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Convenient synthesis of α , β -unsaturated γ -butyrolactones and γ -butyrolactams *via* decarboxylative iodination of paraconic acids and β -carboxyl- γ -butyrolactams using 1,3-diiodo-5,5-dimethylhydantoin \dagger

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A convenient synthetic approach to α,β -unsaturated γ -butyrolactones and α,β -unsaturated γ -butyrolactams is developed. The reaction proceeds *via* decarboxylative iodination of paraconic acids and β -carboxyl- γ -butyrolactams, employing 1,3-diiodo-5,5-dimethylhydantoin (DIH) under irradiation, followed by dehydroiodination of β -iodo- γ -butyrolactones and γ -butyrolactams providing good yields of α,β -unsaturated γ -butyrolactones and γ -butyrolactams, which are synthetically useful building blocks in organic synthesis.

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Introduction

Five-membered ring heterocycles are an important class of compounds. In particular, α , β -unsaturated γ -butyrolactones (or butenolides) and α,β -unsaturated γ -butyrolactams are among advanced synthetic intermediates that find major applications in medicinal chemistry and organic synthesis. They have been used in a range of reactions, such as vinylogous additions to electrophiles,¹ conjugate additions,² and pericyclic reactions.³ The ubiquity of α,β -unsaturated γ -butyrolactones and α,β -unsaturated γ -butyrolactams has been highlighted in naturally occurring compounds, for example, rollicosin,^{4c} (-)-incrustoporine,4i spirofragilide, labdadienolide A, and (+)-erysotramidine that possess important biological properties including antibiotic, antitumor, anti-inflammatory, and anti-HIV activities (Fig. 1).⁴ Not surprisingly, the development of efficient synthetic routes for the preparation of α , β -unsaturated y-butyrolactones and y-butyrolactams has been an area of long-standing importance to synthetic chemists.⁵

Among several synthetic routes, the oxidative decarboxylation of aliphatic carboxylic acids is an important and useful conversion to access alkenes. The existing methods usually require $Pb(OAc)_4/Cu(OAc)_2$ for mediating oxidative elimination



Fig. 1 Natural compounds containing α,β -unsaturated γ -butyrolactone and γ -butyrolactam units.

of the corresponding radical intermediates.⁶ Recently, catalytic decarbonylative or decarboxylative elimination of carboxylic acids to yield alkenes has also received much attention.⁷ In modern organic synthesis, metal-free reactions which avoid the use of heavy metals have been of particular interest. As a consequence, the conversion of carboxylic acids to alkenes *via* a simple consecutive decarboxylative halogenation (halodecarboxylation)⁸ followed by dehydrohalogenation leading to the synthesis of α , β -unsaturated γ -butyrolactones and γ -butyrolactams was investigated. On the basis of our recent report on



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Scheme 1 Proposed synthetic scheme for the synthesis of $\alpha\beta$ -unsaturated γ -butyrolactones and γ -butyrolactams from paraconic acids and β -carboxyl- γ -butyrolactams.

decarboxylative fluorination,⁹ it was envisaged that paraconic acids and β-carboxyl-γ-butyrolactams 1 would undergo decarboxylative iodination to give β -iodo- γ -butyrolactones 2 which, after dehydroiodination, would provide α , β -unsaturated γ -butyrolactones and γ -butyrolactams 3 (Scheme 1). Inspired by the work reported by Gandelman and co-workers on decarboxylative iodination, employing commercially available 1,3diiodo-5,5-dimethylhydantoin (DIH)¹⁰ as an initiator and iodine source,¹¹ we describe herein a convenient synthetic approach to α,β -unsaturated γ -butyrolactones and γ -butyrolactams via decarboxylative iodination of paraconic acids and β-carboxyl-γ-butyrolactams using DIH under irradiative conditions followed by base-promoted dehydroiodination (Scheme 1). Notably, DIH has been extensively explored by Togo and others.¹² However, to the best of our knowledge, the use of DIH for decarboxylative iodination followed by base mediated dehydroiodination has never been reported.

Results and discussion

Our study commenced with the optimization of reaction conditions using paraconic acid 1a¹³ as the model substrate. A standard screening of solvents, reaction times and the stoichiometry of reagents was performed (Table 1). Under thermal conditions, 1a was reacted with DIH (1 equiv.) in refluxing ClCH₂CH₂Cl for 8 h to give β -iodo- γ -butyrolactone 2a together with a trace amount of α , β -unsaturated γ -butyrolactone 3a as observed by TLC and ¹H NMR analyses of the crude mixture (68% conversion based on the recovery of 1a). Without purification, the crude mixture was treated with Et₃N (2 equiv.) in CH₂Cl₂ for 3 h leading to 3a in 67% yield after chromatographic purification (Table 1, entry 1). We were pleased to observe an improved conversion of 1a to a mixture of 2a and 3a (2a: 3a, 1: 2, 96% conversion based on the recovery of 1a) when the reaction was carried out in refluxing ClCH2CH2CH2CH under irradiation of a tungsten lamp (100 W) (Table 1, entry 2). Compound 3a was obtained in 95% yield after treatment of the crude mixture with Et₃N. The Et₃N-mediated HI elimination was proved necessary since a prolonged reaction time (from 8 h to 13 h) led to the incomplete conversion of 1a to 3a; a mixture of 2a and 3a was obtained (2a: 3a, 1: 8, 96% conversion based on the recovery of 1a). Gratifyingly, it was found that decarboxylative-iodination of 1a with DIH (1 equiv.) in refluxing ClCH₂CH₂Cl under photolysis reached completion after 30 min and gave 2a in quantitative yield. Subsequent

Table 1 Optimization of the reaction conditions for the decarboxylative iodination followed by Et_3N -promoted dehydroiodination of $1a^a$



^{*a*} Unless stated otherwise, the reaction was carried out using **1a** (0.5 mmol) in a refluxing solvent (2 mL) under irradiation with a 100 W tungsten lamp. ^{*b*} Yield of analytically pure **3a** after column chromatography (SiO₂). ^{*c*} The reaction was carried out at reflux without irradiation.

