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Spectroscopic properties and gelling ability of a set of rod-like 2,3-disubstituted anthracenes[†]

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The UV absorption spectra of 2,3-didecyloxyanthracene (DDOA) recorded in methanol, ethanol, 1-propanol, acetonitrile and methylcyclohexane reveal interesting features: they show a striking contrast between the isotropic solution and the gel state where, at ambient temperature, a fine structure appears with an apparent bathochromic shift ($\Delta \nu \approx -700 \text{ cm}^{-1}$), as observed in the solid state. Such an effect was mimicked by 2,3-dioxydi- (and -tri-) methyleneanthracenes in which the conformational mobility of the two juxtanuclear oxygen atoms is reduced in a manner similar to that assumed in the gel state. The fluorescence emission of DDOA (10^{-5} M) at very low temperature exhibits a loss of fine structure and a bathochromic shift, for the gel state, in agreement with the presence of aggregates; the excitation spectra were found to be superimposable upon the absorption spectra of the isotropic and gel phases, respectively. Solvent screening for DDOA gelling ability has shown that the most efficient solvents are CH₃OH and CH₃CN. From the phase transition diagram (temperatures of gel setting, T_{gel} , and gel melting, T_m , *versus* concentration), thermodynamic parameters were derived: $\Delta H_{gel}^0/kJ \mod^{-1} = -70$ (CH₃OH), -66 (CH₃CN); $\Delta S_{gel}^0/(J \text{ K}^{-1} \mod^{-1} = -147$ (CH₃OH), -140 (CH₃CN) and $\Delta G_{gel}^0/kJ \mod^{-1}$ (at 300 K) ≈ -26 (CH₃OH), -24 (CH₃CN). These parameters attest to the good stability of these gel systems. Finally, the influence of the chain length ($n = C_7H_{15}$ to $C_{12}H_{25}$) on the efficiency of gel formation (or melting) was investigated in methanol, ethanol, acetonitrile and heptane. It emerges that, to form gels, *n*-decyl and *n*-undecyl were found to be the most suitable chains and methanol and ethanol the most efficient solvents. It should be noted that the ability to form gels in methanol at a concentration of 0.6 mM at ambient temperature qualifies DDOA as a supergelator.

Introduction

Gels are heterogeneous soft materials constituted of three dimensional (3D) networks imprisoning a liquid.^{1–4} They are used in everyday life and continuous research efforts are being directed to extend their applications.^{3–8}

For the last two decades, there has been a growing interest in gelation induced by the addition of low molar mass ($M \le 1000$) substrates (the so-called LMOGs^{9,27}) to organic liquids or, more rarely, water.^{9–29} These compounds show a wide structural diversity. For convenience they can be classified into two categories: (1) gelators involving *hydrogen bonds*, ion-pairs or metal coordination and (2) systems involving *no H bonds* and based on van der Waals and weak electrostatic interactions. In the latter category, a comprehensive study has been devoted to the family of gelators containing steroidal and condensed aromatic rings, denoted ALS (aromatic linked steroids;¹⁹ an example is shown in Scheme 1), whose discovery in 1987 gave a strong impetus to the development of the field.^{19–49}

A few years later²⁸ some of us reported the gelling properties of 2,3-di-*n*-decyloxyanthracene (DDOA) for several organic solvents, especially methanol. Similarly to CAB (Scheme 1), it so happens that DDOA contains an anthracene nucleus that confers fluorescing properties onto the gels, but for DDOA, in contrast to the ALS family, the presence of alkoxy substituents



Scheme 1 Cholesteryl-4-(2-anthryloxy)butanoate (CAB), an example of the ALS family (see text); the presence of an anthracene containing tether was found to induce gelling properties and luminescence for a variety of organic liquids.¹⁹

in the 2 and 3 positions was found to be compelling for gelation to occur (Scheme 2). Furthermore, a rheological study²⁹ has shown that DDOA gels formed in 1-octanol could be described as soft solids exhibiting high yield stress values.

The elasticity of the DDOA gel ($G' \approx 10^4$ Pa at 1 wt %) was analysed as resulting from a 3D network of fibers or bundles of fibers with extended interaction zones, as borne out by small angle neutron scattering (SANS) investigations.²⁹ Various aspects of DDOA gel properties have been recently examined: aerogel formation in supercrital CO₂,³⁰ electrochemical properties in propylene carbonate,³¹ porosity templating of sol-gel derived silica^{32a} and alumina,^{32b} gelation of liquid crystals.³³

However, since the preliminary results,²⁸ no detailed spectroscopic study has been published. Such a study is reported here, together with a systematic exploration of the role played by the chain length ($\mathbf{R} = C_n H_{2n+1}$) and an investigation of the

[†] Electronic supplementary information (ESI) available: UV absorption spectra of DDOA in 1-propanol and acetonitrile as a function of temperature; phase transition diagrams and related plots for DDOA in various solvents. See http://www.rsc.org/suppdata/nj/b3/b306961c/



Scheme 2 2,3-Di-n-alkoxyanthracenes (DAOA) 1 and related heterocyclic compounds 2. The compounds will be denoted $1C_7$, $1C_8$, *etc.* and 2_1 , 2_2 , 2_3 , respectively. The di-*n*-decyloxy derivative (DDOA) was first shown to form a gel with alcohols and alkanes.²⁸

thermodynamic parameters of the phase transitions (sol-to-gel and gel-to-sol).

Results and discussion

1. UV absorption and fluorescence spectroscopic properties of DDOA

The gelling property of DDOA was discovered through an unsuccessful crystallization in CH₃OH.²⁸ Subsequent rapid screening of gel-forming solvents showed that alcohol and alkanes are the best-suited. For this reason, the electronic absorption and emission spectra were studied essentially in methanol and methylcyclohexane (MCH); the latter was chosen for its ability to stay liquid at very low temperatures (m.p. −126 °C).

The absorption spectra of DDOA in CH₃OH are represented in Fig. 1. At 26 °C, the fluid isotropic solution exhibits the typical spectrum of 2,3-dialkoxy substituted anthracenes⁵⁰ but, at -15 °C, the medium is a translucent gel at the concentration used $(10^{-3}M)$ and the spectrum appears to be strongly different. The first absorption band (300-400 nm), essentially constituted of the 1La electronic transition, has a nice fine structure with well-defined vibronic maxima at 338, 347, 365 and 385 nm. The latter band, the most intense, shows an apparent bathochromic shift ($\Delta \nu \simeq -700 \text{ cm}^{-1}$); it probably corresponds to the 0-0 transition. This feature, combined with the enhanced fine structure as is the case for low temperature spectra, points to a great loss of freedom of orientation for the molecules in the gel. This statement is reinforced by the examination of the second band (200-300 nm), which, peaking as expected at 260 nm for the solution, shows a strong decrease of intensity in the gel; this change points to some parallelism between the long axes of the molecules due to π - π stacking.⁵

Typical spectra of the right angle fluorescence of a diluted solution of DDOA in methanol as a function of temperature are represented in Fig. 2. At 20 °C the spectrum resembles that



Fig. 1 Isotropic (···) and gel (—) phase absorption spectra of DDOA ($\cong 10^{-3}$ M) in CH₃OH, at 26 °C and -15 °C, respectively.



