): C, 76.21 (76.50); H, 6.40 (6.20); N, 9.87 (9.50).

6.1.3.7. *Ethyl N-(4-([1,3]benzodioxol-4-yl)-3-cyano-7,7-dimethyl-5-oxo-1,4,5,6,7,8hexahydroquinolin-2-yl)formimidate* (4g)

White crystals, yield 60%, m.p. 220-221 °C. ¹H NMR (500 MHz, DMSO- d_6): δ 0.96 (s, 3H, CH₃), 1.04 (s, 3H, CH₃), 1.28 (t, J = 7.5 Hz, 3H, OCH₂CH₃), 2.11 (d, J = 15.5 Hz, 1H, CH), 2.26 (d, J = 16.5 Hz, 1H, CH), 2.58 (d, J = 17.5 Hz, 2H, CH₂), 4.27-4.31 (q, J = 7.5 Hz, 2H, OCH₂CH₃), 4.47 (s, 1H, C₄-H), 5.86 (s, 1H, Dioxole-H), 6.05 (s, 1H, Dioxole-H), 6.69-6.71 (dd,

J = 2.0, 1.5 Hz, 1H, Ar-H), 6.78-6.80 (m, 2H, Ar-H), 8.52 (s, 1H, N=CH). Anal. Calcd. (Found) for C₂₂H₂₃N₃O₄ (393.44): C, 67.16 (67.39); H, 5.89 (5.51); N, 10.68 (10.42).

6.1.3.8. Ethyl N-(4-(anthracen-9-yl)-3-cyano-7,7-dimethyl-5-oxo-1,4,5,6,7,8-hexahydroquinolin-2-yl)formimidate (4h)

Orange crystals, yield 80%, m.p. 250-252 °C. ¹H NMR (500 MHz, DMSO-*d*₆): δ 0.87 (s, 3H, CH₃), 1.04 (s, 3H, CH₃), 1.27 (t, *J* = 7.5 Hz, 3H, OCH₂CH₃), 1.92 (d, *J* = 16.0 Hz, 1H, CH), 2.14 (d, *J* = 16.5 Hz, 1H, CH), 2.63-2.65 (dd, *J* = 9.0, 9.0 Hz, 2H, CH₂), 4.26-4.31 (q, *J* = 7.5 Hz, 2H, OCH₂CH₃), 6.24 (s, 1H, C₄-H), 7.45-7.47 (m, 2H, Ar-H), 7.53-7.61 (m, 2H, Ar-H), 8.09-8.13 (m, 3H, Ar-H), 8.58 (s, 1H, N=CH), 8.67-8.71 (m, 2H, Ar-H). ¹³C NMR: δ 14.1, 26.9, 28.4, 31.6, 35.5, 50.0, 64.2, 82.7, 113.1, 116.8, 124.5, 124.6, 125.1, 128.9, 129.1, 130.9, 131.5, 132.6, 155.7, 161.9, 162.5, 195.6. EI-MS, *m/z*: 451.33 (5.32, M⁺+2), 450.36 (20.40, M⁺+1), 449.50 (20.40, M⁺), 83.08 (100.00). Anal. Calcd. (Found) for C₂₉H₂₇N₃O₂ (449.55): C, 77.48 (77.22); H, 6.05 (6.32); N, 9.35 (9.63).

6.1.4. Synthesis of 4-amino-5-aryl-8,8-dimethyl-5,8,9,10-tetrahydropyrimido[4,5-*b*]quinolin-6(7*H*)-ones 5a-f

A mixture of compound **4a-e** and **4h** (0.005 mol) and ammonia solution 35% (10 mL) in absolute ethanol (15 mL) was refluxed for 10-12 h. The solvent was evaporated under reduced pressure, and the obtained solid was triturated with ice-water, filtered, washed with water, dried and crystallized from ethanol to yield the pyrimidoquinolines **5a-f**.

6.1.4.1. 4-Amino-5-(2,3-dichlorophenyl)-8,8-dimethyl-5,8,9,10-tetrahydropyrimido[4,5b]quinolin-6(7H)-one (5a)

White crystals, yield 89%, m.p. 227-228 °C. IR: 3478, 3300 (NH₂); 3152 (NH); 1659 (C=O). ¹H NMR (500 MHz, DMSO- d_6): δ 0.91 (s, 3H, CH₃), 1.05 (s, 3H, CH₃), 2.08 (d, J = 16.0 Hz, 1H, CH), 2.30 (d, J = 16.5 Hz, 1H, CH), 2.42-2.49 (m, 1H, CH), 2.66 (d, J = 17.5 Hz, 1H, CH), 5.20 (s, 1H, C₅-H), 6.36 (s, 1H, NH, D₂O-exchangeable), 7.02 (t, J = 9.0 Hz, 2H, Ar-H), 7.26-7.32 (m, 1H, Ar-H), 8.11 (s, 1H, Pyrimidine-H). ¹³C NMR: δ 22.7, 26.1, 28.7, 31.9, 49.9, 94.3, 110.6, 117.0, 117.2, 129.5, 129.6, 129.7, 156.6, 159.7, 161.6, 162.8, 165.3, 195.7. EI-MS, *m/z*: 393.10 (1.52, M⁺+4), 391.10 (7.49, M⁺+2), 389.00 (11.43, M⁺), 167.90 (100.00). Anal. Calcd. (Found) for C₁₉H₁₈Cl₂N₄O (389.28): C, 58.62 (58.44); H, 4.66 (4.33); N, 14.39 (14.65).

6.1.4.2. 4-Amino-5-(3,4-dichlorophenyl)-8,8-dimethyl-5,8,9,10-tetrahydropyrimido[4,5b]quinolin-6(7H)-one **(5b)**

White crystals, yield 92%, m.p. 232-233 °C. IR: 3478, 3313 (NH₂); 3107 (NH); 1657 (C=O). ¹H NMR (500 MHz, DMSO- d_6): δ 0.89 (s, 3H, CH₃), 1.03 (s, 3H, CH₃), 2.12 (d, J = 15.5 Hz, 1H, CH), 2.31 (d, J = 16.0 Hz, 1H, CH), 2.55-2.65 (dd, J = 16.5, 16.0 Hz, 2H, CH₂), 4.92 (s, 1H, C₅-H), 7.05 (s, 2H, NH₂, D₂O-exchangeable), 7.15 (d, J = 6.0 Hz, 1H, Ar-H), 7.50 (d, J = 4.0 Hz, 1H, Ar-H), 7.72 (s, 1H, Ar-H), 8.09 (s, 1H, Pyrimidine-H). ¹³C NMR: δ 26.9, 29.0, 31.8, 32.4, 50.4, 96.3, 113.9, 128.5, 129.8, 130.8, 130.9, 142.5, 144.3, 157.2, 161.5, 163.1, 164.9, 196.3. EI-MS, *m*/*z*: 390.60 (16.60, M⁺+2), 388.87 (29.82, M⁺), 167.51 (100.00). Anal. Calcd. (Found) for C₁₉H₁₈Cl₂N₄O (389.28): C, 58.62 (58.46); H, 4.66 (4.39); N, 14.39 (14.79).

