

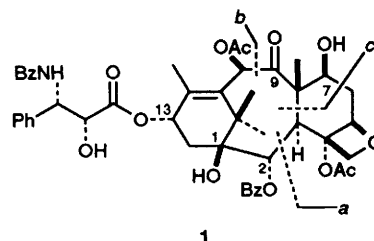
## Enantioselective Synthesis of a 9,10-*seco*-Taxane Derivative *via* Electrophilic Epoxy-allylsilane Ring Closure

Lars Pettersson and Torbjörn Frejd\*†

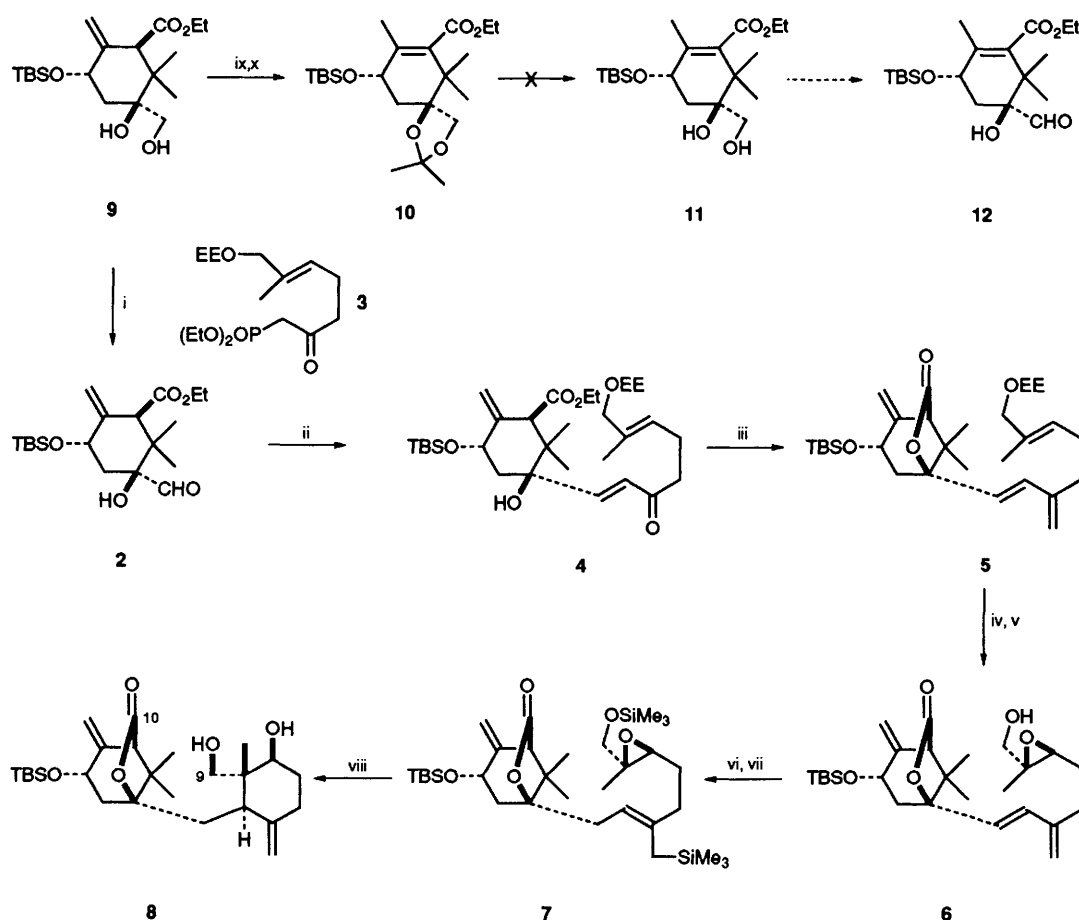
Department of Organic Chemistry 2, Chemical Center, Lund University, PO Box 124, S-221 00 Lund, Sweden

*seco*-Taxane **8**, a potential starting material for further development into highly substituted taxanes, is synthesized by  $\text{BF}_3 \cdot \text{OEt}_2$  treatment of epoxy-allylsilane **7**, in its turn synthesized by joining an *A*-ring derivative **2** with a *C*-ring precursor chain **3**.

We previously reported syntheses of Taxol *A*- and *C*-ring units of high optical purity potentially useful for the construction of functionalized taxoids and perhaps also Taxol.<sup>1–5</sup> Our attempts at the convergent strategy of directly joining the *A*- and the *C*-ring units *via* a C(2)–C(3) bond formation followed



† Present address: Department of Organic Chemistry, Umeå University, S-901 87 Umeå, Sweden.