Fig. 2 Fluorescence and excitation spectra (arbitrary scale) of DDOA in degassed CH₃OH (10⁻⁴ M). Fluorescence λ_{exc} : 370 nm. Excitation: λ_{em} : 435 nm. 25 °C isotropic (···) and -70 °C gel (—) phases, respectively. Insert: Fluorescence spectrum of 2,3-dimeth-oxyanthracene in degassed ethanol ($\approx 3 \times 10^{-5}$ M) at -65 °C (λ_{exc} : 365 nm), for comparison.

of 2,3-dimethoxyanthracene taken as reference and its excitation spectrum is in line with that of Fig. 1. When the temperature is lowered, the gel forms and the fluorescence spectra are red-shifted, as usually seen for aggregates^{52,53} and consistently the excitation spectrum becomes that of the gel as in Fig. 1.

UV spectra of DDOA recorded in ethanol and 1-propanol vs. temperature [-15°C (gel phase) to 20-50°C (isotropic solution)] were found to show the same features as in methanol. An example of the spectra (300-400 nm) in 1-propanol (-15° to 26°C) exhibiting clear isosbestic points is given as supplementary material (Fig. S1 in the ESI).

In methylcyclohexane (MCH), the fluorescence was studied from +20 °C to -104 °C. The fluorescence and excitation spectra of DDOA in degassed MCH at very low concentration (10^{-5} M) are represented in Fig. 3. The excitation spectrum of the gel shows a remarkably fine structure. The maximum wavelengths do not differ from those in CH₃OH. The emission spectrum of the gel is composed of "free" molecules dissolved in the imprisoned solvent and molecular aggregates immobilized within the fibrils, filaments and fibers, which experience different environments and therefore exhibit a variety of spectra as shown in the time-resolved spectra of Fig. 3 (insert). This is borne out by the results of a transient kinetic analysis, reported in Table 1. The wavelength dependent fluorescence decay can be fitted with a sum of two exponentials at all the wavelengths investigated. These decay times are all longer than that of the isolated molecule (\cong 4 ns) and are in agreement with the occurrence of different excited species displaying slightly different emission spectra, as recently discussed by De Schryver and coworkers^{52,53} for anthracene chromophores



Fig. 3 Excitation (left) and fluorescence (right) spectra of DDOA (10^{-5} M) in degassed methylcyclohexane: solution (\cdots) and gel (-). $\lambda_{\rm exc}$ 370 nm; $\lambda_{\rm obs}$ 410 nm (+20 °C) and 480 nm (-100 °C). Insert: time-resolved fluorescence spectra in MCH: $+20 \degree C (\cdots) 0$ ns delay; -104 °C (---) 0 ns delay, -104 °C (—) 40 ns delay. The population of emitting aggregates changes with time.

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Table 1 Fluorescence decay of DDOA ($\cong 8 \times 10^{-5}$ M) in degassed methanol at $-45 \,^{\circ}$ C. τ_1 , τ_2 in ns; A_1 and A_2 are the preexponential factors of the fluorescence intensity decay: $I(t) = A_1 \exp(-t/\tau_1) + A_2 \exp(-t/\tau_2)$

$\lambda_{\rm obs}/nm$	420	440	460	480	
$\tau_1(A_1)$	5 (0.4)	8 (0.4)	9.5 (0.4)	11.5 (0.4)	
$\begin{array}{c} \tau_2 \left(A_2 \right) \\ \chi^2 \end{array}$	14 (0.14) 1.44	20 (0.07) 1.09	25 (0.05) 1.05	37 (0.04) 1.32	

in Langmuir–Blodgett films. The non-structured fluorescence spectra showing maximum intensity at 460 nm suggest the formation of excited aggregates from overlapping monomers as displayed by some non-sandwich overlapping excimers.⁵⁴

The mutual arrangements of the anthracene nuclei in the gel are reminiscent of those in solid DDOA, as reflected in the compared UV spectra (Fig. 4). One might speculate about the correspondence between the crystal structure and the structure of aggregates in the gel as shown for instance by Terech,^{9b} Weiss *et al.*^{27a,b,41b} and Dastidar *et al.*^{27c} but no distinct X-ray diffractograms could be obtained on pure DDOA gels and we never could grow any single crystals of DDOA. In the series the only single crystals suitable for X-ray determination were obtained for di-*n*-hexyloxyanthracene (DHOA),⁵⁵ but no X-ray studies were performed on the gels of DHOA. The question of the structure at the molecular level has been addressed in a recent paper showing some unique features of the gel phase.⁵⁶

The UV absorption spectrum of the gel phase does not depend on the gelling solvent and it was used to follow-up the gel formation of DDOA in a matrix of silica gel.^{32*a*} Such a gel could be formed from Si(OCH₃)₄ (TMOS), methanol, water and a base (to obtain pH 12.7) by adding a methanolic solution of DDOA (10^{-3} M) and heating at 50 °C for several minutes, then allowing to cool down to ambient temperature. Under these conditions, the inorganic gel was formed first, followed by gradual formation of the organic gel. The inclusion of the DDOA gel could be traced by recording the UV spectra as a function of temperature (Fig. 5).

The UV absorption spectra of a solution of DDOA in CH₃CN (6×10^{-4} M) was recorded as a function of temperature (2.4 °C to 50 °C) and showed the same characteristic changes (including isosbestic points) as observed in methanol and 1-propanol (Fig. S2 in the ESI).

The difference in the vibronic structure between the isotropic solutions (in methanol, MCH or acetonitrile) and the gel (and the solid state) might be in line with the ability of oxygen lone pairs to align parallel with the π electrons of the anthracene nucleus. A qualitative piece of evidence to support this hypothesis comes from a comparison of the UV absorption spectra of the heterocyclic derivatives $2_{1,2,3}$ with those of the acyclic compounds 1_{CH3} and 1_{C10} (see Fig. 6). The acyclic compounds exhibit λ_{max} 374–376 nm, irrespective of the solvent and the chain length. It thus reflects an average orientation of the



Fig. 4 UV absorption spectra of DDOA in a KBr pellet (\cdots) and of its gel (-) in methanol at ambient temperature. The diffused light is more important in KBr.



