Synthesis and structure of tetraols with convergent and divergent arrays of hydroxy groups

Hideki Takagi,^a Takashi Hayashi,^a Tadashi Mizutani,^{*a} Hideki Masuda^b and Hisanobu Ogoshi^a

^a Department of Synthetic Chemistry and Biological Chemistry, Graduate School of Engineering, Kyoto University, Sakyo-ku, Kyoto 606-8501, Japan

^b Department of Applied Chemistry, Nagoya Institute of Technology, Showa-ku, Nagoya 466-8555, Japan

Received (in Cambridge) 17th December 1998, Accepted 10th May 1999



As hydrogen-bonding hosts with a partially flexible framework, two types of tetrahydroxy compounds with respectively convergent and divergent arrays of hydroxy groups were prepared. The structures of the tetraol with a diethyleneoxy bridge (1a), and that with a *m*-phenylenedioxy bridge (*syn*-13a and *anti*-13b) were determined by X-ray crystallographic studies. *syn*-13a forms an inclusion crystal with 1,4-dioxane, one molecule of which is bound in the cleft of *syn*-13a and the other in the intermolecular cavity.

During the recent development of host-guest chemistry, it was recognized that the direction of recognition groups and their fluctuation are important factors to determine the selectivity of molecular recognition and the ability of self-assembly formation. Attaching interaction groups to a scaffold in a convergent fashion leads to a host that binds guest in solution, while attaching them in a divergent fashion leads to a tecton¹ that forms self-assembly either in solution or in the solid state. For such chemistry both in the liquid phase and in the crystalline phase, a number of host molecules with a rigid framework have been synthesized, such as crown ethers, cryptands, carcerands, cyclodextrins, cyclophanes, calixalenes, molecular cleft, and porphyrins.² In addition to the ability of a host to display expected intermolecular interactions to bind a guest, conformational changes triggered by intermolecular forces are another important feature of functions of host molecules as exemplified in induced-fit binding of substrates by enzymes.³ For the design of these host molecules, partial flexibility should be given to the host structure.⁴ However, controlling flexibility/rigidity of a molecule requires extensive knowledge of potential energies of all conformers, and we are still far away from our ultimate goal of fine tuning of the flexibility of the host framework structure.

As part of our programme aimed at developing a new host with controlled flexibility, we describe here the synthesis and the structural characterization of novel host molecules, tetrahydroxylic compounds, where the arrangement and dynamics of the hydroxy groups can be controlled by bridging groups with varying flexibility.

Results and discussion

Two types of tetraols were prepared, both of which have a *trans*-cyclohexane-1,3-diol unit in common. Tetraol **1** has ethyleneoxy bridging units while tetraol **13** has a *m*-phenylene unit in addition to the ethyleneoxy unit to provide flexibility to the dimeric structure. Scheme 1 shows the synthesis of the common precursor 7.5

Nucleophilic addition of an enolate anion, generated from ethyl acetate and lithium diisopropylamide, to the carbonyl group of cyclohexane-1,3-dione gave the cyclohexane-1,3diol 5 in four steps. An attempt to perform the addition reaction of two enolate anions to cyclohexane-1,3-dione in one step to directly obtain diester 5 was unsuccessful. Thus, the reaction between cyclohexane-1,3-dione and an excess of the ester enol-



Scheme 1 Reagents and conditions: i, ethylene glycol, TsOH(cat.), benzene, reflux; ii, EtOAc, LDA, THF, -78 °C; then H₃O⁺; iii, H⁺ THF; iv, (CH₂O)_n, EtOH, TsOH(cat.), benzene; then *m*-xylene; v, LiAlH₄, THF; vi, H⁺ MeOH.

ate anion afforded only the corresponding stable enolate from the β -diketone, which was unreactive toward the nucleophile. The addition reactions were, therefore, carried out stepwise *via*



Fig. 1 ¹H NMR spectra of 8 in CDCl₂CDCl₂ in the temperature range of 23.5–135 °C.

cyclic ketal protection. The ¹H NMR spectrum of 5 indicated that this diester was a 1:1 mixture of *cis* and *trans* isomers: the two sets of α -methylene proton signals at δ 2.43, 2.42 and δ 2.70, 2.60 were ascribed to the cis and trans isomers, respectively. The $R_{\rm f}$ -values (TLC) of these isomers were so close that the separation of these isomers by silica gel column chromatography on a large scale was, in practice difficult. In order to protect the two hydroxy groups of diester 5, direct ketalization was examined by using 2,2-dimethoxypropane, but failed possibly due to the strained 1,3-dioxacyclane products. The mixture of cis/trans isomers 5 was treated with paraformaldehyde under acidic conditions to give a mixed acetal as an intermediate. Subsequent thermal decomposition of the mixed acetal afforded cyclic acetal 6 selectively from the cis isomer of 5, while the trans isomer of 5 was converted to a lactone during pyrolysis. Cyclic acetal 6 and the lactone were successfully separated by silica gel column chromatography. A mixture of cis- and trans-5 was recovered from the lactone by treating it with dilute sulfuric acid. Reduction of acetal 6 with lithium aluminium hydride gave diol 7, which adopts the chair-boat conformation as confirmed by observation of ¹H NMR nuclear Overhauser enhancement (NOE) between the acetal methylene protons and the cyclohexane ring protons. After hydrolysis of diol 7 with aq. HCl, white crystalline solids were obtained; recrystallization from CHCl₃ gave 8 in 84% yield.

Tetraol 8 was characterized by ¹H and ¹³C NMR, mass spectroscopy, and elemental analysis. In the ¹H NMR spectrum, the two methylene protons at the α -positions to the primary hydroxy groups appear as magnetically inequivalent protons in CDCl₃, while they become equivalent in CD₃OD. Similar behaviour was observed for the β -methylene protons to the primary hydroxy groups. These observations indicate that intramolecular hydrogen bonding between the primary OH group and the tertiary OH group is formed in CDCl₃ and it breaks in CD₃OD. The variable-temperature ¹H NMR spectra of 8 in $CDCl_2CDCl_2$ (Fig. 1) indicated that the methylene protons α to the primary hydroxy group appear as multiplets centred at δ 3.9 at 23.5 °C and as a doublet of triplets at 135 °C. The intramolecular hydrogen bonding in an apolar solvent is thus cleaved at high temperature. In contrast, the axial and equatorial protons of the cyclohexane ring appeared separately, and this spectral pattern did not change upon heating, showing that the chair conformation with hydroxyethyl groups in the equatorial position is rather rigid. Conformational analysis by ¹H NMR of 1,3-disubstituted cyclohexanes has been reported.⁶

Vapour-pressure osmometry showed that the molecular weight of **8** was 235 in a 0–0.023 M solution in CHCl₃, suggesting that **8** is monomeric in CHCl₃.

