Spiroadamantyl 1,2,4-trioxolane, 1,2,4-trioxane, and 1,2,4-trioxepane pairs: Relationship between peroxide bond iron(II) reactivity, heme alkylation efficiency, and antimalarial activity

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ABSTRACT

These data suggest that iron(II) reactivity for a set of homologous spiroadamantyl 1,2,4-trioxolane, 1,2,4-trioxane, and 1,2,4-trioxepane peroxide heterocycles is a necessary, but insufficient, property of antimalarial peroxides. Heme alkylation efficiency appears to give a more accurate prediction of antimalarial activity than $FeSO_4$-mediated reaction rates, suggesting that antimalarial activity is not merely dependent on peroxide bond cleavage, but also on the ability of reactive intermediates to alkylate heme or other proximal targets.

The semisynthetic artemisinins rapidly reduce parasite burden and are particularly effective when used in a 3-day artemisinin combination treatment (ACT) regimen.[1] One hypothesis[2–4] that accounts for the antimalarial specificity of artemisinin[5] is that the peroxide bond undergoes reductive activation by heme released by parasite hemoglobin digestion to produce carbon-centered free radicals or carbocations that alkylate heme[6] or parasite proteins. Although the 1,2,4-trioxane heterocycle in artemisinin (Fig. 1) is the critical pharmacophore, its presence alone[7] is insufficient for high antimalarial activity as most synthetic 1,2,4-trioxanes are less active than artemisinin.[8] The discovery[9] of ozonide (1,2,4-trioxolane) drug development candidate OZ277 (arterolane) (Fig. 1), now entering Phase III clinical trials in the form of an arterolane maleate + piperaquine phosphate combination, prompted us to investigate a set of structurally related 1,2,4-trioxolanes 1, 1,2,4-trioxanes 2, and 1,2,4-trioxepanes 3[10] (Fig. 2) to better understand the relationship between peroxide bond iron(II) reactivity and antimalarial activity for these homologous peroxide heterocycles.

Trioxolanes 1b[11] and 1c were obtained by Griesbaum coozonolysis[12] of the corresponding oxime ether/ketone[13,14] pairs 4/5 and 6/7 in low yields (Scheme 1). Trioxolane 1a was similarly obtained as previously described.[15]

Trioxanes 2b and 2c were prepared as shown in Scheme 2. In the presence of catalytic p-toluenesulfonic acid (PTSA), 2b was readily formed by condensation of β-hydroperoxy alcohol 8[16] with 5. Attempts to perform the corresponding reaction of β-hydroperoxy alcohol 9[16] with 5 provided only small quantities of trioxane 2c accompanied by a complex mixture of decomposition products.
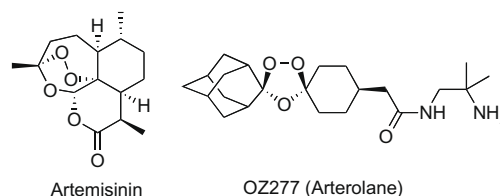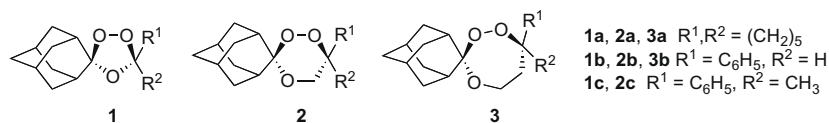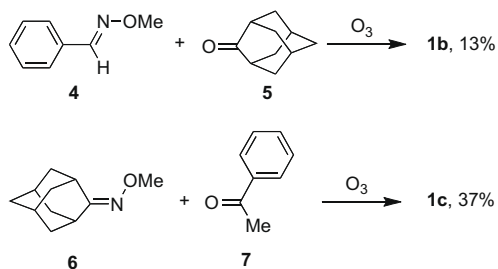

Figure 1. Artemisinin and OZ277.

**Figure 2.** Trioxolanes **1**, trioxanes **2**, and trioxepanes **3**.

Scheme 1.