Fig. 5 UV absorption spectra of DDOA in a silica gel matrix in a 1 mm quartz cell as a function of temperature: (—) $25 \,^{\circ}$ C (organogel), (---) $40 \,^{\circ}$ C (mixture of organogel and isotropic phase), (···) $45 \,^{\circ}$ C (the same mixture with a higher proportion of isotropic phase).

freely rotating oxygen atoms; this is strongly inhibited in the methylenedioxy compound $\mathbf{2}_1$, which results in an hypsochromic shift (370 nm). In contrast, the propylenedioxy substituent in $\mathbf{2}_3$ induces a bathochromic shift (382 nm); another red shift (390 nm) is exhibited by the six-membered ring compound $\mathbf{2}_2$ in which the oxygen atoms appear to be maintained in the best geometry for conjugation. From these data, it emerges that, in the gel, the oxygen atoms should have a geometry intermediate between those of 2,3-propylenedioxyanthracene ($\mathbf{2}_3$) and 2,3-ethylenedioxyanthracene ($\mathbf{2}_2$).

2. Photochemical reactivity of DDOA

Anthracenes are known to have photochromic properties⁵⁷ but our attempts to confer this property to the DDOA gels met with failure. No clear change was noted after 2 h irradiation of a methanolic gel in a Pyrex tube with a medium pressure mercury lamp. The photoreactivity of DDOA was thus investigated in isotropic solution. Irradiation of a degassed benzene solution (10^{-3} M) with a medium pressure mercury lamp, using a Pyrex filter, was followed by UV spectroscopy. The



Fig. 6 UV absorption spectra, at ambient temperature, in MCH $(c \approx 10^{-4}6 \text{ M})$ of 2,3-methylenedioxyanthracene (**2**₁), 2,3-ethylenedioxyanthracene (**2**₂) and 2,3-trimethylenedioxyanthracene (**2**₃). The onset wavelength increases from 370 to 382 to 390 nm for **2**₁, **2**₃ and **2**₂, respectively.. Compare these UV spectra with those of Figs. 1–4.

3. Solvent screening for gel-forming ability

DDOA was found to form gels in methanol at ambient tem*perature* at a concentration as low as 6×10^{-4} M (0.04 wt %). Most gels are known to be formed at a concentration of ca. 1 wt %; the efficiency of DDOA in methanol is therefore outstanding (comparable values for supergelators were reported by others^{35e,46a}) and systematic tests of gel-forming solvent ability were undertaken using the inverted tube method described in the Experimental. Some salient results are collected in Table 2; the molar concentration was chosen with a view to obtaining thermodynamic data (see Section 4) and as a common basis for comparison between the different 2,3dialkoxyanthracenes. It is remarkable that, in many cases, gels can be formed at concentrations as low as 10^{-3} M. One observes that alcohols and alkanes are less easily gelified by DDOA as the molecular weight increases. Among alcohols, gel-forming ability decreases with branching (e.g. butane-2-ol vs. 1-butanol); however, 2,3-butanediol behaves like ethanol. Acetonitrile shows good performance but the solubility is limited to 10^{-2} M. For potential applications to electrochemistry, propylene carbonate was found to deserve special study³¹ and the methacrylates were tested for possible polymerization using DDOA gel as a template.^{11,12} It is also worth noting that supercritical CO₂ (not reported in Table 2) was explored as a solvent to produce aerogels,30 as mentioned in the Introduction.

DDOA is not soluble in water but addition of some water to alcohols was found to be possible, as shown in silica sol

Table 2 Organic solvents tested for gelation of DDOA at low concentration^a by the inverted tube method:. (+) denotes gel formation and (-) no gel formation observed

Solvent	Concentration/M	Ambient temperature	−18°C
Methanol	6×10^{-4}	+	
Ethanol	1×10^{-3}	$+ (8 - 10 \circ C)$	
1-Propanol	1.7×10^{-3}	$+ (0^{\circ}C)$	
2-Propanol	1.2×10^{-3}	-	+
1-Butanol	2×10^{-3}	$+ (0 ^{\circ} C)$	
2-Butanol	1.3×10^{-2}	-	+
2,3-Butanediol	1.1×10^{-3}	+	
1-Octanol	10^{-2}	+	
Propylene carbonate	10^{-2}	+	
<i>n</i> -Pentane	2.8×10^{-3}	_	+
<i>n</i> -Hexane	5×10^{-3}	+	
n-Heptane	10^{-2}	+	
Isooctane	10^{-2}	+	
Acetonitrile ^b	$7.5 imes 10^{-4}$	+	
Acetone	3×10^{-3}	_	+
DMF	1.1×10^{-3}	_	+
DMSO	1.4×10^{-3}	+	
Methyl methacrylate	10^{-2}	+	
Lauryl methacrylate	10^{-2}	+	

^{*a*} Concentration in mol (M) was adopted as it is used in thermodynamics (see phase transition diagrams below). [DDOA] in CH₃OH (molar mass: 490): 5×10^{-4} M = 0.03 wt %; 10^{-3} M = 0.06 wt %; 10^{-2} M = 0.62 wt % ^{*b*} Solubility limited to 10^{-2} M gels using DDOA gel as a template³² and as experienced with other gelators^{58,59} (see Section 1). DDOA was found to be very soluble in CH_2Cl_2 and $CHCl_3$ at ambient temperature; addition of methanol or ethanol to such solutions allows formation of gels; however, this method was not explored further.

4. Phase transition diagrams and thermodynamic parameters

In order to gain more insight into the efficiency of gel formation and the melting of DDOA, phase transition temperatures were measured as a function of concentration in methanol, ethanol, heptane, acetonitrile and propylene carbonate. The temperatures [T_m (gel melting) and T_{gel} (gel formation)] were measured with an accuracy of $\pm 0.5 \,^{\circ}$ C, using a thermostatted bath (heating and cooling rates of $1.5 \,^{\circ}$ C min⁻¹) and an automated apparatus allowing the registration of transmittance of the diffused light. (see Experimental). As the dimensions of the fibers are of the same order of magnitude as the visible light wavelengths, the diffused light intensity increases with their concentration. These measurements seem to be less arbitrary than those based on mechanical tests of gelification.

Phase transition diagrams. A typical phase transition diagram, that of DDOA in methanol, is presented in Fig. 7, accompanied by the thermodynamic correlation expressed in eqns (1) and (2) below (see Fig. 8). In this solvent, the maximum temperature is limited by the boiling point of methanol (64.5 °C); it is thus useless to increase the DDOA concentration to increase $T_{\rm m}$. In principle, the maximum gelification temperature should not exceed the melting point of the gelator (m.p. DDOA 87 °C). One notes a clear hysteresis between the melting and gelification processes; this phenomenon depends on the temperature variation rate and points to the difficulty in characterizing a reversible process dealing with partial heterogeneity. The temperatures of gelification (T_{gel}) were observed to vary very little between different experiments whereas the melting temperatures (T_m) depend on the thermal history of the sample.

Gel formation can be assumed to resemble a crystallization; thus, it will be described by the following thermodynamic equations (see Appendix):

Gel melting :
$$\ln \phi = -\Delta H^0 / R \times 1 / T_m + \Delta S^0 / R$$
 (1)

Gel formation : $\ln \phi = +\Delta H^0 / R \times 1 / T_{gel} - \Delta S^0 / R$ (2)

 ΔH^0 and ΔS^0 are positive for the melting and negative for the gelification processes, respectively; ϕ denotes the molar fraction of the solute in the appropriate solvent (see Appendix).