The synthetic route to 1a is illustrated in Scheme 2. For the



Scheme 2 Reagents and conditions: i, p-TsCl, DMAP, Et₃N, CH₂Cl₂; ii, KO'Bu, DME–DMSO; iii, H⁺, MeOH.

dimer preparation, the two hydroxy groups of 7 were converted to toluene-*p*-sulfonate, methanesulfonate, chloride, bromide, or iodide as leaving groups, and the coupling reactions with 7 were attempted. Various base–solvent combinations were tried to obtain dimers **10a** and **10b**. The combination of ditosyl compound **9** and potassium *tert*-butoxide in Me₂SO gave dimers **10a** and **10b** in low yield. When other leaving groups (halogens) were used, α , β -elimination took place in preference to the coupling reaction. Treatment of a mixture of **10a** and **10b** with HCI in methanol afforded the desired dimer **1a** having *cisoid* tetrahydroxy groups. *syn* Isomer **1a** was isolated and purified by medium-pressure liquid chromatography (MPLC), and characThe *syn* geometry of **1a** was confirmed by X-ray crystallographic analysis. The two cyclohexane rings assumed a chair conformation and the hydrogen-bonding network among the four hydroxy groups stabilized the rigid conformation with a collapsed cleft between the two cyclohexane rings.⁷ The ¹H NMR spectrum of **1a** in CDCl₃ was similar to that in CD₃OD, indicating that **1a** has a rigid framework as compared to **8**. The methylene protons β to the ether oxygen were shifted downfield as the solvent changes from CDCl₃ to CD₃OD, indicating some conformational changes. In the variable-temperature ¹H NMR spectra in CDCl₂CDCl₂ from 24 to 130 °C, almost no changes in coupling constants were observed. The methylene protons β to the ether oxygen were shifted downfield upon heating, which can be attributed to changes in conformation or solvation.

For the synthesis of dimers **13a** and **13b**, dimesyl compound **11** was employed as a precursor (Scheme 3). The reaction of



Scheme 3 Reagents and conditions: i, MsCl, Et₃N, CH₂Cl₂; ii, Ph₃C⁺- BF_4^- , CH₂Cl₂; then aq. NaHCO₃; iii, Cs₂CO₃, DMF.

cyclic acetal **11** with resorcinol in the presence of caesium carbonate in the DME–DMF mixed solvent system also gave corresponding cyclic acetal dimers in 10% yield, but various attempts to deprotect the methylenedioxy group were unsuccessful. This is presumably due to a side reaction in which the generated formyl cation reacts with the resorcinol group under acidic conditions. Therefore, the cyclic acetal had to be removed prior to the coupling of the dimesyl unit with resorcinol to prevent the foregoing reaction. The cyclic acetal protecting group of **11** was removed by trityl tetrafluoroborate–

Table 1Crystal data for 13a · 1.5C4H8O2

| Formula | C ₃₈ H ₅₆ O ₁₁ |
|---|--|
| Formula weight | 688.85 |
| Space group | C2/c |
| aĺÅ | 21.458(2) |
| b/Å | 16.518(1) |
| c/Å | 21.437(2) |
| β/° | 95.581(6) |
| Vol./Å ³ | 7562.5(8) |
| Ζ | 8 |
| ρ (calc.)/g cm ⁻³ | 1.21 |
| Temp. T/°C | 23 |
| Crystal dimensions/mm | $0.25 \times 0.25 \times 0.25$ |
| Radiation | graphite-monochromated Cu- K_{α} radiation and a 12 kW rotating anode generator |
| Linear absorption coeff./cm ⁻¹ | 7.21 |
| Detector aperture | 9.0 mm horizontal |
| | 13.0 mm vertical |
| Scan type | ω –2 θ |
| Scan rate | $16.0^{\circ} \min^{-1} in \omega$ |
| 2θ limits | $2.0^{\circ} < 2\theta < 120.1^{\circ}$ |
| Reflections measured | 5477 total, 5321 unique |
| Reflections used | 3894 |
| No. of variables | 494 |
| R | 0.062 |
| R _w | 0.071 |

 CH_2Cl_2 to afford 12 in 65% yield. The combination of diol 12, resorcinol and caesium carbonate in DMF gave dimers 13a and 13b in 6% yield. Each isomer was separated successfully by MPLC, and characterized by NMR and MS spectra. The purity of the dimers was higher than 95% as checked by ¹H NMR and ¹³C NMR spectroscopy. The aromatic carbons of 13a and 13b in the ¹³C NMR spectra in CDCl₃ and CD₃OD are diagnostic: syn-13a displayed signals at $\delta_{\rm C}$ 159.6 (C-1, -3) and at $\delta_{\rm C}$ 107.5 (C-4, -6), while anti-13b displayed signals at $\delta_{\rm C}$ 159.71 (C-1), 157.87 (C-3), 108.26 (C-4 or 6), 106.25 (C-6 or 4), whose assignments were made by ¹H-¹³C proton-detected singlequantum coherence (HSQC) experiments. The number of magnetically inequivalent aromatic carbons observed in 13b was consistent with its anti structure. From the ¹H NMR spectra, it was difficult to distinguish between the two isomers owing to their similar coupling patterns. In the variable-temperature ¹H NMR spectra of 13a in CDCl₂CDCl₂, the aromatic 5-H (δ 7.11) was shifted upfield by 0.04 ppm and the aromatic 4-H and 6-H upfield by 0.02 ppm as the temperature was varied from 25 to 100 °C, suggesting that conformational changes of the bridge occurred with increasing temperature.

By X-ray diffraction crystallographic study, the crystal structures of 1a, 13a, and 13b were determined. 13a crystallizes in the monoclinic space group C2/c with eight molecules per unit cell. 13b crystallizes in the monoclinic space group $P2_1/c$ with two molecules per unit cell. Crystal data are listed in Tables 1 and 2. As shown in Fig. 2, both 13a and 13b have a cavity consisting of two benzene rings and two cyclohexane rings. The size of the cavity is 5.9 Å (the H · · · H distance between the closest hydrogens of the cyclohexane rings) \times 6.3 Å (the H \cdots H distance between the closest hydrogens of the benzene rings) for 13a and 5.8×6.4 Å for 13b, as clearly seen in the top views. As the side views show, the benzene rings are almost parallel to the cyclohexane rings for 13a, while they are nearly perpendicular to them for 13b. It is noteworthy that the syn isomer, 13a, forms an inclusion crystal with 1,4-dioxane, while other tetraols, 1a and 13b, do not. Two crystallographically different 1,4-dioxane molecules are included in the crystal. One is bound with the shallow cleft of 13a as shown in Fig. 3. The other is bound in an intermolecular cavity (not shown). In the crystal, 13a assumes a conformation where the cyclohexane ring is nearly coplanar to the neighboring molecule, and the hydroxy groups form hydrogen bonds with those in the next molecule to form a linear array of molecules. This arrangement of 13a makes intermolecular



Fig. 2 Structures of (a) 13a and (b) 13b obtained from X-ray crystallographic studies.