6.1.4.3. 4-Amino-5-(2,6-dichlorophenyl)-8,8-dimethyl-5,8,9,10-tetrahydropyrimido[4,5b]quinolin-6(7H)-one (5c)

White crystals, yield 80%, m.p. 227-228 °C. IR: 3467, 3385 (NH₂); 3164 (NH); 1657 (C=O). ¹H NMR (500 MHz, DMSO- d_6): δ 0.97 (s, 3H, CH₃), 1.05 (s, 3H, CH₃), 2.06 (d, J = 16.0 Hz, 1H, CH), 2.27 (d, J = 16.0 Hz, 1H, CH), 2.51 (d, J = 14.0 Hz, 1H, CH), 2.62 (d, J = 18.0 Hz, 1H, CH), 5.51 (s, 1H, C₅-H), 6.15 (s, 1H, NH, D₂O-exchangeable), 7.28 (t, J = 8.0 Hz, 1H, Ar-H), 7.35-7.36 (m, 1H, Ar-H), 7.52-7.54 (m, 1H, Ar-H), 8.12 (s, 1H, Pyrimidine-H). ¹³C NMR: δ 26.5, 28.6, 31.0, 31.6, 50.0, 93.9, 110.2, 128.6, 131.0, 134.5, 136.0, 156.8, 161.9, 162.5, 165.4, 195.7. Anal. Calcd. (Found) for C₁₉H₁₈Cl₂N₄O (389.28): C, 58.62 (58.32); H, 4.66 (4.29); N, 14.39 (14.67).

6.1.4.4. 4-Amino-5-(2-chloro-6-fluorophenyl)-8,8-dimethyl-5,8,9,10-tetrahydropyrimido[4,5b]quinolin-6(7H)-one (5d)

White crystals, yield 79%, m.p. 254-257 °C. IR: 3468, 3387 (NH₂); 3157 (NH); 1658 (C=O). ¹H NMR (500 MHz, DMSO-*d*₆): δ 0.93 (s, 3H, CH₃), 1.05 (s, 3H, CH₃), 2.08 (d, *J* = 16.5 Hz, 1H, CH), 2.29 (d, *J* = 16.0 Hz, 1H, CH), 2.52 (d, *J* = 14.0 Hz, 1H, CH), 2.65 (d, *J* = 18.0 Hz, 1H, CH), 5.28 (s, 1H, C₅-H), 6.23 (s, 1H, NH, D₂O-exchangeable), 7.17 (s, 1H, Ar-H), 7.28-7.29 (m, 2H, Ar-H), 8.12 (s, 1H, Pyrimidine-H). ¹³C NMR: δ 26.3, 28.6, 30.7, 31.8, 49.9, 94.2, 110.6 (d, *J* = 15.9 Hz), 115.1 (d, *J* = 24.0 Hz), 125.7, 126.2, 128.7, 129.9 (d, *J* = 10.3 Hz), 133.7, 156.7, 161.6, 162.6, 165.2, 195.7. EI-MS, *m/z*: 375.22 (12.91, M⁺+2), 374.25 (9.87, M⁺+1), 373.20 (38.92, M⁺), 338.23 (100.00). Anal. Calcd. (Found) for C₁₉H₁₈ClFN₄O (372.83): C, 61.21 (61.59); H, 4.87 (4.52); N, 15.03 (15.42).

6.1.4.5. 4-Amino-5-(2,6-difluorophenyl)-8,8-dimethyl-5,8,9,10-tetrahydropyrimido[4,5b]quinolin-6(7H)-one **(5e)**

White crystals, yield 86%, m.p. 275-277 °C. IR: 3445, 3332 (NH₂); 3134 (NH); 1659 (C=O). ¹H NMR (400 MHz, DMSO-*d*₆): δ 0.95 (s, 3H, CH₃), 1.07 (s, 3H, CH₃), 2.10 (d, *J* = 16 Hz, 1H, CH), 2.30 (d, *J* = 16.0 Hz, 1H, CH), 2.54-2.69 (m, 2H, CH₂), 5.22 (s, 1H, C₅-H), 6.58 (s, 1H, NH), 7.32 (t, *J* = 8.0 Hz, 1H, Ar-H), 7.51 (d, *J* = 8.0 Hz, 1H, Ar-H), 7.70 (d, *J* = 7.2 Hz, 1H, Ar-H), 8.12 (s, 1H, Pyrimidine-H). ¹³C NMR: δ 27.0, 29.1, 32.2, 33.7, 50.4, 95.3, 111.9 (d, *J* = 25.0 Hz), 128.2 (d, *J* = 11.0 Hz), 129.8, 130.8 (d, *J* = 15.0 Hz), 132.5, 132.9, 141.3, 157.2 (d, *J* = 7.4 Hz), 161.9 (d, *J* = 7.1 Hz), 163.1, 163.2, 165.3, 196.2. EI-MS, *m/z*: 357.17 (1.46, M⁺+1), 356.21 (7.99, M⁺), 55.15 (100.00). Anal. Calcd. (Found) for C₁₉H₁₈F₂N₄O (356.14): C, 64.04 (64.41); H, 5.09 (5.43); N, 15.72 (15.40).

6.1.4.6. 4-Amino-5-(anthracen-9-yl)-8,8-dimethyl-5,8,9,10-tetrahydropyrimido[4,5-b]quinolin-6(7H)-one (5f)

Yellow crystals, yield 83%, m.p. 234-236 °C. ¹H NMR (500 MHz, DMSO-*d*₆): δ 0.80 (s, 3H, CH₃), 1.05 (s, 3H, CH₃), 1.93 (d, *J* = 16.5 Hz, 1H, CH), 2.17 (d, *J* = 16.5 Hz, 1H, CH), 2.63-2.74 (dd, *J* = 17.9, 17.5 Hz, 2H, CH₂), 6.46 (s, 1H, C₅-H), 7.40-7.42 (m, 2H, Ar-H), 7.59 (t, *J* = 8.0 Hz, 1H, Ar-H), 7.72 (t, *J* = 8.0 Hz, 1H, Ar-H), 8.04-8.08 (m, 3H, Ar-H), 8.15 (d, *J* = 8.0 Hz, 1H, Ar-H), 8.60 (s, 1H, Pyrimidine-H), 9.12 (d, *J* = 9.0 Hz, 1H, Ar-H). ¹³C NMR: δ 26.5, 28.5, 28.7, 31.6, 50.1, 98.0, 114.1, 122.8, 124.1, 125.2, 126.8, 127.3, 128.6, 130.6, 132.3, 134.6, 156.4, 160.8, 163.4, 195.9. EI-MS, *m/z*: 421.21 (59.01, M⁺+1), 419.71 (33.16, M⁺), 167.51 (100.00). Anal. Calcd. (Found) for C₂₇H₂₄N₄O (420.52): C, 77.12 (77.43); H, 5.75 (5.40); N, 13.32 (13.70).

6.1.5. Synthesis of 3-amino-4-imino-8,8-dimethyl-5-(substituted phenyl)-4,5,7,8,9,10hexahydropyrimido[4,5-*b*]quinolin-6(3*H*)-ones 6a,b

Hydrazine hydrate 99% (5 mL, excess) in ethanol (10 mL) was added dropwise with vigorous stirring to a cold suspension (5-10 °C) of compound 4a,b (0.005 mol) in ethanol (5 mL). The reaction mixture was stirred at room temperature for 3-5 h. The precipitated solid was filtered and crystallized from ethanol/water (3:1) to produce the pyrimidoquinolines 6a,b.