The diagram for gelification of DDOA in *n*-heptane (Fig. 9) clearly shows that the concentrations used are much higher than in methanol (as qualitatively suggested in Section 3). It is accompanied with the linear plot according to eqn. (2) (see Fig. 10). Other diagrams (Fig. S3a–d) and linear graphs



Fig. 7 Phase transition diagram of DDOA in methanol. $T_{\rm m}$ and $T_{\rm gel}$ denote the gel-to-sol and the sol-to-gel transition temperatures, respectively. A hysteresis with $\Delta T \approx 10$ °C was observed. T accuracy ± 0.5 °C.



Fig. 8 $\ln \phi vs. 1/T$ plots for DDOA in methanol, showing a good linearity ($r^2 = 0.987$). 1 represents the gel-to-sol transition and 2 the sol-to-gel process. See eqns (1) and (2).

(Fig. S4a–c) obtained in *n*-heptane, acetonitrile and propylene carbonate are given as supplementary material.

From these data, the thermodynamic parameters ΔH^0 and ΔS^0 were calculated and are listed in Table 3. Are also included the values obtained from a study of 2,3-dialkoxyanthracenes incorporating chains of different lengths (*vide infra*). From the upper part of Table 3, it emerges that *n*-heptane seems to have a lower ability to form gels with DDOA (the free enthalpy at 27 °C clearly has a lower value) than the other solvents, presumably because of a better solvation power. On the other hand, methanol appears to be the most appropriate solvent.

Influence of the chain length on the gelification efficient. After discovering the gelling ability of DDOA and having shown that it is connected to the presence of two alkoxy chains in positions 2 and 3 of anthracene, an obvious question was that of the optimum chain length. The whole series of derivatives $1(C_6-C_{12})$ was prepared and preliminary tests show that $1C_6$ could not give gels at room temperature but provided single crystals suitable for X-ray structure determination.55 The other compounds formed gels in alcohols, though it is noted that $1C_7$ was clearly borderline and $1C_{12}$ looked less efficient in ethanol. Phase transition diagrams were therefore established for 1C₈-1C₁₁ in ethanol (see Figs. 11 and 12) and the thermodynamic parameters were determined (Table 3). Despite some discrepancies in the enthalpy and entropy values, the free enthalpies show a trend of increasing stability (at 27 °C) from $1C_8$ to $1C_{11}$ but the differences are rather small.

Another appraisal of the chain length influence can be obtained from the melting temperatures (although these are less reliable than the gelification temperatures, they can be easily determined by the inverted tube method) for a fixed concentration (10^{-2} M) in different solvents, as shown in Fig. 13. In methanol, ethanol and acetonitrile, $1C_{10}$ and $1C_{11}$ behave as the most efficient compounds if one uses the melting temperature as the main criterion.

The data of Table 3 are to be compared with the published thermodynamic values of other LMOGs. Some of these have



Fig. 9 Phase transition diagram for the gelification of DDOA in *n*-heptane. Temperature accuracy: ± 0.5 °C.



Fig. 10 $\ln \phi$ vs. 1/T plot for DDOA gelification in *n*-heptane [eqn. (2)].

been selected for a variety of gelators and are listed below; most of them (except for the cholesterol-based systems) are of the H-bonded type. Shinkai and coworkers⁶⁰ determined the melting enthalpy for a number of derivatives (differing in the alkoxy group) of para-alkoxy-substituted azobenzeneappended cholesterol-based gelators (α C-3): 41 < $\Delta H_m^0/kJ$ $mol^{-1} < 160$ in ethanol and $41 < \Delta H_m^0/kJ mol^{-1} < 110$ in 1-butanol. For N-benzoyloxycarbonyl-L-alanine-4-hexadecanoyl-2-nitrophenyl esters (BLAHN), Hanabusa and cowor-kers⁶¹ reported $\Delta H_{gel}^0 = -23$ and -25 kJ mol⁻¹, $\Delta S_{gel}^0 = -7$ and -10 J K⁻¹ mol⁻¹and ΔG_{gel}^0 (at 300 K) = -21 and -22 kJ mol⁻¹, in methanol and cyclohexane, respectively. In addition, a more substituted derivative of the same family exhibited the following parameters in cyclohexane: $\Delta H_{gel}^0 = -42 \text{ kJ}$ mol⁻¹, $\Delta S_{gel}^0 = -56 \text{ J K}^{-1} \text{ mol}^{-1}$ and ΔG_{gel}^0 (at 300 K) = -26 kJ mol⁻¹. Garner, Terech and coworkers¹⁵ have derived the parameters for the trans isomer of 4-tert-butyl-1-phenylcyclohexanol (BACOl) in several solvents: $\Delta H_m^0 = 35.6$ and 28.5 kJ mol⁻¹ and $\Delta S_m^0 = 75.5$ and 66 J K⁻¹ mol⁻¹; hence the free enthalpies at 300 K are $\Delta G_{\rm m}^0 = 13.5$ and 9.2 kJ mol⁻¹, in n-heptane and dichloromethane, respectively. Typical values of gel formation enthalpies for hexose derivatives were derived by Shinkai and coworkers:⁶² for α -1-*O*-methyl-4,6-*O*-benzyli-dene-D-glucose $\Delta H_{gel}^0/kJ$ mol⁻¹ = -43 (toluene), -40 (tetrachloromethane) whereas for the D-galactose isomer $\Delta H_{gel}^0/kJ$ $mol^{-1} = -40$ (toluene), -45 (tetrachloromethane). Interestingly, for some dithienylcyclopentenediamides in toluene $\Delta H_{\rm m}^0$ values of up to 98 kJ mol⁻¹ were found, presumably due to a strong H-bond network.⁶³

The enthalpy values reflect the network strength and the importance of the entropy in the phase transition is clearly

Table 3 Thermodynamic parameters $(\Delta H^0, \Delta S^0)$ obtained from the phase transition diagrams (T_{gel} or T_m vs. concentration) and free enthalpy (ΔG^0) at 300 K in CH₃OH, *n*-heptane, CH₃CN and propylene carbonate for DDOA. In EtOH, the parameters were determined for compounds with different chain lengths. Accuracy on ΔH^0 and ΔS^0 is $\pm 10\%$

Solvent and compound	$\Delta H^0/\mathrm{kJ}~\mathrm{mol}^{-1}$		$\Delta S^0/\mathrm{J}\mathrm{K}^{-1}\mathrm{mol}^{-1}$		$\Delta G^0 (27 ^\circ\mathrm{C})/\mathrm{kJ} \mathrm{mol}^{-1}$	
	Gelif.	Melting	Gelif.	Melting	Gelif.	Melting
Methanol						
1C ₁₀	-70	+86	-147	+188	-25.9	+29.5
b-Heptane						
1C ₁₀	-60	+92	-155	+246	-13.5	+18.2
Acetonitrile						
1C ₁₀	-66	+72	-140	+152	-24.0	+26.4
Propylene carbonate						
1C ₁₀	-58	_	-115	_	-24.5	_
Ethanol						
1C ₈	-52	+74	-103	+165	-21.1	+24.5
1C ₉	-53	+85	-104	+190	-21.8	+28.0
1C ₁₀	-64	+93	-139	+218	-22.3	+27.6
1C ₁₁	-48	+81	-82	+171	-23.4	+29.7



Fig. 11 Phase transition diagram for the gelification of $1(\oplus C_8, \oplus C_9, \oplus C_{10}, \blacktriangle C_{11})$ in EtOH. Temperature accuracy: $\pm 0.5 \,^{\circ}\text{C}$.

apparent. The data listed in Table 3 compare well with those reported above and attest to a good stability of DDOA gels in methanol, ethanol and acetonitrile.

