Synthesis of $(1 \rightarrow 4)$ -linked galacturonic acid trisaccharides, a proposed plant wound-hormone and a stereoisomer*

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ABSTRACT

Syntheses of *a*-GalA-(1→4)-*a*-GalA-(1→4)-GalA, a proposed plant wound-hormone, and its stereoisomer β -GalA-(1→4)-*a*-GalA-(1→4)-GalA are described. The key intermediates, *tert*-butyldiphenylsilyl *O*-(6-*O*-acetyl-2,3,4-tri-*O*-benzyl-*a*-D-galactopyranosyl)-(1→4)-*O*-(6-*O*-acetyl-2,3-di-*O*-benzyl-*a*-D-galactopyranosyl)-(1→4)-6-*O*-acetyl-2,3-di-*O*-benzyl- β -D-galactopyranosyl)-(1→4)-6-*O*-acetyl-2,3-di-*O*-benzyl- β -D-galactopyranosyl)-(1→4)-6-*O*-acetyl-2,3-di-*O*-benzyl- β -D-galactopyranosyl)-(1→4)-6-*O*-acetyl-2,3-di-*O*-benzyl- β -D-galactopyranoside, were prepared by stereoselective *a*-glycosylation of *tert*-butyldiphenylsilyl 6-*O*-acetyl-2,3-di-*O*-benzyl- β -D-galactopyranosyl)-(1→4)-6-*O*-acetyl-2,3-di-*O*-benzyl- β -D-galactopyranosyl)-(1→4)-6-*O*-acetyl-2,3-di-*O*-benzyl-*D*-galactopyranosyl)-(1→4)-6-*O*-acetyl-2,3-di-*O*-benzyl-D-galactopyranosyl)-(1→4)-6-*O*-acetyl-2,3-di-*O*-benzyl-D-galactopyranosyl)-(1→4)-6-*O*-acetyl-2,3-di-*O*-benzyl-D-galactopyranosyl)-(1→4)-6-*O*-acetyl-2,3-di-*O*-benzyl-D-galactopyranosyl)-(1→4)-6-*O*-acetyl-2,3-di-*O*-benzyl-D-galactopyranosyl)-(1→4)-6-*O*-acetyl-2,3-di-*O*-benzyl-D-galactopyranosyl)-(1→4)-6-*O*-acetyl-2,3-di-*O*-benzyl-D-galactopyranosyl)-(1→4)-6-*O*-acetyl-2,3-di-*O*-benzyl-D-galactopyranosyl)-(1→4)-6-*O*-acetyl-2,3-di-*O*-benzyl-D-galactopyranosyl]-D-galactopyranosyl]-D-galactopyranosyl]-D-galactopyranosyl]-D-galactopyranosyl]-D-galactopyranosyl]-D-galactopyranosyl]-D-galactopyranosyl]-D-galactopyranosyl]-D-galactopyranosyl]-D-galactopyranosyl]-D-galactopyranosyl]-D-galactopyranosyl]-D-galactopyranosyl]-D-galactopyrano

INTRODUCTION

Small fragments of plant cell-wall pectic polysaccharides, such as the galacturonic acid trisaccharide **1a**, have been proposed to behave as hormone-like substances which may induce a second messenger that would be systemically transported throughout the plant tissues in the tomato and induce the synthesis and accumulation of proteinase inhibitor I, one of the defence substances produced by plants against invading microbes or insects².

As part of a project on the synthesis of plant cell-wall glycans, we have achieved^{1,3} a stereocontrolled total synthesis of the decasaccharide propyl glycoside 1c, a synthetic model for the phytoalexin elicitor-active galacturonic acid oligosaccharides $1b^4$. The allyl galactodecaoside 2 was a key intermediate which was transformed into the corresponding uronic acid glycoside 1d. However, liberation of the anomeric hydroxyl group was not achieved. In a preliminary study of the tris(triphenylphosphine)rhodium (I) chloride-mediated deallylation of allyl galactosides, undesired oxidation of the allyl to the 2-oxopropyl group was encountered.

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In order to synthesize galacturonic acid oligosaccharides of the type released by the degradation of pectin, an alternative to the allyl group for protecting HO-1 was necessary. Galactobioses have been synthesised by way of the stable *tert*-butyldiphenylsilyl glycosides⁵, and the mild conditions used for desilylation⁶ make feasible the regeneration of HO-1 in a late stage of the syntheses of galacturonic acid oligosaccharides.

We now describe a practical approach to the synthesis of such oligosaccharides, using 1-O-*tert*-butyldiphenylsilylated intermediates, which led to a total synthesis of a-GalA-(1 \rightarrow 4)-a-GalA-(1 \rightarrow 4)-GalA (1a), a putative plant wound-hormone, and its stereoisomer, β -GalA-(1 \rightarrow 4)-a-GalA-(1 \rightarrow 4)-GalA (3).



RESULTS AND DISCUSSION

The *tert*-butyldiphenylsilyl galactobiosides 7 and 9 were prepared by stereoselective glycosylation using the galactopyranosyl fluorides 4 and 5 as the donors^{1,3,5}. Under the conditions (SnCl₂, AgClO₄, MS4A, ether) described by Mukaiyama *et al.*⁷, glycosylation of 6 with 5 afforded the *a*-linked galactobioside derivative 9 as the sole coupling product. On the other hand, the reaction of 6 with 4 gave a mixture of the *a*-linked galactobioside derivative 7 and the β isomer 8 in the ratio 4.3–5:1.



As the stereochemical outcome of the glycosylation reaction strongly depends on the choice of the promoter and the protecting groups in the donors, tin(II) trifluoromethanesulfonate⁸ and tributyltin trifluoromethanesulfonate⁹ were examined as alternative promoters, since each has been reported to activate a glycosyl fluoride¹⁰. These stannyl salts smoothly promoted the reaction of **5** with **6** to yield, exclusively, the *a*-linked product **9**. In contrast, they displayed lower stereoselectivity for the reaction of **4** with **6**. Thus, the use of tin(II) trifluoromethanesulfonate gave 96.5% of a mixture of **7** and **8** in the ratio 2.6:1, and the use of tributyltin trifluoromethanesulfonate gave 93.4% of a 1.6:1 mixture. The *a* and β isomers were distinguished on the basis of the n.m.r. data. The configurations at C-1 were assignable from either of the $J_{C-1,H-1}$ values¹¹ or the chemical shifts of the resonances of the anomeric carbons¹². Thus, for the $(1 \rightarrow 4)$ -linked galacto-oligosaccharides, the signals for C-1*a* appeared at ~100 p.p.m. and those for C-1 β at ~103 p.p.m.

