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# Intramolecular cascade rearrangements of enynamine derived ketenimines: Access to acyclic and cyclic amidines

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Dinesh Pratapsinh Chauhan,<sup>a</sup> Sreejith J. Varma,<sup>a,b</sup> Mahesh Gudem,<sup>a</sup> Nihar Panigrahi,<sup>a</sup> Khushboo Singh,<sup>a,c</sup> Anirban Hazra,<sup>a</sup> and Pinaki Talukdar<sup>\*a</sup> Copper-catalyzed reaction of enynamines with sulfonylazides provides acyclic and cyclic amidines. Nucleophilic addition of

the tethered amino group on the *in situ* generated ketenimine forms a six-membered cyclic zwitterionic intermediate which facilitates migration of the tethered amino group to the  $C_s$ -center giving the acyclic amidine. On the other hand, migration of a substituent on the amino group to  $C_{2^-}$  and  $C_4$ -centers results in the formation of cyclic amidines. Computational studies were carried out to validate the mechanism which indicate that the product distribution of the process depends on the substitutions on the enynamine backbone.

### Introduction

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Since the dawn of modern chemistry, reactive intermediates have served as an inspiration for the development of new reactions, molecules, and materials. Indeed, the strategies involving reactive intermediates have made possible the construction of numerous complex organic frameworks in easier and shorter routes. In general, reactive intermediates readily undergo chemical transformations, and their reactions are typically exothermic.<sup>1</sup> Among several reactive entities in organic chemistry, ketenimines represent an important class of reactive intermediates which has been applied widely as a powerful synthon. During last two decades, several methodologies were developed towards the formation of ketenimine, and to explore its synthetic applications.<sup>2</sup> Because the remarkable reactivity, the ketenimine was of comprehensively applied in the synthesis of many complex compounds.<sup>3</sup> Interception of this reactive species with other functional groups led to novel strategies for the reaction discovery.<sup>4</sup> Going through literature, it is observed that ketenimine undergoes nucleophilic addition,<sup>2i</sup> cycloaddition,<sup>5</sup> radical addition,<sup>6</sup> electrocyclic ring-closure reactions,<sup>7</sup> and  $\sigma$ 

<sup>a</sup> Department of Chemistry, Indian Institute of Science Education and Research Pune, Dr. Homi Bhabha Road, Pashan, Pune 411008, Maharashtra, India. *Email: ptalukdar@iiserpune.ac.in*  rearrangements.<sup>3b,3c,8</sup> For example, *N*-sulfonyl/phosphoryl ketenimine undergoes addition reactions with external nucleophiles *e.g.* amines, alcohols, pyrroles, indoles, ammonium salts, etc.<sup>2i</sup>

In 2014, we reported a novel rearrangement of ketenimine intermediate, generated from the substituted propargylamine I, by an internal amine nucleophile forming amidine Ia via the [1,3]-amino group migration (Scheme 1A).<sup>9</sup> Our success in this area prompted us to investigate the possibilities of new rearrangement reactions of ketenimine, envisaged through the incorporation of an additional alkene moiety at the C<sub>2</sub>-C<sub>3</sub> position. Here, we report that such structural modification leads to not only the formation of acyclic amidine IIa but also allows the access to highly functionalized cyclic amidines IIb and IIc from II (Scheme 1B).



Scheme 1. Reaction of propargylamine I with sulfonylazide under Cu(I) catalyzed conditions giving amidine Ia as the [1,3]-amino group migration product (A). Reaction enynamine II with sulfonylazide under Cu(I) catalyzed conditions in the formation of acyclic amidines IIa-IIc (B).

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### **Results and discussion**

We started our preliminary investigation with the conjugated enynamine substrate 1, where the amino moiety is the 4morpholinyl group. The reaction of 1 with TsN<sub>3</sub> (1.1 equiv) in presence of catalytic Cul (10 mol%) and Et<sub>3</sub>N (1.5 equiv) in CHCl<sub>3</sub> provided the [1,5]-amino group migration product 1a in 67% yield (Table 1, Entry 1). When substrates 2-4 were used by introducing phenyl group at either C<sub>2</sub>- or C<sub>3</sub>- or both positions, corresponding [1,5]-amino group migration products 2a-4a were formed (Entries 2-4). The observations were comparable when enynamine substrates were decorated with acyclic amino groups, and C2- and C3-positions were also randomly substituted with a phenyl group (Entries 5-8).

An interesting twist in the product formation was observed when the envne was decorated with the -NBn<sub>2</sub> group. Such a substrate **9** with  $R^3 = R^4 = H$  when reacted with TsN<sub>3</sub> under the Cul catalytic conditions, cyclic amidines 9b (51%) and 9c (16%) were formed apart from the expected acyclic amidine 9a (23%) (Table 2, entry 1). The general plausible mechanism of the process is depicted in Scheme 2. Reaction of enynamine II with a sulfonylazide under Cu(I) catalytic conditions provides the triazole intermediate IId which upon releasing a N2 molecule gives ketenimine IIe.<sup>10</sup> Capture of IIe by the internal amino group leads to the 6-exo-dig cyclization<sup>11</sup> to form the zwitterionic intermediate IIf which has canonical forms IIg and IIh. The C<sub>1</sub>-N bond cleavage of the cyclic intermediate IIf via E1cB process leads to the formation of [1,5]-amino group migration product IIa. The electrophilic migration of the R<sub>1</sub>

|                |                             | DOI: 10.1039/C7OB00                |  |        |  |  |
|----------------|-----------------------------|------------------------------------|--|--------|--|--|
|                |                             | Cul, Et<br>rt,                     | Et <sub>3</sub> N, CHCl <sub>3</sub> ,<br>, 30 min |        | TsN                                      |  |
| R <sup>4</sup> | ~ N <sup>R1</sup>           | + TsN <sub>3</sub>                 |  | →<br>( |  |  |
| R <sup>3</sup> | <sup>8</sup> R <sup>2</sup> |                                    |  |        | $\sim R^2$                               |  |
| 1              | -8                          |                                    |  |        | 1a-8a                                    |  |
| Entry          | 1-8                         | $-NR^{1}R^{2}$                     | R <sup>3</sup>                                     | $R^4$  | <b>1a-8a</b> (Yield) <sup><i>a</i></sup> |  |
| 1              | 1                           | -(4-morpholinyl)                   | н  | н      | <b>1a</b> (67%)                          |  |
| 2              | 2                           | -(4-morpholinyl)                   | Ph   | Н      | <b>2a</b> (76%)                          |  |
| 3              | 3                           | -(4-morpholinyl)                   | н  | Ph     | <b>3a</b> (70%)                          |  |
| 4              | 4                           | -(4-morpholinyl)                   | Ph   | Ph     | <b>4a</b> (65%)                          |  |
| 5              | 5                           | -N(Et) <sub>2</sub>                | Н  | Ph     | <b>5a</b> (67%)                          |  |
| 6              | 6                           | -N( <sup>'</sup> Pr) <sub>2</sub>  | н  | Н      | <b>6a</b> (57%)                          |  |
| 7              | 7                           | -N( <sup>n</sup> Hex) <sub>2</sub> | н  | н      | <b>7a</b> (62%)                          |  |
| 8              | 8                           | -N( <sup>n</sup> Hex) <sub>2</sub> | Н  | Ph     | <b>8a</b> (62%)                          |  |

<sup>a</sup>All yields are isolated yields.

group of the intermediate **IIf** to its C<sub>2</sub>-center gives the cyclic amidine IIb. On the other hand, an electrophilic migration of the R<sub>1</sub> group to the C<sub>4</sub>-center facilitates the formation of adduct IIc.