Characterization of the reaction products indicated that β-hydroperoxy alcohol **9** underwent acid-catalyzed O–O bond (pathway I) and C–O bond (pathway II) heterolysis followed by rearrangement. Formation of ketone **7** (pathway I) can be rationalized by a Grob-type fragmentation or by migration of the hydroxymethyl group.[17] This contrasts with the phenyl migration seen in the acid-catalyzed decomposition of cumene hydroperoxide to form phenol and acetone.[18] Aldehyde **11** may arise from the formation and rearrangement of a relatively stable benzylic tertiary carbocation in the transition state for fragmentation, with the co-generated hydrogen peroxide producing lactone **12** via an acid-catalyzed Baeyer–Villiger oxidation of adamantanone Thus, acid-catalyzed heterolytic peroxide bond fragmentation (pathway II) was the dominant reaction course for **9**. Therefore, we tried protecting the hydroperoxide functional group of **9** to suppress the fragmentation.[19] Treatment of **9** with N,O-bis-(trimethylsilyl)acetamide (BSA) in CH₂Cl₂ gave the corresponding bis(trimethylsilylated) product **10** in high yield. Subsequent condensation of **10** with **5**

in the presence of 10–20% CSA in CH₂Cl₂ afforded the desired **2c** (Scheme 2). Trioxane **2a** was prepared as previously described.[20]

Trioxepanes **3a** and **3b** were prepared via parallel routes (Scheme 3) beginning with alcohol **13**[21] and cinnamyl alcohol (**14**). The corresponding acetates (**15**, **16**) underwent successive Co-mediated dioxygenation[22] and chemoselective reduction to furnish triethylsilyl peroxyalcohols **17** and **18**.[23] The peroxyalcohols underwent HF-mediated condensation with ketone **5** to produce **3a** and **3b**.

In vitro and in vivo antimalarial activities[9] (Table 1) were measured using the chloroquine-resistant K1 and chloroquine-sensitive NF54 strains of *Plasmodium falciparum* and *Plasmodium berghei*-infected mice. Several observations arise from the antimalarial data. First, trioxolane **1b** was more than two orders of magnitude less potent than trioxolanes **1a** and **1c**. Because it has a α-H atom, **1b** may lack the necessary chemical stability[24] required for antimalarial activity. This contrasts with the good antimalarial
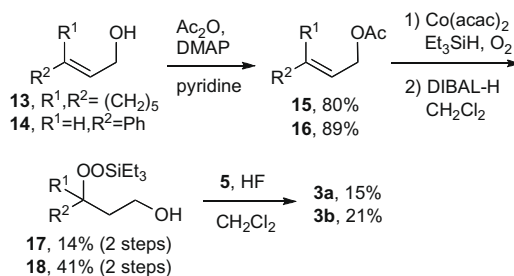
Scheme 3.
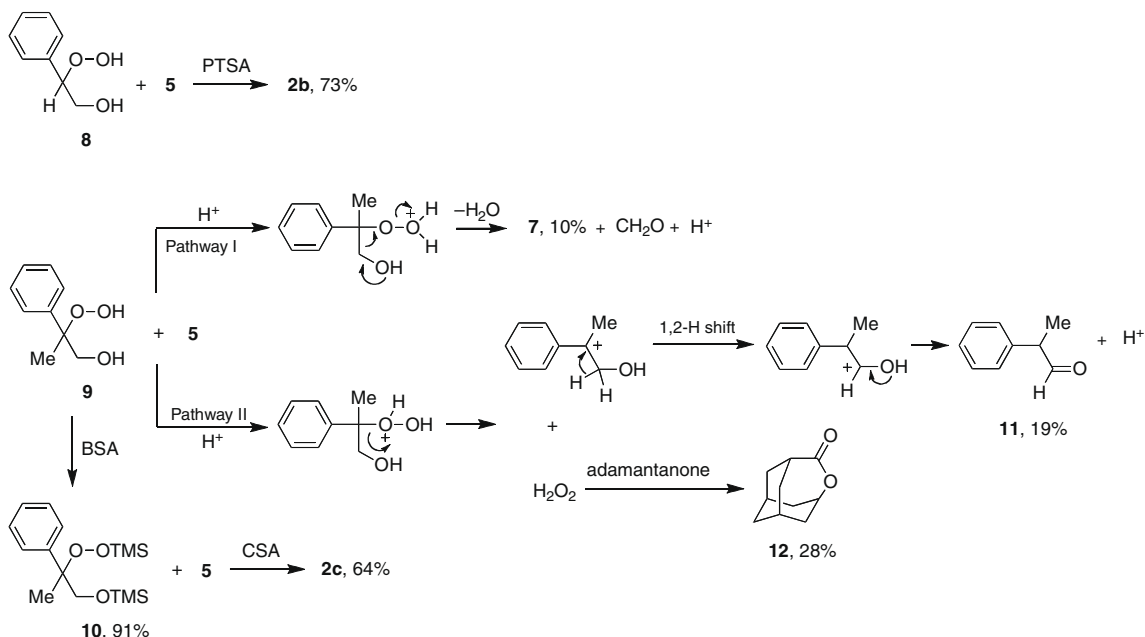
Scheme 2.