treatment of the crude product with Et_3N cleanly provided **3a** in 92% isolated yield (Table 1, entry 3). The yield was drastically lower when DIH was employed in a lesser amount (0.5 equiv.) (Table 1, entry 4). Finally, other simple organic solvents including CH₂Cl₂, EtOAc and MeCN were screened and ClCH₂CH₂Cl was proved to be the optimal solvent (Table 1, entry 3 *vs.* entries 5–7). At this stage, the optimal reaction conditions for decarboxylative iodination followed by base-promoted dehydroiodination of paraconic acids **1** were established (Table 1, entry 3).¹⁴

With the optimized reaction conditions in hand, we next examined the substrate scope and the results are summarized in Table 2. In general, the reactions of paraconic acids **1a–1m**, containing mono- and dialkyl groups at the γ -position, smoothly underwent DIH-mediated decarboxylative iodination to yield the corresponding β -iodo- γ -butyrolactones **2a–2m**. Following Et₃N promoted HI elimination of the primarily formed β -iodo- γ -butyrolactones **2a–2m** yielded their corresponding α , β -unsaturated γ -butyrolactones **3a–3m** in good yields (81–92% yields) (Table 2, entries 1–13). Among these, the trifluoromethylated butenolide analogue¹⁵ **3k** could also be synthesized (82% yield) (Table 2, entry 11).

For paraconic acid **1n**, bearing a phenyl substituent at the γ -position, treatment of its β -iodo- γ -butyrolactone **2n** with Et₃N according to the standard procedure led to a complex mixture of unidentified products and no trace of **3n** could be detected. These observed results may result from the polymerization of product **3n** under basic conditions. Fortunately, it was found

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Table 2Synthesis of α,β -unsaturated γ -butyrolactones and γ -butyrolactams via DIH-mediated decarboxylative iodination followed by dehydroiodi-nation of paraconic acids and β -carboxyl- γ -butyrolactams





^{*a*} Isolated yield. ^{*b*} Compounds 3 were obtained after purification of 2 by column chromatography (SiO₂, see the Experimental section). The ratio of products was determined by ¹H NMR analysis. ^{*c*} A mixture of diastereomers (dr = 8:1; 3,4-*cis*-4,5-*trans*/3,4-*trans*-4,5-*trans*). ^{*d*} A single isomer (3,4-*cis*-4,5-*trans*).

that compound 2n readily underwent dehydroiodination during column chromatography (SiO₂, see the Experimental section) to afford an inseparable mixture of the expected α,β -unsaturated γ -butyrolactone **3n** together with its deconjugated isomer 3n' in 62% combined yield $(3n:3n' = 2:1, {}^{1}H)$ NMR analysis) (Table 2, entry 14). Similarly, the reaction of paraconic acids 10 and 1p, containing 4-MePh- and 4-ClPhgroups at the γ -positions, also provided the expected butenolides 30 and 3p together with their deconjugated butenolides **3o'** and **3p'** in 61% (**3o**: **3o'**; 10:1) and 74% (**3p**: **3p'**; 1:4) yields, respectively (Table 2, entries 15 and 16). The formation of the deconjugated butenolides 3n', 3o', and 3p' was presumably through the corresponding dienols generated from the enolization of α , β -unsaturated γ -butyrolactones **3n**, **3o**, and **3p**, respectively. Notably, deconjugated butenolides have been widely used in several enantioselective vinylogous additions to various Michael acceptors.¹ In contrast to 1p, α -methylated paraconic acid 1q underwent the reaction to give only the desired α,β -unsaturated γ -butyrolactone **3q** as the sole product in 45% yield (Table 2, entry 17).

As an extension of the present protocol, DIH-promoted decarboxylative iodination followed by Et_3N -promoted dehydroiodination can also be applied to the β -carboxyl- γ -butyrolactams leading to α , β -unsaturated γ -butyrolactams in moderate to good yields. Thus, phenyl- and benzyl protected β -carboxyl amides **1r** and **1s** yielded α , β -unsaturated γ -butyrolactams **3r** and **3s** in 46% and 75% yields, respectively (Table 2, entries 18 and 19). Similarly, β -carboxyl amide **1t**, bearing a phenyl substituent at the γ -position gave a mixture of α , β -unsaturated γ -butyrolactam **3t** and its deconjugated isomer **3t'** (76% yield, **3t** : **3t'** = 3 : 2, ¹H NMR analysis) (Table 2, entry 20). The reaction of **1u** and **1v**, having an electron donating group, gave the desired α , β -unsaturated γ -butyrolactams **3u** (74% yield) and **3v** (62% yield), respectively, each as the sole product (Table 2, entries 21 and 22). α -Methylated β -carboxyl- γ -butyrolactams **1w** and **1x** also gave the corresponding α , β -unsaturated γ -butyrolactams **3w** and **3x** in 33% and 45% yields, respectively (Table 2, entries 23 and 24).

A radical-type mechanism for decarboxylative iodination^{11*a*} of **1** was proposed as shown in Scheme 2. Acyl hypoiodite **A** was formed from the reaction of **1** with DIH. The homolytic cleavage of the O–I bond of the intermediate **A** followed by decarboxylation of the resulting carboxyl radical **B** gave the carbon radical **C** which was subsequently trapped by an iodine to give β -iodo- γ -butyrolactones or lactams **2**. Finally, dehydroiodination of **2** provided the desired products **3**.



Scheme 2 Proposed mechanism for decarboxylative iodination-dehydroiodination of paraconic acids and β -carboxyl- γ -butyrolactams.

Conclusions

In conclusion, we have successfully developed a convenient synthetic approach to α , β -unsaturated γ -butyrolactones and γ -butyrolactams from readily available paraconic acids and β -carboxyl- γ -butyrolactams. Decarboxylative iodination of the carboxylic acid precursors using commercially available 1,3-diiodo-5,5-dimethylhydantoin (DIH) under irradiation followed by dehydroiodination gave good yields of the desired α , β -unsaturated γ -butyrolactone and γ -butyrolactam products, which are synthetically useful building blocks in organic synthesis and valuable core structures found in biologically active natural compounds.