The galactobiosides 7–9 were converted into the *tert*-butyldiphenylsilyl galactotrioside derivatives 15 and 17, suitable precursors for the $a-(1\rightarrow 4)$ -linked galacturonic acid trisaccharide 1a and its stereoisomer as follows.

Transformation of 7 into the galactobiosyl fluoride 10 has been reported^{1,5}. In a similar manner, the stereoisomer 12 was prepared from 8 by desilylation with tetrabutylammonium fluoride-acetic acid in tetrahydrofuran⁶ and treatment of the resulting hemiacetal 11 with diethylaminosulfur trifluoride (DAST) in tetrahydrofuran¹³. The product 12 was obtained as a 1:3 $\alpha\beta$ -mixture. Compound 9 was converted into the galactobiosyl acceptor 14 by acid-catalyzed *O*-deisopropylidenation followed by selective acetylation of the resulting diol 13.

Glycosylation of 6 with 10 gave 76.8% of the galactotrioside derivative 15 as the sole product by using stannous chloride and silver perchlorate in ether. When the same reaction was carried out in 1,2-dichloroethane, lower stereoselectivity was observed and 3.9% of the β -linked isomer 16 was obtained in addition to 68.3% of 15. In contrast, glycosylation of 6 with the β -(1 \rightarrow 4)-linked galactobiosyl donor 12 gave both the *a*- (17,



73.3%) and β -linked (18, 4.6%) trisaccharide derivatives, even in ether. Apparently, the *a*-D-galactopyranosyl residue at C-4a in the donor 10 promotes higher *a*-stereoselectivity than the β -D-galactopyranosyl residue at C-4a in the donor 12. The galactotrioside derivative 17 was also produced as a minor product (11.6%) in the reaction of 4 with the galactobiosyl acceptor 14, in which 15 was the major product (63.3%). The structural assignment of these products was based on comparison of their ¹H- and ¹³C-n.m.r. data^{11,12}.

The presence of a characteristic signals for 'Bu and three OAc groups in the 'H-n.m.r. spectra of 15-18 demonstrated that they were stereoisomeric trisaccharide derivatives. The anomeric configurations of 15 and 17 were assignable on the basis of the $J_{C-1,H-1}$ values, and the structures of 16 and 18 were assigned by the values of chemical

shift of the resonances for the anomeric carbon atoms (see Experimental). The isomers (16–18) that involved at least one β linkage exhibited one or two doublets for benzylic hydrogen at lower field than δ 5.00 [*e.g.*, 16 δ 5.20 (*J* 11.0 Hz), 17 δ 5.09 (*J* 11.6 Hz), 18 δ 5.00 (*J* 11.0 Hz) and 5.14 (*J* 11.0 Hz)], whereas no such low-field signal was observed for the isomer 15. Although the benzylic proton which is deshielded in each of 16–18 cannot be specified, this characteristic feature helped to confirm the configuration of the glycosidic linkages.

Among the above galactotrioside derivatives, 15 and 17 were obtained in amounts sufficient for further transformations to the corresponding uronides. Deacetylation of 15 with methanolic sodium methoxide gave 19 (96%). Oxidation of 19 to the uronic acid 21 was carried out in two steps^{1,3}. Swern oxidation¹⁴ converted 19 into the aldehyde 20 which, without purification, was further oxidized¹⁵ with aqueous sodium chlorite to give 21 (86% after chromatography). The structure of 21 was established after treatment with ethereal diazomethane to give the methyl ester 22, the ¹H-n.m.r. spectrum of which contained three singlets for methyl groups at δ 3.26, 3.38, and 3.48. Purification of such a large molecule as the decacarboxylic acid 1d was laborious, as reported¹ in the synthesis of 1c. A pure sample of 1d was obtained by methyl esterification, chromatography, and hydrolysis. This procedure was applied also to purify the tricarboxylic acid 21. For the regeneration of 21 from 22, an S_N2 type demethylation procedure¹⁶ (LiI, pyridine) gave an acceptable yield (69%), whereas saponification was unsuitable probably because of β -elimination.

Desilylation of 21 with tetrabutylammonium fluoride in the presence of acetic acid⁶ afforded 23 (86%), which was then hydrogenated (10% Pd–C) to complete the total synthesis of the a-(1 \rightarrow 4)-linked galacturonic acid trisaccharide 1a. Purification by anion-exchange chromatography gave a homogeneous sample of 1a, the ¹H-n.m.r. spectral data of which were coincident with that of the sample derived from natural pectins.

Likewise, the stereoisomeric galactotrioside derivative 17 was transformed into the uronic acid 3. Deacetylation of 17 (79%) was followed by the two-step oxidation to give the tricarboxylic acid 26 (80%). The signals for the corresponding trimethyl ester 27 appeared at δ 3.23, 3.33, and 3.43. Regeneration of 26 from 27 was also examined. On heating 27 with lithium iodide in dry pyridine for 7 h, 57% of 26 was obtained together with the elimination product 29. Desilylation of 26 followed by hydrogenolysis afforded the galacturonic acid trisaccharide 3, the anion-exchange chromatographic behavior and ¹H-n.m.r. spectrum of which were compatible with the assigned structure.

The results reported demonstrate a practical method for synthesizing $a \cdot (1 \rightarrow 4)$ linked galacturonic acid oligosaccharides. A total synthesis of a galacturonic acid dodecasaccharide, a phytoalexin-elictor active oligomer, using this methodology, will be reported elsewhere.

