1236 Short Papers SYNTHESIS

Use of Hydrazine Acetate as a Mild and Efficient Reagent for Anomeric Denitration of Carbohydrates

Tatsushi Toyokuni,* Shaopei Cai, Barbara Dean
The Biomembrane Institute and University of Washington, 201 Elliott Ave. W., Seattle, Washington 98119, USA
Received 13 April 1992; revised 27 May 1992

Treatment of 3,4,6-tri-O-acetyl-2-azido-2-deoxy- α , β -D-galactopyranosyl nitrate (1), 3,6-di-O-acetyl-2-azido-2-deoxy-4-O-(2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)- α , β -D-glucopyranosyl nitrate (3), and 6-O-acetyl-2-azido-2-deoxy-3,4-O-isopropylidene- α , β -D-galactopyranosyl nitrate (5) with a slight excess of hydrazine acetate in anhydrous dimethylformamide at room temperature for 2 hours gave the corresponding hemiacetals, 2, 4, and 6 in the range of an 87-92% yield.

The 2-azido-2-deoxy derivatives of galactopyranose, glucopyranose, and lactose are versatile building blocks for the synthesis of biologically important galactosamine-and glucosamine-containing glycosides, including cell-surface carbohydrate ligands of the vascular selectins, tumor-associated carbohydrate antigens, human blood-group determinants, and bacterial cell-wall peptidoglycans. The azidonitration of O-protected glycals, introduced by Lemieux and Ratcliffe in 1977, continues to provide a useful route to 2-azido-2-deoxy sugars. The reaction yields 2-azido-1-nitrate adducts, which are sub-

sequently converted into various glycosyl donors by either direct displacement of an anomeric nitrate with halide ions⁶ and potassium *O*-ethyl dithiocarbonate,⁷ or denitration followed by trichloroacetimidation⁸ and fluorination.⁹

In the course of our study on a synthetic tumor vaccine ¹⁰ based on Tn (GalNAc α 1 \rightarrow *O*-Ser/Thr), T (Gal β 1 \rightarrow 3Gal NAc α 1 \rightarrow *O*-Ser/Thr), and their sialylated antigens, ³ we have found that hydrazine acetate effectively cleaves nitrate groups from 2-azido-2-deoxy- α , β -glycopyranosyl nitrates in mild and near-neutral conditions, and hence can be used in the presence of both acid- and basesensitive protecting groups. Hydrazine acetate has been used for regioselective 1-*O*-deacetylation of peracetylated glycopyranoses ¹¹ and for deprotection of *N*-phthaloyl groups in the synthesis of base-sensitive peptides. ¹² There have been some reports in the literature ¹³ that hydrazine reduces sugar nitrates into the corresponding sugar

December 1992 SYNTHESIS 1237

alcohol in the presence of a platinum or palladium catalyst.

Our method involves simply stirring the azido nitrates, which were prepared by azidonitration of properly protected glycals, in anhydrous dimethylformamide with a slight excess of hydrazine acetate at room temperature and then extracting the corresponding hemiacetals with ethyl acetate (Scheme 1). The reaction is often complete within 2 hours and the yield is, after column chromatography on silica gel, in the range of 87-92%. The method proved especially useful for conversion of acid-sensitive derivative 5 into hemiacetal 6. Usual denitration⁸ of 5 with sodium nitrite in aqueous 1,4-dioxane at 80°C for 6 hours was accompanied by extensive de-O-isopropylidenation. A similar decomposition of an acid-sensitive protecting group has been reported for denitration of 2-azido-6-O-benzyl-2-deoxy-3,4-O-isopropylidene-α-Dgalactopyranosyl nitrate with sodium nitrite. 14

Scheme 1

10

N₃

5

Scheme 2

Compound 5, which is a key intermediate for the synthesis of sialyl Tn antigen as well as for the modification of Tn antigen, ¹⁵ was readily prepared from D-galactal ¹⁶ 7 through 8 and 10^{17} in a total yield of 47% as an α and β mixture (Scheme 2). A modified Liptak method ¹⁸ of isopropylidenation produced no significant amount of degradation products of acid-sensitive ¹⁹ 7. However, the longer reaction time led to the formation of methanol-adduct ¹⁹ 9.