We further investigated the effects of R and  $R^1-R^4$  groups on product distribution to obtain greater insight on observed rearrangement processes. All conjugated enynamine substrates were prepared according to three general sequences outlined in Schemes S1, S3, S5 and S7.<sup>12</sup> The -NBn<sub>2</sub> substituted enynamine **10** ( $R^3 = Ph$ ,  $R^4 = H$ ), when treated with TsN<sub>3</sub>, cyclic amidines **10b** (50%) and **10c** (40%) were

|       | R    | $ \begin{array}{c}                                     $ | RSO <sub>2</sub> N <sub>3</sub> , Cul<br>CHCl <sub>3</sub> , rt, 30 | , Et <sub>3</sub> N,<br>0 min RS0<br>→ R <sup>1</sup><br>F | $\begin{array}{c} \text{RSO}_2 N \\ R \\ R \\ N \\ R^2 \\ R^2 \end{array} \xrightarrow{\begin{array}{c} R^3 \\ 2 \\ 3 \\ R^4 \\ R^4 \end{array}} $ |        | + R <sup>4</sup> 3 2<br>R <sup>3</sup> R <sup>1</sup> | + $R^{4}$ $R^{3}$ $R^{2}$ $R^{2}$ $R^{3}$ |   |
|-------|------|--|---|--|--|--------|---|---|---|
| 9-24  |      |  |   | 9a-26a   |  | 9b-26b | 9c-26c  |   |   |
| Entry | 9-24 | R  | $R^1$   | R <sup>2</sup>   | R <sup>3</sup>   | $R^4$  | <b>9a-26a</b> (Yield) <sup>a</sup>                    | <b>9b-26b</b> (Yield) <sup><i>a</i></sup> | <b>9c-26c</b> (Yield) <sup><i>a</i></sup> |
| 1     | 9    | $4-MeC_6H_4$   | Bn <sup>b</sup>   | Bn   | н  | н      | <b>9a</b> (23%)                                       | <b>9b</b> (51%)                           | <b>9c</b> (16%)                           |
| 2     | 10   | $4-MeC_6H_4$   | Bn  | Bn   | Ph   | н      | <b>10</b> a (0%)                                      | <b>10b</b> (50%)                          | <b>10c</b> (40%)                          |
| 3     | 11   | $4-MeC_6H_4$   | Bn  | Bn   | н  | Ph     | <b>11a</b> (84%)                                      | <b>11b</b> (5%)                           | <b>11c</b> (0%)                           |
| 4     | 12   | $4-MeC_6H_4$   | Bn  | Bn   | Ph   | Ph     | <b>12a</b> (48%)                                      | <b>12b</b> (30%)                          | <b>12c</b> (0%)                           |
| 5     | 13   | $4-MeC_6H_4$   | 4- <i>CN</i> Bn <sup>c</sup>  | 4- <i>CN</i> Bn  | н  | н      | <b>13a</b> (13%)                                      | <b>13b</b> (37%)                          | <b>13c</b> (23%)                          |
| 6     | 14   | $4-MeC_6H_4$   | 4-CNBn  | 4- <i>CN</i> Bn  | Ph   | н      | <b>14a</b> (0%)                                       | <b>14b</b> (51%)                          | <b>14c</b> (32%)                          |
| 7     | 15   | $4-MeC_6H_4$   | 4- <i>CN</i> Bn   | 4- <i>CN</i> Bn  | н  | Ph     | <b>15a</b> (77%)                                      | <b>15b</b> (0%)                           | <b>15c</b> (0%)                           |
| 8     | 16   | $4-MeC_6H_4$   | 4- <i>CN</i> Bn   | 4- <i>CN</i> Bn  | Ph   | Ph     | <b>16a</b> (73%)                                      | <b>16b</b> (18%)                          | <b>16c</b> (0%)                           |
| 9     | 17   | $4-MeC_6H_4$   | 4- <i>MeO</i> Bn <sup>d</sup>                                       | 4- <i>MeO</i> Bn   | н  | н      | <b>17a</b> (0%)                                       | <b>17b</b> (63%)                          | <b>17c</b> (12%)                          |
| 10    | 18   | $4-MeC_6H_4$   | 4- <i>MeO</i> Bn  | 4- <i>MeO</i> Bn   | Ph   | н      | <b>18a</b> (0%)                                       | <b>18b</b> (60%)                          | <b>18c</b> (11%)                          |
| 11    | 19   | $4-MeC_6H_4$   | 4- <i>MeO</i> Bn  | 4- <i>MeO</i> Bn   | н  | Ph     | <b>19a</b> (8%)                                       | <b>19b</b> (70%)                          | <b>19c</b> (6%)                           |
| 12    | 20   | $4-MeC_6H_4$   | 4- <i>MeO</i> Bn  | 4- <i>MeO</i> Bn   | Ph   | Ph     | <b>20</b> a (0%)                                      | <b>20b</b> (68%)                          | <b>20c</b> (0%)                           |
| 13    | 21   | $4-MeC_6H_4$   | Bn  | 4- <i>CN</i> Bn  | н  | н      | <b>21a</b> (14%)                                      | <b>21b</b> (39%)                          | <b>21c</b> (23%)                          |
| 14    | 22   | $4-MeC_6H_4$   | Bn  | 4- <i>CN</i> Bn  | Ph   | н      | <b>22a</b> (0%)                                       | <b>22b</b> (50%)                          | <b>22c</b> (25%)                          |
| 15    | 23   | $4-MeC_6H_4$   | Bn  | 4- <i>CN</i> Bn  | н  | Ph     | <b>23a</b> (86%)                                      | <b>23b</b> (0%)                           | <b>23c</b> (0%)                           |
| 16    | 24   | $4-MeC_6H_4$   | Bn  | 4- <i>CN</i> Bn  | Ph   | Ph     | <b>24a</b> (46%)                                      | <b>24b</b> (29%)                          | <b>24c</b> (0%)                           |
| 17    | 10   | Me   | Bn  | Bn   | Ph   | н      | <b>25a</b> (0%)                                       | <b>25b</b> (52%)                          | <b>25c</b> (34%)                          |
| 18    | 11   | $4-NO_2C_6H_4$   | Bn  | Bn   | н  | Ph     | <b>26a</b> (49%)                                      | <b>26b</b> (25%)                          | <b>26c</b> (0%)                           |

#### Table 1. Formation of [1,5]-amino group migration products from envnamines.



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obtained as rearranged products (Entry 2). Corresponding acyclic amidine **10a** could not be traced from the reaction mixture. On the contrary, the substrate **11** ( $R^3 = H$ ,  $R^4 = Ph$ ) under similar reaction conditions provided **11a** (84%) as the major isomer (Entry 3). In this case, the isomer **11b** was isolated as the minor product, and formation of **11c** could not be detected. Similar reaction of **12** ( $R^3 = R^4 = Ph$ ) gave two products **12a** (48%) and **12b** (30%) (Entry 4). Apart from one dimensional <sup>1</sup>H and <sup>13</sup>C NMR studies, the molecular structures acyclic and cyclic amidines were further confirmed using either single crystal X-ray diffraction (*e.g.* Fig. 1 for **10b** and **11a**) or two-dimensional NMR spectroscopic (*e.g.* <sup>1</sup>H-<sup>1</sup>H COSY and <sup>1</sup>H-<sup>13</sup>C HETCOR for **9c, 21c** and **25c**) analysis (see ESI).