6.1.5.1. *3-Amino-5-(2,3-dichlorophenyl)-4-imino-8,8-dimethyl-4,5,7,8,9,10hexahydropyrimido*[4,5-b]quinolin-6(3H)-one **(6a)**

White powder, yield 96%, m.p. 206-207 °C. IR: 3304, 3285 (NH₂); 3196 (NH); 1655 (C=O). ¹H NMR (500 MHz, DMSO- d_6): δ 0.90 (s, 3H, CH₃), 1.03 (s, 3H, CH₃), 2.05 (d, *J* = 16.5 Hz, 1H,

CH), 2.27 (d, J = 16.0 Hz, 1H, CH), 2.51 (d, J = 18.0 Hz, 1H, CH), 2.61 (d, J = 18.0 Hz, 1H, CH), 5.08 (s, 1H, C₅-H), 5.66 (s, 2H, NH₂, D₂O-exchangeable), 6.56 (s, 1H, NH, D₂O-exchangeable), 6.73 (s, 1H, NH, D₂O-exchangeable), 7.28-7.45 (m, 3H, Ar-H), 8.06 (s, 1H, Pyrimidine-H). ¹³C NMR: δ 26.5, 28.6, 31.8, 32.9, 50.0, 98.9, 112.1, 127.2, 128.2, 129.2, 130.7, 131.9, 142.3, 150.3, 152.1, 155.0, 164.1, 195.8. EI-MS, *m*/*z*: 408.00 (1.31, M⁺+4), 406.00 (1.90, M⁺+2), 404.00 (3.05, M⁺), 77.10 (100.00). Anal. Calcd. (Found) for C₁₉H₁₉Cl₂N₅O (404.30): C, 56.45 (56.80); H, 4.74 (4.41); N, 17.32 (17.60).

6.1.5.2. *3-Amino-5-(3,4-dichlorophenyl)-4-imino-8,8-dimethyl-4,5,7,8,9,10hexahydropyrimido*[4,5-b]quinolin-6(3H)-one **(6b)**

White powder, yield 90%, m.p. 200-202 °C. IR: 3303, 3275 (NH₂); 3197 (NH); 1654 (C=O). ¹H NMR (500 MHz, DMSO- d_6): δ 0.91 (s, 3H, CH₃), 1.03 (s, 3H, CH₃), 2.10 (d, J = 16.5 Hz, 1H, CH), 2.28 (d, J = 16.0 Hz, 1H, CH), 2.57-2.63 (m, 2H, CH₂), 4.77 (s, 1H, C₅-H), 5.64 (s, 2H, NH₂, D₂O-exchangeable), 6.81 (s, 1H, NH, D₂O-exchangeable), 7.11-7.49 (m, 3H, Ar-H), 7.76 (s, 1H, NH, D₂O-exchangeable), 8.04 (s, 1H, Pyrimidine-H). ¹³C NMR: δ 26.9, 29.1, 32.3, 39.4, 50.5, 90.4, 111.9, 124.1, 127.8, 129.2, 134.2, 136.4, 137.8, 150.2, 153.3, 155.5, 164.5, 196.3. EI-MS, *m/z*: 404.86 (16.64, M⁺+1), 379.68 (100.00). Anal. Calcd. (Found) for C₁₉H₁₉Cl₂N₅O (404.30): C, 56.45 (56.71); H, 4.74 (4.39); N, 17.32 (17.59).

6.1.6. Synthesis of 5-(2,3-dichlorophenyl)-4-imino-8,8-dimethyl-3-(substituted benzylideneamino)-4,5,7,8,9,10-hexahydropyrimido[4,5-*b*]quinolin-6(3*H*)-ones 7a-d

A mixture of compound **6a** (2.02 g, 0.005 mol), the appropriate benzaldehyde (0.005 mol) and a catalytic amount of piperidine (0.5 mL) in ethanol (10 mL) was refluxed for 6-8 h. The mixture was concentrated, and the precipitated solid was filtered, washed several times with cold ethanol and crystallized from ethanol to yield the pyrimidoquinolines **7a-d**.

6.1.6.1. 3-(Benzylideneamino)-5-(2,3-dichlorophenyl)-4-imino-8,8-dimethyl-4,5,7,8,9,10hexahydropyrimido[4,5-b]quinolin-6(3H)-one (7a)

Dark yellow crystals, yield 85%, m.p. 226-227 °C. ¹H NMR (500 MHz, DMSO- d_6): δ 0.94 (s, 3H, CH₃), 1.06 (s, 3H, CH₃), 2.11 (d, J = 16.5 Hz, 1H, CH), 2.33 (d, J = 16.0 Hz, 1H, CH), 2.54 (d, J = 18.0 Hz, 1H, CH), 2.68 (d, J = 18.0 Hz, 1H, CH), 5.79 (s, 1H, C₅-H), 7.27 (d, J = 6.0 Hz, 1H, Ar-H), 7.39-7.46 (m, 4H, Ar-H), 7.71 (s, 3H, Ar-H), 8.12 (s, 1H, Pyrimidine-H), 8.34 (s, 1H, N=CH), 10.56 (s, 1H, NH, D₂O-exchangeable). ¹³C NMR: δ 26.4, 28.7, 31.8, 32.1, 49.9, 96.0, 110.9, 126.6, 126.7, 128.9, 129.2, 129.8, 132.6, 133.4, 134.2, 134.9, 137.5, 144.7, 156.7, 158.4,

162.5, 164.5, 195.8. EI-MS, *m/z*: 492.19 (31.26, M⁺), 222.00 (100.00). Anal. Calcd. (Found) for C₂₆H₂₃Cl₂N₅O (492.40): C, 63.42 (63.09); H, 4.71 (4.31); N, 14.22 (14.60).

6.1.6.2. 3-((4-Chlorobenzylidene)amino)-5-(2,3-dichlorophenyl)-4-imino-8,8-dimethyl-4,5,7,8,9,10-hexahydropyrimido[4,5-b]quinolin-6(3H)-one **(7b)**

Yellow crystals, yield 82%, m.p. 224-226 °C. ¹H NMR (500 MHz, DMSO- d_6): δ 0.89 (s, 3H, CH₃), 1.05 (s, 3H, CH₃), 2.18 (d, J = 16.0 Hz, 1H, CH), 2.34 (d, J = 16.0 Hz, 1H, CH), 2.58-2.67 (dd, J = 16.0, 16.0 Hz, 2H, CH₂), 5.76 (s, 1H, C₅-H), 7.11 (d, J = 8.0 Hz, 1H, Ar-H), 7.47-7.52 (m, 4H, Ar-H), 7.71 (d, J = 8.0 Hz, 2H, Ar-H), 8.18 (s, 1H, Pyrimidine-H), 8.38 (s, 1H, N=CH), 11.18 (s, 1H, NH, D₂O-exchangeable). ¹³C NMR: δ 26.5, 28.5, 31.0, 32.0, 49.9, 97.2, 113.6, 127.9, 128.3, 129.0, 129.4, 130.0, 130.6, 130.8, 133.2, 134.2, 143.6, 144.4, 156.8, 158.3, 162.5, 164.3, 196.0. Anal. Calcd. (Found) for C₂₆H₂₂Cl₃N₅O (525.09): C, 59.27 (59.52); H, 4.21 (4.61); N, 13.29 (13.60).