Table 2 Crystal data for 13b

| Formula | C:nH40. |
|---|---|
| Formula weight | 556.69 |
| Space group | $P2_1/c$ |
| a/Å | 11.727(4) |
| b/Å | 10.915(4) |
| c/Å | 12.109(4) |
| β/° | 112.25(2) |
| Vol./Å ³ | 1434.6(8) |
| Z | 2 |
| ρ (Calc.)/g cm ⁻³ | 1.289 |
| Temp. T/°C | 23 |
| Crystal dimensions/mm | $0.40 \times 0.30 \times 0.20$ |
| Radiation | graphite-monochromated |
| | $Mo-K_a$ radiation and a 12 kW |
| | rotating anode generator |
| Linear absorption coeff./cm ⁻¹ | 0.91 |
| Detector aperture | 6.0 mm horizontal |
| | 6.0 mm vertical |
| Scan type | $\omega - 2\theta$ |
| Scan rate | $16.0^{\circ} \operatorname{min}^{-1} \operatorname{in} \omega$ |
| 2θ limits | $2.0^{\circ} < 2\theta < 60.0^{\circ}$ |
| Reflections measured | 4584 total, 4395 unique |
| Reflections used | 2939 |
| No. of variables | 270 |
| R | 0.036 |
| R _w | 0.052 |
| | |

cavities in the crystal, in which the second 1,4-dioxane is included (Fig. 4).

Experimental

General remarks

¹H NMR spectra were recorded on a JEOL A-500 (500 MHz), a JEOL GX-400 (400 MHz), and a JEOL JNM FX 90Q (90 MHz) FT NMR spectrometer. Chemical shifts are reported in parts per million and referenced to internal CHCl₃ (¹H δ 7.25) or CDCl₃ (¹³C δ 77.0). Coupling constants are reported in hertz (Hz) and splitting patterns are designated as s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), AB q (AB quartet), br (broad). IR spectra were recorded on a BIO-RAD FTS-7 FT-IR spectrometer. Mass spectra were obtained with a JEOL JMS DX-300 mass spectrometer. Elemental analyses were



Fig. 3 Structures of a complex between **13a** and 1,4-dioxane obtained from X-ray crystallographic studies.

performed at the Microanalysis Center of the Department of Pharmaceutical Sciences, Kyoto University. Analytical TLC was performed on plates coated with a 0.25 mm thickness of silica gel 60-F254 (Merck). Column chromatography was performed using Merck Kieselgel 60 (70–230 mesh) or Pharma basic aluminium oxide activity grade I. Gas–liquid chromatography was conducted on a Shimadzu GC-4B gas chromatograph, equipped with a 3 m and a 1 m glass column packed with 30% silicone DC550 on Celite 545 and silicone SE30. MPLC was performed with a silica gel pre-packed column [Lobar Größe A(240-10) LiChroprep Si 60 (40–63 µm, Merck)]. Single-crystal X-ray diffraction was performed on a Rigaku AFC7R diffractometer.

Materials

Unless otherwise noted, materials were obtained from commercial sources and used after distillation under nitrogen. THF, diethyl ether, *n*-hexane, triethylamine, DME and diisopropylamine were distilled over sodium benzophenone ketyl and stored under nitrogen. Ethyl acetate, DMSO, DMF and 1,2-dichloroethane were distilled from CaH_2 , and 1,4-dioxane from LiAlH₄.

1,4-Dioxaspiro[4.5]decan-7-one 2

This compound was prepared according to the reported method.⁸ A mixture of cyclohexane-1,3-dione (75.0 g, 0.669



Fig. 4 Perspective view of the crystal structure of 13a-1.5 1,4-dioxane.

mol), ethylene glycol (41.5 g, 0.669 mol) and toluene-*p*-sulfonic acid monohydrate (TsOH) (1.27 g, 6.69 mmol) in 1.8 l of benzene was efficiently stirred and refluxed for 4.5 h. Slightly more than the theoretical amount of water was collected in a Dean–Stark trap. The benzene solution was washed successively with saturated aq. NaHCO₃ (60 ml) and saturated aq. NaCl (50 ml × 2). After drying of the solution over anhydrous MgSO₄ the solvent was distilled off and the product **2** was purified by vacuum distillation at 62–75 °C/0.15 mmHg (71.5 g, 60%); ¹H NMR (90 MHz, CDCl₃) δ 1.75–2.46 (m, 6H), 2.58 (s, 2H), 3.95 (s, 4H, OCH₂CH₂O).

7-Ethoxycarbonylmethyl-7-hydroxy-1,4-dioxaspiro[4.5]decane 3

To a solution of lithium diisopropylamide (LDA) (0.521 mol) in dry THF (320 ml) was added dropwise ethyl acetate (45.9 g, 0.521 mol) at -78 °C under nitrogen with stirring. Stirring was continued for an additional hour. A solution of ketal 2 (71.5 g, 0.401 mol) in dry THF (100 ml) was added dropwise to the reaction mixture. After being stirred for 30 min, the reaction mixture was hydrolyzed by adding 4 M hydrochloric acid (200 ml) in one portion. The organic layer was separated. The aqueous layer was neutralized and extracted several times with diethyl ether. The combined organic extracts were washed with saturated aq. NaCl and dried over anhydrous MgSO4. The solvent was evaporated under reduced pressure to give 109.4 g of a pale yellow oil, consisting mainly of the desired product as determined by GLC. This material was used directly in the next step without further purification. For analysis, a small portion of 3 was purified by column chromatography (SiO₂, hexane-EtOAc = 2:1); TLC (silica gel) $R_f 0.24$ (hexane-EtOAc = 2:1); ¹H NMR (90 MHz, CDCl₃) δ 1.26 (t, J = 7.2 Hz, 3H, CO₂CH₂CH₃), 1.37–2.07 (m, 8H), 2.53 (s, 2H, CH₂CO₂Et), 3.97 $(s, 4H, OCH_2CH_2O), 4.18 (q, J = 7.2 Hz, 2H, CO_2CH_2CH_3); IR$ (liquid film, cm⁻¹) 3512m, 2945w, 2882w, 1731s (C=O).