Table 1
Activity of **1**–**3** against *P. falciparum* in vitro and *P. berghei* in vivo

Compd	IC$_{50}$ (ng/ml)[a]		Activity[b] (%) po
	K1	NF54	
NONE	—	—	0
1a[c]	0.97	1.4	>99.99
1b	400	>1000	25
1c	2.9	3.4	99.0
2a[d]	54	44	0
2b	9.1	19	96
2c	19	22	0
3a	ND[e]	>1000	0
3b	ND[e]	>1000	ND[e]
ART[c,f]	1.6	2.8	98

[a] Mean from *n* = 2–3 against chloroquine-resistant (K1) and chloroquine-sensitive (NF54) strains of *P. falciparum*.
[b] Groups of three *P. berghei*-infected NMRI mice were treated orally one day post-infection with trioxolanes (100 mg/kg) dissolved or suspended in 3% ethanol and 7% Tween 80. Antimalarial activity was measured by percent reduction in parasitemia on day 3 post-infection. Individual measurements generally differed by less than 10%.
[c] Data from Dong et al.[15]
[d] Data from Tang et al.[20]
[e] Not determined.
[f] ART = artemisinin.

activity of the corresponding and presumably more chemically stable trioxane **2b**. Trioxanes **2a** and **2c** were less potent than the corresponding trioxolanes **1a** and **1c**; neither **2a** nor **2c** had any activity in vivo. The less accessible peroxide bond in the trioxane vs trioxolane heterocycle[20] may explain why trioxanes **2a** and **2c** are less active than their trioxolane chemical cousins **1a** and **1c**. Similarly, the complete lack of activity of trioxepanes **3a** and **3b** may arise from peroxide bonds that are too sterically hindered (vide infra). These data suggest that antimalarial activity for these spiroadamantyl trioxolanes, trioxanes, and trioxepanes depends on a finely tuned balance between peroxide bond shielding and accessibility. To provide mechanistic insight into this hypothesis, we examined the reactivity of several of these compounds with iron(II) in the form of FeSO$_4$ and heme.

The reactivity of **1**–**3** with inorganic ferrous iron was determined using previously established standardized conditions.[25] In these experiments, pseudo-first order reaction rate constants for **1b**, **2a**, **2b**, **3a**, and **3b** (0.03 mM) with FeSO$_4$ (3 mM) in 50% acetonitrile/water at 37 °C under an argon atmosphere were determined. These data (*n* = 3) were corrected for non-specific degradation in iron-free controls. Under these experimental conditions, the previously determined pseudo-first order rate constants (*k*) for **1a** and artemisinin were 0.41 ± 0.02 h^{-1}[26] and 0.054 ± 0.006 h^{-1}.[25] The weakly active **1b** reacted considerably more rapidly (*k* = 1.77 ± 0.06 h^{-1}) with iron(II) than did **1a** confirming our previous observations[26,27] that chemically reactive peroxides do not necessarily possess high antimalarial activities. Trioxane **2b** reacted with iron(II) at a rate (*k* = 0.12 ± 0.03 h^{-1}) between that of **1a** and artemisinin, whereas trioxepane **3b** reacted much more slowly (*k* = 0.021 ± 0.006 h^{-1}), and neither **2a** nor **3a** underwent statistically significant degradation over the 24 h time course studied (*k* = 0.011 ± 0.013 h^{-1} and 0.017 ± 0.012 h^{-1}, respectively).