6.1.6.3. 3-((4-Bromobenzylidene)amino)-5-(2,3-dichlorophenyl)-4-imino-8,8-dimethyl-4,5,7,8,9,10-hexahydropyrimido[4,5-b]quinolin-6(3H)-one (7c)

Yellow crystals, yield 78%, m.p. 293-295 °C. ¹H NMR (500 MHz, DMSO-*d*₆): δ 0.94 (s, 3H, CH₃), 1.06 (s, 3H, CH₃), 2.13 (d, *J* = 16.0 Hz, 1H, CH), 2.32 (d, *J* = 15.5 Hz, 1H, CH), 2.57 (d, *J* = 18.0 Hz, 1H, CH), 2.69 (d, *J* = 18.0 Hz, 1H, CH), 5.83 (s, 1H, C₅-H), 7.22 (t, *J* = 8.5 Hz, 1H, Ar-H), 7.45 (d, *J* = 7.0 Hz, 1H, Ar-H), 7.58-7.69 (m, 5H, Ar-H), 8.09 (s, 1H, Pyrimidine-H), 8.36 (s, 1H, N=CH), 10.57 (s, 1H, NH, D₂O-exchangeable), 12.19 (s, 1H, NH, D₂O-exchangeable). ¹³C NMR: δ 26.5, 28.6, 31.8, 33.0, 49.9, 96.3, 111.1, 122.9, 127.4, 128.5, 129.5, 130.4, 131.9, 132.2, 132.7, 133.5, 141.0, 143.4, 156.8, 158.3, 162.5, 164.7, 195.8. EI-MS, *m/z*: 570.39 (31.68, M⁺-1), 316.80 (100.00). Anal. Calcd. (Found) for C₂₆H₂₂BrCl₂N₅O (571.30): C, 54.66 (54.45); H, 3.88 (3.61); N, 12.26 (12.60).

6.1.6.4. 5-(2,3-Dichlorophenyl)-4-imino-3-((4-(dimethylamino)benzylidene)amino)-8,8-dimethyl-4,5,7,8,9,10-hexahydropyrimido[4,5-b]quinolin-6(3H)-one (7d)

Dark yellow crystals, yield 70%, m.p. 302-304 °C. ¹H NMR (500 MHz, DMSO-*d*₆): δ 0.94 (s, 3H, CH₃), 1.06 (s, 3H, CH₃), 2.12 (d, *J* = 16.5 Hz, 1H, CH), 2.32 (d, *J* = 16.5 Hz, 1H, CH), 2.56 (d, *J* = 18.0 Hz, 1H, CH), 2.68 (d, *J* = 18.0 Hz, 1H, CH), 2.95 (s, 6H, N(CH₃)₂), 5.89 (s, 1H, C₅-H), 6.75 (d, *J* = 9.0 Hz, 2H, Ar-H), 7.18-7.21 (t, *J* = 8.0 Hz, 1H, Ar-H), 7.44 (d, *J* = 7.0 Hz, 1H, Ar-H), 7.51 (d, *J* = 8.0 Hz, 2H, Ar-H), 7.68 (d, *J* = 7.5 Hz, 1H, Ar-H), 7.94 (s, 1H, Pyrimidine-H), 8.28 (s, 1H, N=CH), 10.21 (s, 1H, NH, D₂O-exchangeable). ¹³C NMR: δ 26.5, 28.7, 31.8, 33.2, 42.7, 50.0, 95.3, 111.6, 111.9, 121.4, 127.3, 128.1, 129.0, 129.4, 130.5, 132.6, 141.3, 145.7, 151.3, 156.6, 158.3, 162.4, 164.7, 195.8. EI-MS, *m/z*: 535.84 (13.34, M⁺+1), 533.78 (3.90, M⁺-

1), 258.58 (100.00). Anal. Calcd. (Found) for C₂₈H₂₈Cl₂N₆O (535.47): C, 62.81 (62.51); H, 5.27 (5.55); N, 15.69 (15.83).

6.1.7. Synthesis of 1-(5-(2,3-dichlorophenyl)-4-imino-8,8-dimethyl-6-oxo-5,6,7,8,9,10hexahydropyrimido[4,5-*b*]quinolin-3(4*H*)-yl)-3-(4-substituted phenyl)thioureas 8a,b

A mixture of **6a** (0.81 g, 0.002 mol) and the appropriate phenyl isothiocyanate (0.005 mol) in ethanol (10 mL) was refluxed for 3-4 h. The solvent was evaporated under reduced pressure, and the remained residue was crystallized from ethanol/water (1:1) to yield the thiourea derivatives **8a,b**.

6.1.7.1. 3-(4-Chlorophenyl)-1-(5-(2,3-dichlorophenyl)-4-imino-8,8-dimethyl-6-oxo-5,6,7,8,9,10hexahydropyrimido[4,5-b]quinolin-3(4H)-yl)thiourea **(8a)**

Pale green powder, yield 62%, m.p. 256-258 °C. ¹H NMR (500 MHz, DMSO-*d*₆): δ 0.93 (s, 3H, CH₃), 1.04 (s, 3H, CH₃), 2.06 (d, *J* = 15.5 Hz, 1H, CH), 2.28 (d, *J* = 16.0 Hz, 1H, CH), 2.52 (d, 18.0 Hz, 1H, CH), 2.64 (d, *J* = 17.0 Hz, 1H, CH), 5.10 (s, 1H, C₅-H), 6.64 (s, 2H, 2NH, D₂O-exchangeable), 7.36-7.46 (m, 3H, Ar-H), 7.68 (d, *J* = 8.5 Hz, 2H, Ar-H), 7.79 (d, *J* = 8.0 Hz, 2H, Ar-H), 8.09 (s, 1H, Pyrimidine-H), 9.39 (s, 1H, NH, D₂O-exchangeable), 9.97 (s, 1H, NH, D₂O-exchangeable). ¹³C NMR: δ 26.4, 28.7, 31.7, 31.8, 49.9, 94.5, 111.1, 126.9, 127.0, 128.5, 129.5, 132.5, 133.1, 134.9, 138.8, 139.4, 151.3, 152.8, 156.7, 161.4, 162.7, 181.1, 195.7. EI-MS, *m/z*: 573.79 (9.85, M⁺), 523.18 (100.00). Anal. Calcd. (Found) for C₂₆H₂₃Cl₃N₆OS (573.92): C, 54.41 (54.79); H, 4.04 (4.36); N, 14.64 (14.89).

6.1.7.2. *1-(5-(2,3-Dichlorophenyl)-4-imino-8,8-dimethyl-6-oxo-5,6,7,8,9,10hexahydropyrimido[4,5-b]quinolin-3(4H)-yl)-3-(4-methoxyphenyl)thiourea* **(8b)**

Pale yellow powder, yield 67%, m.p. 238-240 °C. ¹H NMR (500 MHz, DMSO-*d*₆): δ 0.93 (s, 3H, CH₃), 1.04 (s, 3H, CH₃), 2.06 (d, *J* = 16.0 Hz, 1H, CH), 2.28 (d, *J* = 16.0 Hz, 1H, CH), 2.51 (d, *J* = 18.0 Hz, 1H, CH), 2.64 (d, *J* = 17.5 Hz, 1H, CH), 3.76 (s, 3H, OCH₃), 5.10 (s, 1H, C₅-H), 6.70 (s, 1H, NH, D₂O-exchangeable), 6.98-7.15 (m, 4H, Ar-H), 7.36-7.38 (dd, *J* = 2.5, 8.5 Hz, 1H, Ar-H), 7.45 (d, *J* = 2.5 Hz, 1H, Ar-H), 7.77 (d, *J* = 8.0 Hz, 1H, Ar-H), 8.09 (s, 1H, Pyrimidine-H), 9.33 (s, 1H, NH, D₂O-exchangeable), 9.57 (s, 1H, NH, D₂O-exchangeable). ¹³C NMR: δ 26.4, 28.7, 31.7, 32.2, 49.9, 55.3, 94.5, 111.1, 115.1, 126.9, 127.4, 129.5, 132.5, 133.1, 134.9, 137.1, 139.2, 156.7, 158.4, 161.4, 162.7, 164.8, 181.9, 195.7. EI-MS, *m/z*: 569.19 (54.98, M⁺), 119.38 (100.00). Anal. Calcd. (Found) for C₂₇H₂₆Cl₂N₆O₂S (569.51): C, 56.94 (57.23); H, 4.60 (4.93); N, 14.76 (14.95).