EXPERIMENTAL

General. — Optical rotations were determined with a Perkin–Elmer Model 241 MC polarimeter, for solutions in CHCl₃ at 25°, unless noted otherwise. Column chromatography was performed on Silica Gel (Merck 70–230 mesh). Flash chromatography was performed on columns of Wako Gel C-300 (200–300 mesh). T.l.c. and h.p.t.l.c. were performed on Silica Gel 60 F_{254} (Merck). N.m.r. spectra were recorded with a JEOL GX500 [¹H (500 MHz)] or FX90Q [¹³C (22.50 MHz)] spectrometer. Chemical shifts are expressed in p.p.m. downfield from the signal for internal Mc₄Si, for solutions in CDCl₃, unless noted otherwise, and, for solutions in D₂O, in p.p.m. downfield from the signal for Me₄Si, by reference to internal Me₃COH (1.230).

tert-Butyldiphenylsilyl O-(6-O-acetyl-2,3,4-tri-O-benzyl-a- and - β -D-galactopyranosyl)-(1 \rightarrow 4)-6-O-acetyl-2,3-di-O-benzyl- β -D-galactopyranoside (7 and 8). — (a) With Sn(OTf)₂ as the promoter. A mixture of 4 (ref. 1) (88 mg, 0.18 mmol) and 6 (ref. 1) in dry ether (5 mL) was added to a stirred mixture of Sn(OTf)₂⁸ (150 mg, 0.36 mmol) and powdered molecular sieves 4 Å (220 mg) under Ar at -10° . After stirring for 1 h at -10° to -4° , the reaction was stopped by quenching with pyridine (1 mL). The mixture was filtered through Celite and extracted with EtOAc-ether (1:1), and the extract was washed with water and brine, dried (Na₂SO₄), and concentrated *in vacuo*. Column chromatography of the residue on silica gel (20 g) with hexane–EtOAc (3:1) gave 7 (87 mg, 69.4%) and then 8 (34 mg, 27.1%). The physical data of 7 and 8 were identical with those described¹.

(b) With n-Bu₃SnOTf as the promoter. To a mixture of **4** (88 mg, 0.18 mmol), **6** (72 mg, 0.11 mmol), and powdered molecular sieves 4 Å (230 mg) in dry ether (3 mL) was added ethereal 0.16M *n*-Bu₃SnOTf⁹ (3 mL) with stirring under Ar at -10° . The stirred mixture was allowed to warm to room temperature overnight, then filtered through Celite, and stirred with saturated aq. KF (10 mL) for 4 h. The precipitate was removed, the filtrate was extracted with EtOAc, and the extract was washed with water and brine, dried (Na₂SO₄), and concentrated *in vacuo*. Column chromatography of the residue on silica gel (20 g) with hexane–EtOAc (4:1) gave **7** (72 mg, 57.5%) and **8** (45 mg, 35.9%).

tert-Butyldiphenylsilyl O-(2,3-di-O-benzyl-4,6-O-isopropylidene-a-D-galactopyranosyl)- $(1\rightarrow 4)$ -6-O-acetyl-2,3-di-O-benzyl- β -D-galactopyranoside (9). — (a) With $Sn(OTf)_2$ as the promoter. Reaction of 5 (ref. 1) (81 mg, 0.20 mmol) with 6 (73 mg, 0.11 mmol), as described above, gave 9 (72 mg, 61.8%) and its O-deisopropylidenated derivative 13 (10 mg, 8.9%) after chromotagraphy on silica gel (20 g) with hexane– EtOAc-pyridine (60:40:1). The physical data of 9 and 13 were identical with those described¹.

(b) With n-Bu₃SnOTf as the promoter. Reaction of 5 (81 mg, 0.20 mmol) with 6 (73 mg, 0.11 mmol) was carried out in dry ether (3 mL) with an ethereal solution of 0.16M *n*-Bu₃SnOTf (4 mL) and powdered molecular sieves 4 Å (200 mg) at -7° for 2 h. The mixture was worked-up as described above and chromatography on silica gel (20 g) with hexane–EtOAc–pyridine (60:40:1) afforded 9 (88 mg, 75.5%) as the sole product.

 $O-(6-O-Acetyl-2,3,4-tri-O-benzyl-\beta-D-galactopyranosyl)-(1\rightarrow 4)-6-O-acetyl-2,3-$

di-O-benzyl-D-galactopyranose (11). — To a mixture of 8 (295 mg, 0.26 mmol) and AcOH (110 μ L, 1.92 mmol) in dry tetrahydrofuran (8mL) was added M *n*-Bu₄NFtetrahydrofuran (1.1 mL, 1.1 mmol). The mixture was stirred at room temperature for 3 days. Most of the solvent was evaporated *in vacuo* and the residue was extracted with ether-EtOAc (1:1). The extract was washed with water and brine, dried (Na₂SO₄), and concentrated *in vacuo*. Column chromatography of the crude product on silica gel (25 g) with hexane-EtOAc (1:1) gave 11 (200 mg, 86.2%), $R_{\rm F}$ 0.25 and 0.21 (1:1 hexane-EtOAc). ¹H-N.m.r. data: δ 1.97 (1.96) (s, 3 H, Ac), 2.04 (2.05) (s, 3 H, Ac).

Anal. Calc. for C₅₁H₅₆O₁₃: C, 69.85; H, 6.45. Found: C, 69.47; H, 6.40.

O-(6-O-Acetyl-2,3,4-tri-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-6-O-acetyl-2,3di-O-benzyl-D-galactopyranosyl fluoride (12). — To a stirred solution of 11 (166 mg, 0.19 mmol) in dry tetrahydrofuran (1 mL) at -10° was added diethylaminosulfur trifluoride (DAST, 40 μ L, 0.30 mmol). The mixture was allowed to warm to room temperature, a few drops of MeOH were added, the mixture was concentrated *in vacuo*, the residue was extracted with ether–EtOAc (1:1), and the extract was washed with water and brine, dried (Na₂SO₄), and concentrated *in vacuo*. Column chromatography of the residue on silica gel (25 g) with hexane–EtOAc (7:3) gave 12 (162 mg, 97.4%) as a 1:3 $\alpha\beta$ -mixture, $R_{\rm F}$ 0.47 and 0.41 (1:1, hexane–EtOAc). ¹H-N.m.r. data: δ 1.97 (s, 3 H, Ac), 2.05 (2.06) (s, 3 H, Ac).

Anal. Calc. for C₅₁H₅₅FO₁₂: C, 69.69; H, 6.31. Found: C, 69.89; H, 6.31.