**Scheme 1** Reagents and conditions: i, 4 equiv. of dimethyl sulfoxide, 2 equiv. of  $(\text{COCl})_2$ , 8 equiv. of  $\text{EtN}(\text{Pr})_2$ ,  $\text{CH}_2\text{Cl}_2$ ,  $-60^\circ\text{C}$  for 20 min then  $+20^\circ\text{C}$  1 h, 90%; ii, 1.3 equiv. of  $\text{NaH}$ , 1.5 equiv. of **3**, 1,2-dimethoxyethane,  $0^\circ\text{C}$ , 20 min then **2** at  $85^\circ\text{C}$ , 10 min, 93%; iii, 2 equiv. of  $1.4 \text{ mol dm}^{-3}$   $\text{BuLi}$  in hexane, 2 equiv. of  $\text{Ph}_3\text{PCH}_2\text{Br}$ , tetrahydrofuran,  $+20^\circ\text{C}$ , then add **4** rapidly at  $50^\circ\text{C}$ , 15 min, 90%; iv, 0.1 equiv. of pyridinium toluene-*p*-sulfonate (PPTS),  $\text{EtOH}$ ,  $+20^\circ\text{C}$ , 12 h, 98%; v, 0.32 equiv. of L-(+)-DET, 0.7 g of 4 Å molecular sieves, 1.53 equiv. of *tert*-butylhydroperoxide (TBHP) (anhydrous), 0.27 equiv. of  $\text{Ti}(\text{OPr})_4$ ,  $\text{CH}_2\text{Cl}_2$ ,  $-40^\circ\text{C}$ , 1.5 h, 97%; vi, 2 equiv. of  $\text{EtN}(\text{Pr})_2$ , 2 equiv. of  $\text{Me}_3\text{SiCl}$ ,  $0^\circ\text{C}$ , 5 min, 99%; vii, 0.3 ml of  $\text{Me}_3\text{SiH}$ , toluene, 230 mg of **6** ( $\text{R} = \text{SiMe}_3$ ), 1.5 mg of  $(\text{Ph}_3\text{P})_3\text{RhCl}$ ,  $+20^\circ\text{C}$ , 16 h, occasional opening of the closed reaction vessel at  $0^\circ\text{C}$ , then PPTS in  $\text{EtOH}$ , 5 min, 35%; viii, **7** in  $\text{CH}_2\text{Cl}_2$  slowly added to 15 equiv. of  $\text{BF}_3\cdot\text{OEt}_2$ ,  $0^\circ\text{C}$ , then add aq.  $\text{NaHCO}_3$ , 30%; ix, acetone,  $\text{BF}_3\cdot\text{OEt}_2$ ,  $\text{CH}_2\text{Cl}_2$ ,  $0^\circ\text{C}$ , 3 min, 96%; x, DBU (neat, 5 times the mass of the acetone),  $185^\circ\text{C}$ , 1 h, 60%. EE = ethoxyethyl, DET = diethyltartrate, DBU = 1,8-diaza-[5.4.0]bicycloundec-7-ene. Yields are given for isolated products pure by  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectroscopy, TLC analysis and having correct elemental analysis. All reactions were performed under  $\text{N}_2$  in dry solvents.

by the C(9)–C(10) bond formation for the synthesis of the taxane core (formation of bond *a* followed by *b*, Fig. 1) have hitherto been unsuccessful.<sup>3,4</sup> Steric crowding at C-2 is probably too great, which suggested that a sterically less demanding chain-shaped C-ring precursor be attached to C-2 of a suitable A-ring structure (bond formation order *a*, *c*, *b*). As a continuation of this work we now report the synthesis of 9,10-*seco*-taxane **8** (Scheme 1), which may serve as a starting material for taxanes.<sup>7–9</sup> Both of the C-9 and C-10 terminals† could conceivably be modified to allow testing of several different C–C bond formation reactions. It was demonstrated several years ago by Kende *et al.*<sup>8</sup> and more recently by Nicolaou *et al.*<sup>9</sup> that the 9,10-*seco*-taxane route may be of great importance for the synthesis of taxanes. Syntheses of other optically active A-ring derivatives similar to **2** have been published.<sup>10</sup>