Experimental

General information

The ¹H NMR spectra were recorded on a Bruker-400 (400 MHz) or a Bruker-500 (500 MHz) spectrometer in CDCl₃ using tetramethylsilane as an internal standard. The ¹³C NMR spectra were recorded on a Bruker-400 (100 MHz) or a Bruker-500 (125 MHz) spectrometer in CDCl₃ using solvent peaks as an internal standard. The IR spectra were recorded on an ALPHA FTIR spectrometer. The mass spectra were recorded using a Thermo Finnigan Polaris Q mass spectrometer. The high resolution mass spectra were recorded on a HR-TOF-MS Micromass model VQ-TOF2 mass spectrometer. Melting points were recorded using a M-565 Büchi melting point apparatus and are uncorrected. Column chromatography was performed using Merck silica gel 60 (0.063-0.200 mm) (Art 7734). 1,2-Dichloroethane (ClCH₂CH₂Cl) and dichloromethane (CH₂Cl₂) were distilled over calcium hydride and stored over activated molecular sieves (4 Å). Other common solvents [acetone, CH_2Cl_2 , hexanes, and ethyl acetate (EtOAc)] were distilled before use. Compounds 1 were synthesized according to the literature procedures.13

General procedure

A mixture of paraconic acid or β -carboxyl- γ -butyrolactam 1 (0.5 mmol) and DIH (1 equiv.) in 1,2-dichloroethane (2 mL) was heated to reflux under irradiation by a tungsten-filament lamp (100 W) for 30 minutes. The reaction mixture was then cooled down to room temperature, washed with saturated Na₂S₂O₃ (20 mL), and extracted with CH_2Cl_2 (3 × 10 mL). The combined organic layers were washed with saturated NaHCO3 and brine, dried over anhydrous Na₂SO₄, and concentrated to give β-iodo- γ -butyrolactones 2. The obtained crude compounds 2a-2m and 2r-2x were dissolved in dry CH₂Cl₂ (2 mL) and treated with Et_3N (2 equiv.). The progress of the reaction was monitored by TLC. After the complete consumption of the starting material, the reaction mixture was passed through a short column (SiO_2 , CH₂Cl₂) to give the products 3a-3m and 3r-3x, respectively. Without treatment with Et₃N, the crude compounds 2n-2p were converted into 3n-3p and 3n'-3p' during purification by column chromatography (SiO₂).

1-Oxaspiro[4.5]**dec-3-en-2-one** (3a). Pale yellow oil; $R_{\rm f} = 0.44$ (CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.47$ (d, J = 5.6 Hz, 1H, CH), 6.01 (d, J = 5.6 Hz, 1H, CH), 1.85–1.32 (m, 10H, 5 × CH₂). ¹³C NMR (100 MHz, CDCl₃): $\delta = 172.5$ (C), 160.6 (CH), 120.1 (CH), 88.5 (C), 34.6 (2 × CH₂), 24.6 (CH₂), 22.4 (2 × CH₂). IR (neat): $\tilde{\nu} = 1744$, 1602, 1448, 1209, 1137, 969, 816. MS: m/z (%) = 153 (4) [M + H]⁺, 152 (6) [M]⁺, 149 (100), 135 (12), 121 (16), 95 (38), 81 (32), 57 (30). HRMS (ESI-TOF): calcd for C₉H₁₂O₂Na [M + Na]⁺ 175.0735; found 175.0733.

1-Oxaspiro[**4.4**]**non-3-en-2-one (3b).** Pale yellow oil; $R_{\rm f} = 0.44$ (CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.39$ (d, J = 5.6 Hz, 1H, CH), 6.01 (d, J = 5.6 Hz, 1H, CH), 2.08–1.74 (m, 8H, 4 × CH₂). ¹³C NMR (100 MHz, CDCl₃): $\delta = 172.5$ (C), 159.0 (CH), 120.2 (CH), 96.8 (C), 36.8 (2 × CH₂), 24.6 (2 × CH₂). IR (neat): $\tilde{\nu} = 1738$, 1600, 1434, 1153, 934, 817. MS: m/z (%) = 139 (8) [M + H]⁺, 138 (10) [M]⁺, 134 (30), 126 (100), 112 (63), 91 (72), 81 (93), 67 (60), 55 (75). HRMS (ESI-TOF): calcd for C₈H₁₀O₂Na [M + Na]⁺ 161.0578; found 161.0580.

1-Oxaspiro[**4.6**]**undec-3-en-2-one** (**3c**). Pale yellow oil; $R_f = 0.44$ (CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.48$ (d, J = 5.6 Hz, 1H, CH), 5.95 (d, J = 5.6 Hz, 1H, CH), 1.96–1.49 (m, 12H, $6 \times CH_2$). ¹³C NMR (100 MHz, CDCl₃): $\delta = 172.6$ (C), 161.5 (CH), 119.1 (CH), 92.0 (C), 37.6 ($2 \times CH_2$), 28.9 ($2 \times CH_2$), 22.7 ($2 \times CH_2$). IR (neat): $\tilde{\nu} = 1744$, 1604, 1460, 1240, 1162, 939, 815. MS: m/z (%) = 126 (62), 91 (100), 79 (89), 66 (51), 55 (45). HRMS (ESI-TOF): calcd for C₁₀H₁₄O₂Na [M + Na]⁺ 189.0891; found 189.0892.

1-Oxaspiro[4.7]**dodec-3-en-2-one** (**3d**). Pale yellow oil; $R_f = 0.40 \text{ (CH}_2\text{Cl}_2$). ¹H NMR (400 MHz, CDCl}3): $\delta = 7.53 \text{ (d, } J = 5.6 \text{ Hz}, 1\text{H}, CH$), 5.99 (d, J = 5.6 Hz, 1H, CH), 2.04–1.43 (m, 14H, 7 × CH₂). ¹³C NMR (100 MHz, CDCl}3): $\delta = 172.5 \text{ (C)}, 161.1 \text{ (CH)}, 120.0 \text{ (CH)}, 92.1 \text{ (CH)}, 33.4 (2 × CH_2), 27.9 (2 × CH_2), 24.4 (CH_2), 22.4 (2 × CH_2). IR (neat): <math>\tilde{\nu} = 1745$, 1600, 1465, 1161, 1023, 818. MS: m/z (%) = 180 (11) [M]⁺, 133 (36), 109 (52), 95 (78), 91 (84), 81 (89), 79 (100), 67 (55), 55 (65). HRMS (ESI-TOF): calcd for C₁₁H₁₆O₂Na [M + Na]⁺ 203.1048; found 203.1049.