6.1.8. Synthesis of 9,9-dimethyl-12-(substituted phenyl)-8,9,10,12-tetrahydro-[1,2,4]triazolo[1',5':1,6]pyrimido[4,5-*b*]quinolin-11(7*H*)-ones 9a,b

A mixture of compound **6a,b** (0.002 mol) and triethyl orthoformate (10 mL) was refluxed for 8–10 h. The reaction mixture was cooled and diluted with cold ethanol (10 mL). The precipitated solid was filtered and crystallized from ethanol to yield the triazolopyrimidoquinolines **9a,b**.

6.1.8.1. *12-(2,3-Dichlorophenyl)-9,9-dimethyl-8,9,10,12-tetrahydro-*[1,2,4]triazolo[1',5':1,6]pyrimido[4,5-b]quinolin-11(7H)-one **(9a)**

White powder, yield 79%, m.p. 180-182 °C. ¹H NMR (500 MHz, DMSO-*d*₆): δ 1.04 (s, 3H, CH₃), 1.07 (s, 3H, CH₃), 2.21 (d, *J* = 16.5 Hz, 1H, CH), 2.32 (d, *J* = 16.0 Hz, 1H, CH), 2.67-2.79 (dd, *J* = 18.0, 18.0 Hz, 2H, CH₂), 5.25 (s, 1H, C₁₂-H), 7.32-7.34 (dd, *J* = 2.0, 2.0 Hz, 1H, Ar-H), 7.49 (d, *J* = 8.5 Hz, 1H, Ar-H), 7.58 (d, *J* = 2.5 Hz, 1H, Ar-H), 8.58 (s, 1H, Pyrimidine-H), 9.66 (s, 1H, Triazole-H). ¹³C NMR: δ 26.9, 28.4, 32.1, 34.1, 50.0, 102.6, 111.5, 128.9, 129.6, 130.4, 130.5, 130.8, 141.0, 143.5, 151.7, 152.3, 156.8, 164.7, 196.1. EI-MS, *m/z*: 418.29 (2.81, M⁺+4), 416.25 (12.82, M⁺+2), 414.16 (23.78, M⁺), 69.06 (100.00). Anal. Calcd. (Found) for C₂₀H₁₇Cl₂N₅O (414.29): C, 57.98 (57.64); H, 4.14 (4.54); N, 16.90 (16.67).

6.1.8.2. *12-(3,4-Dichlorophenyl)-9,9-dimethyl-8,9,10,12-tetrahydro-*[1,2,4]triazolo[1',5':1,6]pyrimido[4,5-b]quinolin-11(7H)-one **(9b)**

White powder, yield 82%, m.p. 185-186 °C. ¹H NMR (500 MHz, DMSO-*d*₆): δ 1.02 (s, 3H, CH₃), 1.07 (s, 3H, CH₃), 2.22 (d, *J* = 16.0 Hz, 1H, CH), 2.32 (d, *J* = 16.5 Hz, 1H, CH), 2.69 (d, *J* = 18.0 Hz, 1H, CH), 2.77 (d, *J* = 17.5 Hz, 1H, CH), 5.25 (s, 1H, C₁₂-H), 7.32-7.34 (dd, *J* = 2.0, 2.0 Hz, 1H, Ar-H), 7.50 (d, *J* = 8.5 Hz, 1H, Ar-H), 7.58 (d, *J* = 2.0 Hz, 1H, Ar-H), 8.58 (s, 1H, Pyrimidine-H), 9.67 (s, 1H, Triazole-H). ¹³C NMR: δ 26.9, 28.3, 32.0, 34.1, 49.9, 102.6, 111.4, 128.8, 129.6, 130.4, 130.5, 130.7, 140.9, 143.5, 151.6, 152.2, 156.8, 164.6, 195.9. EI-MS, *m/z*: 414.30 (0.09, M⁺), 93.10 (100.00). Anal. Calcd. (Found) for C₂₀H₁₇Cl₂N₅O (414.29): C, 57.98 (57.59); H, 4.14 (4.39); N, 16.90 (16.76).

6.2. Biology

All biological screening methods are presented in details in the Supplementary file.

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Fig. 1. The ATP-binding site of EGFR-TK.



Fig. 2. Examples of first-generation (gefitinib and erlotinib), second-generation (pelitinib and neratinib) and third-generation (osimertinib and olmutinib) EGFR-TKIs, and quinolines **A-D** with reported antitumor and EGFR-TK inhibitory activities.



Fig. 3. The designed new series of hexahydroquinolines 2a-j, 3a-c and 4a-h as prospective antitumor agents and EGFR-TKIs.


Fig. 4. Examples of pyrimidines **E-G**, benzylideneamines **H**, phenylthioureas **I** and [1,2,4]triazoles **J,K** with reported antitumor and EGFR-TK inhibitory activities.



Fig. 5. The designed new hybrid compounds 5a-f, 6a,b, 7a-d, 8a,b and 9a,b as prospective antitumor agents and EGFR-TKIs.



Fig. 6. The ORTEP plots of compounds 2c (A) and 2f (B).



Fig. 7. % Inhibition of EGFR^{WT} by compounds 2e, 2h, 5b, 5c, 6a, 7d and 9b (at 10 µM).



Fig. 8. % Inhibition of kinase activity induced by compound 7d (at 10 μ M).



Fig. 9. Bar diagram showing % cell distribution in G0/G1, S, G2/M and Pre-G1 phases for HepG2, HCT-116 and MCF-7 cells treated with vehicle control and **7d** (at the IC₅₀ of the corresponding cell line).



Fig. 10. Flow cytometry analysis of cell cycle phase distribution in HepG2, HCT-116 and MCF-7 cells after treatment with vehicle control and **7d** (at the IC_{50} of the corresponding cell line).



Annexin V-FITC

Fig. 11. Annexin V-FITC/PI double staining for detection of apoptosis in HepG2, HCT-116 and MCF-7 cells after treatment with **7d** (at the IC₅₀ of the corresponding cell line). Q1 quadrant represents dead (necrotic) cells; Q2 quadrant represents late apoptosis; Q3 quadrant represents live cells; Q4 quadrant represents early apoptosis. Summation of early and late apoptosis represents total apoptosis.



Fig. 12. Bar diagram showing % apoptosis in HepG2, HCT-116 and MCF-7 cells treated with vehicle control and 7d (at the IC₅₀ of the corresponding cell line).



Fig. 13. Assessment of the effect of compound **7d** on Bax and Bcl-2 levels in HepG2, HCT-116 and MCF-7 cells.



Fig. 14. (A) 2D Interaction of **7d** with the binding site of EGFR^{WT}. **(B)** 3D Interaction of **7d** with the binding site of EGFR^{WT}. Atoms are colored as follows: cyan for carbon, light gey for hydrogen, blue for nitrogen, red for oxygen and green for chlorine. (PDB code: 3W2S).