tert-Butyldiphenylsilyl O-(2,3-di-O-benzyl-a-D-galactopyranosyl)- $(1 \rightarrow 4)$ -6-Oacetyl-2,3-di-O-benzyl-a-D-galactopyranoside (13). — A mixture of 9 (1.37 g, 1.34 mmol), tetrahydrofuran (5 mL), and aq. 80% AcOH (30 mL) was stirred at room temperature for 2 days, then concentrated. Column chromatography of the residue on silica gel (120 g) with toluene–EtOAc (3:2) gave 13 (1.20 g, 91.2%), $[a]_{p}^{27} + 54^{\circ}$ (c 1.4). N.m.r. data: 1 H, $\delta 1.14$ (s, 9 H, 1 Bu), 1.88 (s, 3 H, Ac), 3.27 (bt, 1 H, J6.34 Hz, H-5a), 3.31 (dd, 1 H, J2.7 and 10.0 Hz, H-3a), 3.52 (m, 1 H, H-6b), 3.62 (dd, 1 H, J5.1 and 11.5 Hz, H-6b), 3.69 (dd, 1 H, J7.3 and 10.0 Hz, H-2a), 3.78 (bd, 1 H, J 2.4 Hz, H-4a), 3.88 (dd, 1 h, J 3.4 and 9.8 Hz, H-2b), 3.97 (dd, 1 H, J 3.2 and 9.8 Hz, H-3b), 4.07 (bt, 1 H, H-5b), 4.09 (b, 1 H, H-4b), 4.15 (dd, 1 H, J 6.6 and 11.2 Hz, H-6a), 4.24 (dd, 1 H, J 6.4 and 11.2 Hz, H-6a), 4.60 (d, 1 H, J 11.7 Hz, CH, Ph), 4.61 (d, 1 H, J 7.3 Hz, H-1a), 4.68 (d, 1H, J 11.7 Hz, CH₂Ph), 4.71 (d, 1 H, J11.9 Hz, CH₂Ph), 4.76 (d, 1 H, J11.7 Hz, CH₂Ph), 4.80 (d, 1 H, J11.9 Hz, CH₂Ph), 4.83 (d, 1 H, J11.7 Hz, CH₂Ph), 4.84 (d, 1 H, J11.0 Hz, CH₂Ph), 4.91 (d, 1 H, J 3.4 Hz, H-1b), 4.96 (d, 1 H, J 11.0 Hz, CH₂Ph), 7.23-7.42 (m, 26 H, aromatic H), 7.70–7.74 (m, 4 H, aromatic H); ${}^{13}C$, $\delta 19.3$ (CMe₃), 20.7 (CH₃CO), 27.1 $[C(CH_3)_3], 62.7, 63.1, 69.5, 70.1, 72.6, 73.1, 73.8, 75.1, 76.1, 76.3, 77.5, 80.7, 98.3 (C-1a),$ 100.2 (C-1b), 127.3, 127.4, 127.5, 127.8, 127.9, 128.1, 128.3, 128.4, 128.5, 130.0, 133.7, 135.8, 136.0, 138.0, 138.1, 138.6, 170.2 (C=O).

Anal. Calc. for C₅₄H₆₆O₁₂Si: C, 70.85; H, 6.77. Found: C, 70.47; H, 6.74.

tert-Butyldiphenylsilyl O-(6-O-acetyl-2,3-di-O-benzyl-a-D-galactopyranosyl)-(1 \rightarrow 4)-6-O-acetyl-2,3-di-O-benzyl- β -D-galactopyranoside (14). — To an ice-cooled solution of 13 (1.38 g, 1.40 mmol) in dry pyridine (15 mL) was added acetyl chloride (125 μ L, 1.76 mmol) with stirring. After stirring at 0–10° for 3 h, a few drops of water were

added, the mixture was concentrated *in vacuo*, the residue was extracted with EtOAc. and the extract was washed with water and brine, dried (Na_2SO_4) , and concentrated. Column chromatography of the residue on silica gel (150 g) with toluene-EtOAc (3:1) gave a triacetate (0.10 g, 6.7%) and then 14 (1.25 g, 86.9%), $[a]_{p}^{23} + 55^{\circ}$ (c 0.7), R_{x} 0.35 (7:3 toluene-hexane). N.m.r. data: ¹H, δ 1.12 (s, 9 H, ¹Bu), 1.88 (s, 3 H, Ac), 1.90 (s, 3 H, Ac), 3.26 (t, 1 H, J6.6 Hz, H-5a), 3.30 (dd, 1 H, J2.8 and 9.8 Hz, H-3a), 3.71 (dd, 1 H, J7.3 and 9.8 Hz, H-2a), 3.81 (d, 1 H, J 2.4 Hz, H-4a), 3.89 (dd, 1 H, J 3.4 and 9.8 Hz, H-2b), 4.01 (dd, 1 H, J 3.1 and 9.8 Hz, H-3b), 4.04 (dd, 1 H, J 6.1 and 11.0 Hz, H-6b). 4.05 (bs, 1 H, H-4b), 4.13 (dd, 1 H, J7.0 and 11.3 Hz, H-6a), 4.19 (dd, 1 H, J7.0 and 11.3 Hz, H-6a or H-6b), 4.23 (dd, 1 H, J 6.1 and 11.3 Hz, H-6a or H-6b), 4.35 (t, 1 H, J 6.4 Hz, H-5b), 4.60 (d, 1 H, J7.3 Hz, H-1a), 4.65 (d, 1 H, J12.8 Hz, CH₂Ph), 4.70 (d, 1 H, J11.9 Hz, CH₂Ph), 4.72 (d, 1 H, J12.8 Hz, CH₂Ph), 4.78 (d, 1 H, J11.3 Hz, CH₂Ph), 4.80 (d, 1 H, J11.9 Hz, CH₂Ph), 4.83 (d, 1 H, J11.9 Hz, CH₂Ph), 4.86 (d, 1 H, J11.0 Hz, CH, Ph), 4.94 (d, 1 H, J 3.4 Hz, H-1b), 4.96 (d, 1 H, J 11.0 Hz, CH, Ph), 7.22-7.42 (m, 26 H, aromatic H), 7.69–7.73 (m, 4 H, aromatic H); ¹³C, δ 19.3 (CMe₃), 20.7 (CH₃CO), 20.8 (CH₃CO), 27.1 [C(CH₃)₃], 62.7, 63.0, 67.6, 68.1, 72.3, 72.6, 73.9, 75.0, 75.5, 76.2, 77.4, 80.2, 80.8, 98.1 (C-1a), 99.8 (C-1b), 127.2, 127.4, 127.5, 127.9, 128.1, 128.3, 129.5, 135.8, 136.0, 138.2, 138.3, 138.7, 170.5 (C=O), 170.2 (C=O).

Anal. Calc. for C₆₀H₆₈O₁₃Si: C, 70.29; H, 6.69. Found: C, 70.27; H, 6.69.