5,5-Dimethylfuran-2(5*H***)-one (3e).** Pale yellow oil; $R_{\rm f} = 0.33$ (CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.41$ (d, J = 5.6 Hz, 1H, CH), 5.99 (d, J = 5.6 Hz, 1H, CH), 1.49 (s, 6H, $2 \times CH_3$). ¹³C NMR (100 MHz, CDCl₃): $\delta = 172.4$ (C), 161.2 (CH), 119.9 (CH), 86.6 (C), 25.3 ($2 \times CH_3$). IR (neat): $\tilde{\nu} = 1732$, 1461, 1258, 1117, 1016, 795. MS: m/z (%) = 113 (40) [M + H]⁺, 112 (11) [M]⁺, 77 (87), 51 (39). HRMS (ESI-TOF): calcd for C₆H₈O₂Na [M + Na]⁺ 135.0422; found 135.0423.

5,5-Diethylfuran-2(5*H***)-one (3f).** Pale yellow oil; $R_f = 0.44$ (CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.29$ (d, J = 5.7 Hz, 1H, CH), 6.08 (d, J = 5.7 Hz, 1H, CH), 1.88 (dq, $J_2 = 7.3$ Hz, $J_1 = 14.6$ Hz, 2H, CH₂), 1.77 (dq, $J_2 = 7.3$ Hz, $J_1 = 14.6$ Hz, 2H, CH₂), 0.86 (t, J = 7.3 Hz, 6H, 2 × CH₃). ¹³C NMR (100 MHz, CDCl₃): $\delta = 172.9$ (C), 158.9 (CH), 121.6 (CH), 92.0 (C), 29.5 (2 × CH₂), 7.7 (2 × CH₃). IR (neat): $\tilde{\nu} = 1750$, 1603, 1460, 1224, 1133, 950, 820. MS: m/z (%) = 140 (19) [M]⁺, 133 (47), 126 (89), 95 (75), 81 (100), 67 (88), 55 (81). HRMS (ESI-TOF): calcd for C₈H₁₂O₂Na [M + Na]⁺ 163.0735; found 163.0734.

5-Ethyl-5-methylfuran-2(5*H***)-one (3g).** Pale yellow oil; $R_f = 0.40$ (CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.36$ (d, J = 5.6 Hz, 1H, C*H*), 6.03 (d, J = 5.6 Hz, 1H, C*H*), 1.86 (dq, $J_2 = 7.3$ Hz, $J_1 = 14.6$ Hz, 1H, CH*H*), 1.76 (dq, $J_2 = 7.3$ Hz, $J_1 = 14.6$ Hz, 1H, CH*H*), 1.76 (dq, $J_2 = 7.3$ Hz, $J_1 = 14.6$ Hz, 1H, CH*H*), 1.47 (s, 3H, CH₃), 0.88 (t, J = 7.3 Hz, 3H, CH₃). ¹³C NMR (100 MHz, CDCl₃): $\delta = 172.6$ (C), 160.2 (CH), 120.7 (CH), 89.3 (C), 31.2 (CH₂), 23.5 (CH₃), 8.0 (CH₃). IR (neat): $\tilde{\nu} = 1745$, 1603, 1459, 1235, 1130, 950, 819. MS: m/z (%) = 127 (21) [M + H]⁺, 126 (100) [M]⁺, 112 (48), 95 (76), 81 (90), 67 (78), 55 (84). HRMS (ESI-TOF): calcd for C₇H₁₀O₂Na [M + Na]⁺ 149.0578; found 149.0579.

5-Isopropyl-5-methylfuran-2(5*H*)**-one** (3**h**). Pale yellow oil; $R_{\rm f} = 0.42$ (CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.34$ (d, J = 5.7 Hz, 1H, CH), 5.97 (d, J = 5.7 Hz, 1H, CH), 1.91 (hept, J = 6.9 Hz, 1H, CH), 1.37 (s, 3H, CH₃), 0.91 (d, J = 6.9 Hz, 3H, CH₃), 0.88 (d, J = 6.9 Hz, 3H, CH₃). ¹³C NMR (100 MHz, CDCl₃): $\delta = 172.7$ (C), 159.7 (CH), 120.8 (CH), 91.7 (C), 35.5 (CH), 21.3 (CH₃), 17.7 (CH₃), 17.2 (CH₃). IR (neat): $\tilde{\nu} = 1744$, 1603, 1457, 1249, 1120, 948, 819. MS: m/z (%) = 141 (28) [M + H]⁺, 140 (100) [M]⁺, 138 (22). HRMS (ESI-TOF): calcd for C₈H₁₂O₂Na [M + Na]⁺ 163.0735; found 163.0732.

5-Ethyl-5-phenylfuran-2(*5H*)**-one** (3i). Pale yellow oil; $R_f = 0.53$ (CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.63$ (d, J = 5.6 Hz, 1H, CH), 7.42–7.28 (m, 5H, ArH), 6.08 (d, J = 5.6 Hz, 1H, CH), 2.19 (dq, $J_2 = 7.3$ Hz, $J_1 = 14.6$ Hz, 1H, CHH), 2.03 (dq, $J_2 = 7.3$ Hz, $J_1 = 14.6$ Hz, 1H, CHH), 0.89 (t, J = 7.3 Hz, 3H, CH₃). ¹³C NMR (100 MHz, CDCl₃): $\delta = 172.4$ (C), 159.2 (CH), 138.8 (C), 128.8 (2 × CH), 128.1 (CH), 125.0 (2 × CH), 120.0 (CH), 91.9 (C), 32.7 (CH₂), 8.0 (CH₃). IR (neat): $\tilde{\nu} = 3085$, 3066, 1752, 1601, 1448, 1210, 1121, 1024, 816, 760, 698. MS: m/z (%) = 189 (7) [M + H]⁺, 188 (12) [M]⁺, 159 (100), 131 (53), 103 (63), 77 (58), 67 (14), 55 (9). HRMS (ESI-TOF): calcd for C₁₂H₁₂O₂Na [M + Na]⁺ 211.0735; found 211.0733.