Ethyl (1-hydroxy-3-oxocyclohexyl)acetate 4

To the crude product **3** (109.4 g) obtained above were added THF (80 ml) and 10.5 M hydrochloric acid (80 ml) and the mixture was stirred at ambient temperature for 9 h. To this reaction mixture was added aq. NaHCO₃ until the aqueous layer was neutralized. The aqueous layer was extracted several times with diethyl ether, and the combined extracts were washed with saturated aq. NaCl and dried over Na₂SO₄. The solvent was removed and the residue was distilled under reduced pressure to yield a liquid (69.1 g, 68% based on monoketal **2**), bp 100–136 °C/0.25–0.3 mmHg; ¹H NMR (90 MHz, CDCl₃) δ 1.27 (t, *J* = 7.3 Hz, 3H, CO₂CH₂CH₃), 1.45–2.54 (m, 8H), 2.54 (s, 2H, CH₂CO₂Et), 4.21 (q, *J* = 7.3 Hz, 2H, CO₂CH₂CH₃); IR (liquid film, cm⁻¹) 3488m, 2959w, 1714s (C=O).

Diethyl 1,3-dihydroxycyclohexane-1,3-diacetate 5

To a solution of LDA (0.346 mol) in dry THF (250 ml) was added dropwise dry ethyl acetate (30.5 g, 0.346 mol) at -78 °C under nitrogen with stirring. Stirring was continued for 1 h. A solution of the hydroxycyclohexanone 4 (23.1 g, 0.115 mol) in dry THF (150 ml) was added dropwise to the reaction mixture. After being stirred for 1 h, the reaction mixture was hydrolyzed by adding 4 M hydrochloric acid (170 ml) in one portion, and the stirred solution was warmed to room temperature. The organic layer was separated, the aqueous layer was neutralized, and extracted several times with diethyl ether, and the combined extracts were washed with saturated aq. NaCl and dried over anhydrous Na₂SO₄. The solvent was removed and the residue was distilled under reduced pressure to yield (31.3 g, 94%) of a pale yellow liquid (mixture of cis and trans isomers); bp 138-151 °C/0.25 mmHg. The ¹H NMR spectrum of the product revealed that the ratio of the two isomers was ca. 1:1. For analysis, the two isomers were separated by column chromatography (SiO₂, hexane-EtOAc = 3:1): cis isomer; TLC (silica

gel) $R_{\rm f}$ 0.26 (hexane-EtOAc = 2:1); ¹H NMR (400 MHz, CDCl₃) δ 1.26 (t, J = 7.1 Hz, 6H, CO₂CH₂CH₃), 1.32 (ddd, ${}^{2}J = 13.7$ Hz, ${}^{3}J = 4.3$, 13.7 Hz, 2H), 1.57 (d, J = 14.0 Hz, 1H), 1.52–1.60 (m, 1H), 1.85–1.79 (m, 2H), 1.95 (dt, ${}^{2}J = 14.0$ Hz, ${}^{4}J = 2.6$ Hz, 1H), 2.01 (dtt, ${}^{2}J = 13.8$ Hz, ${}^{3}J = 3.7$, 13.8 Hz, 1H), 2.42 ($\frac{1}{2}$ AB q, J = 15.0 Hz, 2H, CH₂CO₂Et), 2.43 ($\frac{1}{2}$ AB q, J = 15.0 Hz, 2H, CH₂CO₂Et), 4.15 (q, J = 7.1 Hz, 4H, CO₂-CH₂CH₃); ¹³C NMR (100 MHz, CDCl₃) δ 14.15, 16.67, 36.31, 44.32, 46.87, 60.58, 71.47, 171.80; IR (liquid film, cm⁻¹) 3454 (br), 2981m, 2939m, 2875m, 1732s (C=O), 1196m: trans isomer; TLC (silica gel) $R_f 0.29$ (hexane–EtOAc = 2:1); ¹H NMR (400 MHz, CDCl₃) δ 1.27 (t, J = 7.1 Hz, 6H, CO₂CH₂CH₃), 1.49– 1.68 (m, 6H), 2.60 ($\frac{1}{2}$ AB q, J = 16.0 Hz, 2H, CH₂CO₂Et), 2.70 $(\frac{1}{2}AB q, J = 16.0 Hz, 2H, CH_2CO_2Et), 4.17 (q, J = 7.1 Hz, 4H)$ $CO_2CH_2CH_3$); IR (liquid film, cm⁻¹) 3501br, 2982m, 2940m, 2874m, 1727s, 1712s (C=O), 1192m.

Diethyl 2,4-dioxabicyclo[3.3.1]nonane-1,5-diacetate 6

The cyclohexane-1,3-diol 5 (5.03 g, 0.017 mol, a mixture of cis and trans) was mixed with paraformaldehyde (3.14 g, 0.105 mol) in a mixture of ethanol (10.1 ml) and benzene (30 ml). After addition of TsOH (0.066 g), the mixture was heated at 50 °C (bath temp.) for 1 h and refluxed for 3 h with a Dean-Stark trap. Evaporation of the solvent afforded the mixed acetal. To this residue was added m-xylene (20 ml) and the reaction mixture was heated at 150 °C for 2 h. The solvent was removed under reduced pressure, and the residue was purified by column chromatography (SiO₂, hexane–EtOAc = 4:1) to give 2.18 g (42%) of a colorless oil 6; TLC (silica gel) $R_{\rm f}$ 0.31 (hexane–EtOAc = 4:1); MS m/z 300 (M⁺), 299 (M⁺ – H); ¹H NMR (400 MHz, CDCl₃) δ 1.26 (t, J=7.1 Hz, 6H, $CO_2CH_2CH_3$), 1.31 (ddd, ²J = 13.5 Hz, ³J = 4.9, 13.5 Hz, 2H), 1.50 (d, J = 14.3 Hz, 1H), 1.50–1.56 (m, 1H), 1.70–1.76 (m, 2H), 2.14 (dtt, ${}^{2}J = 13.4$ Hz, ${}^{3}J = 4.8$, 13.4 Hz, 1H), 2.44 (${}^{1}_{2}AB$ q, J = 14.6 Hz, 2H, CH₂CO₂Et), 2.54 ($\frac{1}{2}$ AB q, J = 14.6 Hz, 2H, CH_2CO_2Et), 2.916 (dt, ²J = 14.4 Hz, ⁴J = 2.7 Hz, 1H), 4.12 $(\frac{1}{2}AB q, J = 7.2, 10.8 Hz, 2H, CH_2CO_2Et), 4.14 (\frac{1}{2}AB q, 1)$ ${}^{2}J = 10.8$ Hz, ${}^{3}J = 7.2$ Hz, 2H, CH₂CO₂Et), 4.70 (d, J = 5.7 Hz, 1H, OCH₂O), 5.12 (d, *J* = 5.7 Hz, 1H, OCH₂O).