Very recently, thiophenol has been employed in the presence of N,N-diisopropylethylamine to achieve anomeric denitration. ¹⁴ Although the thiophenol method is effective for both acid- and base-sensitive compounds, its malodor is undesirable from a practical standpoint. We, therefore, feel that our method with hydrazine acetate should be of general utility for the denitration of azido nitrates, particularly for those possessing acid-sensitive protecting groups.

Melting points were measured with a Fisher-Johns melting point apparatus and are uncorrected. ¹H NMR spectra were recorded on a Bruker WM-500 spectrometer in CDCl₃ with TMS as an internal standard. High resolution mass spectrometry (HRMS) was obtained on a JEOL JMS-HX 110 mass spectrometer, operating in the FAB mode, using a matrix of *m*-nitrobenzyl alcohol. Optical rotations were determined at 23 °C with a Perkin-Elmer 241MC polarimeter. Silica gel used for flash column chromatography²⁰ was purchased from EM Science (Gibbstown, NJ; particle mesh size 0.040–0.063 mm). Compounds 1 and 3 were prepared according to the method developed by the Lemieux's group. ⁶ Hydrazine acetate is commercially available from Aldrich (Milwaukee, WI) and is also readily prepared by mixing equivalent amounts of hydrazine and AcOH under N₂. The structures of the known compounds were verified by ¹H NMR spectroscopy.

General Procedure for Denitration:

Five 8 mmol portions of hydrazine acetate (total 40 mmol) were added to a stirred solution of azido nitrates (37 mmol) in dry DMF (120 mL) at 4 min intervals at r.t. in N_2 . The mixture was stirred at r.t. for 2 h, poured into saturated brine (800 mL), and extracted with EtOAc (3 × 300 mL). The combined EtOAc layer was washed with H_2O (3 × 100 mL) and dried (Na_2SO_4). After concentration, the residue was purified by flash column chromatography.

3,4,6-Tri-O-acetyl-2-azido-2-deoxy- α , β -D-galactopyranose²² (2): Denitration of 3,4,6-tri-O-acetyl-2-azido-2-deoxy- α -D-galactopyranosyl nitrate⁶ (1 α) and its β -anomer⁶ 1 β gave, after flash column chromatography (hexane/EtOAc, 4:3), 2 in 90% and 92% yields, respectively.

3,6-Di-O-acetyl-2-azido-2-deoxy-4-O-(2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)- α , β -D-glucopyranose⁸ (4):

Denitration of 3,6-di-O-acetyl-2-azido-2-deoxy-4-O-(2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)- α -D-glucopyranosyl nitrate²¹ (3 α) and its β -anomer²¹ 3 β gave, after flash column chromatography (hexane/EtOAc, 1:1), 4 in 90% and 89% yields, respectively.

6-O-Acetyl-2-azido-2-deoxy-3,4-O-isopropylidene-α,β-D-galactopy-ranose (6):

Denitration of 5α and 5β (see below for the preparation) gave, after flash column chromatography (hexane/EtOAc, 3:2), 6 in 88% and 87% yields, respectively.

¹H NMR: for the α-isomer: $\delta = 1.36$ (s, 3 H) and 1.52 (s, 3 H), (CMe₂), 2.09 (s, 3 H), OAc), 2.86 (d, 1 H, J = 3.7 Hz, OH), 3.49 (dd, 1 H, J = 7.9, 3.3 Hz, 2-H), 4.22 (dd, 1 H, J = 5.4, 2.4 Hz, 4-H), 4.28 (dd, 1 H, J = 11.9, 7.8 Hz, 6a-H), 4.38 (dd, 1 H, J = 11.9, 3.6 Hz, 6b-H), 4.49 (dd, 1 H, J = 7.9, 5.4 Hz, 3-H), 4.45 (m, 1 H, 5-H), 5.30 (dd, 1 H, and J = 3.7, 3.3 Hz, 1-H);