Quantum chemical calculations were done to validate the proposed mechanism and rationalize the product distribution. For a simplified version of enynamine III with  $R^1 = R^2 = Me$ ,  $R^3 = R^4 = H$  (Fig. S7), optimized structures of the cyclic intermediate IIIf (Fig. 2), and corresponding products IIIa, IIIb, and IIIc were obtained using the MP2 and DFT (B3LYP) methods with the 6-31G basis set (Fig. S7). Transition state structures IIIa', IIIb' and IIIc' corresponding to these products (Fig. 2), and the reaction barrier of each process were also obtained (Table S1). Small differences in the barrier heights between MP2 and DFT for a given product were observed. The MP2 method is known to be more reliable than DFT (B3LYP) for calculating reaction barriers,<sup>[13]</sup> but the former is computationally more expensive than the latter.



Fig. 1 Single crystal XRD structures of cyclic amidine 10b (A) and acyclic amidine 11a (B) presented as stick model.

Calculations on the enynamine II with  $R^1 = R^2 = Bn$ , and different combinations of H and Ph for R<sup>3</sup>, R<sup>4</sup> corresponding to substrates 9-12 were performed using DFT (B3LYP/6-31G). In these cases, obtaining the transition state structures using the MP2 method, which is significantly more computationally demanding than DFT, is impractical. The transition state structures and consequently barrier heights using DFT corresponding to products IIa and IIb were obtained (Table S2). The transition state for IIc proved elusive in spite of using several techniques and initial structures to search for it. Since the MP2 barriers are more reliable, we applied a correction to the DFT barriers to estimate them better using the results of the simplified enynamine III where both MP2 and DFT barriers were calculated. Assuming that there are similar differences in MP2 and DFT calculated barrier heights in the present case as in III, we applied the calculated difference in III as a correction to the present case (Table S2). The corrected barrier heights uniformly predict the trends in the observed product distribution, thus supporting the proposed mechanism. Inspection of the intermediate structures suggests that a phenyl group at the C<sub>2</sub>-center (*i.e.* for substrate **10**) increases the contributions of IIg and IIh through further delocalization of the negative charge. Enhancement of negative charge at C<sub>2</sub>and  $C_4$ -centers further favors the attacks on the R<sup>1</sup> group from these sites thereby, stabilizing corresponding transition states. In case of 11, the phenyl group at C<sub>3</sub>-center does not contribute significantly to the stabilization of the negative charge on the intermediate IIf. Therefore, IIf facilitates a C<sub>1</sub>-N bond cleavage leading to the formation of [1,5]-migration product. For **12**, the substitution of phenyl groups at C<sub>2</sub>-center again opens the scope of R<sup>1</sup> group migration.

Having established the effect of substituents at  $C_2$ - and  $C_3$ centers, we next evaluated the effects of different *p*substituted arylmethylene groups at  $R^1$  and  $R^2$  positions in controlling the product distribution. For enynamines **13-16**,

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designed with  $R^1 = R^2 = 4$ -CNBn, and varying  $R^3$  and  $R^4$  groups among H and Ph, the distributions of products were comparable to that observed with 9-12 (Entries 5-8). However, for substrates **17-20** with of  $R^1 = R^2 = 4$ -MeOBn, an interesting bias of product distribution towards cyclic systems was observed (Entries 9-12). For example, the substrate  $17 (R^3 = R^4)$ = H) upon reaction with TsN<sub>3</sub> gave only cyclic products 17b (63%) and 17c (12%). Therefore, the electron rich aromatic moiety of 4-MeOBn contributes significantly in shifting the course of rearrangement towards cyclic products due to the higher migratory aptitude of the 4-OMeBn group. Subsequently, the migratory aptitudes of Bn and 4-CNBn groups were compared by considering enynamines 21-24. Compound **21** ( $R^3 = R^4 = H$ ) when reacted with TsN<sub>3</sub>, compounds 21a-21c were formed in 14%, 39% and 23% yields, respectively (Entry 13). Structural analysis of 21b and 21c clearly indicated the migration of the benzyl group to either C2- or C4-position (see ESI). For the substrates 21-24, the trends in product distribution are similar to the corresponding substrates 9-12 as might have been expected (Entries 14, 16). Substrate **23** ( $R^3 = H$  and  $R^4 = Ph$ ) however, gave only **23a** (86%) as expected (Entry 15). As anticipated, the substitutions on the sulfonyl group did not have any contribution in changing the product distribution for substrates 10 and 11 (Entries 17, 18).

To get an experimental evidence of the zwitterionic cyclic intermediate, a 2-vinylpiperidine substituted enynamine **27** was designed. Reaction of **27** with TsN<sub>3</sub> gave bicycloamidine **28** as single product (Scheme 3A) and this structure was unambiguously confirmed by X-ray crystallographic data analysis (Fig. 3A). Therefore, the ketenimine intermediate **27a** forms the cyclic intermediate **27b** which subsequently undergoes aza-Claisen rearrangement<sup>14</sup> by an attack from C<sub>4</sub>-to C<sub>4</sub>-position followed by migration of the double bond from C<sub>4</sub>-=C<sub>3'</sub> to the C<sub>3'</sub>=C<sub>2'</sub>, and cleavage of the C<sub>2'</sub>--N bond (Scheme S9). Compound **11a**, a representative conjugated acyclic diene,



Fig. 2 Structures of the intermediate IIIf (A), and transition states IIIa', IIIb' and IIIc' (B-D) for the formation of IIIa, IIIb and IIIc calculated at the MP2/6-31G level.



Scheme 3. Synthesis of bridged bicycloamidine 28 (A). [2+2] Cycloaddition of 11a with in situ generated benzyne (B).



Fig. 3 Single crystal XRD structures of cyclic amidine 28 (A) and acyclic amidine 30 (B) presented as stick model.

when reacted with highly reactive benzyne, generated from **29**, compound **30** was formed as an exclusive [2+2] cycloaddition<sup>15</sup> product (Scheme 3B). Such observation is unique because a thermal [4+2] cycloaddition<sup>16</sup> is more common under the applied conditions. The structure of **30** was confirmed by X-ray crystallographic data analysis (Fig. 3B).