Differential scanning calorimetry. This technique is generally used to determine phase transition temperatures and enthalpies. Measurements effectuated on pure DDOA powder for the melting and resolidification led to a m.p. of 87.7 °C and $\Delta H_{\rm m}^0 = 42 \text{ kJ mol}^{-1}$ while the solidification point is 87.1 °C with $\Delta H_{\rm solid}^0 = 37 \text{ kJ mol}^{-1}$. Because methanol has a relatively low melting point, propylene carbonate (boiling point at 1 bar of 240 °C) was chosen to measure gel transition temperatures. The thermograms are shown in Fig. 14. The transition enhalping were determined to be $\Delta H_{\rm m}^0 = 65 \text{ kJ mol}^{-1}$ and $\Delta H_{\rm gel}^0 = -37 \text{ kJ mol}^{-1}$. Although one finds a hysteresis gap of about 10 °C, as in the preceding measurements based on diffused light, the gelification temperature (≈ 55 °C) is different from that found with the automated apparatus ($\approx 60^{\circ}$ C, Fig. S₃d in ESI). The discrepancy observed for the ΔH_{d} values in propylene carbonate $(-58 \text{ kJ mol}^{-1} \text{ reported in Table})$ 3) and the above value $(-37 \text{ kJ mol}^{-1} \text{ obtained by DSC})$ is not clearly understood. Moreover, we cannot offer a satisfactory explanation for the fact that ΔH_m^0 was found to be 42 kJ mol⁻ (DDOA powder) and 65 kJ mol⁻¹ (gel in propylene carbonate). The DSC method should be applied to all samples under various heating and cooling rates. These thermodynamic data should be considered as preliminary.

Summary and conclusions

In exploring further the spectroscopic properties of DDOA ($1C_{10}$), this study has revealed that the UV spectra of the gel state exhibits a clear fine structure and an apparent bathochromic shift (\approx -700 cm⁻¹) irrespective of the solvent used (methanol, ethanol, 1-propanol, acetonitrile, methylcyclohexane or *n*-heptane). It is suggested that these features are induced by a reduced mobility of the conformation of the juxtanuclear oxygen atoms, as in 2,3-dioxydi- (and -tri-) methyleneanthracenes. The remarkable gel-forming properties of DDOA (in methanol, at 20 °C, at 6 × 10⁻⁴ M = 0.04 wt %)



Fig. 12 Phase transition diagram for the melting of $1(\oplus C_8, \oplus C_9, \oplus C_{10})$ and $\blacktriangle C_{11}$ in EtOH. Temperature accuracy: $\pm 0.5 \,^{\circ}C$.



Fig. 13 Gel melting temperatures (T_m) as a function of the chain length in $1(C_7-C_{12})$ at $c = 10^{-2}$ M in \bullet methanol, \bullet ethanol, \blacksquare acetonitrile and $\blacktriangle n$ -heptane.

were also observed in other members of the family with $R = C_7 H_{15}$ to $C_{12} H_{25}$, in ethanol, acetonitrile and heptane; IC_{10} and IC_{11} were found to be the most efficient gelators and *n*-heptane the least appropriate solvent. A test of solvent efficiency was obtained by plotting the phase transition temperature diagram *versus* concentration; from these diagrams, the thermodynamic parameters ΔH_m^0 , ΔS_{gel}^0 and ΔS_m^0 (gel denotes gel formation and m is for melting) that have been determined are in keeping with a good stability of these systems in alcohols and acetonitrile and a fair stability in *n*-heptane. The present results confirm and extend previous work on DDOA. Other recent studies dealing with infrared spectra and polarized fluorescence spectra of aerogels help in getting a deeper insight into the gel structure at the molecular level.⁵⁶

Experimental

General methods and materials

Melting points were determined in capillary tubes on a Buchi 510 apparatus and are uncorrected. FTIR spectra were recorded on a Perkin Elmer Paragon 1000 spectrophotometer. ¹H NMR spectra at 250 MHz and ¹³C NMR spectra at 62.9 MHz were recorded on a Bruker 250 AC instrument for solutions in CDCl₃ unless stated otherwise. Chemical shifts are given in ppm and J values are given in Hz. Chromatographic separations were performed on SDS silica chromagel (70–210 μ m). Mass spectra were obtained on an AutoSpeq EQ spectrometer. Elemental analyses were performed by the Microanalytical Service, Institut du Pin, University Bordeaux I. Electronic absorption spectra and transmission spectra were measured on



Fig. 14 Differential scanning calorimetry of a 10^{-2} M solution of DDOA in propylene carbonate: heating (\rightarrow) and cooling (\leftarrow) rates of 5 °C min⁻¹.

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an Hitachi U-3300 spectrophotometer equipped with a thermocontrolled cell monitored by a Lauda thermostat. The measurements were realized using cells having 10 mm optical pathlength and a platinum temperature sensor. For differential scanning calorimetry measurements, a Perkin Elmer DSC 7 equipped with a stainless steel capsule having a rubber O-ring was used.

The temperature-dependent fluorescence emission and excitation spectra were obtained on a Hitachi F-4500 fluorescence spectrophotometer corrected for excitation and emission using quartz cells with a 1 cm optical pathlength, which were placed in a copper holder inside a quartz Dewar flushed with cold nitrogen gas. The temperature was adjusted by the nitrogen flow. The fluorescence decays were measured by single photon counting (Applied Photophysics) as described elsewhere.⁶⁴ Spectroscopic grade solvents were used for spectrophotometric measurements. No fluorescent contaminants were detected upon excitation in the wavelength region of experimental interest. The samples were degassed by freeze-pump-thaw cycles on a high-vacuum line and sealed under vacuum.

Solvents used for synthesis and gelation tests (alcohols, alkanes, acetonitrile, acetone, DMF, DMSO, propylene carbonate, methyl and laurylmethacrylate, *etc.*) were purchased from SDS (Peypin, Bouches du Rhône, France) and are of synthesis grade. Solvents of spectroscopic grade (methanol, ethanol, 1-propanol, acetonitrile, heptane, methylcyclohexane, *etc.*) from Aldrich were used for UV and fluorescence studies.