5-Methyl-5-phenylfuran-2(5*H*)-one (3j). Pale yellow oil; R_f = 0.38 (CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃): δ = 7.57 (d, *J* = 5.6 Hz, 1H, CH), 7.35–7.29 (m, 4H, ArH), 7.29–7.22 (m, 1H, ArH), 5.99 (d, *J* = 5.6 Hz, 1H, CH), 1.76 (s, 3H, CH₃). ¹³C NMR (100 MHz, CDCl₃): δ = 172.4 (C), 160.4 (CH), 139.2 (C), 128.8

 $(2 \times CH)$, 128.3 (CH), 124.7 (2 × CH), 119.3 (CH), 88.9 (C), 26.3 (CH₃). IR (neat): $\tilde{\nu} = 3087$, 1747, 1447, 765, 697. MS: m/z (%) = 175 (4) [M + H]⁺, 174 (9) [M]⁺, 159 (39), 131 (100), 103 (78), 77 (51). HRMS (ESI-TOF): calcd for $C_{11}H_{10}O_2Na$ [M + Na]⁺ 197.0578; found 197.0578.

5-Phenyl-5-(trifluoromethyl)furan-2(5*H***)-one (3k). White solid; mp 70–72 °C (CH₂Cl₂); R_f = 0.61 (CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃): \delta = 7.82 (d, J = 5.7 Hz, 1H, C***H***), 7.61–7.52 (m, 2H, Ar***H***), 7.51–7.40 (m, 3H, Ar***H***), 6.39 (d, J = 5.7 Hz, 1H, C***H***). ¹³C NMR (100 MHz, CDCl₃): \delta = 169.7 (C), 150.8 (CH), 130.9 (C), 130.2 (CH), 130.0 (3 × CH), 126.6 (CH), 124.2 (CH), 122.4 (q, J = 280.3 Hz, CF₃), 87.4 (q, J = 32.2 Hz, C). ¹⁹F NMR (376 MHz, CDCl₃): \delta = -76.9 (s, 3 F). IR (neat): \tilde{\nu} = 3117, 1797, 1775, 1292, 1171, 761, 694. MS: m/z (%) = 228 (1) [M]⁺, 159 (100), 131 (48), 103 (43), 77 (26). HRMS (ESI-TOF): calcd for C₁₁H₇F₃O₂Na [M + Na]⁺ 251.0296; found 251.0296.**

5-Butylfuran-2(5*H***)-one (31).** Pale yellow oil; $R_{\rm f} = 0.44$ (CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.47$ (dd, $J_2 = 1.3$ Hz, $J_2 = 5.7$ Hz, 1H, CH), 6.11 (dd, $J_3 = 1.9$ Hz, $J_2 = 5.7$ Hz, 1H, CH), 5.08–5.01 (m, 1H, CH), 1.84–1.81 (m, 2H, CH₂), 1.50–1.30 (m, 4H, 2 × CH₂), 0.92 (t, J = 7.1 Hz, 3H, CH₃). ¹³C NMR (100 MHz, CDCl₃): $\delta = 173.2$ (C), 156.3 (CH), 121.5 (CH), 83.4 (C), 32.8 (CH₂), 27.0 (CH₂), 22.4 (CH₂), 13.8 (CH₃). IR (neat): $\tilde{\nu} = 1747$, 1601, 1466, 1162, 1024, 818. MS: m/z (%) = 140 (17) [M]⁺, 133 (40), 95 (90), 81 (100), 67 (89), 55 (98). HRMS (ESI-TOF): calcd for C₈H₁₂O₂Na [M + Na]⁺ 163.0735; found 163.0729.

5-Phenethylfuran-2(5*H***)-one (3m).** Pale yellow oil; $R_f = 0.48$ (CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.40$ (dd, $J_2 = 1.4$ Hz, $J_2 = 5.7$ Hz, 1H, CH), 7.34–7.27 (m, 2H, ArH), 7.25–7.17 (m, 3H, ArH), 6.11 (dd, $J_3 = 2.0$ Hz, $J_2 = 5.7$ Hz, 1H, CH), 5.05–4.98 (m, 1H, CH), 2.90–2.72 (m, 2H, CH₂), 2.14–2.03 (m, 1H, CHH), 2.00–1.88 (m, 1H, CHH). ¹³C NMR (100 MHz, CDCl₃): $\delta = 173.0$ (C), 156.1 (CH), 140.2 (C), 128.6 (2 × CH), 128.5 (2 × CH), 126.4 (CH), 121.6 (CH), 82.4 (C), 34.9 (CH₂), 31.3 (CH₂). IR (neat): $\tilde{\nu} = 3085$, 3062, 1744, 1601, 1454, 1159, 1102, 815, 752, 699. MS: m/z (%) = 189 (9) [M + H]⁺, 188 (10) [M]⁺, 178 (14), 105 (24), 97 (38), 91 (100), 77 (34), 65 (24), 55 (16). HRMS (ESI-TOF): calcd for C₁₂H₁₂O₂Na [M + Na]⁺ 211.0735; found 211.0738.

5-Phenylfuran-2(5H)-one (3n) and 5-phenylfuran-2(3H)-one (3n'). A 2:1 mixture of 3n and 3n' was obtained as an orange semi-solid after column chromatography (SiO₂, 20% EtOAc in hexanes). 3n; $R_f = 0.20$ (20% EtOAc in hexanes), 3n'; $R_f = 0.38$ (20% EtOAc in hexanes); ¹H NMR (400 MHz, CDCl₃, integrated equally for both isomers): δ = 7.56–7.50 (m, 2H, ArH of 3n'), 7.46 (dd, $J_2 = 1.6$ Hz, $J_2 = 5.6$ Hz, 1H, CH of 3n), 7.37–7.26 (each m, 3H, ArH of 3n and 3H, ArH of 3n'), 7.23-7.16 (m, 2H, ArH of 3n), 6.16 (dd, $J_3 = 2.1$ Hz, $J_2 = 5.6$ Hz, 1H, CH of 3n), 5.97–5.92 (m, 1H, CH of 3n), 5.72 (t, J = 2.7 Hz, 1H, CH of 3n'), 3.35 (d, J = 2.7 Hz, 2H, CH_2 of 3n'). ¹³C NMR (100 MHz, CDCl₃): δ = 175.9 (C of 3n'), 173.1 (C of 3n), 155.8 (CH of 3n), 153.9 (C of 3n'), 134.2 (1C of 3n and 1C of 3n'), 129.3 (CH of 3n), 129.6 (CH of 3n'), 128.6 (2 × CH of 3n'), 129.0 (2 × CH of 3n), 126.5 (2 × CH of 3n), 124.7 (2 × CH of 3n'), 120.9 (CH of 3n), 97.6 (CH of 3n'), 84.3 (CH of 3n), 34.6 (CH₂ of 3n'). IR (neat): $\tilde{\nu}$ = 3090, 3034, 1746, 1454, 1154, 1028, 760, 697. MS: m/z (%) = 161 (15) [M + H]⁺, 160 (87) [M]⁺, 131 (99), 103 (70), 77

(100). HRMS (ESI-TOF): calcd for $C_{10}H_8O_2Na [M + Na]^+$ 183.0422; found 183.0430.