#### Conclusions

In conclusion, we have demonstrated the reaction of substituted enynamines with sulfonylazides under Cu(I) catalytic conditions providing acyclic and cyclic amidines. Structures of these products were confirmed by NMR and single crystal X-ray diffraction studies. The central carbon of in situ formed ketenimine was attacked by tethered amino group in the 6-exo-dig cyclization mode followed by breaking of C-N bond in the E1cB fashion gives acyclic amidines. On the other hand, the delocalization of negative charge on the cyclic zwitterionic intermediate facilitated the transfer of a nitrogen substituent to C3- and C5-centers leading to the formation of cyclic amidines. The mechanism and the product distribution of these reactions were evaluated by computational studies. Two interesting applications of the strategy were illustrated through the formation of a bridged bicycloamidine and a [2+2] cycloaddition reaction.

#### Experimental

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**General Information** All the chemicals were purchased from commercial sources and used as received unless stated otherwise. All reactions were carried out under the nitrogen atmosphere. Solvents: dichloromethane (DCM), ethyl acetate (EtOAc), methanol (MeOH) and petroleum ether, were distilled prior to thin layer and column chromatography. Column chromatographic purification was performed on silica gel (100–200 mesh). TLC was carried out with silica gel 60-F-254 plates.

The <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on 400 MHz Jeol 400 MHz (or 100 MHz for <sup>13</sup>C) spectrometers using either residual solvent signal as an internal reference or from internal tetramethylsilane on the  $\delta$  scale (CDCl<sub>3</sub>:  $\delta_{\rm H}$  7.24 ppm,  $\delta_{\rm C}$  77.0 ppm). The chemical shifts ( $\delta$ ) are reported in ppm and coupling constants (*J*) in Hz. The following abbreviations are used: m (multiplet), s (singlet), br s (broad singlet), d (doublet), t (triplet) dd (doublet of doublet), dt (doublet of triplet), q (quartet), sex (sextet), and sept (septet). High-resolution mass spectra were recorded using Micro Mass ESTOF MS spectrometer. (FT-IR) spectra were recorded using Bruker:  $\alpha$  ALPHA spectrophotometer (neat) and reported in cm<sup>-1</sup>. Crystal structures were recorded on a Bruker single crystal X-Ray diffractometer.

### General procedure for the copper catalyzed reaction of 1 with tosyl azide

The reaction conditions and results are shown in Tables 1. A typical procedure is given for the reaction of **1** with tosyl azide (Table 1, entry 1). To the solution of enynamine **1** (100 mg, 0.6 mmol) in chloroform (3.0 mL) was added tosyl azide (156 mg, 0.8 mmol), Et<sub>3</sub>N (108 mg, 1.0 mmol) followed by Cul (12 mg, 10 mol%) and stirred for 30 minutes at room temperature. The reaction was quenched by saturated NH<sub>4</sub>Cl and compound was extracted in chloroform (15 mL  $\times$  2). Solvent was evaporated and obtained crude product was purified by column chromatography (*Eluent*: 30% EtOAc in hexane) to afford the desired acyclic amidine product **1a** (142 mg, 67% yield) as a colorless semi-solid.

**1a:** IR (neat):  $v_{max}/cm^{-1}$  3743, 3678, 3648, 3619, 2967, 2921, 2861, 2319, 1740, 1693, 1643, 1595, 1519, 1476, 1444, 1346, 1273, 1190, 1141, 1114, 1087; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.71 (d, J = 8.2 Hz, 2H), 7.17 (d, J = 8.2 Hz, 2H), 6.30 (t, J = 11.4 Hz, 1H), 6.16 (d, J = 11.5 Hz, 1H), 5.96 (dt, J = 16.7 Hz, 10.7 Hz, 1H), 5.32 (d, J = 16.9 Hz, 1H), 5.21 (d, J = 10.0 Hz, 1H), 3.81 (t, J = 4.5 Hz, 2H), 3.68 (t, J = 5.0 Hz, 2H), 3.59 (t, J = 5.0 Hz, 2H), 3.46 (t, J = 5.0 Hz, 2H), 2.33(s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  163.1, 142.1, 140.2, 135.4, 131.3, 129.1, 126.9, 123.6, 120.3, 66.7, 66.3, 47.9, 44.7, 21.5; HRMS (ESI): Calc. for C<sub>16</sub>H<sub>21</sub>N<sub>2</sub>O<sub>3</sub>S [M+H]<sup>\*</sup>: 321.1273; Found: 321.1281.

**2a:** Isolated yield: 76%, 132 mg. Colorless semi-solid, IR (neat):  $v_{max}/cm^{-1}$  2967, 2920, 2860, 1721, 1597, 1521, 1443, 1344, 1344, 1275, 1225, 1150, 1113, 1088, 1029; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.83 (d, *J* = 8.2 Hz, 2H), 7.34 (dd, *J* = 6.6, 3.3 Hz, 3H), 7.32 – 7.22 (m, 7H), 6.71 (dd, *J* = 12.4, 0.8 Hz, 1H), 6.48 (d, *J* = 12.5 Hz, 1H), 5.52 (s, 1H), 5.44 (s, 1H), 3.46 (dd, *J* = 11.3, 6.4 Hz, 4H), 3.34 (s, 2H), 2.39 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 162.2, 143.8, 142.2, 140.6, 138.2, 136.4, 129.2, 128.5, 128.4, 126.7, 126.7, 120.7, 120.4, 66.3, 65.7,

 47.6, 44.1, 29.8, 21.6; HRMS (ESI): Calc. for C22H25N2O3StilMotHine

 397.1586; Found: 397.1584.

**3a:** Isolated yield: 70%, 122 mg. Colorless semi-solid, IR (neat):  $v_{max}/cm^{-1}$  2967, 2920, 2860, 1721, 1597, 1521, 1443, 1344, 1344, 1275, 1225, 1150, 1113, 1088, 1029; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.74 (d, *J* = 8.3 Hz, 2H), 7.44 – 7.35 (m, 5H), 7.18 (d, *J* = 7.9 Hz, 2H), 6.25 (s, 1H), 6.20 (dd, *J* = 17.3, 10.8 Hz, 1H), 5.30 (dt, *J* = 10.8, 1.2 Hz, 1H), 5.20 (d, *J* = 17.3 Hz, 1H), 3.96 – 3.89 (m, 2H), 3.80 – 3.75 (m, 2H), 3.72 – 3.67 (m, 2H), 3.62 – 3.56 (m, 2H), 2.38 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  163.3, 145.2, 142.1, 140.4, 137.7, 132.1, 129.1, 128.7, 128.6, 128.5, 126.9, 122.8, 119.7, 66.7, 66.3, 47.8, 44.8, 29.8, 21.5; HRMS (ESI): Calc. for C<sub>22</sub>H<sub>25</sub>N<sub>2</sub>O<sub>3</sub>S [M+H]<sup>+</sup>: 397.1586; Found: 397.1584.