Gelation tests

In a typical gelation experiment, 1 cm^3 of solvent of special grade was added to a weighed amount of gelator in a septum-capped test tube (*ca.* 4 cm length and 0.7 cm diameter) in order to obtain the desired concentration. The mixture was warmed to the boiling point until the solid was dissolved and then was allowed to cool down to room temperature. In those cases in which a gel was formed, which generally occurred within several minutes and was confirmed by observing that the sample did not flow *when the test tube was inverted*, the tube was weighed to quantify the amount of gelatinized solvent. When the solution was turbid but the solvent was not entirely gelatinized, the mixture was warmed again and then cooled in a freezer ($-18 \,^\circ$ C) for 24 h.

Phase transition temperature measurements

The measure of temperatures was based on the variation of diffused visible light intensity as a function of the temperature, using an automated apparatus, as described elsewhere.³¹ The monitoring light was that of a laser diode beam, at 670 nm, a wavelength at which the samples do not absorb, and the transmitted light was measured by a photodiode. The heating/cooling cycles were performed at a rate of $1 \,^{\circ}\text{C} \, \text{min}^{-1}$ and repeat measurements (at least 3 per compound) indicate that the phase transition temperatures [T_{gel} (sol \rightarrow gel) and T_m (gel \rightarrow sol)] could be determined with an accuracy of $\pm 0.5 \,^{\circ}\text{C}$. A hysteresis was systematically observed (see Fig. 15 for a typical diagram), related to the existence of an enthalpy for the sol-gel transition and its magnitude depends on the heating/ cooling rates.

Preparations

Compounds $1(C_1 \text{ and } C_7-C_{12})$ and 2_{1-3} , studied in this article, have been synthesized from a common substrate: 2, 3-dihy-droxyanthraquinone (4), as outlined in Scheme 3.

The preparations of **4** (m.p. 393–394 °C, sublim.), **1**C₁ (m.p. 133 °C), **1**C₁₀ (m.p. 84 °C) and **1**C₁₂ (m.p. 74 °C) were described in a preceding publication.⁵⁰ Compounds **2**₁ (m.p. 250–252 °C) and **2**₂ (m.p. 204–205 °C) are known.⁶⁵ In the



Fig. 15 Typical light transmission diagram as a function of temperature (variation $1 \,^{\circ}$ C min⁻¹) as recorded by the self-monitoring set-up.³¹

following, the preparations of $1(C_7, C_8, C_9 \text{ and } C_{11})$ and 2_3 are detailed.

Compounds $3C_n$: general procedure for the alkylation of 2.3-dihydroxyanthraquinone. In a three-neck round-bottom flask equipped with a reflux condenser and a dropping funnel, the required amount of 4 and potassium carbonate (2.5 molar equiv. per hydroxy group) were added to freshly distilled DMF $(10 \text{ cm}^3 \text{ per mmol of 4})$; the medium was warmed until complete dissolution then the appropriate alkylbromide (2.25 molar equiv. per hydroxy group) was added dropwise over 20 min. The reaction mixture was refluxed for 12 h before being allowed to cool to room temperature. The solvent was evaporated under reduced pressure and the residue was treated with water and thoroughly extracted with dichloromethane. The organic layers were combined, dried (MgSO₄) and filtered; the solvent was evaporated and the raw solid was chromatographed on a column of silica gel after dissolution in one of the following eluting mixtures of CH2Cl2-petroleum ether (v/v): (A) 20:80; (B) 40:60. Average yields were 80-85%.

2,3-Diheptyloxy-9,10-anthraquinone ($3C_7$). Starting from 1.5 g of 4, compound $3C_7$ (2.4 g, 88%), m.p. 92 °C (methanol), was isolated with elution system B. Anal. calcd for C_{28} H₃₆ O₄: C, 77.03; H, 8.31%; found: C, 76.92; H, 8.37%. ν_{max} (KBr): 3076, 2956, 2924, 2854, 1669, 1577, 1514, 1466, 1377, 1332, 1312, 1219, 1111, 1087, 713 and 621 cm⁻¹. ¹H NMR (CDCl₃): δ 8.16 (2H, m, H5 and H8); 7.65 (2H, m, H6 and H7); 7.55 (2H, s, H1 and H4); 4.11 (4H, t, J = 6.7 Hz, OCH₂); 1.85 (4H, m, OCH₂CH₂); 1.47 (4H, m, OCH₂CH₂CH₂); 1.21 (12H, m); 0.87 (6H, t, J = 6.7 Hz, CH₃). ¹³C NMR: δ 182.4 (C9 and C10); 153.7 (C2 and C3); 133.6 (C6 and C7); 133.5 (C8a and C10a); 128.0 (C4a and C9a); 126.8 (C5 and C8); 109.2 (C1 and C4); 69.3 (OCH₂); 31.8 (CH₂CH₂CH₃). ²¹C

2,3-Di-n-octyloxy-9,10-anthraquinone ($3C_8$). Starting from 1.5 g of 4, compound $3C_8$ (2.45 g, 85%), m.p. 97 °C (methanol),



Scheme 3 Synthetic pathways for the preparation of $1(C_1, C_7-C_{12})$ and 2_{1-3} : (a) RBr, DMF, K_2CO_3 ; (b: i) NaBH₄, EtOH; (b: ii) HCl; (c) Br(CH₂)_nBr, DMF, K_2CO_3 ; (d) Zn, NH₄OH.

was isolated with elution system B. ν_{max} (KBr): 3076, 2921, 2851, 1671, 1576, 1515, 1465, 1377, 1332, 1315, 1220, 1111, 1087 and 712 cm⁻¹. ¹H NMR (CDCl₃): δ 8.15 (2H, m, H5 and H8); 7.65 (2H, m, H6 and H7); 7.55 (2H, s, H1 and H4); 4.11 (4H, t, J = 6.7 Hz, OCH₂); 1.85 (4H, m, OCH₂CH₂); 1.47 (4H, m, OCH₂CH₂CH₂); 1.30–1.20 (16H, m, CH₂); 0.86 (6H, t, J = 6.7 Hz, CH₃). ¹³C NMR: δ 182.4 (C9 and C10); 153.7 (C2 and C3); 133.6 (C6 and C7); 133.5 (C8a and C10a); 128.0 (C4a and C9a); 26.8 (C5 and C8); 109.2 (C1 and C4); 69.3 (OCH₂); 31.8 (t); 29.3 (t); 29.0 (t); 25.9 (OCH₂CH₂CH₂); 22.7 (OCH₂CH₃); 14.1 (CH₃). FAB⁺-MS: m/z (%) 464 (100), 353 (44), 240 (85).