Fig. 15. (A) 2D Interaction of **7d** with the binding site of EGFR^{L858R}. **(B)** 3D Interaction of **7d** with the binding site of EGFR^{L858R}. Atoms are colored as follows: cyan for carbon, light gey for hydrogen, blue for nitrogen, red for oxygen and green for chlorine. (PDB code: 4LQM).



Fig. 16. (A) 2D Interaction of **7d** with the binding site of EGFR^{T790M}. **(B)** 3D Interaction of **7d** with the binding site of EGFR^{T790M}. Atoms are colored as follows: cyan for carbon, light gey for hydrogen, blue for nitrogen, red for oxygen and green for chlorine. (PDB code: 2JIV).

- 48 -



Fig. 17. (A) 2D Interaction of **7d** with the binding site of JAK3. **(B)** 3D Interaction of **7d** with the binding site of JAK3. Atoms are colored as follows: cyan for carbon, light gey for hydrogen, blue for nitrogen, red for oxygen and green for chlorine. (PDB code: 5TTV).

Comp. No.		IC ₅₀ (µM) ^{a,b}		Comp. No.		IC ₅₀ (µM) ^{a,b}	
	HepG2	HCT-116	MCF-7		HepG2	HCT-116	MCF-7
2a	61.60±3.4	55.27±3.5	64.54±3.7	4h	14.29±1.2	9.82±0.9	14.32±1.2
2b	57.48±3.5	28.64±1.9	10.86±0.9	5a	63.69±3.7	56.37±3.6	81.98±4.5
2c	40.93±2.8	44.50±2.9	38.46±2.5	5b	6.03±0.4	7.84±0.6	9.46±0.8
2d	22.53±1.9	30.05±2.3	34.50±2.3	5c	8.53±0.6	7.13±0.7	10.81±0.9
2e	8.13±0.7	6.81±0.3	8.20±0.4	5d	17.12±1.5	16.05±1.4	20.61±1.6
2f	14.66±1.3	13.64±1.4	18.10±1.5	5e	25.06±2.3	23.71±1.8	29.17±2.3
2g	73.46±3.9	74.27±3.8	80.54±4.3	5f	19.50±1.7	14.91±1.3	24.08±1.9
2h	10.35±1.0	8.00±0.6	8.89±0.7	6a	2.91±0.1	5.66±0.4	6.56±0.4
2i	78.37±4.3	75.03±4.2	86.68±4.7	6b	59.18±3.2	62.39±3.9	72.19±4.0
2ј	45.93±3.0	53.91±3.3	52.01±3.2	7a	26.53±2.1	19.32±1.6	21.84±1.8
3 a	87.43±4.5	91.42±4.8	>100.00	7b	9.50±0.8	12.90±1.1	31.65±2.3
3 b	49.53±3.3	53.52±3.5	69.32±3.8	7c	35.21±2.6	54.56±3.1	58.10±3.3
3c	70.65±3.8	59.70±3.6	88.46±4.7	7d	4.46±0.2	5.27±0.5	3.25±0.2
4 a	38.11±2.9	32.65±2.3	52.78±3.4	8a	29.68±2.4	32.04±2.2	43.79±2.7
4b	29.34±2.6	19.18±1.6	33.83±2.5	8b	14.49±1.3	8.27±0.7	12.57±1.1
4 c	41.65±3.0	38.60±2.5	57.82±3.5	9a	50.03±3.4	49.11±3.2	77.63±4.1
4d	12.48 ± 1.4	8.35±0.9	13.60±1.4	9b	6.18±0.5	3.85±0.2	9.13±0.9
4 e	35.84±2.8	42.97±2.7	46.01±3.1	Doxorubicin	4.50±0.2	5.23±0.3	4.17±0.2
4f	43.46±3.1	48.26±2.9	65.10±3.7	Erlotinib	8.19±0.4	11.21±0.6	4.16±0.2
4 g	10.17±0.9	12.10±1.1	7.92±0.6	Osimertinib	8.44±0.5	8.89±0.6	9.72±0.6

Table 1. Results of antitumor screening of the newly synthesized compounds.

 ${}^{a}IC_{50}$ values = mean ± SD of three independent determinations. ${}^{b}IC_{50}$ (µM): strong (1-10); moderate (11-50); weak (51-100); no activity (> 100). Bold values represent the best results.

	IC ₅₀ (µM) ^{a,b}							
Comp. No.	Canc	er cells	Normal cells					
	A431	H1975	WI38	WISH				
2e	9.82±1.0	7.13±0.6	86.40±4.2	57.49±3.3				
2h	6.43±0.5	2.89±0.3	67.35±3.6	58.17±3.3				
5b	7.89±0.6	8.75±0.6	55.08±3.2	19.78±1.7				
5c	9.10±1.1	4.46±0.4	48.75±3.0	26.14±1.9				
6a	10.62±1.5	12.31±1.7	39.26±2.5	29.53±2.3				
7d	4.96±0.3	1.32±0.2	64.27±3.4	46.38±2.8				
9b	9.67±1.3	10.11±1.3	51.49±3.1	63.37±3.5				
Doxorubicin	2.73±0.2	4.19±0.2	6.72±0.5	3.18±0.2				
Erlotinib	3.76±0.2	8.52±0.5						
Osimertinib	5.63±0.3	0.69±0.1						

Table 2. Results of cytotoxicity against A431 and H1975 cancer cells, as well as WI38 and WISH normal cells.

 ${}^{a}IC_{50}$ values = mean ± SD of three independent determinations. ${}^{b}IC_{50}$ (μ M): strong (1-20); moderate (21-50); weak (51-100); non-cytotoxic (> 100). Bold values represent the best results.

Table 3. Cytotoxic selectivity against HepG2	, HCT-116, MCF-7,	A431 and H1975	cancer cells
versus WI38 and WISH normal cells.			

	SI ^a								
Comp. No.	Selectivity against HepG2 versus WI38 (WISH)	Selectivity against HCT-116 versus WI38 (WISH)	Selectivity against MCF-7 versus WI38 (WISH)	Selectivity against A431 versus WI38 (WISH)	Selectivity against H1975 versus WI38 (WISH)				
2e	10.63 (7.07)	12.69 (8.44)	10.54 (7.01)	8.80 (5.85)	12.12 (8.06)				
2h	6.51 (5.62)	8.42 (7.27)	7.58 (6.54)	11.65 (9.05)	23.30 (20.13)				
5b	9.13 (3.28)	7.03 (2.52)	5.82 (2.09)	6.98 (2.51)	6.29 (2.26)				
5c	5.72 (3.06)	6.84 (3.67)	4.51 (2.42)	5.36 (2.87)	10.93 (5.86)				
6a	13.49 (10.15)	6.94 (5.22)	5.98 (4.50)	3.70 (2.78)	3.19 (2.40)				
7d	14.41 (10.40)	12.19 (8.80)	19.77 (14.27)	12.96 (9.35)	48.69 (35.14)				
9b	8.33 (10.25)	13.37 (16.46)	5.64 (6.94)	5.32 (6.55)	5.09 (6.27)				
Doxorubicin	1.49 (0.71)	1.28 (0.61)	1.61 (0.76)	2.46 (1.16)	1.60 (0.65)				

^aSI: Selectivity index = IC_{50} calculated for normal cell/ IC_{50} calculated for cancer cell. Bold values represent the highest SI toward each cancer cell line.