5-(p-Tolyl)furan-2(5H)-one (30) and 5-(p-tolyl)furan-2(3H)one (30'). A 10:1 mixture of 30 and 30' was obtained as an orange solid after column chromatography (SiO₂, 20% EtOAc in hexanes). **30**; $R_f = 0.33$ (20% EtOAc in hexanes), **30**'; $R_f =$ 0.47 (20% EtOAc in hexanes); mp 49-52 °C (CH₂Cl₂). ¹H NMR (500 MHz, CDCl₃, integrated equally for both isomers): δ = 7.49 (dd, $J_2 = 1.6$ Hz, $J_2 = 5.6$ Hz, 1H, CH of 3o), 7.20 (each d, J = 8.1 Hz, 2H, ArH of **30** and 2H, ArH of **30**'), 7.14 (each d, J = 8.1 Hz, 2H, ArH of **30** and 2H, ArH of **30**′), 6.20 (dd, J₃ = 2.1 Hz, J₂ = 5.6 Hz, 1H, CH of **30**), 5.97 (br s, 1H, CH of **30**), 5.70 (t, J = 2.7 Hz, 1H, CH of 3o'), 3.39 (d, J = 2.7 Hz, 2H, CH₂ of 3o'), 2.37 (s, 3H, CH₃ of 3o'), 2.36 (s, 3H, CH₃ of 3o). ¹³C NMR (125 MHz, CDCl₃): δ = 173.1 (C of **30**), 173.0 (C of **30**'), 155.8 (CH of **30**), 155.7 (C of 3o'), 139.4 (C of 3o), 138.9 (C of 3o'), 131.3 (C of 30'), 131.2 (C of 30), 129.7 (2 × CH of 30), 129.4 (2 × CH of 30'), 126.5 (2 × CH of 30), 124.7 (2 × CH of 30'), 121.0 (CH of 30), 96.6 (CH of 30'), 84.4 (CH of 30), 34.6 (CH₂ of 30'), 21.2 (CH₃ of **30**), 21.1 (CH₃ of **30**'). IR (neat): $\tilde{\nu}$ = 3080, 3038, 1795, 1748, 1512, 1306, 1158, 819. MS: m/z (%) = 175 (10) $[M + H]^+$, 174 (52) [M]⁺, 145 (100), 115 (94), 91 (80), 65 (34). HRMS (ESI-TOF): calcd for $C_{11}H_{10}O_2Na [M + Na]^+$ 197.0578; found 197.0572.

5-(4-Chlorophenyl)furan-2(5H)-one (3p) and 5-(4-chlorophenyl)furan-2(3H)-one (3p'). A 1:4 mixture of 3p and 3p' was obtained as an orange semi-solid after column chromatography (SiO₂, 20% EtOAc in hexanes). **3p**; $R_f = 0.16$ (20% EtOAc in hexanes), 3p'; $R_f = 0.33$ (20% EtOAc in hexanes); ¹H NMR (400 MHz, CDCl₃, integrated equally for both isomers): δ = 7.47 (d, J = 8.6 Hz, 2H, ArH of 3p'), 7.44 (dd, $J_2 = 1.6$ Hz, $J_2 =$ 5.6 Hz, 1H, CH of 3p), 7.31 (each d, J = 8.6 Hz, 2H, ArH of 3p and 2H, ArH of 3p'), 7.14 (d, J = 8.4 Hz, 2H, ArH of 3p), 6.18 (dd, *J*₃ = 2.1 Hz, *J*₂ = 5.6 Hz, 1H, CH of **3p**), 5.92 (br s, 1H, CH of 3p), 5.72 (t, J = 2.7 Hz, 1H, CH of 3p'), 3.36 (d, J = 2.7 Hz, 2H, CH₂ of 3p'). ¹³C NMR (100 MHz, CDCl₃): δ = 175.5 (C of 3p'), 172.7 (C of 3p), 155.3 (CH of 3p), 153.0 (C of 3p'), 135.5 (C of 3p'), 135.3 (C of 3p), 132.8 (C of 3p), 129.3 (2 × CH of 3p), 129.0 (2 × CH of 3p'), 127.8 (2 × CH of 3p), 126.8 (C of 3p'), 126.0 (2 × CH of 3p'), 121.3 (CH of 3p), 98.2 (CH of 3p'), 83.5 (CH of **3p**), 34.6 (CH₂ of **3p**'). IR (neat): $\tilde{\nu} = 3031$, 1746, 1682, 1514, 1157, 812. MS: m/z (%) = 195 (13) $[M + H]^+$, 194 (86) $[M]^+$, 159 (100), 139 (77), 103 (53), 74 (58). HRMS (ESI-TOF): calcd for $C_{10}H_7ClO_2Na [M + Na]^+$ 217.0032; found 217.0028.

5-(4-Chlorophenyl)-3-methylfuran-2(5*H***)-one (3q). A white solid was obtained after column chromatography (SiO₂, 50% CH₂Cl₂ in hexanes). R_{\rm f} = 0.29 (20% EtOAc in hexanes); mp 83–86 °C (CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃): \delta = 7.29 (d, J = 8.5 Hz, 2H, Ar***H***), 7.14 (d, J = 8.4 Hz, 2H, Ar***H***), 7.03 (dd, J_3 = 1.7 Hz, J_2 = 1.7 Hz, 1H, C***H***), 5.77 (dd, J_4 = 1.7 Hz, J_2 = 1.7 Hz, 1H, C***H***), 5.77 (dd, J_4 = 1.7 Hz, J_2 = 1.7 Hz, 1H, C***H***), 1.93 (dd, J_4 = 1.7 Hz, J_3 = 1.7 Hz, 3H, C***H***₃). ¹³C NMR (400 MHz, CDCl₃): \delta = 174.0 (C), 147.8 (CH), 135.0 (C), 133.6 (C), 129.9 (C), 129.2 (2 × CH), 127.8 (2 × CH), 81.3 (CH), 10.6 (CH₃). IR (neat): \tilde{\nu} = 3064, 1743, 1492, 1042, 823. MS: m/z (%) = 210 (11) [M + H]⁺, 179 (100), 150 (46), 122 (33), 96 (40), 82 (76), 55 (56). HRMS (ESI-TOF): calcd for C₁₁H₉ClO₂Na [M + Na]⁺ 231.0189; found 231.0182.**