1238 Short Papers SYNTHESIS

for the β -isomer: $\delta = 1.35$ (s, 3 H) and 1.54 (s, 3 H) (CMe₂), 2.09 (s, 3 H, OAc), 3.29 (d, 1 H, J = 5.8 Hz, OH), 3.41 (dd, 1 H, J = 7.8, 7.7 Hz, 2-H), 4.01 (m, 1 H, 5-H), 4.02 (dd, 1 H, J = 7.8, 5.7 Hz, 3-H), 4.11 (dd, 1 H, J = 5.7, 2.2 Hz, 4-H), 4.30 (dd, 1 H, J = 12.9, 7.6 Hz, 6a-H), 4.39 (dd, 1 H, J = 12.9, 3.4 Hz, 6b-H), 4.58 (dd, 1 H, J = 7.8, 5.8 Hz, 1-H).

HRMS: calc. for $C_{11}H_{17}N_3O_6 + Na$: 310.1015; found: 310.1015.

1,5-Anhydro-2-deoxy-3,4-*O*-isopropylidene-D-*lyxo*-hex-1-enitol¹⁷ (8):

p-TsOH · H₂O (5.0 mg, 26.3 μmol) was added to a stirred suspension of 1,5-anhydro-2-deoxy-D-lyxo-hex-1-enitol (D-galactal)¹⁶ (7) (1.0 g, 6.8 mmol) in 2,2-dimethoxypropane (DMP) (10 mL, 79.7 mmol) at r.t. The mixture became homogeneous in 2 min. After an additional 2 min, H₂O (10 mL) was added and stirring was continued for another 30 s. The mixture was then diluted with sat. aq NaHCO₃ (10 mL) and extracted with CH₂Cl₂ (3 × 30 mL). The combined CH₂Cl₂ layer was washed with H₂O (3 × 50 mL), dried (Na₂SO₄), and concentrated to a pale yellow syrup. Flash column chromatography (hexane/EtOAc, 2:1) afforded 8 (854 mg, 67 %) as a colorless syrup.

 $[\alpha]_{D}$ + 37.6° (c = 1.0, CHCl₃) [Lit.¹⁷ $[\alpha]_{D}$ + 28° (c = 1.9, CHCl₃)].

When the acetalation mixture was stirred at r.t. for 1 h, methyl 2-deoxy-3,4-O-isopropylidene- α -D-lyxo-hexopyranoside (9) was obtained in a 38% yield as a colorless syrup along with 8 (27%). [α]_D + 37.7° (c = 1.3, CHCl₃).

¹H NMR: δ = 1.33 (s, 3 H) and 1.48 (s, 3 H) (CMe₂), 1.74 (ddd, 1 H, J = 14.8, 5.8, 4.0 Hz, 2ax-H), 2.18 (ddd, 1 H, J = 14.8, 5.3, 5.2 Hz, 2eq-H), 3.39 (s, 3 H, OMe), 3.79 (dd, 1 H, J = 10.4, 4.3 Hz, 6a-H), 3.81 (ddd, 1 H, J = 5.4, 4.3, 2.0 Hz, 5-H), 3.91 (dd, 1 H, J = 10.4, 5.4 Hz, 6b-H), 4.17 (dd, 1 H, J = 7.2, 2.0 Hz, 4-H), 4.48 (ddd, 1 H, J = 7.2, 5.2, 4.0 Hz, 3-H), 4.87 (dd, 1 H, J = 5.8, 5.3 Hz, 1-H). HRMS: calc. for C₁₀H₁₈O₅ + Na: 241.1051; found: 241.1059.