As has been previously demonstrated for **1a** versus **2a**,[20] the difference in iron-mediated reactivity between **1a** and **3a** can be explained by considering peroxide bond accessibility of the more reactive equatorial peroxide bond conformers[28] as determined by molecular modeling (Spartan'08, Wavefunction, Inc.). As shown in Figure 3, the peroxide bond LUMO in **1a** is easily accessible from the cyclohexane side, whereas the peroxide bond LUMO in **3a** is
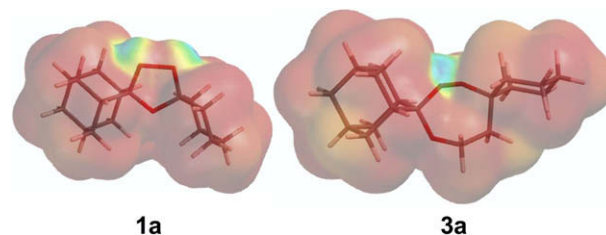
Figure 3. LUMO (blue) mapped on isodensity surfaces of **1a** and **3a**.

not accessible from either direction, consistent with the low reactivity of **3a** with iron(II). The greater steric crowding in **3a** is due to the methylene groups of the trioxepane ring decreasing the angle between the adamantane and cyclohexane rings. This lack of iron-mediated reactivity for trioxepane **3a** is consistent with its lack of antimalarial activity; similar arguments have been put forth to explain the low antimalarial activity of other sterically congested 1,2,4-trioxepanes.[29]

We have previously shown[30] a good correlation between in vitro antimalarial activity and heme alkylation efficiency for a series of 22 trioxolanes. We measured the efficiency of heme alkylation for these trioxolanes, trioxanes and trioxepanes using these same experimental conditions.[30] Significant heme alkylation was observed within 30 s for the active **1c** (67 ± 2%) and moderately active **2b** (55 ± 4%), and to a slightly lesser extent for the less active **1b** (50 ± 1%) and **2a** (45 ± 1%); however the extent of heme alkylation by these compounds was far lower than previously reported for the highly active trioxolanes **1a** (86 ± 0.2%) and OZ277 (83 ± 2%).[30] Trioxepanes **3a** and **3b** mediated very little heme alkylation (25 ± 1% and 1 ± 0.4%), in agreement with their lack of antimalarial activity. The investigation of heme alkylation appears to give a more accurate prediction of antimalarial activity than investigation of iron-mediated reaction rates, suggesting that antimalarial activity is not merely dependant on peroxide bond cleavage, but also on the ability of reactive intermediates to alkylate heme or other proximal targets. This further supports a likely role for heme alkylation in the antimalarial mechanism of action of peroxide antimalarials.[6]

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10. *Adamantane-2-spiro-3′-5′-phenyl-1′,2′,4′-trioxolane* (**1b**):[11] ^{1}H NMR (500 MHz, CDCl$_3$) δ 1.70–1.93 (m, 9H), 2.03–2.16 (m, 4H), 2.22–2.26 (m, 1H), 6.09 (s, 1H), 7.36–7.46 (m, 3H), 7.50–7.56 (m, 2H). ^{13}C NMR (125.7 MHz, CDCl$_3$) δ 26.5, 26.8, 34.2, 34.6, 34.9, 35.7, 35.8, 36.78, 36.8, 103.7, 113.0, 127.9, 128.5, 130.3, 132.8. Anal. Calcd for C$_{17}$H$_{20}$O$_3$: C, 74.97; H, 7.40. Found: C, 74.94; H, 7.19. Adamantane-2-spiro-3′-5′-methyl-5′-phenyl-1′,2′,4′-trioxolane (**1c**): mp 70–