2,4-Dioxabicyclo[3.3.1]nonane-1,5-diethanol 7

To a suspension of lithium aluminium hydride (1.913 g, 0.050 mol) in dry THF (45 ml) was added dropwise a solution of diester 6 in dry THF (15 ml) at 0 °C under nitrogen, and the mixture was stirred at ambient temperature for 1 h. To the mixture were added water (1.9 ml), 15% aq. sodium hydroxide (1.9 ml), and water (5.7 ml) in this order with vigorous stirring. The precipitate was filtered off. After concentration of the filtrate, the residue was purified by column chromatography on silica gel (EtOAc-ethanol = 10:1) to afford a white solid 7. (4.19 g, 92%); TLC (silica gel) $R_f 0.33$ (hexane-EtOAc = 10:1); MS m/z 216 (M⁺), 215 (M⁺ – H); ¹H NMR (400 MHz, CDCl₃) δ 1.16 (ddd, ²J = 13.5 Hz, ³J = 5.0, 13.5 Hz, 2H), 1.23 (d, J = 14.4 Hz, 1H), 1.53–1.61 (m, 1H), 1.66 ($\frac{1}{2}$ AB q dd, $^{2}J = 14.6$ Hz, ${}^{3}J = 4.3$, 6.0 Hz, 2H, CH₂CH₂OH), 1.81 ($\frac{1}{2}$ AB q dd, ${}^{2}J = 14.6$ Hz, ${}^{3}J = 4.9$, 8.2 Hz, 2H, CH₂CH₂OH), 1.85–1.90 (m, 2H), 2.09 (dtt, ${}^{2}J = 13.3$ Hz, ${}^{3}J = 4.8$, 13.3 Hz, 1H), 2.43 (dt, ${}^{2}J = 14.3$ Hz, ${}^{3}J = 2.7$ Hz, 1H), 3.76 ($\frac{1}{2}$ AB q dd, ${}^{2}J = 11.2$ Hz, ${}^{3}J = 4.9, 6.1$ Hz, 2H, CH₂CH₂OH), 3.89 ($\frac{1}{2}$ AB q dd, ${}^{2}J = 11.2$ Hz, ³*J* = 4.3, 8.2 Hz, 2H, CH₂CH₂OH), 4.73 (d, *J* = 5.7 Hz, 1H, OCH₂O), 5.11 (d, J = 5.7 Hz, 1H, OCH₂O); ¹³C NMR (100 MHz, CDCl₃) δ 17.75, 35.88, 37.64, 43.09, 58.64, 75.22, 84.14; IR (liquid film, cm⁻¹) 3385br, 2934m, 2772w, 1155m, 1087m, 1014m.

1,3-Dihydroxycyclohexane-1,3-diethanol 8

Acetal **7** (0.088 g, 0.407 mmol) was dissolved in methanol (2 ml) and 2.0 M HCl-methanol (2 ml) was added. The mixture was

stirred at ambient temperature for 3 days. The solvent was evaporated under reduced pressure. The residue was chromatographed on a silica gel column with EtOAc-ethanol (10:1) to give 0.070 g (84%) of 8. An analytical sample was prepared by recrystallization from EtOAc-ethanol to give white crystals; mp 101–104 °C; TLC (silica gel) $R_{\rm f}$ 0.32 (EtOAc–ethanol = 10:1); MS m/z 186 (M⁺ – H₂O), 168 (M⁺ – 2H₂O); ¹H NMR (400 MHz, CDCl₃) δ 1.20 (d, J = 13.8 Hz, 1H), 1.23 (ddd, ²J = 13.7 Hz, ${}^{3}J = 4.4$, 13.7 Hz, 2H), 1.52–1.59 (m, 1H), 1.63 ($\frac{1}{2}$ AB q dd, ${}^{2}J = 14.8$ Hz, ${}^{3}J = 3.8$, 6.3 Hz, 2H, CH₂CH₂OH), 1.73 ($\frac{1}{2}$ AB q dd, ${}^{2}J = 14.8$ Hz, ${}^{3}J = 4.0$, 8.1 Hz, 2H, $CH_{2}CH_{2}OH$), 1.89–1.83 (m, 2H), 1.99 (dtt, ${}^{2}J = 13.8$ Hz, ${}^{3}J = 3.7$, 13.8 Hz, 1H), 2.12 (dt, $^{2}J = 13.9$ Hz, $^{4}J = 2.6$ Hz, 1H), 3.09 (t, J = 3.8 Hz, 2H, primary OH), 3.85–4.00 (m, 4H, CH₂CH₂OH), 4.781 (s, 2H, tertiary OH); ¹³C NMR (22.5 MHz, CDCl₃) δ 16.71, 37.64, 43.58, 44.58, 59.16, 74.08; IR (liquid film, cm⁻¹) 3338br, 2939m, 1449w, 1422m, 1150m, 1029m, 852m (Calc. for C₁₀H₂₀O₄: C, 58.80; H, 9.87. Found: C, 58.59; H, 9.82%).

Ditosyl unit 9

The procedure of Marshall⁹ was modified. To a stirred, cooled (0 °C) solution of diol 7 (4.501 g, 0.021 mol), 4-(dimethylamino)pyridine (DMAP) (130 mg), and triethylamine (8.7 ml, 0.062 mol) in CH₂Cl₂ (60 ml) was added toluene-p-sulfonyl chloride (p-TsCl) (11.11 g, 0.058 mol). The reaction mixture was stirred for 8.5 h at ambient temperature. The mixture was treated with diethyl ether (50 ml) and filtered, and the filter cake was washed with diethyl ether. The filtrate was evaporated under reduced pressure. The residue was purified by column chromatography on silica gel (hexane-EtOAc = 2:1) to afford 9.997 g (92%) of ditosyl compound 9 as a viscous pale yellow oil; ¹H NMR (90 MHz, CDCl₃) δ 1.79 (t, J = 7.4 Hz, 4H, CH2CH2O), 0.97-2.20 (m, 8H), 2.44 (s, 6H, aryl CH3), 4.17 (t, J = 6.8 Hz, 4H, CH₂CH₂O), 4.56 ($\frac{1}{2}$ AB q, J = 5.9 Hz, 1H, OCH₂O), 4.75 ($\frac{1}{2}$ AB q, J = 5.9 Hz, 1H, OCH₂O), 7.39 ($\frac{1}{2}$ AB q, J = 8.7 Hz, 4H, ArH), 7.82 ($\frac{1}{2}$ AB q, J = 8.7 Hz, 4H, ArH); IR (liquid film, cm⁻¹) 2950m, 2769w 1358s, 1190s, 1176s; MS m/z 524 (M⁺), 481 (M⁺ - 43).