2,3-Di-n-nonyloxy-9,10-anthraquinone ($3C_9$). Starting from 1.0 g of **4**, compound $3C_9$ (1.75 g, 85%), m.p. 103—104 °C (methanol), was isolated with elution system B. ν_{max} (KBr): 3078, 2957, 2920, 2850, 1670, 1575, 1514, 1466, 1332, 1220, 1111, 712 et 621 cm⁻¹. ¹H NMR (CDCl₃): δ 8.17 (2H, m, H5 and H8); 7.66 (2H, m, H6 and H7); 7.59 (2H, s, H1 and H4); 4.13 (4H, t, J = 6.7 Hz, OCH₂); 1.82 (4H, m, OCH₂CH₂); 1.45 (4H, m, OCH₂CH₂CH₂); 1.30–1.20 (20H, m, CH₂); 0.83 (6H, t, J = 6.7 Hz, CH₃). ¹³C NMR: δ 183.1 (C9 and C10); 153.5 (C2 and C3); 133.6 (C6 and C7); 129.5 (C8a and C10a); 127.7 (C4a and C9a); 126.9 (C5 and C8); 109.3 (C1 and C4); 69.4 (OCH₂); 31.9 (t); 29.6 (t); 29.4 (t); 29.3 (t); 28.9 (t); 25.9 (OCH₂CH₂CH₂); 22.7 (OCH₂CH₂); 14.1 (CH₃). FAB⁺-MS: m/z (%) 492 (100), 367 (24), 240 (53).

2,3-Di-n-undecyloxy-9,10-anthraquinone ($3C_{11}$). Starting from 1.0 g of **4**, compound $3C_{11}$ (1.8 g, 79%), m.p. 97°C (heptane–benzene), was obtained with elution system A. ν_{max} (KBr): 2955, 2922, 2849, 1669, 1576, 1515, 1466, 1378, 1329, 1221, 1113, 713 and 621 cm⁻¹. ¹H NMR (CDCl₃): δ 8.26 (2H, m, H5 and H8); 7.74 (2H, m, H6 and H7); 7.67 (2H, s, H1 and H4); 4.18 (4H, t, J = 6.9Hz, OCH₂); 1.89 (4H, m, OCH₂CH₂); 1.48 (4H, m, OCH₂CH₂CH₂); 1.25 (28H, m, CH₂); 0.87 (3H, t, J = 6.6 Hz, CH₃). ¹³C NMR: δ 182.7 (C9 and C10); 153.5 (C2 and C3); 133.7 (C6 and C7); 133.5 (C8a and C10a); 128.0 (C4a and C9a); 126.9 (C5 and C8); 109.3 (C1 and C4); 69.4 (OCH₂); 32.0 (t); 29.8 (t); 29.7 (t); 29.6 (t); 29.4 (t); 28.9 (t); 26.0 (t); 22.7 (CH₂CH₃); 14.2 (CH₃). FAB⁺-MS: m/z (%) 548 (100); 389 (37); 240 (59).

Compounds $1C_n$: general procedue for the reduction of anthraquinones $3C_n$. In a three-neck round-bottom flask equipped with a reflux condenser, a drying column and a magnetic stirrer, the required amount of 2,3-dialkoxy-9,10-anthraquinone and 2-propanol (20 cm³ per mmol) were placed and a large excess of sodium borohydride (ca. 20 molar equiv.) was added in small portions at such a rate as to avoid a rapid temperature rise. The medium was refluxed under nitrogen for 3 h and allowed to cool to room temperature. The mixture was then hydrolyzed with 35% hydrochloric acid in crushed ice and extracted repeatedly with dichloromethane. The organic layers were combined, washed with a sodium hydroxide solution (pH \approx 9) and water until neutral, dried (MgSO₄) and filtered. The solvent was evaporated and the solid residue (which is the anthrone intermediate), which was not purified further, was treated in the same way, using the same set-up as above, except that the reflux was maintained for ca. 12 h. The final residue was chromatographed on a column of silica gel after dissolution in a solvent mixture of CH₂Cl₂-petroleum ether (v/v 30:70). Average yields were 50-60%.

2,3-Di-n-heptyloxyanthracene (1C₇). Starting from 3C₇ (0.75 g), 0.39 g (56%) of 1C₇, m.p. 110 °C, was isolated. Anal. calcd for C₂₈H₃₈O₂: C, 82.76; H, 9.36%; found: C, 82.66; H, 9.47%. $\nu_{\rm max}$ (KBr): 2951, 2929, 2855, 1627, 1568, 1493, 1465, 1387, 1291, 1261, 1228, 1193, 1166, 1007, 889, 832, 743, 595 and 474 cm⁻¹. ¹H NMR: δ 8.24 (s, H9 and H10); 7.96 (m, H5 and H8); 7.45 (m, H6 and H7); 7.22 (s, H1 and H4); 4.17 (t, J = 6.4Hz, OCH₂); 2.00 (m, OCH₂CH₂); 1.58 (m,

OCH₂CH₂CH₂); 1.43 (m, CH₂); 1.02 (t, J = 6.4 Hz, CH₃). ¹³C NMR: δ 150.2 (C2 and C3); 130.9 (C8a and C10a); 128.9 (C4a and C9a); 127.7 (C5 and C8); 124.4 (C6 and C7); 123.8 (C9 and C10); 106.0 (C1 and C4); 68.8 (OCH₂); 32.0 (t); 29.3 (t); 29.2 (t); 26.2 (CH₂CH₃); 14.2 (CH₃).

2,3-Di-n-octyloxyanthracene (1C₈). Starting from 0.81 g of $3C_8$, 0.37 g (49%) of $1C_8$, m.p. 105 °C, was obtained. ν_{max} (KBr): 2952, 2930, 2855, 1627, 1568, 1493, 1465, 1387, 1291, 1261, 1230, 1193, 1166, 1006, 889, 832, 743, 595 and 474 cm^{-1.} ¹H NMR: δ 8.22 (s, H9 and H10); 7.95 (m, H5 and H8); 7.43 (m, H6 and H7); 7.20 (s, H1 and H4); 4.17 (t, J = 6.4Hz, OCH₂); 1.97 (m, OCH₂CH₂); 1.58 (m, OCH₂CH₂CH₂); 1.39 (m, CH₂); 0.96 (t, J = 6.4 Hz, CH₃). ¹³C NMR: δ 150.1 (C2 and C3); 130.8 (C8a and C10a); 128.8 (C4a and C9a); 127.7 (C5 and C8); 124.4 (C6 and C7); 123.8 (C9 and C10); 106.0 (C1 and C4); 68.8 (OCH₂); 31.9 (t); 29.5 (t); 29.4 (t); 29.2 (t); 26.2 (CH₂CH₃); 22.8 (t); 14.2 (CH₃). HRMS: calcd for C₃₀H₄₂O₂: 434.3184; found: 434.3148.