1-Phenyl-1*H***-pyrrol-2(5***H***)-one (3r**). Pale yellow solid; mp 81–85 °C (CH₂Cl₂); $R_{\rm f} = 0.24$ (10% EtOAc in hexanes). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.68-7.61$ (m, 2H, Ar*H*), 7.36–7.28 (m, 2H, Ar*H*), 7.12 (dt, $J_2 = 1.9$ Hz, $J_2 = 6.0$ Hz, 1H, C*H*), 7.10–7.04 (m, 1H, Ar*H*), 6.22 (dt, $J_3 = 1.9$ Hz, $J_2 = 6.0$ Hz, 1H, C*H*), 4.39 (dd, $J_3 = 1.9$ Hz, $J_2 = 1.9$ Hz, 2H, C*H*₂). ¹³C NMR (100 MHz, CDCl₃): $\delta = 170.2$ (C), 142.2 (CH), 139.0 (C), 129.3 (CH), 129.1 (2 × CH), 124.2 (CH), 118.9 (2 × CH), 53.2 (CH₂). IR (neat): $\tilde{\nu} = 3350, 3079, 3043, 1681, 1596, 753, 689.$ MS: m/z (%) = 160 (6) [M + H]⁺, 159 (38) [M]⁺, 130 (100), 77 (33). HRMS (ESI-TOF): calcd for C₁₀H₉NONa [M + Na]⁺ 182.0582; found 182.0582.

1-Benzyl-1*H***-pyrrol-2(5***H***)-one (3s).** Pale yellow oil; $R_{\rm f} = 0.22$ (20% EtOAc in hexanes). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.37-7.19$ (m, 5H, Ar*H*), 7.06 (dt, $J_2 = 1.7$ Hz, $J_2 = 6.0$ Hz, 1H, C*H*), 6.23 (dt, $J_3 = 1.8$ Hz, $J_2 = 6.0$ Hz, 1H, C*H*), 4.64 (s, 2H, CH₂Ph), 3.88 (br s, 2H, CH₂). ¹³C NMR (100 MHz, CDCl₃): $\delta = 171.4$ (C), 142.8 (CH), 137.2 (C), 128.7 (2 × CH), 127.9 (3 × CH), 127.5 (CH), 52.2 (CH₂), 45.9 (CH₂). IR (neat): $\tilde{\nu} = 3368, 3063, 3030, 1659, 1450, 717, 699.$ MS: m/z (%) = 174 (6) [M + H]⁺, 173 (50) [M]⁺, 172 (72), 106 (40), 91 (100), 77 (27), 65 (34). HRMS (ESI-TOF): calcd for C₁₁H₁₁NONa [M + Na]⁺ 196.0738; found 196.0734.

1-Benzyl-5-phenyl-1H-pyrrol-2(5H)-one (3t) and 1-benzyl-5-phenyl-1H-pyrrol-2(3H)-one (3t'). A 3:2 mixture of 3t and 3t' was obtained as a pale brown viscous oil. 3t; $R_f = 0.36$ (30%) acetone in hexanes), 3t'; $R_f = 0.42$ (30% acetone in hexanes); ¹H NMR (400 MHz, CDCl₃, integrated equally for both isomers): δ = 7.42–7.24 (each, m, 5H, ArH of 3t and 5H, ArH of 3t'), 7.23-7.14 (each, m, 1H, ArH of 3t and 3H, ArH of 3t'), 7.14–7.09 (m, 2H, ArH of 3t), 7.09–7.04 (m, 2H, ArH of 3t), 6.99–6.94 (m, 2H, ArH of 3t'), 7.01 (dd, J₂ = 1.6 Hz, J₂ = 5.8 Hz, 1H, CH of 3t), 6.28 (dd, J₃ = 1.6 Hz, J₂ = 5.8 Hz, 1H, CH of 3t), 5.27 (t, J = 2.6 Hz, 1H, CH of 3t'), 5.19 (d, J = 15.0 Hz, 1H, CHHPh of 3t), 4.88 (br s, 1H, CH of 3t), 4.68 (s, 2H, CH₂Ph of 3t'), 3.60 (d, J = 15.0 Hz, 1H, CHHPh of 3t), 3.27 (d, J = 2.6 Hz, 2H, CH₂ of 3t'). ¹³C NMR (100 MHz, CDCl₃): δ = 178.4 (C of 3t'), 171.3 (C of 3t), 148.1 (CH of 3t), 146.2 (C of 3t'), 137.5 (C of 3t'), 137.3 (C of 3t), 134.4 (C of 3t), 131.7 (C of 3t'), 129.2 (2 × CH of 3t), 128.9 (CH of 3t'), 128.8 (2 × CH of 3t'), 128.7 (2 × CH of 3t), 128.4 (4 × CH of 3t'), 128.3 (CH of 3t), 128.2 (2 × CH of **3t**), 127.5 (2 × CH of **3t** and 2 × CH of **3t**'), 127.2 (CH of 3t), 127.1 (CH of 3t'), 126.2 (CH of 3t), 102.4 (CH of 3t'), 65.8 (CH of 3t), 44.3 (CH₂ of 3t'), 43.4 (CH₂ of 3t), 37.3 (CH₂ of 3t'). IR (neat): $\tilde{\nu}$ = 3306, 3061, 3029, 1677, 1453, 843, 768, 698. MS: m/z (%) = 250 (4) [M + H]⁺, 249 (22) [M]⁺, 144 (100), 115 (43), 91 (18), 77 (8). HRMS (ESI-TOF): calcd for $C_{17}H_{15}NONa [M + Na]^{+}$ 272.1051; found 272.1053.