Dimesyl unit 11

To a stirred, cooled (0 °C) solution of diol 7 (0.956 g, 4.42 mmol) and triethylamine (1.54 ml, 11.05 mmol) in CH₂Cl₂ (15 ml) was added methanesulfonyl chloride (MsCl) (1.114 g, 9.73 mmol). The reaction mixture was stirred for 4 h at ambient temperature. A small amount of white solid was precipitated. The mixture was filtered, and the mother liquor was evaporated under reduced pressure. The residue was purified by column chromatography on silica gel (CHCl₃–EtOAc = 6:1) to afford dimesyl compound **11**, as a colorless oil, which slowly solidified upon storage at room temperature *in vacuo* to give a white solid (1.390 g, 84%); ¹H NMR (90 MHz, CDCl₃) δ 1.02–2.28 (m, 7H), 1.94 (t, *J* = 6.6 Hz, 4H, CH₂CH₂O), 2.33–2.61 (m, 1H), 3.04 (s, 6H, CH₃), 4.29–4.57 (m, 4H), 4.80 ($\frac{1}{2}$ AB q, *J* = 5.9 Hz, 1H, OCH₂O), 5.09 ($\frac{1}{5}$ AB q, *J* = 5.9 Hz, 1H, OCH₂O).

cis-1,3-Bis(2-mesylethyl)cyclohexane-1,3-diol 12

Acetal 7 (5.358 g, 14.39 mmol) and trityl tetrafluoroborate (5.894 g, 17.85 mmol) were stirred in dichloromethane (125 ml) for 7 h at ambient temperature. To the reaction mixture was added saturated aq. NaHCO₃ (50 ml) and the two-phase mixture was stirred for 15 min. The organic phase was washed with water, dried over anhydrous Na₂SO₄, and evaporated *in vacuo*. The residue was chromatographed over silica gel (CHCl₃–EtOAc = 1:1) to afford 3.394 g (65%) of **12** as an oil; ¹H NMR (90 MHz, CDCl₃) δ 1.13–2.11 (m, 8H), 1.91 (t, *J* = 6.4 Hz, 4H, CH₂CH₂O); IR (liquid film, cm⁻¹) 3485br, 2941m, 1350s, 1173s.

Acetal dimers 10a and 10b

A mixture of diol 7 (0.433 g, 2.0 mmol) and potassium tertbutoxide (0.494 g, 4.4 mmol) in dry DME (15 ml) was efficiently stirred and heated at 80 °C (bath temp.) for 1 h. A solution of ditosyl compound 9 (1.049 g, 2.0 mmol) in Me₂SO (5 ml) was added, and the reaction mixture was heated to reflux and stirred for 13 h. The suspension was cooled to 25 °C, filtered, and washed with diethyl ether. The solvent was removed under reduced pressure. The residue was partitioned between 20 ml of CH₂Cl₂ and 10 ml of water, the two layers were separated, and the aqueous layer was extracted again with two 20 ml portions of CH2Cl2. The combined organic phases were dried over Na₂SO₄. The solvent was evaporated, leaving an oil, which was chromatographed on silica gel (hexane-EtOAc = 4:1) to give 16 mg (2.1%) of a mixture of syn and anti isomers as a white solid. Analytical samples of each isomer were purified by MPLC (hexane-EtOAc = 10:1). First fraction: TLC (silica gel) $R_f 0.40$ (hexane-EtOAc = 2:1); MS m/z 395 (M⁺ -H), 393 (M⁺ - 3H), 368 (M⁺ - C_2H_4), 320; ¹H NMR (400 MHz, CDCl₃) δ 1.32 (ddd, ²J = 13.3 Hz, ³J = 4.8, 13.3 Hz, 4H), 1.51 (d, J = 15.1 Hz, 2H), 1.40–1.53 (m, 6H), 1.65 ($\frac{1}{2}$ AB q dd, ${}^{2}J = 15.0 \text{ Hz}, {}^{3}J = 2.1, 7.2 \text{ Hz}, 4\text{H}, \text{C}H_{2}\text{C}H_{2}\text{O}), 1.79 (\frac{1}{2}\text{AB q dd},$ $^{2}J = 15.0$ Hz, $^{3}J = 2.6$, 8.5 Hz, 4H, CH₂CH₂O), 2.14 (dtt, ${}^{2}J = 13.1$ Hz, ${}^{3}J = 4.8$, 13.1 Hz, 2H), 2.65 (dt, ${}^{2}J = 15.1$ Hz, ${}^{4}J = 2.5$ Hz, 2H), 3.36 (ddd, ${}^{2}J = 10.2$ Hz, ${}^{3}J = 2.1$, 8.5 Hz, 4H, CH₂CH₂O), 3.59 (ddd, ${}^{2}J = 10.2$ Hz, ${}^{3}J = 2.6$, 7.2 Hz, 4H, CH₂CH₂O), 4.74 (d, J = 5.6 Hz, 2H, OCH₂O), 5.16 (d, J = 5.6 Hz, 2H, OCH₂O); ¹³C NMR (22.5 MHz, CDCl₃) δ 18.10, 36.13, 36.60, 42.37, 67.59, 74.23, 85.22. Second fraction: TLC (silica gel) $R_f 0.40$ (hexane-EtOAc = 2:1); MS m/z 395 (M⁺ - H), 393 $(M^+ - 3H)$, 368 $(M^+ - C_2H_4)$, 320; ¹H NMR (400 MHz, $CDCl_3$) δ 1.29 (ddd, ²J = 13.4 Hz, ³J = 4.8, 13.4 Hz, 4H), 1.43 (d, J = 15.1 Hz, 2H), 1.40–1.52 (m, 6H), 1.64 ($\frac{1}{2}$ AB q dd, $^{2}J = 14.9$ Hz, ${}^{3}J = 2.3$, 7.9 Hz, 4H, CH₂CH₂O), 1.77 (${}^{1}_{2}$ AB q dd, ${}^{2}J = 14.9$ Hz, ${}^{3}J = 2.4$, 7.5 Hz, 4H, CH₂CH₂O), 2.14 (dtt, ${}^{2}J = 13.2$ Hz, ${}^{3}J = 4.6, 13.2 \text{ Hz}, 2\text{H}), 2.76 (dt, {}^{2}J = 15.1 \text{ Hz}, {}^{4}J = 2.5 \text{ Hz}, 2\text{H}),$ 3.38 (ddd, ${}^{2}J = 10.2$ Hz, ${}^{3}J = 2.4$, 7.5 Hz, 4H, CH₂CH₂O), 3.59 $(ddd, {}^{2}J = 10.2 \text{ Hz}, {}^{3}J = 2.3, 7.9 \text{ Hz}, 4\text{H}, \text{CH}_{2}\text{C}H_{2}\text{O}), 4.77 \text{ (d},$ *J* = 5.6 Hz, 2H, OCH₂O), 5.18 (d, *J* = 5.6, 2H, OCH₂O).