**4a:** Isolated yield: 65%, 101 mg. Colorless semi-solid. IR (neat):  $v_{max}/cm^{-1}$  2967, 2920, 2860, 1721, 1597, 1521, 1443, 1344, 1344, 1275, 1225, 1150, 1113, 1088, 1029; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.72 (d, *J* = 8.3 Hz, 2H), 7.43 – 7.39 (m, 2H), 7.34 (m, 3H), 7.26 (dd, *J* = 6.7, 3.3 Hz, 2H), 7.23 – 7.19 (m, 3H), 7.17 (d, *J* = 8.0 Hz, 2H), 6.79 (s, 1H), 5.86 (d, *J* = 0.6 Hz, 1H), 5.34 (s, 1H), 3.58 (s, 4H), 3.40 (s, 2H), 3.25 (s, 2H), 2.40 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 162.49, 148.81, 143.98, 142.01, 140.37, 139.06, 137.34, 129.04, 128.76, 128.65, 128.54, 128.18, 128.06, 127.96, 127.41, 126.88, 126.56, 119.30, 119.03, 66.11, 65.68, 48.12, 44.53, 21.49; HRMS (ESI): Calc. for C<sub>28</sub>H<sub>29</sub>N<sub>2</sub>O<sub>3</sub>S [M+H]<sup>+</sup>: 473.1899; Found: 473.1894.

### General procedure for the copper catalyzed reaction of 9 with tosyl azide

The reaction conditions and results are shown in Tables 2. To the solution of enynamine **9** (100 mg, 0.3 mmol) in chloroform (3.0 mL) was added tosyl azide (90 mg, 0.4 mmol),  $Et_3N$  (50 mg, 0.5 mmol) followed by CuI (6.0 mg, 10 mol%) and stirred for 30 minutes at room temperature. The reaction was quenched by sat. NH<sub>4</sub>Cl and compound was extracted in chloroform (15 mL × 2). Solvent was evaporated and obtained crude product was purified by column chromatography (*Eluent*: 20% EtOAc in hexane) to afford amidine products **9a** (38 mg, 23% yield), **9b** (84 mg, 51% yield) and **9c** (26 mg, 16% yield) as colorless semi-solids.

**9a:** Isolated yield: 23%, 38 mg. Colorless semi-solid, IR (neat):  $v_{max}/cm^{-1}$  3744, 3027, 2925, 2864, 1739, 1692, 1639, 1533, 1484, 1383, 1342, 1273, 1139, 1083, 1026; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.76 (d, *J* = 8.2 Hz, 2H), 7.40 – 7.31 (m, 6H), 7.24 (m, 2H), 7.20 (d, *J* = 8.0 Hz, 2H), 7.12 (d, *J* = 6.5 Hz, 2H), 6.33 (m, 2H), 6.08 (m, 1H), 5.35 (dt, *J* = 16.7 Hz, 0.7 Hz, 1H), 5.22 (d, *J* = 10 Hz, 1H), 4.72 (br.s, 2H), 4.5 (s, 2H), 2.4 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  165.3, 141.8, 137.1, 135.2, 130.2, 129.1, 128.7, 128.1, 128.1, 126.5, 126.2, 121.9, 53.2, 48.2, 21.5; HRMS (ESI): Calc. for C<sub>26</sub>H<sub>27</sub>N<sub>2</sub>O<sub>2</sub>S [M+H]<sup>+</sup>: 431.1793.

**9b:** Isolated yield: 51%, 84 mg. Colorless semi-solid, IR (neat):  $v_{max}/cm^{-1}$  3742, 3029, 2924, 2863, 1740, 1691, 1638, 1531, 1481, 1382, 1340, 1270, 1137, 1081, 1025; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.85 (d, *J* = 8.2 Hz, 2H), 7.33 (m, 3H), 7.24 – 7.20 (m, 9H), 6.84 (dd, *J* = 7.7 Hz, 2.3 Hz, 2H), 6.58 (dd, *J* = 9.8 Hz, 4.16 Hz, 1H), 4.92 (d, *J* = 14.4 Hz, 1H), 4.51 (d, *J* = 14.4 Hz, 1H), 3.37 (dd, *J* = 12.8 Hz, 5.6 Hz, 1H), 3.17 (dd, *J* = 12.7 Hz, 5.8 Hz, 1H), 2.67 (m, 2H), 2.41 (dd, *J* = 11.4, 4.6 Hz, 1H), 2.40 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  157.9, 144.0, 142.1, 141.2, 137.7, 135.9, 129.2, 128.9, 128.9, 128.7, 128.6,

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128.1, 126.8, 126.4, 119.9, 52.8, 48.9, 37.3, 35.7, 21.6; HRMS (ESI): Calc. for  $C_{26}H_{27}N_2O_2S\left[M\!+\!H\right]^+\!\!:431.1793;$  Found: 431.1790.

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**9c:** Isolated yield: 16%, 26 mg. Colorless semi-solid. IR (neat):  $v_{max}/cm^{-1}$  3743, 3030, 2925, 2859, 1708, 1647, 1597, 1552, 1516, 1495, 1450, 1341, 1270, 1141, 1084, 1029; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.80 (d, *J* = 8.24 Hz, 2H), 7.28 – 7.21 (m, 10H), 7.09 (dd, *J* = 7.16 Hz, 1.6 Hz, 2H), 5.74 (m, 2H), 4.91 (d, *J* = 14.54 Hz, 1H), 4.74 (m, 1H), 4.26 (d, *J* = 14.6 Hz, 1H), 3.50 (m, 1H), 3.35 (dd, *J* = 13 Hz, 7.7 Hz, 1H), 3.06 (d, *J* = 17.5 Hz, 1H), 2.41 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 165.3, 141.7, 141.6, 137.0, 135.2, 130.1, 129.1, 128.7, 128.0, 128.0, 127.8, 126.6, 126.4, 126.1, 121.7, 53.1, 48.1, 40.4, 40.2, 21.4; HRMS (ESI): Calc. for C<sub>26</sub>H<sub>27</sub>N<sub>2</sub>O<sub>2</sub>S [M+H]<sup>+</sup>: 431.1793; Found: 431.1792.

**10b:** Isolated yield: 50%, 75 mg. Colorless semi-solid. IR (neat): v/cm<sup>-1</sup> 3008, 2931, 2838, 1621, 1577, 1534, 1514, 1467, 1441, 1381, 1275, 1246, 1174, 1140, 1113, 1084, 1032; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.79 (d, *J* = 8.3 Hz, 2H), 7.40 (d, *J* = 10.2 Hz, 1H), 7.28 – 7.08 (m, 11H), 7.04 (d, *J* = 6.9 Hz, 2H), 6.98 (dd, *J* = 8.1, 1.5 Hz, 2H), 6.79 (d, *J* = 10.2 Hz, 1H), 6.65 (d, *J* = 6.8 Hz, 2H), 4.81 (d, *J* = 14.6 Hz, 1H), 4.54 (d, *J* = 14.6 Hz, 1H), 3.58 (s, 2H), 3.04 (d, *J* = 13.5 Hz, 1H), 2.92 (d, *J* = 13.5 Hz, 1H), 2.42 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  157.8, 146.6, 142.0, 141.2, 140.8, 135.3, 135.2, 130.4, 129.2, 128.7, 128.4, 128.1, 127.8, 127.4, 127.0, 126.5, 126.4, 120.1, 56.2, 52.9, 44.7, 44.0, 21.6; HRMS (ESI): Calc. for C<sub>32</sub>H<sub>31</sub>N<sub>2</sub>O<sub>2</sub>S [M+H]<sup>+</sup>: 507.2106; Found: 507.2115.