Triacetate: $R_{\rm F}$ 0.56 (7:3 toluene–EtOAc). ¹H-N.m.r. data: δ 1.13 (s, 9 H, ¹Bu), 1.88 (s, 3 H, Ac), 1.92 (s, 3 H, Ac), 2.08 (s, 3 H, Ac), 5.56 (m, 1 H, H-4b).

tert-Butyldiphenylsilyl O-(6-O-acetyl-2,3,4-tri-O-benzyl-a-D-galactopyranosyl)-(1 \rightarrow 4)-O-(6-O-acetyl-2,3-di-O-benzyl-a- and - β -D-galactopyranosyl)-(1 \rightarrow 4)-6-O-acetyl-2,3-di-O-benzyl- β -D-galactopyranoside (15 and 16).— (a) With 1,2-dichloroethane as the solvent. To a mixture of SnCl₂ (72 mg, 0.38 mmol), AgClO₄ (87 mg, 0.42 mmol), and powdered molecular sieves 4 Å (500 mg) was added a solution of 10 (ref. 1) (106 mg, 0.12 mmol) and 6 (77 mg, 0.12 mmol) in 1,2-dichloroethane (8 mL) with stirring at -10° . Stirring was continued from -10° to room temperature during 7.5 h, pyridine (0.5 mL) was added, the mixture was filtered through Celite and extracted with CHCl₃, and the extract was washed with water and brine, dried (Na₂SO₄), and concentrated *in vacuo*. Column chromatography of the residue on silica gel (25 g) with hexane–EtOAc (3:1) gave 15 (114 mg, 63.3%), and a mixture (28 mg, 15.5%) of 15 and 16. Preparative t.l.c. (hexane–EtOAc, 3:2) of the latter mixture gave 15 (9 mg) and 16 (7 mg).

Compound **15** had $[a]_{D}^{20}$ + 40° (*c* 0.5), $R_{\rm F}$ 0.51. N.m.r. data: ¹H, δ 1.12 (s, 9 H, ¹Bu), 1.84 (s, 3 H, Ac), 1.86 (s, 3 H, Ac), 1.91 (s, 3 H, Ac), 3.22 (bt, 1 H, *J* 6.6 Hz, H-5a), 3.25 (dd, 1 H, *J* 2.7 and 10.0 Hz, H-3a), 3.65 (dd, 1 H, *J* 7.3 and 9.8 Hz, H-2a), 4.55 (d, 1 H, *J* 7.3 Hz, H-1a); ¹³C, δ 19.2 (*C*Me₃), 20.7 (*C*H₃CO), 27.0 [C(*C*H₃)₃], 98.0 (*J*_{C-1,H-1} 155.0 Hz, C-1a), 99.5 (*J*_{C-1,H-1} 167.2 Hz, C-1b or C-1c), 99.6 (*J*_{C-1,H-1} 168.5 Hz, C-1b or C-1c), 169.8 (C=O), 169.9 (C=O), 170.2 (C=O).

Anal. Calc. for C₈₉H₉₈O₁₉Si: C, 71.27; H, 6.59. Found: C, 70.89; H, 6.57.

Compound **16** had $[a]_{p}^{20} + 37^{\circ} (c \ 0.4), R_{F} \ 0.45$. N.m.r. data: ¹H, $\delta \ 1.09 (s, 9 \ H, ^{t}Bu)$, 1.82 (s, 3 H, Ac), 1.85 (s, 3 H, Ac), 1.98 (s, 3 H, Ac), 5.20 (d, 1 H, *J* 11.0 Hz, *CH*₂Ph); ¹³C, δ 1.93 (*C*Me₃), 20.8 (*C*H₃CO), 27.0 [C(*C*H₃)₃], 98.1 (C-1a), 100.4 (C-1c), 102.5 (C-1b), 170.2 (C = O), 170.3 (C = O), 170.4 (C = O). Anal. Calc. for C₈₉H₉₈O₁₉Si·H₂O: C, 70.43; H, 6.64. Found: C, 70.39; H, 6.48.

(b) With ether as the solvent. Reaction of **10** (270 mg, 0.31 mmol) and **6** (200 mg, 0.31 mmol) was carried out in dry ether (20 mL) by using SnCl₂ (185 mg, 0.96 mmol), AgClO₄ (220 mg, 1.06 mmol), and powdered molecular sieve 4 Å (1.5 g) at -10° to -5° for 1.5 h. After work-up as described above, column chromatography of the crude product gave **15** (354 mg, 76.8%) and no detectable amount of **16**.

tert-Butyldiphenylsilyl O-(6-O-acetyl-2,3,4-tri-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-O-(6-O-acetyl-2,3-di-O-benzyl- α - and - β -D-galactopyranosyl)-(1 \rightarrow 4)-6-O-acetyl-2,3-di-O-benzyl- β -D-galactopyranoside (17 and 18). — In the manner described for 15 and 16, 12 (149 mg, 0.16 mmol) and 6 (102 mg, 0.16 mmol) were reacted in dry ether to afford a mixture of 6, 17, and 18 after 3 h. Column chromatography of the crude product gave 17 (145 mg) and a mixture (73 mg) of 6, 17, and 18, t.l.c. (toluene–EtOAc, 4:1) of which gave 17 (30 mg, total 73.3%), 6 (19 mg), and 18 (11 mg, 4.6%).

Compound 17 had $[a]_{D}^{20} + 39^{\circ}$ (c 1.3), R_{F} 0.65 (7:3 toluene–EtOAc). N.m.r. data: ¹H, δ 1.11 (s, 9 H, ¹Bu), 1.84 (s, 3 H, Ac); 1.85 (s, 3 H, Ac), 1.98 (s, 3 H, Ac), 3.24 (bt, 1 H, J 6.4 Hz, H-5a), 3.29 (dd, 1 H, J 2.7 and 9.8 Hz, H-3a), 3.43 (bt, 1 H, J 6.3 Hz, H-5c), 3.48 (dd, 1 H, J 2.7 and 9.8 Hz, H-3c), 3.71 (dd, 1 H, J 7.3 and 9.8 Hz, H-2a), 3.73 (d, 1 H, J 2.1 Hz, H-4c), 3.80 (dd, 1 H, J 7.6 and 9.5Hz, H-2c), 3.83 (d, 1 H, J 2.1 Hz, H-4a), 4.58 (d, 1 H, J 7.3 Hz, H-1a), 4.89 (d, 1 H, J 7.6 Hz, H-1c), 5.01 (d, 1 H, J 3.4 Hz, H-1b), 5.09 (d, 1 H, J 11.6 Hz, CH₂Ph); ¹³C, δ 19.2 (CMe₃), 20.8 (CH₃CO), 27.0 [C(CH₃)₃], 98.0 (J_{C-1,H-1} 155.0 Hz, C-1a), 99.6 (J_{C-1,H-1} 168.5 Hz, C-1b), 102.8 (J_{C-1,H-1} 162.4 Hz, C-1c), 170.2 (C=O), 170.4 (C=O).