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Comp. No.	IC ₅₀ (µM) ^a				
	EGFR ^{WT}				
2e	0.153±0.003				
2h	0.129 ± 0.003				
5b	0.143 ± 0.003				
5c	0.216±0.005				
6a	0.347 ± 0.007				
7d	$0.083{\pm}0.002$				
9b	0.459 ± 0.009				
Erlotinib	0.067 ± 0.002				
Osimertinib	0.480 ± 0.009				

Table 4. EGFR^{WT} inhibitory activity of the most potent antitumor compounds.

 ${}^{a}IC_{50}$ values = mean \pm SD of three independent determinations. Bold value represents the best result.

									_
Comp. No. IC ₅₀ (µM) ^a									
	Tyrosine kinases contain SH2 Domain	Tyrosine kinases contain 1 SH2 and SH3 Domains		Other human protein or lipid kinases					
	TYK2	JAK3	BTK	EGFR ^{L858R}	EGFR ^{T790M}	VEGFR-2	mTOR	HER-4	_
7d	0.089±0.002	0.069±0.003	0.183±0.006	0.053±0.002	0.026±0.001	0.150±0.005	0.166±0.005	0.298±0.009	_
Erlotinib	0.034±0.002	0.146±0.005	0.077±0.003	0.045±0.001	0.443±0.002	0.055±0.001	0.062 ± 0.002	0.162±0.004	
Osimertinib				0.025±0.001	0.022±0.001	Co			

Table 5. Kinase profile assay of compound 7d against eight human kinases.

 ${}^{a}IC_{50}$ values = mean \pm SD of three independent determinations. Bold values represent the best results.

Table 6. Summary of residues involved in arene-H interaction, number of hydrogen bonds, residues involved in hydrogen bonding interaction and docking interaction energy (kcal/mol) of compound **7d** with EGFR^{WT}, EGFR^{L858R}, EGFR^{T790M} and JAK3.

Enzyme	Residues involved in arene-H interaction	No. of H- bonds	Residues involved in H- bonding interaction	Docking interaction energy (kcal/mol)
EGFR ^{WT}	Leu844	2	Asp800, Ser720	-6.73
EGFR ^{L858R}	Leu718, Val726	1	Met793	-6.03
EGFR ^{T790M}	Gly796	2	Met793	-6.82
JAK3	Arg953	2	Arg911, Lys830	-5.01

Comp. No.	Molecular properties						
	TPSA ^a	Nrotb ^b	miLogP ^c	nHBD ^d	nHBAe	M. wt.	<i>n</i> Vs ^f
2a	78.91	1	2.83	3	4	362.26	0
2b	78.91	1	3.33	3	4	362.26	0
2c	78.91	1	2.83	3	4	362.26	0
2d	78.91	1	2.32	3	4	345.81	0
2e	78.91	1	1.83	3	4	329.35	0
2f	78.91	1	1.80	3	4	329.35	0
2g	78.91	2	3.84	3	4	369.47	0
2h	97.38	1	1.44	3	6	337.38	0
2i	78.91	1	3.23	3	4	343.43	0
2j	78.91	1	3.87	3	4	393.49	0
3 a	92.42	2	2.79	4	5	381.26	0
3 b	92.42	2	2.29	4	5	381.26	0
3c	92.42	2	1.26	4	5	348.35	0
4 a	74.49	4	3.63	1	5	412.32	0
4b	74.49	4	4.13	1	5	418.32	0
4c	74.49	4	3.63	l	5	418.32	0
4d	/4.49	4	3.12	l	5	401.87	0
4e	74.49	4	2.60	I	5	385.41	0
4f	74.29	5	4.64	1	5	425.53	0
4 g	92.96	4	2.24	1	7	393.44	0
4h	74.49	4	4.67	1	5	449.55	0
5a	80.91	1	3.73	3	5	389.29	0
5b	80.91	1	3.75	3	5	389.29	0
5c	80.91	1	3.73	3	5	389.29	0
5d	80.91	1	3.21	3	5	372.83	0
5e 5f	80.91	1	2.70	3	5	350.38	0
51	06.91	1	4.70	5	5	420.32	0
0a 6h	90.80	1	1.92	4	6	404.30	0
00 7a	83.14	3	3 97	2	6	492 41	0
7 a 7 b	83 14	3	4 65	2	6	526.86	ů 1
76	83.14	3	1.09	2	6	571.31	1
70 7d	86 38	<u>з</u>	4.78	2	0 7	535.48	1
8a	94.83	5	4 23	4	7	573 94	1
8b	104.06	6	3 60	4	8	569 52	1
9a	72.19	1	3.78	1	6	414.30	0
9b	72.19	1	3.75	1	6	414.30	0
Doxorubicin	207.70	5	-1.64	8	12	544.53	3
Erlotinib	74.75	10	2.79	1	7	393.44	0
Osimertinib	87.55	10	4.08	2	9	499.62	0

Table 7. Calculated Veber's and Lipinski's rule parameters for the new compounds.

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aTPSA: Topological polar surface area, bNrotb: Number of rotatable bonds, cmiLogP: The parameter of lipophilicity,

^d*n*HBD: Number of hydrogen bond donor sites, ^e*n*HBA: Number of hydrogen bond acceptor sites, ^f*n*Vs: Number of violations.

Comp. No.		ADME properties			ADME properties		
	BBB ^a	%PPB ^b	%HIA ^c		BBB ^a	%PPB ^b	%HIA ^c
2a	1.13	99.51	94.94	4h	0.13	94.66	97.22
2b	0.70	100.00	94.94	5a	1.13	95.50	95.32
2c	1.13	95.71	94.94	5b	0.69	99.56	95.32
2d	0.82	87.95	94.44	5c	1.13	92.09	95.32
2e	0.15	83.83	94.02	5d	0.89	86.67	94.77
2 f	0.64	81.31	94.02	5e	0.75	87.23	94.40
2g	0.17	87.97	94.97	5f	0.12	92.05	96.43
2h	0.55	62.55	94.15	6a	1.55	86.90	88.79
2i	0.11	90.69	94.96	6b	1.45	89.96	88.79
2j	0.13	90.01	96.14	7a	4.93	88.79	93.09
3a	0.78	95.31	95.29	7b	6.08	96.02	93.65
3 b	0.23	98.34	95.29	7c	6.24	100.00	93.94
3c	0.63	81.63	93.68	7d	3.35	84.97	93.37
4 a	0.24	89.35	96.41	8a	4.26	94.80	92.98
4 b	0.39	91.17	96.41	8b	2.71	89.10	91.88
4 c	0.24	88.75	96.41	9a	1.05	90.58	96.72
4d	0.19	88.11	96.23	9b	0.37	91.39	96.72
4e	0.17	88.41	96.37	Doxorubicin	0.03	32.79	31.95
4 f	0.05	91.22	96.42	Erlotinib	0.04	93.15	96.29
4 g	0.24	71.26	97.09	Osimertinib	0.18	85.18	96.34

Table 8. In silico ADME properties of the new compounds.

^a BBB: Blood-brain barrier penetration, ^b %PPB: % Plasma protein binding, ^c %HIA: % Human intestinal absorption.