72 °C. ^{1}H NMR (500 MHz, CDCl$_3$) δ 1.72 (s, 3H), 1.56–2.20 (m, 14H), 7.28–7.38 (m, 3H), 7.50–7.56 (m, 2H). ^{13}C NMR (125.7 MHz, CDCl$_3$) δ 25.9, 26.5, 26.9, 34.5, 34.75, 34.78, 35.3, 35.4, 36.6, 36.8, 108.7, 112.9, 125.2, 128.06, 128.11, 142.6. Anal. Calcd for C$_{18}$H$_{22}$O$_3$: C, 75.50; H, 7.74. Found: C, 75.58; H, 7.63. Adamantane-2-spiro-3′-6′-phenyl-1′,2′,4′-trioxane (**2b**): mp 110–111 °C. ^{1}H NMR (500 MHz, CDCl$_3$) δ 1.60–2.16 (m, 13H), 3.07 (br, s, 1H), 3.83 (dd, J_1 = 12 Hz, J_2 = 3.5 Hz, 1H), 4.12 (dd, J_1 = 12 Hz, J_2 = 10.5 Hz, 1H), 5.30 (dd, J_1 = 10.5 Hz, J_2 = 3.5 Hz, 1H), 7.30–7.40 (m, 5H). ^{13}C NMR (125.7 MHz, CDCl$_3$) δ 27.16, 27.17, 29.4, 33.0, 33.3, 33.5, 33.7, 36.3, 37.2, 62.7, 81.3, 104.5, 127.3, 128.7, 129.1, 135.3. Anal. Calcd for C$_{18}$H$_{22}$O$_3$: C, 75.50; H, 7.74. Found: C, 75.76; H, 7.60; Adamantane-2-spiro-3′-6′-methyl-6′-phenyl-1′,2′,4′-trioxane (**2c**): mp 61–62 °C, ^{1}H NMR (500 MHz, CDCl$_3$) δ 1.44–2.04 (br m, 16H), 3.02 (br s 1H), 3.75 (br s 1H), 4.16 (br s 1H), 7.25–7.60 (m, 5H). ^{13}C NMR (125.7 MHz, CDCl$_3$) δ 21.9 (br), 25.4 (br), 27.14, 27.17 (br), 29.7 (br), 33.2 (br), 33.3, 33.5, 36.2 (br), 37.2, 65.7 (br), 79.8 (br), 104.4, 124.7 (br), 127.6 (br), 128.3, 141.6 (br). Anal. Calcd for C$_{19}$H$_{24}$O$_3$: C, 75.97; H, 8.05. Found: C, 76.16; H, 8.24; Adamantane-2-spiro-3′-7′-cyclohexyl-1′,2′,4′-trioxepane (**3a**): ^{1}H NMR (400 MHz, CDCl$_3$) δ 1.30–1.86 (20H), 1.89–2.02 (m, 3H), 2.02–2.15 (m, 2H), 2.40 (br s 1H), 3.69 (dt, J_{Ha} = 3.7, J_{Hb} = 3.7, 1H), 3.69 (dt, J = 12.4, 3.7, 1H), 3.90 (apparent pentet, J = 6.2, probably dt, J = 12.2, 6.2, 1H), ^{13}C NMR (100.6 MHz, CDCl$_3$) δ 22.0, 22.4, 26.0, 27.3, 32.9, 33.6, 33.7, 33.8, 33.9, 34.2, 34.6, 35.7, 37.5, 42.3, 58.1, 81.7, 107.5, HRMS-FAB calcd for C$_{18}$H$_{29}$O$_3$ (MH$^+$): 293.2117; found 293.2103. Adamantane-2-spiro-3′-7′-phenyl-1′,2′,4′-trioxepane (**3b**): ^{1}H NMR (400 MHz, CDCl$_3$) δ 1.59–1.75 (m, 6H), 1.79–1.86 (m, 2H), 1.89–2.14 (m, 6H), 2.22–2.42 (m, 2H), 3.85 (dt, J = 12.2, 3.4, 1H), 4.13 (app t, J = 11.3, 1H), 5.14 (dd, J = 11.5, 3.5, 1H), 7.29–7.40 (m, 5H), ^{13}C NMR (75.5 MHz, CDCl$_3$) δ 27.1, 27.2, 33.1, 33.6, 33.9, 34.1, 34.7, 35.0, 37.5, 39.4, 60.0, 76.6, 86.1, 108.6, 127.3, 128.4, 128.5, 138.5, HRMS-FAB calc for C$_{19}$H$_{25}$O$_3$ (MH$^+$): 301.1804; found 301.1809.

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28. The equatorial peroxide conformer provides greater access to the peroxide bond from the cyclohexane side. For **1a** and **3a**, the energy difference between the two chair conformers ($\Delta E = E_{eq} - E_{ax}$) is 0.280 and 1.71 kcal/mol, respectively.
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