Anal. Calc. for C₈₉H₉₈O₁₉Si: C, 71.27; H, 6.59. Found: C, 70.94; H, 6.59.

Compound 18 had $[a]_{p}^{24}$ + 22.5° (c 0.5), R_{p} 0.55. N.m.r. data: ¹H, δ 1.08 (s, 9 H, ¹Bu), 1.89 (s, 3 H, Ac), 1.96 (s, 3 H, Ac), 2.01 (s, 3 H, Ac), 4.59 (d, 1 H, J 7.3 Hz, H-1a), 4.86 (d, 1 H, J 7.6 Hz, H-1b or H-1c), 4.88 (d, 1 H, H-1b or H-1c), 5.00 (d, 1 H, J 11.0 Hz, CH_2 Ph), 5.14 (d, 1 H, J 11.0 Hz, CH_2 Ph); ¹³C, δ 1.93 (CMe₃), 20.9 (CH₃CO), 27.0 [C(CH₃)₃], 98.1 (C-1a), 102.7 (C-1b or C-1c), 103.1 (C-1b or C-1c), 170.5 (C=O), 170.7 (C=O).

Anal. Calc. for C₈₉H₉₈O₁₉Si·H₂O: C, 70.42; H, 6.51. Found: C, 70.61; H, 6.62.

Alternative preparation of 15 and 17. — Reaction of 4 (145 mg, 0.29 mmol) and 14 (230 mg, 0.22) with SnCl₂ (112 mg, 0.59 mmol), AgClO₄ (122 mg, 0.59 mmol), and powdered molecular sieve 4 Å (1 g) in dry ether (12 mL) was performed at -10° to room temperature for 4 h. Column chromatography (CHCl₃–EtOAc, 95:5) of the product on silica gel (35 g) gave 15 (213 mg, 63.3%), $R_{\rm F}$ 0.58, and 17 (39 mg, 11.6%), $R_{\rm F}$ 0.46.

tert-Butyldiphenylsilyl O-(2,3,4-tri-O-benzyl-a-D-galactopyranosyl)- $(1 \rightarrow 4)$ -O-(2,3-di-O-benzyl-a-D-galactopyranosyl)- $(1 \rightarrow 4)$ -2,3-di-O-benzyl- β -D-galactopyranoside (19). — A mixture of 15 (200 mg, 0.13 mmol), tetrahydrofuran (0.5 mL), and methanolic 0.1M NaOMe (2 mL) was stirred at room temperature overnight, then neutralised with Amberlyst 15 (H⁺) resin, and concentrated. Column chromatography of the residue on silica gel (25 g) with toluene–EtOAc (3:2) gave 19 (175 mg, 95.5%), $[a]_{p}^{20}$ +47° (c 0.6). N.m.r. data: ¹H, δ 1.09 (s, 9 H, ¹Bu), 4.55 (d, 1 H, J 7.4 Hz, H-1a), 4.88 (d, 1 H, J 3.2 Hz, H-1b or H-1c), 4.91 (d, 1 H, J 3.2 Hz, H-1b or H-1c; ¹³C, δ 19.2 (CMe₃), 27.1 [C(CH₃)₃], 98.3 (C-1a), 99.8 (C-1b or C-1c), 100.8 (C-1b or C-1c). *Anal.* Calc. for C₈₃H₉₂O₁₆Si: C, 72.57; H, 6.75. Found: C, 72.59; H, 6.78. tert-*Butyldiphenylsilyl* O-(2,3,4-tri-O-benzyl-β-D-galactopyranosyl)-(1→4)-O-(2, 3-di-O-benzyl-a-D-galactopyranosyl)-(1→4)-2,3-di-O-benzyl-β-D-galactopyranoside (24). — Deacetylation of 17 (165 mg, 0.11 mmol), as described above, gave 24 (119 mg, 78.7%), $[a]_{p}^{24}$ +45° (*c* 0.6). N.m.r. data: ¹H, δ 1.10 (s, 9 H, ¹Bu), 3.29 (dd, 1H, J 3.0 and 9.8 Hz, H-3a), 3.67 (dd, 1 H, J 7.3 and 9.8 Hz, H-2a), 3.90 (dd, 1 H, J 7.6 and 9.8 Hz, H-2c), 4.58 (d, 1 H, J 7.3 Hz, H-1a), 4.70 (d, 1 H, J 7.6 Hz, H-1c), 4.88 (d, 1 H, J 3.1 Hz, H-1b); ¹³C, δ 19.2 (*C*Me₃), 27.1 [C(*C*H₃)₄], 98.3 (C-1a), 100.1 (C-1b), 104.5 (C-1c).

Anal. Calc. for C₈₃H₉₂O₁₆Si: C, 72.57; H, 6.75. Found: C, 72.03; H, 6.76.

tert-Butyldiphenylsilyl O-(2,3,4-tri-O-benzyl-a-D-galactopyranosyluronic acid)- $(1 \rightarrow 4)$ -O-(2,3-di-O-benzyl-a-D-galactopyranosyluronic acid)- $(1 \rightarrow 4)$ -2,3-di-O-benzyl- β -D-galactopyranosiduronic acid (21). — To a solution of oxalyl chloride (116 μ L, 1.33 mmol) in dry CH₂Cl₂ (3 mL) was added dry methyl sulfoxide (200 μ L, 2.81 mmol) at -78° under Ar. After 15 min, a solution of **19** (123 mg, 0.09 mmol) in dry CH₂Cl₂ (2 mL) was added, stirring was continued for 30 min, N,N-di-isopropylethylamine (1 mL, 5.74 mmol) was added, and, after 5 min, the cooling bath was removed. The mixture was stirred for another 30 min, diluted with CHCl₃, washed successively with dil HCl, water, and brine, dried (Na_2SO_4) , and concentrated to give the crude trialdehyde 20, a solution of which in 'BuOH (3.7 mL) and 2-methyl-2-butene (1.4 mL, 16.7 mmol) was stirred overnight with a solution of NaClO₂ (230 mg, 2.54 mmol) and NaH₂PO₄ (230 mg) in water (2.3 mL). The mixture was concentrated in vacuo, diluted with water, and extracted with hexane. Then the aqueous layer was acidified with M HCl and extracted with EtOAc. The extract was washed with water and brine, dried (Na₂SO₄), and concentrated in vacuo. Column chromatography of the residue on silica gel (25 g) with CHCl₃-AcOH (97:3) gave **21** (109 mg, 86%), $[a]_{p}^{20} + 93^{\circ}$ (c 0.3). N.m.r. data: ¹H, δ 1.17 (s, 9 H, 'Bu), 3.25 (dd, 1 H, J 2.7 and 10.0 Hz, H-3a), 3.57 (dd, 1 H, J 7.3 and 10.0 Hz, H-2a), 4.59 (d, 1 H, J 7.3 Hz, H-1a), 5.08 (m, 2 H, H-1b,c); ${}^{13}C$, δ 19.1 (CMe₃), 26.9 [C(CH₃)₃], 98.3, 98.7, 99.4 (C-1a, 1b, 1c), 167.8 (C = O), 169.0 (C = O).