6-*O*-Acetyl-1,5-anhydro-2-deoxy-3,4-*O*-isopropylidene-D-*lyxo*-hex-1-enito1¹⁷ (10):

The conventional acetylation of **8** (404 mg, 2.2 mmol) with Ac_2O (2 mL) and pyridine (4 mL) gave, after flash column chromatography (hexane/EtOAc, 3:1), **10** (494 mg, 100 %) as a colorless syrup. $[\alpha]_D + 18.0^\circ$ (c = 1.0, CHCl₃) [Lit.¹⁷ $[\alpha]_D + 16^\circ$ (c = 1.2, CHCl₃].

6-*O*-Acetyl-2-azido-2-deoxy-3,4-*O*-isopropylidene- α , β -D-galactopyranosyl Nitrate (5 α ,5 β):

A solution of 10 (686 mg, 3 mmol) in anhydr. MeCN (15 mL) was added dropwise to a vigorously stirred mixture of NaN₃ (300 mg, 4.5 mmol) and cerium(IV) ammonium nitrate (CAN) (5 g, 9 mmol) at $-20\,^{\circ}$ C under N₂, and stirring was continued at $-20\,^{\circ}$ C for 7 h. The mixture was then diluted with cold Et₂O (150 mL), washed with cold H₂O (3 × 100 mL) unit it became neutral, and dried (Na₂SO₄). After concentration, the residue was subjected to flash column chromatography (hexane/EtOAc, 2:1, containing 0.3 % Et₃N). The first fraction gave 5 α (490 mg, 49%) as a colorless solid. [α]_D + 144.5° (c = 1.01, CHCl₃).

¹H NMR: δ = 1.37 (s, 3 H) and 1.54 (s, 3 H) (CMe₂), 2.08 (s, 3 H, OAc), 3.87 (dd, 1 H, J = 7.5, 4.0 Hz, 2-H), 4.27 (dd, 1 H, J = 5.8, 2.3 Hz, 4-H), 4.31 (dd, 1 H, J = 13.3, 4.0 Hz, 6a-H), 4.33 (dd, 1 H, J = 13.3, 4.1 Hz, 6b-H), 4.37 (ddd, 1 H, J = 4.1, 4.0, 2.3 Hz, 5-H), 4.39 (dd, 1 H, J = 7.5, 5.8 Hz, 3-H), 6.23 (d, 1 H, J = 4.0 Hz, 1-H). HRMS: calc. for C₁₁H₁₆N₄O₈ + Na: 355.0865; found: 355.0868.

The second fraction afforded 5β (209 mg, 21%) as a colorless solid, which crystallized from Et₂O. mp 91–92°C. [α]_D + 66.9° (c = 1.3, CHCl₃).

¹H NMR: δ = 1.37 (s, 3 H) and 1.56 (s, 3 H) (CMe₂), 2.08 (s, 3 H, OAc), 3.55 (dd, 1 H, J = 8.8, 7.3 Hz, 2-H), 4.15 (ddd, 1 H, J = 7.5, 4.3, 2.2 Hz, 5-H), 4.16 (dd, 1 H, J = 7.3, 5.8 Hz, 3-H), 4.19 (dd, 1 H, J = 5.8, 2.2 Hz, 4-H), 4.31 (dd, 1 H, J = 11.9, 7.5 Hz, 6a-H), 4.39 (dd, 1 H, J = 11.9, 4.3 Hz, 6b-H), 5.47 (d, 1 H, J = 8.8 Hz, 1-H). HRMS: calc. for C₁₁H₁₆N₄O₈ + Na: 355.0865; found: 355.0854.

We thank Ms. Mary Ellen Salyan for obtaining the mass spectra. This study was supported by funds from The Biomembrane Institute.