The ideal A-ring unit to start with would be the C-2 aldehyde **12**. This requires that the acetonide protection is removed from **10**,<sup>2,5</sup> which was obtained from **9**<sup>2,5</sup> by acetonide formation followed by heating in neat DBU at  $185^\circ\text{C}$  for 1 h. However, efforts to cleave the acetonide were all unsuccessful (*e.g.* acidic solvolysis, or treatment with  $\text{Me}_2\text{BBr}$ ,<sup>11</sup>  $\text{PdCl}_2(\text{MeCN})_2$ <sup>12</sup> and  $\text{Ph}_3\text{CBF}_4$ <sup>13</sup>); either the acetonide was too stable or the material was destroyed. Instead, the primary hydroxy group of the unconjugated A-ring derivative **9** was oxidized to give the  $\alpha$ -hydroxy aldehyde **2** in 90% yield using the Swern procedure.<sup>14</sup> This method is well suited for 1,2-diols containing one tertiary hydroxy group, whereas other oxidizing agents may cleave the diol.<sup>15</sup>

The Horner–Emmons–Wadsworth (HEW) reaction<sup>16</sup> of **2** with the sodium salt of **3**<sup>17</sup> was very efficient producing the  $\alpha,\beta$ -unsaturated ketone **4** in 93% yield. As indicated by the  $^1\text{H}$  NMR spectrum ( $J_{2,3}$  15.5 Hz) the C(2)–C(3) double bond in **4** was of pure *E* configuration. The conjugated diene **5** was prepared by a Wittig reaction ( $\text{Ph}_3\text{P}=\text{CH}_2$  in tetrahydrofuran) of **4** in 90% yield. Lactonisation took place during this

† Taxane numbering is used throughout.

reaction. Deprotection of **5** followed by a catalytic Sharpless epoxidation<sup>18</sup> gave the epoxy alcohol **6** (R = H) in excellent yield (97%).§

The hydrosilylation was then performed with the trimethylsilyl-protected derivative **6** (R = SiMe<sub>3</sub>) using (Ph<sub>3</sub>P)<sub>3</sub>RhCl as a catalyst, which produced the epoxy allylsilane **7** (R = H after hydrolytic work-up) in 35% yield along with some side products (the regioisomeric allyl silane and the 1,4-hydrogenation product). The C-ring cyclization to give **8**¶ (30% isolated yield) was accomplished by adding **7** to 15 equiv. of BF<sub>3</sub>·OEt<sub>2</sub> in CH<sub>2</sub>Cl<sub>2</sub> (not *vice versa*) at 0 °C, followed by column chromatography and recrystallization for heptane. Unfortunately, the crystals were not suitable for X-ray analysis.

The structure of **8** was verified by NMR techniques, including DEPT, HETCOR, HOM2DJ, COSY, and NOESY. Some important changes in the <sup>1</sup>H and <sup>13</sup>C NMR spectra in going from **7** to **8** are (i) loss of the SiMe<sub>3</sub> group, the C-3 double bond and the epoxide; (ii) a pronounced upfield shift of the C-19 methyl group (both in the <sup>1</sup>H and in the <sup>13</sup>C NMR spectrum); (iii) new <sup>1</sup>H signals for the C-20 *exo*-methylene group, the >CH(OH) moiety at C-7 and the <sup>13</sup>C signal for C-3. The relative stereochemistry at C-3, C-7 and C-8 and the assignment of H-3 were determined by 2D-techniques, including NOESY. In particular NOE effects were obtained for 2-H ↔ 19-H, 19-H ↔ 7-OH, 3-H ↔ 7-H

indicating that the three former groupings were *cis* related on one side of the ring system and the latter two *cis* related on the other. This information together with the stereospecificity of the Sharpless epoxidation to give **6** strongly suggest that the C-ring precursor chain in **7** gave a six-membered ring with the correct stereochemistry at C-3, C-8 and C-7 as compared to Taxol. Thus, we now have control over five of the ten stereocentres related to Taxol (C-1, C-3, C-7, C-8, C-13) not counting those of the side chain. Obviously, the yields in the hydrosilylation and the ring closure steps need improving and further work along these lines is in progress.

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§ The diastereoisomeric excess of the epoxide **6** (R = H) was not determined. However, the set of signals corresponding to only one diastereoisomer could be detected in its <sup>1</sup>H NMR spectrum.