**10c:** Isolated yield: 40%, 60 mg. Colorless semi-solid. IR (neat): v/cm<sup>-1</sup> 3018, 2935, 2831, 1624, 1575, 1535, 1511, 1471, 1444, 1382, 1271, 1244, 1184, 1148, 1121, 1094, 1038; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.88 (d, *J* = 8.3 Hz, 2H), 7.35 – 7.30 (m, 3H), 7.29 – 7.17 (m, 11H), 7.13 (m, 4H), 6.05 (dd, *J* = 5.6, 2.7 Hz, 1H), 4.94 (d, *J* = 14.5 Hz, 2H), 4.33 (d, *J* = 14.6 Hz, 1H), 3.77 (dd, *J* = 17.1, 1.7 Hz, 1H), 3.56 (dd, *J* = 13.1, 7.2 Hz, 1H), 3.29 (dd, *J* = 13.1, 3.5 Hz, 1H), 3.20 (dt, *J* = 17.1, 2.9 Hz, 1H), 2.43 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  165.0, 141.8, 141.6, 136.9, 135.0, 133.4, 130.2, 129.1, 128.7, 128.7, 128.2, 128.1, 128.0, 127.8, 126.7, 126.2, 124.9, 122.3, 53.3, 49.9, 40.8, 40.4, 21.4; HRMS (ESI): Calc. for C<sub>32</sub>H<sub>31</sub>N<sub>2</sub>O<sub>2</sub>S [M+H]<sup>+</sup>: 507.2106; Found: 507.2115.

**11a:** Isolated yield: 84%, 126 mg. Colorless semi-solid. IR (neat): v/cm<sup>-1</sup> 3740, 3025, 2922, 2862, 1733, 1690, 1635, 1530, 1480, 1381, 1341, 1271, 1135, 1081, 1022; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.77 (d, *J* = 8.2 Hz, 2H), 7.39 – 7.28 (m, 13H), 7.20 (d, *J* = 8.0 Hz, 2H), 7.16 (dd, *J* = 8.1 Hz, 1.7 Hz, 2H), 6.33 (s, 1H), 6.22 (dd, *J* = 17.2 Hz, 10.8 Hz, 1H), 5.21 (dt, *J* = 10.8 Hz, 1.2 Hz, 1H), 5.13 (dd, *J* = 17.2 Hz, 0.84 Hz, 1H), 4.59 (br.s, 2H), 2.39 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  164.8, 144.8, 142.0, 140.7, 137.9, 135.8, 135.2, 132.1, 129.2, 129.1, 128.9, 128.9, 128.7, 128.5, 128.4, 128.3, 128.1, 127.4, 126.9, 122.5, 120.5, 51.9, 50.0, 21.5; HRMS (ESI): Calc. for C<sub>32</sub>H<sub>31</sub>N<sub>2</sub>O<sub>2</sub>S [M+H]<sup>+</sup>: 507.2106; Found: 507.2106.

**11b:** Isolated yield: 5%, 8 mg. Colorless semi-solid. IR (neat):  $v_{max}/cm^{-1}$  3740, 3025, 2921, 2861, 1743, 1693, 1635, 1535, 1485, 1384, 1342, 1273, 1135, 1085, 1021; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.91 (d, *J* = 8.2 Hz, 2H), 7.60 (m, 3H), 7.43 (m, 3H), 7.32 (m, 3H), 7.26 (m, 5H), 7.11 (m, 3H), 6.53 (dd, *J* = 7.2 Hz, 3.7 Hz, 2H), 5.09 (d, *J* = 14.3 Hz, 1H), 4.39 (d, *J* = 14.3 Hz, 1H), 3.47 (dd, *J* = 13.2 Hz, 5.0 Hz, 1H), 3.23 (d, *J* = 13.2 Hz, 1H), 3.02 (m, 1H), 2.59 (dd, *J* = 14.0 Hz, 1H), 3.02 (m, 2H), 2.59 (dd, *J* = 14.0 Hz, 1H), 3.02 (m, 2H), 2.59 (dd, *J* = 14.0 Hz, 1H), 3.02 (m, 2H), 2.59 (dd, *J* = 14.0 Hz, 1H), 3.21 (m, 2H), 3.

3.16 Hz, 1H), 2.39 (s, 3H), 2.31 (dd, J = 14.0 Hz, 11.0 Hz, 11.0 Hz, 11.0 Hz, 11.0 MZ, 11.0 MHz, 11.

**12a:** Isolated yield: 48%, 68 mg. Colorless semi-solid. IR (neat): v/cm<sup>-1</sup> 3744, 3035, 2933, 2866, 1723, 1688, 1645, 1538, 1481, 1382, 1342, 1275, 1138, 1088, 1021; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.77 (d, *J* = 8.3 Hz, 2H), 7.40 – 7.34 (m, 6H), 7.31 (m, 6H), 7.19 (d, *J* = 8.0 Hz, 2H), 7.17 – 7.01 (m, 10H), 6.90 (s, 1H), 5.69 (d, *J* = 0.6 Hz, 1H), 5.37 (d, *J* = 0.5 Hz, 1H), 2.41 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>):  $\delta$  163.9, 148.0, 142.7, 141.9, 140.6, 138.8, 137.1, 135.5, 135.3, 129.1, 129.1, 129.0, 128.9, 128.6, 128.4, 128.3, 128.1, 127.8, 127.7, 127.2, 127.1, 126.8, 126.7, 119.6, 119.3, 52.2, 49.6, 21.5; HRMS (ESI): Calc. for C<sub>38</sub>H<sub>35</sub>N<sub>2</sub>O<sub>2</sub>S [M+H]<sup>+</sup>: 583.2419; Found: 583.2430.