2, 3-Di-n-nonyloxyanthracene (1C₉). Starting from $3C_9$ (1.23 g), 0.64 g (55%) of $1C_9$, m.p. 100 °C, was isolated. Anal. calcd for $C_{32}H_{46}O_2$: C, 83.12; H, 9.96%; found: C, 83.29; H, 9.94%. ν_{max} (KBr): 2953, 2919, 2851, 1629, 1568, 1530, 1491, 1481, 1466, 1384, 1287, 1223, 1165, 1005, 889, and 739 cm⁻¹. ¹H NMR: δ 8.24 (s, H9 and H10); 7.98 (m, H5 and H8); 7.47 (m, H6 and H7); 7.21 (s, H1 and H4); 4.17 (t, J = 6.6Hz, OCH₂); 1.99 (m, OCH₂CH₂); 1.58 (m, OCH₂CH₂CH₂); 1.25 (m, 20H, CH₂); 0.98 (t, J = 6.6 Hz, CH₃). ¹³C NMR: δ 150.0 (C2 and C3); 130.7 (C8a and C10a); 128.7 (C4a and C9a); 127.6 (C5 and C8); 124.4 (C6 and C7); 123.7 (C9 and C10); 105.9 (C1 and C4); 68.7 (OCH₂); 32.0 (CH₂CH₂CH₃); 29.7 (t); 29.5 (t); 29.4 (t); 29.1 (t, OCH₂CH₂); 26.2 (t, OCH₂CH₂CH₂); 22.8 (CH₂CH₃); 14.2 (CH₃).

2, 3-Di-n-undecyloxyanthracene ($1C_{II}$). Starting from $3C_{11}$ (0.73 g), 0.43 g (62%) of $1C_{11}$, m.p. $102-103 \,^{\circ}$ C (pentane), was obtained. ν_{max} (KBr): 2917, 2849, 1630, 1586, 1491, 1481, 1466, 1384, 1287, 1222, 1164, 890 and 739 cm⁻¹. ¹H NMR: δ 8.25 (s, H9 and H10); 7.98 (m, H5 and H8); 7.45 (m, H6 and H7); 7.23 (s, H1 and H4); 4.20 (t, J = 6.8Hz, OCH₂); 2.00 (m, OCH₂CH₂); 1.55 (m, OCH₂CH₂CH₂); 1.37 (m, CH₂); 0.98 (t, J = 6.8 Hz, CH₃). ¹³C NMR: δ 150.1 (C2 and C3); 130.8 (C8a and C10a); 128.8 (C4a and C9a); 127.6 (C5 and C8); 124.4 (C6 and C7); 123.8 (C9 and C10); 106.0 (C1 and C4); 68.8 (OCH₂); 32.0 (CH₂CH₂CH₃); 29.7 (t); 29.5 (t); 29.4 (t); 29.1 (t); 26.2 (t); 22.8 (CH₂CH₃); 14.2 (CH₃). FAB⁺-MS: m/z (%) 518 (100); 329 (33); 307 (45); 289 (52); 210 (68). HRMS: calcd for C₃₆H₅₄O₂: 518.4123; found 518.4116.

2,3-Trimethylenedioxyanthracene (2₃). 2,3-Dihydroxy-9,10-anthraquinone (**4**; 3 g, 12.5 mmol),⁵⁰ potassium carbonate (5 g, 36 mmol) and 100 cm³ DMF were placed in a three-necked round-bottom flask equipped with a dropping funnel and a reflux condenser, under a nitrogen atmosphere. A solution of 1,3-dibromopropane (3 g, 15 mmol) in 50 cm³ DMF was added slowly through the dropping funnel to the mixture heated at 100 °C. After the addition, the medium temperature was raised to 160 °C and maintained for 17 h. Another aliquot of the anthraquinone solution (0.5 g, 3.6 mmol, in 25 cm³ DMF) was then added and allowed to react for a further 2 h. After cooling, the reaction mixture was diluted with 300 cm³ water, which generated a yellowish precipitate that was filtered off and dissolved in CH₂Cl₂, washed with water and dried on Na₂SO₄. After filtration and evaporation of the solvent, the pale brown raw material $(5_3, 2.4 \text{ g})$ was collected to be used in the next step without further purification.

In a round-bottom flask equipped with a reflux condenser, a ground mixture of the crude anthraquinone derivative (5_3) and zinc powder (5 g, 7.7 mmol) was added to a 20% ammonia

aqueous solution (100 cm³). The mixture, which rapidly turned dark red, was stirred at 90 °C until the coloration vanished (*i.e.* for about 10 h). The mixture was filtered and the solid was washed with boiling ethanol, dissolved in dichloromethane and chromatographed on a column of silica gel (eluant: CH₂Cl₂-petroleum ether, v/v 50:50). Compound **2**₃ was isolated as a pale yellow powder (1.1 g, overall yield 35%), m.p. 218 °C. Anal. calcd for C₁₇H₁₄O₂: C, 81.60; H, 5.60; O, 12.80%; found: C, 82.22; H, 5.60; O, 12.73%. ν_{max} (KBr): 3040, 2950, 2860, 1475, 1455, 1380, 1300, 1275, 1210, 1135, 1040, 935, 885 and 730 cm⁻¹. ¹H NMR (CDCl₃): δ 2.28 (m, 2H); 4.33 (t, 4H); 7.0–8.26 (m, 8H). MS: m/z (%) 250 (M⁺, 100), 222 (25), 181 (28), 165 (23), 164 (25), 152 (17), 28 (31).

Appendix: determination of equilibrium constants between solution and gel phase from the phase transition diagrams

At a given temperature, the equilibrium constant of the reversible reaction: gel \rightleftharpoons liquid (solution) is expressed by K =activity (liquid)/activity (gel) $\approx a/1$.

The melting of a gel into the liquid phase is assumed to resemble the melting of a solid⁶⁶ whose activity is 1. As the concentration of DDOA is low, the activity of the liquid phase will be represented by that of its molar fraction denoted ϕ ; therefore:

$$\ln K = \ln \phi \tag{A1}$$

where

$$\ln\phi = -\Delta G^0 / RT \tag{A2}$$

and

$$\ln\phi = -\Delta H^0/R \times 1/T_{\rm m} + \Delta S^0/R \tag{A3}$$

with $T_{\rm m}$ being the melting temperature.

Conversely, the reversible formation of a gel from the liquid phase (similar to a crystallization) will be represented by $K \approx 1/a$, which leads to:

$$\ln\phi = +\Delta H^0 / R \times 1 / T_{gel} - \Delta S^0 / R \tag{A4}$$

where T_{gel} is the gelification temperature. ΔH^0 and ΔS^0 (standard conditions) will be positive for the melting and negative for the gelification. The molar fraction $\phi = x_{solute}/(x_{solute} + x_{solvent})$. As $x_{solute} << x_{solvent}$, $\phi \approx x_{solute}/x_{solvent}$ and therefore, $\phi = (m/M)/(m'/M')$ where *m* and *m'* are the masses of solute and solvent, respectively while *M* and *M'* are the molar masses.

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