Tetraol dimer I 1a

Acetal 10 (mixture of syn and anti isomers) (38.8 mg, 0.098 mmol) was dissolved in 2.0 M HCl in methanol (2 ml), and the mixture was stirred at room temperature for 20 h. After evaporation of the solvent under reduced pressure, the residue was chromatographed on a silica gel column with EtOAc-CHCl₃ (4:1). The fractions containing product were combined and rechromatographed at medium pressure (EtOAc-CHCl₃ = 1:6) to give 8.4 mg (46%) of 1a (syn isomer) as a white solid. An analytical sample was prepared by recrystallization from ethyl acetate to give white crystals, mp 210-215 °C; TLC (silica gel) $R_{\rm f}$ 0.41 (EtOAc-CHCl₃ = 4:1); MS m/z 372 (M⁺), 354 $(M^{+} - H_{2}O), 336 (M^{+} - 2H_{2}O), 329, 318 (M^{+} - 3H_{2}O); {}^{1}H$ NMR (400 MHz, CDCl₃) δ 1.13 (ddd, ²*J* = 13.5 Hz, ³*J* = 4.3, 13.5 Hz, 4H), 1.20 (d, J = 13.2 Hz, 2H), 1.34 (dt, ${}^{2}J = 14.8$ Hz, ${}^{3}J = 2.2$ Hz, 4H, CH₂CH₂O), 1.36–1.44 (m, 2H), 1.69–1.76 (m, 4H), 1.97 (dtt, ${}^{2}J = 13.8$ Hz, ${}^{3}J = 3.7$, 13.8 Hz, 2H), 2.07 (dtd, ${}^{2}J = 14.8$ Hz, ${}^{3}J = 4.6$, 12.9 Hz, 4H, CH₂CH₂O), 2.17 (dt, ${}^{2}J = 13.2$ Hz, ${}^{4}J = 2.4$ Hz, 2H, CH₂CH₂O), 3.55 ($\frac{1}{2}$ AB q dd, ${}^{2}J = 9.3$ Hz, ${}^{3}J = 2.3$, 4.5 Hz, 4H, CH₂CH₂O), 3.71 ($\frac{1}{2}$ AB q dd, ${}^{2}J = 9.3$ Hz, ${}^{3}J = 2.3$, 12.8 Hz, 4H, CH₂CH₂O), 5.35 (s, 4H, tertiary OH); ¹³C NMR (22.5 MHz, CDCl₃) δ 16.67, 39.64, 39.88, 41.55, 67.31, 71.75; IR (KBr, cm⁻¹) 3397s, 2935m, 2866m, 1175w, 1128w (Calc. for C₂₀H₃₆O₆·1/4H₂O: C, 63.72; H, 9.76. Found: C, 63.72; H, 9.92%).

Dimer II 13a and 13b

A suspension of Cs₂CO₃ (0.733 g, 2.250 mmol) in DMF (70 ml)

was stirred at room temperature under N2. A solution containing dimesyl compound 12 (0.676 g, 1.875 mmol) and resorcinol (0.206 g, 1.875 mmol) in DMF (10 ml) was added, and the mixture was stirred vigorously at 50 °C for 2 days. The solvent was removed in vacuo. The residue was taken up in THF-ethanol (10:1), and applied to an alumina column, which was eluted with THF-ethanol (10:1). After evaporation of the solvent, the residual mixture was chromatographed on a silica gel column with THF-CHCl₃ (1:5). The fractions containing products were combined and rechromatographed at medium pressure (EtOAc-CHCl₃ = 1:6) to give dimers 13a and 13b as a white solid, total yield 30 mg (6%) (anti isomer 16 mg, 3%; syn isomer 14 mg, 3%). 13a (syn isomer): TLC (silica gel) R_f 0.60 (EtOAc-CHCl₃ = 4:1); MS m/z 556 (M⁺), 538 (M⁺ - H₂O), 520 (M⁺ - 2H₂O), 502 (M⁺ - 3H₂O), 484 (M⁺ - 4H₂O); ¹H NMR (500 MHz, CDCl₃ + CD₃OD) δ 1.34 (ddd, ²J = 13.6 Hz, ³J = 4.1, 13.6 Hz, 4H), 1.37 (d, J = 14.3 Hz, 2H), 1.47–1.53 (m, 2H), 1.59– 1.65 (m, 4H), 1.76 ($\frac{1}{2}$ AB q t, ^{2}J = 14.6 Hz, ^{3}J = 6.0 Hz, 4H, CH₂CH₂O), 1.88 (m, 4H, CH₂CH₂O), 1.90–1.95 (m, 4H), 3.98 ($\frac{1}{2}$ AB q dd, ^{2}J = 9.6 Hz, ^{3}J = 5.5, 6.8 Hz, 4H, CH₂CH₂O), 4.07 $(\frac{1}{2}AB q dd, {}^{2}J = 9.6 Hz, {}^{3}J = 6.3, 6.3 Hz, 4H, CH_{2}CH_{2}O),$ 6.31 (t, J = 2.2 Hz, 2H, ArH), 6.37 (dd, J = 2.2, 8.3, 4H, ArH), 7.01 (t, J = 8.3 Hz, 2H, ArH); ¹³C NMR (125 MHz, $CDCl_3 + CD_3OD$) δ 16,63, 37.85, 41.88, 42.00, 63.87, 72.13, 100.11, 107.45, 129.78, 159.63; 13b (anti isomer): TLC (silica gel) $R_f 0.60$ (EtOAc–CHCl₃ = 4:1); MS m/z 556 (M⁺), 538 $(M^+-H_2O), \ \ 520 \ \ (M^+-2H_2O), \ \ 502 \ \ (M^+-3H_2O), \ \ 484$ $(M^+ - 4H_2O)$; ¹H NMR (500 MHz, CDCl₃ + CD₃OD) δ 1.34 $(ddd, {}^{2}J = \bar{1}3.5 \text{ Hz}, {}^{3}J = 3.9, 13.5 \text{ Hz}, 4\text{H}), 1.38 (d, J = 14.5 \text{ Hz}, 4\text{H})$ 2H), 1.49–1.55 (m, 2H), 1.70–1.76 (m, 4H), 1.84 (t, J = 7.0 Hz, 8H, CH₂CH₂O), 1.85-1.96 (m, 4H), 4.03-4.11 (m, 8H, CH_2CH_2O), 6.32–6.40 (m, 6H), 7.00–7.04 (m, 2H); ¹³C NMR (125 MHz, CDCl₃ + CD₃OD) δ 16.85, 37.11, 41.93, 44.55, 63.91, 72.51, 101.98, 106.25, 108.26, 129.90, 157.87, 159.71.