**12c:** Isolated yield: 30%, 43 mg. Colorless semi-solid. IR (neat):  $v/cm^{-1}$  3738, 3032, 2925, 2867, 1722, 1687, 1638, 1531, 1483, 1382, 1344, 1277, 1133, 1077, 1032; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.73 (d, J = 8.0 Hz, 4H), 7.63 (s, 2H), 7.42 – 7.31 (m, 7H), 7.28 – 7.13 (m, 23H), 7.13 – 7.02 (m, 8H), 6.76 (d, J = 7.5 Hz, 4H), 6.61 (d, J = 7.4 Hz, 4H), 4.92 (d, J = 14.7 Hz, 2H), 4.06 (d, J = 14.7 Hz, 2H), 3.91 (d, J = 13.0 Hz, 2H), 3.46 (d, J = 14.3 Hz, 2H), 3.39 (d, J = 13.0 Hz, 2H), 3.46 (d, J = 14.3 Hz, 2H), 3.39 (d, J = 13.0 Hz, 2H), 3.46 (d, J = 14.3 Hz, 2H), 13.0 Hz, 2H), 3.26 (d, J = 14.3 Hz, 2H), 2.41 (s, 6H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>):  $\delta$  157.52, 156.04, 141.74, 141.46, 141.31, 137.96, 135.91, 134.88, 130.55, 129.29, 129.01, 128.73, 128.49, 128.46, 128.37, 128.02, 128.01, 127.45, 127.33, 127.14, 126.90, 126.37, 121.50, 57.74, 52.28, 47.61, 42.00, 21.46; HRMS (ESI): Calc. for C<sub>38</sub>H<sub>35</sub>N<sub>2</sub>O<sub>2</sub>S [M+H]<sup>+</sup>: 583.2419; Found: 583.2420.

**13a:** Isolated yield: 13%, 20 mg. Colorless semi-solid. IR (neat): v/cm<sup>-1</sup> 3022, 2918, 2218, 1713, 1625, 1545, 1488, 1441, 1412, 1338, 1276, 1225, 1178, 1141, 1083, 1021;<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.73 – 7.67 (m, 4H), 7.60 (d, *J* = 8.2 Hz, 2H), 7.30 (d, *J* = 8.2 Hz, 2H), 7.23 (d, *J* = 7.9 Hz, 4H), 6.43 (t, *J* = 11.4 Hz, 1H), 6.24 (d, *J* = 11.6 Hz, 1H), 6.05 (dt, *J* = 16.8, 10.6 Hz, 1H), 5.42 (d, *J* = 16.7 Hz, 1H), 5.31 (d, *J* = 10.1 Hz, 1H), 4.77 – 4.66 (m, 2H), 4.61 (s, 2H), 2.42 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  164.6, 142.6, 140.8, 140.1, 139.7, 136.1, 133.0, 132.6, 130.9, 129.1, 129.0, 127.8, 126.7, 124.4, 119.9, 52.2, 50.3, 29.7, 21.5; HRMS (ESI): Calc. for C<sub>28</sub>H<sub>25</sub>N<sub>4</sub>O<sub>2</sub>S [M+H]<sup>+</sup>: 481.1698; Found: 481.1702.

**13b:** Isolated yield: 37%, 57 mg. Colorless semi-solid. IR (neat): v/cm<sup>-1</sup> 3013, 2927, 2238, 1711, 1609, 1552, 1491, 1452, 1422, 1345, 1273, 1218, 1171, 1141, 1086, 1021; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.76 (d, *J* = 8.3 Hz, 2H), 7.59 (dd, *J* = 8.4, 3.4 Hz, 4H), 7.29 (dd, *J* = 13.3, 8.4 Hz, 6H), 7.10 (d, *J* = 8.3 Hz, 2H), 6.56 (dd, *J* = 10.0, 4.0 Hz, 1H), 4.77 (d, *J* = 15.0 Hz, 1H), 4.64 (d, *J* = 15.0 Hz, 1H), 3.44 (dd, *J* = 12.9, 5.5 Hz, 1H), 3.18 (dd, *J* = 12.8, 6.6 Hz, 1H), 2.86 – 2.75 (m, 2H), 2.63 (td, *J* = 10.6, 3.9 Hz, 1H), 2.44 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 157.70, 143.30, 142.87, 141.22, 140.65, 138.71, 132.67, 132.63, 129.67, 129.34, 128.90, 128.67, 126.34, 120.38, 118.74, 111.20, 55.88, 52.91, 50.07, 37.67, 35.18, 21.59; HRMS (ESI): Calc. for C<sub>28</sub>H<sub>25</sub>N<sub>4</sub>O<sub>2</sub>S [M+H]<sup>+</sup>: 481.1698; Found: 481.1699.

**13c:** Isolated yield: 23%, 35 mg. Colorless semi-solid. IR (neat): v/cm<sup>-1</sup> 3018, 2925, 2226, 1702, 1601, 1554, 1491, 1447, 1414, 1344, 1269, 1209, 1174, 1149, 1088, 1011; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.03 (s, 1H), 7.71 (d, *J* = 8.0 Hz, 2H), 7.57 (d, *J* = 7.9 Hz, 2H), 7.51 (d, *J* 

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= 8.0 Hz, 2H), 7.45 (d, J = 8.0 Hz, 2H), 7.23 (d, J = 7.9 Hz, 2H), 7.14 (d, J = 8.0 Hz, 2H), 5.83 (dd, J = 10.0, 4.1 Hz, 1H), 5.77 (dd, J = 8.6, 4.0 Hz, 1H), 4.94 (d, J = 15.1 Hz, 1H), 4.77 (s, 1H), 4.18 (d, J = 15.1 Hz, 1H), 3.64 (d, J = 15.7 Hz, 1H), 3.38 (m, 3H), 2.98 (s, 3H), 2.90 (s, 3H), 2.43 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  164.5, 142.7, 142.4, 140.8, 140.5, 132.5, 131.9, 130.7, 129.2, 128.2, 126.1, 126.0, 121.9, 118.8, 118.3, 111.7, 110.8, 53.1, 49.0, 40.5, 39.9, 21.5; HRMS (ESI): Calc. for C<sub>28</sub>H<sub>25</sub>N<sub>4</sub>O<sub>2</sub>S [M+H]<sup>+</sup>: 481.1698; Found: 481.1697.

**14b:** Isolated yield: 51%, 73 mg. Colorless semi-solid. IR (neat): v/cm<sup>-1</sup> 3019, 2926, 2228, 1703, 1605, 1555, 1498, 1446, 1415, 1343, 1270, 1215, 1173, 1140, 1083, 1019; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.67 (d, *J* = 8.2 Hz, 2H), 7.45 (d, *J* = 10.2 Hz, 1H), 7.39 (d, *J* = 8.1 Hz, 2H), 7.34 (d, *J* = 8.2 Hz, 2H), 7.29 – 7.15 (m, 6H), 6.92 (t, *J* = 7.4 Hz, 4H), 6.76 (d, *J* = 8.2 Hz, 2H), 6.69 (d, *J* = 10.3 Hz, 1H), 5.02 (d, *J* = 15.1 Hz, 1H), 4.21 (d, *J* = 15.1 Hz, 1H), 2.96 (d, *J* = 13.4 Hz, 1H), 2.40 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 157.5, 145.3, 142.4, 140.5, 140.5, 139.8, 132.3, 131.9, 130.9, 129.2, 128.9, 128.5, 127.9, 126.4, 126.3, 120.7, 118.4, 111.5, 111.2, 57.5, 52.7, 44.53, 44.1, 21.5; HRMS (ESI): Calc. for  $C_{34}H_{29}N_4O_2S$  [M+H]<sup>+</sup>: 557.2011; Found: 557.2012.