Comp. No.	Toxic	Drug-likeness		
-	Tumorigenicity	Irritancy	Mutagenicity	
2a	-	-	-	-3.42
2b	-	-	-	-3.42
2c	-	-	-	-3.42
2d	-	-	-	-4.93
2e	-	-	-	-6.69
2f	-	-	-	-6.69
2g	-	-	-	-3.00
2h	-	-	-	-3.44
2i	-	-	-	-6.17
2j	++	++	++	-7.77
3a	-	-	-	1.35
3 b	-	-	-	1.35
3c	-	-	-	-0.43
4 a	-	-	-	-4.68
4b	-	-	-	-4.68
4 c	-	-	-	-4.68
4d	-	-	-	-6.18
4 e	-	-	-	-6.47
4f	-	-	-	-4.25
4g	-	-	-	-4.73
4h	++	++	++	-9.01
5a	-		-	0.66
5b	-		-	0.66
50	-	_	-	0.66
5d	-	-	_	-2.22
5e	_	_	_	-2.46
50 5f	++	++	++	-5.01
6a	-		-	0.27
6b	_	<u> </u>	-	0.27
7a		-	-	4.18
7b	_	-	-	4.90
7c		-	-	2.36
7d	<u> </u>	-	_	3.30
9 a 8 a				3 70
0a 8h		-	-	J.17 7 NQ
00	-	-	-	2.00 3.21
98 06	-	-	-	J.41 2 21
90 Dovomukisir	-	- -	-	J.21 7 10
Evioti-ih	-	++	-	1.19
Eriotinib	-	-	-	-0./3

Table 9. In silico prediction of toxicity risks in human and drug-likeness of the new compounds.

-low risk, + moderate risk, ++ high risk. Bold values represent the best results.



Scheme 1. Synthesis of compounds 2a-j.



Comp. No.	Ar	Comp. No.	Ar	Comp. No.	Ar
3a	CI CI	4e	F	5d	
3b		4f	\rightarrow	5e	F
3c	F F	4g	⊂⊂⊂° o	5f	
4a	CI CI	4h		6a	
4b	-CI	5a	CI CI	6b	
4c		5b			
4d		5c			

Scheme 2. Synthesis of compounds 3a-c, 4a-h, 5a-f and 6a,b.



Scheme 3. Synthesis of compounds 7a-d, 8a,b and 9a,b.

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Captions of Figures, Tables and Schemes

Fig. 1. The ATP-binding site of EGFR-TK.

Fig. 2. Examples of first-generation (gefitinib and erlotinib), second-generation (pelitinib and neratinib) and third-generation (osimertinib and olmutinib) EGFR-TKIs, and quinolines **A-D** with reported antitumor and EGFR-TK inhibitory activities.

Fig. 3. The designed new series of hexahydroquinolines 2a-j, 3a-c and 4a-h as prospective antitumor agents and EGFR-TKIs.

Fig. 4. Examples of pyrimidines **E-G**, benzylideneamines **H**, phenylthioureas **I** and [1,2,4]triazoles **J,K** with reported antitumor and EGFR-TK inhibitory activities.

Fig. 5. The designed new hybrid compounds 5a-f, 6a,b, 7a-d, 8a,b and 9a,b as prospective antitumor agents and EGFR-TKIs.

Fig. 6. The ORTEP plots of compounds 2c (A) and 2f (B).

Fig. 7. % Inhibition of EGFR^{WT} by compounds 2e, 2h, 5b, 5c, 6a, 7d and 9b at concentration of $10 \mu M$.

Fig. 8. % Inhibition of kinase activity induced by compound 7d (at 10μ M).

Fig. 9. Bar diagram showing % cell distribution in G0/G1, S, G2/M and Pre-G1 phases for HepG2, HCT-116 and MCF-7 cells treated with vehicle control and **7d** (at the IC₅₀ of the corresponding cell line) for 24 h.

Fig. 10. Flow cytometry analysis of cell cycle phase distribution in HepG2, HCT-116 and MCF-7 cells after treatment with vehicle control and **7d** (at the IC_{50} of the corresponding cell line) for 24 h.

Fig. 11. Annexin V-FITC/PI double staining for detection of apoptosis in HepG2, HCT-116 and MCF-7 cells after treatment with **7d** (at the IC_{50} of the corresponding cell line) for 24 h. Q1 quadrant represents dead (necrotic) cells; Q2 quadrant represents late apoptosis; Q3 quadrant represents live cells; Q4 quadrant represents early apoptosis. Summation of early and late apoptosis represents total apoptosis.

Fig. 12. Bar diagram showing % apoptosis in HepG2, HCT-116 and MCF-7 cells treated with vehicle control and **7d** (at the IC_{50} of the corresponding cell line) for 24 h.
Fig. 13. Assessment of the effect of compound **7d** on Bax and Bcl-2 levels in HepG2, HCT-116 and MCF-7 cells.

Fig. 14. (A) 2D Interaction of **7d** with the binding site of EGFR^{WT}. **(B)** 3D Interaction of **7d** with the binding site of EGFR^{WT}. Atoms are colored as follows: cyan for carbon, light gey for hydrogen, blue for nitrogen, red for oxygen and green for chlorine. (PDB code: 3W2S).

Fig. 15. (A) 2D Interaction of **7d** with the binding site of EGFR^{L858R}. **(B)** 3D Interaction of **7d** with the binding site of EGFR^{L858R}. Atoms are colored as follows: cyan for carbon, light gey for hydrogen, blue for nitrogen, red for oxygen and green for chlorine. (PDB code: 4LQM).

Fig. 16. (A) 2D Interaction of **7d** with the binding site of EGFR^{T790M}. **(B)** 3D Interaction of **7d** with the binding site of EGFR^{T790M}. Atoms are colored as follows: cyan for carbon, light gey for hydrogen, blue for nitrogen, red for oxygen and green for chlorine. (PDB code: 2JIV).

Fig. 17. (A) 2D Interaction of **7d** with the binding site of JAK3. **(B)** 3D Interaction of **7d** with the binding site of JAK3. Atoms are colored as follows: cyan for carbon, light gey for hydrogen, blue for nitrogen, red for oxygen and green for chlorine. (PDB code: 5TTV).

Table 1. Results of antitumor screening of the newly synthesized compounds.

 Table 2. Results of cytotoxicity against A431 and H1975 cancer cells, as well as WI38 and WISH normal cells.

Table 3. Cytotoxic selectivity against HepG2, HCT-116, MCF-7, A431 and H1975 cancer cells versus WI38 and WISH normal cells.

Table 4. EGFR^{WT} inhibitory activity of the most active antitumor compounds.

Table 5. Kinase profile assay of compound 7d against eight human kinases.

Table 6. Summary of hydrophobic interaction, number of hydrogen bonds, residues involved in hydrogen bonding interaction and docking interaction energy (kcal/mol) of compound **7d** with EGFR^{WT}, EGFR^{L858R}, EGFR^{T790M} and JAK3.

Table 7. Calculated Veber's and Lipinski's rule parameters for the new compounds.

Table 8. In silico ADME properties of the new compounds.

Table 9. In silico prediction of toxicity risks in human and drug-likeness of the new compounds.

Scheme 1. Synthesis of compounds 2a-j.

Scheme 2. Synthesis of compounds 3a-c, 4a-h, 5a-f and 6a,b.

Scheme 3. Synthesis of compounds 7a-d, 8a,b and 9a,b.

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Design, synthesis and biological evaluation of new series of hexahydroquinoline and fused quinoline derivatives as potent inhibitors of wild-type EGFR and mutant EGFR (L858R and T790M)

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- New hexahydroquinolines and fused quinolines were prepared.
- The new analogs were screened for their antitumor activity
- Analog 7d showed potent and safe antitumor activity
- Analog **7d** showed obvious inhibition of EGFR^{WT}, EGFR^{T790M}, EGFR^{L858R} and JAK3
- Analog 7d induced cell cycle arrest in G2/M and pre-G1 phases
- Analog 7d induced cell death through apoptosis supported by increased Bax/Bcl-2 ratio