¶ Physical data for selected compounds. For **7**: *R*<sub>f</sub> = 0.24 (silica, heptane : ethyl acetate 2 : 1); [α]<sub>D</sub><sup>20</sup> + 101 (c 0.99, CDCl<sub>3</sub>); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>); δ 0.04 (s, 9H, -SiMe<sub>3</sub>), 0.05, 0.06 [2s, 6H, -Si(CH<sub>3</sub>)<sub>2</sub>-], 0.90 (s, 9H, Bu<sup>t</sup>), 0.99, 1.10 (2s, 6H, H-16, H-17), 1.28 (s, 3H, H-19), 1.52, 1.58 (2d, 2H, *J*<sub>AB</sub> 13.6 Hz, H-20), 1.63–1.75 (m, 4H, H-14, H-6, -OH), 2.13 (m, 2H, H-5), 2.24 (dd, 1H, *J*<sub>AB</sub> 15.3, *J* 8.5 Hz, H-2), 2.32 (dd, 1H, *J*<sub>AB</sub> 14.0, *J* 7.8 Hz, H-14), 2.38 (dd, 1H, *J*<sub>AB</sub> 15.3, *J* 5.6 Hz, H-2), 2.85 (s, 1H, H-11), 3.03 (t, 1H, *J* 6.1 Hz, H-7), 3.58 (dd, 1H, *J*<sub>AB</sub> 12.2, *J* 8.6 Hz, H-9), 3.68 (dd, 1H, *J*<sub>AB</sub> 12.2, *J* 4.4 Hz, H-9), 4.30 (dddd, 1H, *J* 9.3, 7.8, 2.4, 2.4 Hz, H-13), 4.95 (dd, 1H, *J* 2.4, 1.5 Hz, H-18), 5.12 (dd, 1H, *J* 8.5, 5.6 Hz, H-3), 5.24 (dd, 1H, *J* 2.4, 1.5 Hz, H-18); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ -5.08, -4.84 (-SiMe<sub>2</sub>-), -0.61 (-SiMe<sub>3</sub>), 14.28 (C-19), 18.18 (-CMe<sub>3</sub>), 20.13 (C-16), 21.72 (C-20), 22.46 (C-17), 25.79 (-CMe<sub>3</sub>), 27.38, 31.68, 36.04 (C-6, C-5, C-2), 39.18 (C-14), 45.32 (C-15), 59.55, 60.81, 60.96 (C-7, C-8, C-11), 65.18 (C-9), 67.73 (C-13), 91.41 (C-1), 111.19 (C-18), 114.98 (C-3), 140.11 (C-4), 142.64 (C-12), 175.77 (C-10).

For **8**: *R*<sub>f</sub> = 0.19 (silica, heptane : ethyl acetate 1 : 1); [α]<sub>D</sub><sup>20</sup> + 90 (c 0.30, CDCl<sub>3</sub>); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 0.06, 0.06 (2s, 6H, -SiMe<sub>2</sub>-), 0.71 (s, 3H, H-19), 0.91 (s, 9H, Bu<sup>t</sup>), 0.98, 1.13 (2s, 6H, H-16, H-17), 1.50–1.62 (m, 3H, H-6, H-2, H-14), 1.81 (d, 1H, *J* 4.8 Hz, >CHOH, disappears with D<sub>2</sub>O), 1.88 (m, 1H, H-6), 2.04 (dd, 1H, *J*<sub>AB</sub> 12.8, *J* 4.5 Hz, broad, H-5), 2.10 (dd, 1H, *J* 4.2, 6.1 Hz, -CH<sub>2</sub>OH, disappears with D<sub>2</sub>O), 2.13 (dd, 1H, *J*<sub>AB</sub> 14.9, *J* 9.6 Hz, H-2), 2.22 (d, 1H, *J* 9.6 Hz, broad, H-3), 2.39 (ddd, 1H, *J*<sub>AB</sub> 12.8, *J* 4.2, 4.2 Hz, H-5), 2.47 (dd, 1H, *J*<sub>AB</sub> 14.0, *J* 7.7 Hz, H-14), 2.84 (s, 1H, H-11), 3.50 (dd, 1H, *J*<sub>AB</sub> 11.0, *J* 4.2 Hz, H-9), 3.76 (dd, 1H, *J*<sub>AB</sub> 11.0, *J* 6.1 Hz, H-9), 3.91 (ddd, 1H, *J* 4.8, 10.5, 4.7 Hz, H-7), 4.24 (dddd, 1H, *J* 9.4, 7.7, 2.5, 2.5 Hz, H-13), 4.84 (s, 1H, H-20), 4.94 (dd, 1H, *J* 2.5, 1.5 Hz, H-18), 5.01 (s, 1H, H-20), 5.23 (dd, 1H, *J* 2.5, 1.5 Hz, H-18); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ -5.09, -4.78 (-SiMe<sub>2</sub>-), 10.57 (C-19), 18.20 (-CMe<sub>3</sub>), 19.61 (C-16), 22.54 (C-17), 25.82 (-CMe<sub>3</sub>), 26.73 (C-2), 32.29 (C-6), 33.70 (C-5), 39.34 (C-14), 40.72 (C-3), 44.84, 46.46 (C-8, C-15), 60.55 (C-11), 67.59 (C-13), 67.72 (C-9), 73.51 (C-7), 91.09 (C-1), 110.57 (C-20), 111.00 (C-18), 142.52 (C-12), 146.49 (C-4), 175.87 (C-10).