1-Benzyl-5-(*p***-tolyl)-1***H***-pyrrol-2(5***H***)-one** (**3u**). Pale brown viscous oil; $R_{\rm f} = 0.26$ (30% EtOAc in hexanes). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.26-7.15$ (m, 3H, Ar*H*), 7.10 (d, *J* = 7.8 Hz, 2H, Ar*H*), 7.05 (d, *J* = 6.6 Hz, 2H, Ar*H*), 6.91 (dd, *J*₂ = 1.5 Hz, *J*₂ = 5.9 Hz, 1H, C*H*), 6.88 (d, *J* = 8.0 Hz, 2H, Ar*H*), 6.19 (dd, *J*₃ = 1.7 Hz, *J*₂ = 5.9 Hz, 1H, C*H*), 5.10 (d, *J* = 14.9 Hz, 1H, CHHPh), 4.77 (br s, 1H, C*H*), 3.51 (d, *J* = 14.9 Hz, 1H, CHHPh) 2.29 (s, 3H, CH₃). ¹³C NMR (100 MHz, CDCl₃): $\delta = 171.3$ (C), 148.2 (CH), 138.7 (C), 137.4 (C), 131.2 (C), 129.8 (2 × CH), 128.6

(2 × CH), 128.2 (2 × CH), 127.4 (3 × CH), 126.1 (CH), 65.6 (CH), 43.3 (CH₂), 21.2 (CH₃). IR (neat): $\tilde{\nu}$ = 3306, 3029, 1665, 1437, 816, 756, 700. MS: m/z (%) = 264 (4) [M + H]⁺, 263 (12) [M]⁺, 158 (100), 129 (33), 115 (26), 91 (18), 77 (5). HRMS (ESI-TOF): calcd for C₁₈H₁₇NONa [M + Na]⁺ 286.1208; found 286.1215.

1-Benzyl-5-(4-methoxyphenyl)-1*H*-**pyrrol-2**(5*H*)-**one** (**3v**). Pale brown viscous oil; $R_{\rm f} = 0.32$ (40% EtOAc in hexanes). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.35-7.19$ (m, 1H of C*H* and 2H of Ar*H*), 7.15–7.08 (m, 2H, Ar*H*), 7.03–6.95 (m, 3H, Ar*H*), 6.92–6.86 (m, 2H, Ar*H*), 6.26 (dd, $J_3 = 1.7$ Hz, $J_2 = 5.8$ Hz, 1H, C*H*), 5.16 (d, J = 15.0 Hz, 1H, CH*H*Ph), 4.84 (br s, 1H, C*H*), 3.82 (s, 3H, OC*H*₃), 3.58 (d, J = 15.0 Hz, 1H, C*H*HPh). ¹³C NMR (100 MHz, CDCl₃): $\delta = 171.2$ (C), 159.9 (C), 148.3 (CH), 137.4 (C), 128.8 (2 × CH), 128.6 (2 × CH), 128.2 (2 × CH), 127.4 (CH), 126.1 (CH), 125.9 (C), 114.5 (2 × CH), 65.3 (CH), 53.3 (CH₃), 43.3 (CH₂). IR (neat): $\tilde{\nu} = 3306$, 3104, 3064, 3023, 1670, 1510, 1238, 1023, 830, 765, 699. MS: m/z (%) = 280 (16) [M + H]⁺, 279 (87) [M]⁺, 188 (67), 174 (100), 131 (26), 91 (47). HRMS (ESI-TOF): calcd for $C_{18}H_{17}NO_2Na$ [M + Na]⁺ 302.1157; found 302.1161.

3-Methyl-1-phenyl-1*H***-pyrrol-2(5***H***)-one** (**3w**). White solid; mp 81–83 °C (EtOAc/hexanes); $R_{\rm f} = 0.33$ (30% EtOAc in hexanes). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.78-7.71$ (m, 2H, Ar*H*), 7.44–7.33 (m, 2H, Ar*H*), 7.16–7.08 (m, 1H, Ar*H*), 6.82–6.76 (m, 1H, C*H*), 4.33–4.27 (m, 2H, C*H*₂), 1.99–1.93 (m, 3H, C*H*₃). ¹³C NMR (100 MHz, CDCl₃): $\delta = 170.8$ (C), 139.5 (C), 136.8 (C), 134.7 (CH), 129.0 (2 × CH), 123.9 (CH), 118.4 (2 × CH), 50.8 (CH₂), 11.3 (CH₃). IR (neat): $\tilde{\nu} = 3062$, 3041, 1671, 1595, 1496, 1380, 758, 690. MS: *m/z* (%) = 174 (14) [M + H]⁺, 173 (28) [M]⁺, 144 (65), 121 (29), 91 (40), 79 (54). HRMS (ESI-TOF): calcd for C₁₁H₁₁NONa [M + Na]⁺ 196.0738; found 196.0730.

1-Benzyl-5-(4-methoxyphenyl)-3-methyl-1*H***-pyrrol-2**(5*H*)**-one** (3**x**). Light brown viscous oil; $R_{\rm f} = 0.36$ (30% EtOAc in hexanes). ¹H NMR (400 MHz, CDCl₃): $\delta = 7.33-7.21$ (m, 3H, Ar*H*), 7.15–7.08 (m, 2H, Ar*H*), 6.99–6.93 (m, 2H, Ar*H*), 6.91–6.84 (m, 2H, Ar*H*), 6.63–6.58 (m, 1H, C*H*), 5.15 (d, *J* = 14.9 Hz, 1H, CH*H*Ph), 4.72–4.66 (m, 1H, C*H*), 3.82 (s, 3H, C*H*₃), 3.60 (d, *J* = 14.9 Hz, 1H, C*H*HPh) 2.02–1.98 (m, 3H, C*H*₃). ¹³C NMR (100 MHz, CDCl₃): $\delta = 171.9$ (C), 159.7 (C), 141.2 (CH), 137.6 (C), 133.9 (C), 128.7 (2 × CH), 128.6 (2 × CH), 128.2 (2 × CH), 127.3 (CH), 127.0 (C), 114.4 (2 × CH), 63.1 (CH), 55.3 (CH₃), 43.7 (CH₂), 11.2 (CH₃). IR (neat): $\tilde{\nu} = 3064$, 3031, 1678, 1608, 1510, 1242, 1030, 831, 705. MS: *m*/*z* (%) = 294 (33) [M + H]⁺, 293 (100) [M]⁺, 202 (77), 188 (61), 145 (24), 91 (37). HRMS (ESI-TOF): calcd for C₁₉H₁₉NO₂Na [M + Na]⁺ 316.1313; found 316.1337.

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