X-Ray crystal structure determinations

Diffraction data were collected at 295 K with a Rigaku AFC-7R four-circle diffractometer having a rotating anode and using graphite-monochromated Cu-*Ka* ($\lambda = 1.541$ 78 Å) and Mo-*Ka* ($\lambda = 0.710$ 69 Å) radiations for **13a** · 1.5C₄H₈O₂ and **13b**, respectively. The crystal data and details of the parameters associated with data collection for the two crystals are given in Tables 1 and 2. The unit-cell parameters were derived from least-squares refinement of 25 well centred reflections. The reflection intensities were monitored by three standard reflections for every 150 reflections, and the decay of intensities was within 2% for the two crystals. Reflection data were corrected for Lorentz and polarization effects. An empirical absorption correction, based on ψ scans, was applied.¹⁰

The structures for the compounds were solved by the direct method¹¹ and successive Fourier techniques¹² and refined anisotropically for non-hydrogen atoms by full-matrix least-squares calculations. Refinements were continued until all shifts were smaller than one-third of the standard deviations of the parameters involved. Atomic scattering factors and anomalous dispersion terms were taken from the literature.¹³ All the hydrogen atoms were located from the difference Fourier maps, and their parameters were isotropically refined. The *R*- and *R*_w-values were 0.062 and 0.071 for **13a**·1.5C₄H₈O₂ and 0.036 and 0.052 for **13b**, respectively. The weighting scheme $w^{-1} = \sigma^2(F_o)$ was employed for both crystals. The final difference Fourier maps did not show any significant features in both cases. The calculations were performed on an IRIS Indigo XS-24 computer by using the program system teXsan.¹⁴

CCDC reference number for $13a \cdot 1.5C_4H_8O_2$ and 13b 207/ 329. See http://www.rsc.org/suppdata/p1/1999/1885 for crystallographic files in .cif format.

Acknowledgements

We thank T. Kobatake and H. Fujita for their kind help in the mass and NMR spectroscopy measurements. This work was supported by a Grant-in-Aid for Scientific Research from the Ministry of Education, Science, Sports, and Culture, Japan.

References

- (a) M. Simard, D. Su and J. D. Wuest, J. Am. Chem. Soc., 1991, 113, 4696; (b) J. C. MacDonald and G. M. Whitesides, Chem. Rev., 1994, 94, 2383; (c) K. Endo, T. Ezuhara, M. Koyanagi, H. Masuda and Y. Aoyama, J. Am. Chem. Soc., 1997, 119, 499.
- 2 For reviews, see, (a) J. R. Fredericks and A. D. Hamilton, in *Comprehensive Supramolecular Chemistry*, ed. J. L. Atwood, J. E. Davies, D. D. MacNicol, F. Vögtle and J.-M. Lehn, Elsevier, Oxford, UK, 1996, vol. 9, pp. 565–594; (b) E. E. Simanek, X. Li, I. S. Choi and G. M. Whitesides, in *Comprehensive Supramolecular Chemistry*, ed. J. L. Atwood, J. E. Davies, D. D. MacNicol, F. Vögtle and J.-M. Lehn, Elsevier, Oxford, UK, 1996, vol. 9, pp. 595–621; (c) D. A. Bell and E. V. Anslyn, in *Comprehensive Supramolecular Chemistry*, ed. J. L. Atwood, J. E. Davies, D. D. MacNicol, F. Vögtle and J.-M. Lehn, Elsevier, Oxford, UK, 1996, vol. 2, pp. 439–475.
- 3 (a) D. Koshland, Proc. Natl. Acad. Sci. USA, 1958, 44, 98; (b)
 D. Koshland, J. Yankeelov and J. Thoma, Fed. Proc., 1962, 21, 1031;
 (c) D. Koshland, J. Theor. Biol., 1962, 2, 75.
- 4 Examples of allosteric hosts, see (a) P. D. Beer and A. S. Rothin, J. Chem. Soc., Chem. Commun., 1988, 52; (b) F. Ebmeyer and J. Rebek, Jr., Angew. Chem., 1990, 102, 1191; Angew. Chem., Int. Ed. Engl., 1990, 29, 1148; (c) H.-J. Schneider and D. Ruf, Angew. Chem., 1990, 102, 1192; Angew. Chem., Int. Ed. Engl., 1990, 29, 1159; (d) R. P. Sijbesma and R. J. M. Nolte, J. Am. Chem. Soc., 1991, 113,

6695; (e) M. Inouye, M. Ueno and T. Kitao, J. Org. Chem., 1992, **57**, 1639; (f) T. Hayashi, T. Asai, H. Hokazono and H. Ogoshi, J. Am. Chem. Soc., 1993, **115**, 12210; (g) T. Mizutani, S. Yagi, A. Honmaru and H. Ogoshi, J. Am. Chem. Soc., 1996, **118**, 5318.

- 5 For a review of design of flexible molecules, see R. W. Hoffmann, *Angew. Chem.*, *Int. Ed. Engl.*, 1992, **31**, 1124.
- 6 N. Morelle, J. Gharbi-Benarous, F. Acher, G. Valle, M. Crisma, C. Toniolo, R. Azerad and J.-P. Girault, J. Chem. Soc., Perkin Trans. 2, 1993, 525.
- 7 For preliminary accounts of preparation and X-ray crystal structure of 1a, see T. Hayashi, H. Takagi, H. Masuda and H. Ogoshi, J. Chem. Soc., Chem. Commun., 1993, 364.
- 8 M. P. Mertes, J. Org. Chem., 1961, 26, 5236.
- 9 J. A. Marshall, B. S. DeHoff and D. G. Cleary, J. Org. Chem., 1986, 51, 1735.
- 10 A. C. T. North, D. C. Phillips and F. S. Mathews, *Acta Crystallogr.*, *Sect. A*, 1968, 24, 351.
- 11 A. Altomare, M. C. Burla, M. Cascarano, C. Giacovazzo, A. Guagliardi and G. Polidori, J. Appl. Crystallogr., 1993, 26, 343.
- 12 DIRDIF92: P. T. Beurskens, G. Admiraal, G. Beurskens, W. P. Bosman, S. Garcia-Granda, R. O. Goulg, J. M. M. Smits and C. Smykalla (1992). The DIRDIF program system, Technical Report of the Crystallography Laboratory, University of Nijmegen, The Netherlands.
- 13 D. Cromer and J. T. Wabber, in *International Tables for X-ray Crystallography*, The Kynoch Press, Birmingham, England, 1974, vol. IV, Table 2.2 A.
- 14 *teXsan*: Crystal Structure Analysis Package, Molecular Structure Corporation, The Woodlands, TX, 1985 and 1992.

Paper 8/09834D