- Schmidt, R. R. Pure Appl. Chem. 1989, 61, 1257.
 Paulsen, H. Angew. Chem. 1990, 102, 851; Angew. Chem., Int. Ed. Engl. 1990, 29, 823.
 Sinay, P. Pure Appl. Chem. 1991, 63, 519.
- (2) Brandley, B. K.; Swiedler, S. J.; Robbins, P.W. Cell 1990, 63, 861.
- Winkelhake, J. L. Glyconjugate J. 1991, 8, 381.
 (3) Hakomori S. Adv. Cancer Res. 1989, 52, 257
- (3) Hakomori, S. Adv. Cancer Res. 1989, 52, 257. Singhal, A.K.; Hakomori, S. BioEssays 1990, 12, 223.
- (4) Watkins, W.M. Pure Appl. Chem. 1991, 63, 561.
- (5) Gabriel, O. In Escherichia coli Salmonella typhimurium; Neidhardt, F.C., Ed.; Am. Soc. Microbiol.: Washington, D.C., 1987; Vol. 1, p. 504.
 Park, J. T. Ibid. p. 663.
- (6) Lemieux, R.U.; Ratcliffe, R.M. Can. J. Chem. 1979, 57, 1244.
- (7) Marra, A.; Gauffeny, F.; Sinay, P. Tetrahedron 1991, 47, 5149.
- (8) Grundler, G.; Schmidt, R. R. Liebigs Ann. Chem. 1984, 1826.
- (9) Rosenbrook, W., Jr.; Riley, D.A.; Lartey, P.A. Tetrahedron Lett. 1985, 26, 3. Posner, G.H.; Haines, S.R. Ibid. 1985, 26, 5.
- (10) Toyokuni, T.; Dean, B.; Hakomori, S. Tetrahedron Lett. 1990, 31, 2673.
 Singhal, A.; Fohn, M.; Hakomori, S. Cancer Res. 1991, 51,
- (11) Excoffier, G.; Gagnaire, D.; Utille, J.-P. Carbohydr. Res. 1975, 39, 368.
- (12) Schwyzer, R.; Costopanagiotis, A.; Sieber, P. Helv. Chim. Acta 1963, 46, 870.
- (13) Boschan, R.; Merrow, R. T.; Van Dolah, R. W. Chem. Rev. 1955, 55, 485. Honeyman, J.; Morgan, J. W. W. Adv. Carbohydr. Chem. 1957, 12, 117.
- (14) Gauffeny, F.; Marra, A.; Shun, L.K.S.; Sinay, P.; Tabeur, C. Carbohydr. Res. 1991, 219, 237.
- (15) Toyokuni, T.; Cai, S. unpublished.
- (16) Shafizadeh, F. In *Methods in Carbohydrate Chemistry*; Whistler, R.L.; Wolfrom, M.L.; Eds.; Academic Press: New York, 1963; Vol. II, p 409.
- (17) Compounds 8 and 10 have also been prepared from 3,4-O-isopropylidene derivatives of phenyl 1-thio-β-D-galactopyranoside and β-D-galactopyranosyl phenyl sulfone by reductive lithiation with lithium naphthalenide: Fernandez-Mayoralas, A.; Trumtel, M.; Veyrieres, A.; Sinay, P. Carbohydr. Res. 1989, 188, 81.
- (18) Liptak, A.; Fuegedi, P.; Kerekgyarto, J.; Nanasi, P. Carbohydr. Res. 1983, 113, 225.
 Rashid, A.; Mackie, W.; Colquhoun, L.J.; Lamba, D. Can. J. Chem. 1990, 68, 1122.
- (19) Ferrier, R.J. Adv. Carbohydr. Chem. 1965, 20, 67.
 Ferrier, R.J. Ibid. 1969, 24, 199.
 Blackburne, I.D.; Frederichs, P.M.; Guthrie, R.D. Aust. J. Chem. 1976, 29, 381.
- (20) Still, W.C.; Kahn, M.; Mitra, A. J. Org. Chem. 1978, 43, 2923.
- (21) Lemieux, R.U.; Ratcliffe, R.M. U.S. Patent 4195174, 1980 *Chem. Abstr.* 1979, 90, 87846.
- (22) Paulsen, H.; Sumfleth, B. Chem. Ber. 1979, 112, 3203.