Anal. Calc. for C₈₃H₈₆O₁₉Si·H₂O: C, 69.54; H, 6.19. Found: C, 69.11; H, 6.07.

tert-Butyldiphenylsilyl O-(2,3,4-tri-O-benzyl-β-D-galactopyranosyluronic acid)-(1→4)-O-(2,3-di-O-benzyl-α-D-galactopyranosyluronic acid)-(1→4)-2,3-di-Obenzyl-β-D-galactopyranosiduronic acid (26). — Oxidation of 24 (96 mg, 0.07 mmol), as described for 21, afforded 26 (79 mg, 79.9%), $[a]_{D}^{24}$ + 75° (c 1.1). N.m.r. data: ¹H, δ 1.10 (s, 9 H, ¹Bu), 3.29 (dd, 1 H, J 2.8 and 10.1 Hz, H-3a), 3.48 (dd, 1 H, J 3.1 and 9.8 Hz, H-3c), 3.64 (dd, 1 H, J 7.6 and 10.1 Hz, H-2a), 3.75 (dd, 1 H, J 7.6 and 9.8 Hz, H-2c), 4.61 (d, 1 H, J 7.6 Hz, H-1a or H-1c), 5.12 (d, 1 H, J 2.7 Hz, H-1b); ¹³C, δ 19.1 (CMe₃), 26.9 [C(CH₃)₃], 98.3 (C-1a), 99.3 (C-1b), 103.1 (C-1c), 167.6 (C=O), 169.2 (C=O).

O-(Methyl 2,3,4-tri-O-benzyl-a-D-galactopyranosyluronate)-($1 \rightarrow 4$)-O-(methyl 2,3-di-O-benzyl-a-D-galactopyranosyluronate)-($1 \rightarrow 4$)-[methyl (tert-butyldiphenylsilyl 2,3-di-O-benzyl- β -D-galactopyranosid)uronate] (22). — A solution of 21 (45 mg, 0.03 mmol) in EtOAc (1 mL) was treated with a large excess of fresh ethereal diazomethane, then concentrated. Column chromatography of the residue on silica gel (5 g) with hexane-EtOAc (7:3) gave 22 (39 mg, 84.2%), $[a]_{\rm D}^{25} + 66^{\circ}$ (c 0.8). N.m.r. data: ¹H, δ 1.17

(s, 9 H, ^tBu), 3.26 (s, 3 H, OMe), 3.38 (s, 3 H, OMe), 3.48 (s, 3 H, OMe), 3.63 (dd, 1 H, J 7.6 and 10.1 Hz, H-2a), 3.78 (dd, 1 H, J 2.7 and 10.1 Hz, H-3a), 4.50 (d, 1 H, J 7.6 Hz, H-1a), 5.05 (d, 1 H, J 3.4 Hz, H-1b or H-1c), 5.28 (d, 1 H, J 3.4 Hz, H-1b or H-1c); ¹³C, δ 19.3 (*C*Me₃), 27.1 [C(*C*H₃)₃], 51.7 (OCH₃), 51.8 (OCH₃), 51.9 (OCH₃), 98.1 (C-1a), 98.8, 99.5 (C-1b, C-1c), 168.1 (C=O), 168.8 (C=O), 169.5 (C=O).

Anal. Calc. for C_{ss}H₀,O₁₀Si·H₂O: C, 69.99; H, 6.42. Found: C, 69.61; H, 6.48.

O-(*Methyl* 2,3,4-tri-O-benzyl-β-D-galactopyranosyluronate)-(1→4)-O-(*methyl* 2,3-di-O-benzyl-a-D-galactopyranosyluronate)-(1→4)-[methyl (tert-butyldiphenylsilyl 2,3-di-O-benzyl-β-D-galactopyranosid)uronate] (**27**). — Esterification of **26** (62 mg, 0.045 mmol) with ethereal diazomethane followed by column chromatography, as described for **22**, afforded **27** (45 mg, 70.5%), $[a]_{D}^{25}$ + 73° (*c* 0.6). N.m.r. data: ¹H, δ 1.16 (s, 9 H, ¹Bu), 3.23 (s, 3 H, OMe), 3.33 (dd, 1 H, J 2.7 and 9.8 Hz, H-3a or H-3c), 3.43 (dd, 1 H, J 3.1 and 9.8 Hz, H-3a or H-3c), 3.53 (s, 3 H, OMe), 3.64 (s, 3 H, OMe), 3.69 (dd, 1 H, J 7.3 and 9.8 Hz, H-2a or H-2c), 3.70 (dd, 1 H, J 7.6 and 9.8 Hz, H-2a or H-2c), 3.96 (dd, 1 H, J 3.4 and 10.1 Hz, H-2b), 4.09 (dd, 1 H, J 2.8 and 10.1 Hz, H-3b), 5.33 (d, 1 H, J 3.4 Hz, H-1b); ¹³C, δ 19.4 (CMe₃), 27.1 [C(CH₃)₃], 51.8 (OCH₃), 51.9 (OCH₃), 52.3 (OCH₃), 98.1 (C-1a), 99.3 (C-1b), 102.1 (C-1c), 168.1 (C=O), 168.4 (C=O), 168.8 (C=0).

Anal. Calc. for C₈₆H₉₂O₁₉Si: C, 70.86; H, 6.36. Found: C, 70.60; H, 6.36.

Regeneration of **21** from **22**. — A mixture of **22** (13.5 mg, 9.3 μ mol) and anhydrous LiI (36 mg, 0.27 mmol) in dry pyridine (3.5 mL) was heated under reflux under argon for 7 h, then concentrated *in vacuo*. A solution of the residue in water was acidified with dil. HCl and extracted with EtOAc, and the extract was washed with water and brine, dried (Na₂SO₄), and concentrated *in vacuo*. Preparative t.l.c. (CHCl₃–AcOH, 97:3) of the residue gave a fraction which was dissolved in EtOAc, and the solution was washed with dil HCl, water, and brine, dried (Na₂SO₄), and concentrated (Na₂SO₄), and concentrated *in vacuo*.

Regeneration of **26** *from* **27.** — As described above, a mixture of **27** (29 mg, 19.9 μmol) and LiI (75 mg, 0.56 mmol) in dry pyridine (7.5 mL) was heated for 7 h. Preparative t.l.c. of the crude product afforded **26** (16 mg, 57%) and **29** (5 mg) as a less polar material. Compound **29**, ¹H-n.m.r. data: δ 1.08 (s, 9 H, ¹Bu), 3.73 (s, 3 H, OMe), 3.78 (bt, 1 H, H-2), 4.09 (bt, 1 H, H-3), 4.57 (d, 1 H, J 11.6 Hz, CH₂Ph), 4.59 (d, 1 H, J 11.6 Hz, CH₂Ph), 4.62 (d, 1 H, J 11.6 Hz, CH₂Ph), 4.69 (d, 1 H, J 11.6 Hz, CH₂Ph), 5.29 (d, 1 H, J 5.2 Hz, H-1), 6.19 (d, 1 H, J 3.7 Hz, H-4).

O-(2,3,4-Tri-O-benzyl-a-D-galactopyranosyluronic acid)- $(1 \rightarrow 4)$ -O-(2,3-di-O-benzyl-a-D-galactopyranosyluronic acid)- $(1 \rightarrow 4)$ -(2,3-di-O-benzyl-D-galactopyranuronic acid (23). — A mixture of 21 (21 mg, 14.8 µmol), AcOH (15 µL, 0.26 mmol), and M tetrabutylammonium fluoride/tetrahydrofuran (150 µL, 0.15 mmol) in dry tetrahydrofuran (1.2 mL) was stirred at room temperature for 4 days, then diluted with water (15 mL), acidified with dil. HCl, and extracted with EtOAc. The extract was washed with water and brine, dried (Na₂SO₄), and concentrated *in vacuo*. T.I.c. (CHCl₃-MeOH-AcOH 18:1:1) of the residue gave a fraction which was dissolved in EtOAc, and the solution was washed with dil. HCl, water, and brine, dried (Na₂SO₄), and concentrated to give 23 (15 mg, 86%).

Anal. Calc. for $C_{67}H_{68}O_{19}$ ·H₂O: C, 67.33; H, 5.90. Found: C, 67.61; H, 5.98. O-(2,3,4-Tri-O-benzyl- β -D-galactopyranosyluronic acid)-(1 \rightarrow 4)-O-(2,3-di-O-benzyl-a-D-galactopyranosyluronic acid)-(1 \rightarrow 4)-2,3-di-O-benzyl-D-galactopyranuronic acid (**28**). — Desilylation of **26** (15 mg, 10.6 μ mol), as described above for **23**, gave **28** (11 mg, 88.2%).

Anal. Calc. for C₆₇H₆₈O₁₉·H₂O: C, 67.33; H, 5.90. Found: C, 67.62; H, 5.86.

O-(a-D-Galactopyranosyluronic acid)- $(1 \rightarrow 4)$ -O-(a-D-galactopyranosyluronicacid)- $(1 \rightarrow 4)$ -D-galactopyranuronic acid (1a). — A solution of 23 (15 mg, 12.7 μ mol) in aq. 80% MeOH (10 mL) was stirred with 10% Pd-C (10 mg) under hydrogen at room temperature for 3 days, then filtered through Celite, and concentrated in vacuo. Shortcolumn (2 mL) chromatography of the residue on Sephadex LH-20 with 50% aq. MeOH removed traces of impurities. Further purification was effected by anionexchange chromatography on Mono-Q (HR 5/5), using a linear NH $_{4}$ ·HCO, gradient $(0.2 \rightarrow 0.4 \text{ m})^1$. Fractions were assayed for uronosyl residue by the *m*-hydroxybiphenyl method¹⁷ (A at 520 nm). The fractions having a maximal content of 1a were combined and concentrated in vacuo at 70°. Water was distilled several times from the residue to leave 1a (4.7 mg). ¹H-N.m.r. data [D₂O, internal 'BuOH (δ 1.23), 80°]; δ 3.49 (dd, J 7.6, 9.8 Hz, H-2aβ), 3.60-3.85 (m, H-2aa,2b,2c, H-3aβ), 3.90 (dd, J 3.4, 10.1 Hz, H-3), 3.97-4.01 (m), 4.05 (s, H-5a β), 4.29 (bs, H-4c), 4.38 (bs, H-5a α), 4.43 (bs, H-4a β), 4.44(bs, H-4b), 4.58 (d, J 7.6 Hz, H-1a β), 4.70 (m, H-5b), 4.71 (bs, H-5c), 5.10 (bs, H-1b or H-lc), 5.13 (bs, H-1b or H-1c), 5.31 (bs, H-1aa).

Anal. Calc. for $C_{18}H_{26}O_{19}\cdot 0.5H_{2}O: C, 38.94$; H, 4.90. Found: C, 38.91; H, 4.83. O-(β -D-Galactopyranosyluronic acid)-($1 \rightarrow 4$)-O-(α -D-galactopyranosyluronic acid)-($1 \rightarrow 4$)-D-galactopyranuronic acid (**3**). — Hydrogenolysis of **28** (10 mg, 8.5 μ mol) with 10% Pd/C (7 mg) in 80% MeOH (7 mL), as described above for **1a**, followed by anion-exchange chromatography (Mono-Q) afforded **3** (4.5 mg). ¹H-N.m.r. data (D₂O, 60°): δ 3.49 (dd, J 7.6 and 10.1 Hz, H-2a β), 3.55 (bt, 1 H, H-2c), 3.67 (dd, 1 H, J 3.4 and 9.8 Hz, H-3c), 3.74 (dd, J 3.1 and 10.1 Hz, H-3a β), 3.82 (dd, J 3.7 and 10.7 Hz, H-2aa), 4.05 (bs, H-5a β), 4.59 (d, J 7.6 Hz, H-1a β), 4.70 (d, 1 H, J 7.6 Hz, H-1c), 4.79 (m, 1 H, H-5b), 5.13 (d, 1 H, J 2.8 Hz, H-1b), 5.31 (d, J 3.7 Hz, H-1aa).

Anal. Calc. for C₁₈H₂₆O₁₉: C, 39.57; H, 4.80. Found: C, 39.25; H, 5.05.

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