**14c:** Isolated yield: 32%, 46 mg. Colorless semi-solid. IR (neat): v/cm<sup>-1</sup> 3024, 2922, 2222, 1706, 1603, 1551, 1495, 1444, 1411, 1347, 1277, 1214, 1173, 1142, 1085, 1012; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.71 (d, *J* = 8.3 Hz, 2H), 7.52 (d, *J* = 8.2 Hz, 2H), 7.48 (d, *J* = 8.3 Hz, 2H), 7.39 (d, *J* = 8.2 Hz, 2H), 7.36 – 7.31 (m, 3H), 7.21 (d, *J* = 8.2 Hz, 2H), 7.17 – 7.10 (m, 4H), 6.02 (dd, *J* = 5.5, 2.6 Hz, 1H), 4.96 (d, *J* = 15.2 Hz, 2H), 4.24 (d, *J* = 15.1 Hz, 1H), 3.89 (dd, *J* = 17.1, 1.7 Hz, 1H), 3.55 (dt, *J* = 17.1, 2.8 Hz, 1H), 3.44 (qd, *J* = 13.0, 5.7 Hz, 2H), 2.41 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): 164.31, 142.84, 142.55, 140.76, 140.42, 136.10, 133.27, 132.62, 132.00, 130.86, 129.30, 129.09, 128.87, 128.33, 126.11, 124.81, 121.66, 118.85, 118.41, 111.92, 111.03, 53.29, 50.71, 40.74, 40.40, 21.58; HRMS (ESI): Calc. for C<sub>34</sub>H<sub>29</sub>N<sub>4</sub>O<sub>2</sub>S [M+H]<sup>+</sup>: 557.2011; Found: 557.2012.

**15a:** Isolated yield: 77%, 110 mg. Colorless semi-solid. IR (neat): v/cm<sup>-1</sup> 3022, 2924, 2226, 1703, 1605, 1555, 1498, 1444, 1414, 1343, 1275, 1210, 1170, 1145, 1085, 1018; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.69 (d, *J* = 8.1 Hz, 4H), 7.62 (d, *J* = 8.1 Hz, 2H), 7.41 – 7.36 (m, 4H), 7.35 (s, 1H), 7.31 (dd, *J* = 6.6, 3.0 Hz, 2H), 7.26 (d, *J* = 8.3 Hz, 2H), 7.21 (d, *J* = 8.2 Hz, 2H), 6.27 (s, 1H), 6.20 (dd, *J* = 17.2, 10.8 Hz, 1H), 5.29 (d, *J* = 10.8 Hz, 1H), 5.23 (d, *J* = 17.3 Hz, 1H), 4.68 (s, 3H), 2.42 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 164.86, 145.69, 142.59, 140.78, 140.31, 139.84, 137.36, 133.01, 132.56, 131.82, 129.16, 128.85, 128.51, 128.45, 127.83, 126.75, 123.41, 119.20, 118.31, 118.07, 112.54, 112.08, 52.41, 50.57, 21.50; HRMS (ESI): Calc. for C<sub>34</sub>H<sub>29</sub>N<sub>4</sub>O<sub>2</sub>S [M+H]<sup>+</sup>: 557.2011; Found: 557.2018.

**16a:** Isolated yield: 73%, 100 mg. Colorless semi-solid. IR (neat): v/cm<sup>-1</sup> 3025, 2921, 2227, 1708, 1601, 1554, 1496, 1432, 1411, 1348, 1275, 1213, 1173, 1141, 1088, 1011; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.69 (d, *J* = 8.3 Hz, 2H), 7.63 (d, *J* = 8.3 Hz, 2H), 7.55 (d, *J* = 8.4 Hz, 2H), 7.34 (m, 6H), 7.23 – 7.13 (m, 10H), 7.10 (d, *J* = 8.3 Hz, 2H), 6.86 (s, 1H), 5.79 (d, *J* = 0.5 Hz, 1H), 5.40 (s, 1H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 164.2, 149.1, 143.1, 142.6, 140.5, 140.4, 139.8, 138.3, 136.6, 132.9, 132.2, 129.2, 129.2, 128.8, 128.6, 127.5, 127.1, 126.7,

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### 126.4, 119.1, 118.5, 112.4, 111.8, 52.7, 50.2, 21.5; HRMS, (ESI): Galce for $C_{40}H_{34}N_4O_2S$ [M+H]<sup>+</sup>: 633.2324; Found: 639.23179.39/C7OB00499K

**16b:** Isolated yield: 18%, 25 mg. Colorless semi-solid. IR (neat): v/cm<sup>-1</sup> 3018, 2920, 2216, 1711, 1612, 1542, 1491, 1438, 1410, 1339, 1268, 1251, 1179, 1133, 1085, 1013;  $\delta$  7.65 (d, *J* = 5.8 Hz, 6H), 7.62 (s, 3H), 7.38 – 7.29 (m, 17H), 7.23 (d, *J* = 8.2 Hz, 8H), 7.22 – 7.16 (m, 19H), 7.14 (d, *J* = 7.9 Hz, 6H), 6.69 (dd, *J* = 8.1, 5.8 Hz, 12H), 5.13 (d, *J* = 15.1 Hz, 3H), 3.95 (d, *J* = 12.7 Hz, 3H), 3.81 (d, *J* = 15.1 Hz, 3H), 3.51 (d, *J* = 14.3 Hz, 3H), 3.37 (d, *J* = 12.8 Hz, 3H), 3.30 (d, *J* = 14.4 Hz, 3H), 2.40 (s, 10H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>):  $\delta$  142.30, 141.37, 140.92, 140.71, 140.43, 137.43, 132.12, 131.72, 131.22, 129.88, 129.14, 129.03, 128.86, 128.26, 128.13, 127.99, 127.01, 126.30, 121.59, 118.53, 118.43, 111.12, 111.00, 58.58, 52.07, 47.83, 42.07, 21.49; HRMS (ESI): Calc. for C<sub>40</sub>H<sub>34</sub>N<sub>4</sub>O<sub>2</sub>S [M+H]<sup>+</sup>: 633.2324; Found: 633.2321.

#### Crystallography

Crystal structure of compound 11a (CCDC 1525177): C<sub>26</sub>H<sub>26</sub>N<sub>2</sub>O<sub>2</sub>S; Compound **11a** was crystallized from slow evaporation of CH<sub>2</sub>Cl<sub>2</sub>/hexane at room temperature. A colorless cubic shaped crystal with approximate dimensions 0.185 x 0.161 x 0.058 mm gave an Orthorhombic with space group C2/c; a = 27.148(6) b = 6.2305(14) c = 27.401(6) Å,  $\alpha =$  $90^{\circ} \beta = 92.154^{\circ} \gamma = 90^{\circ}; V = 4631.5(18) \text{ Å}^{3}; T = 296 (2) \text{ K}; Z = 8;$  $\rho_{calc} = 1.235 \text{ Mgm}^{-3}; \vartheta_{max} = 26.37^{\circ}; MoKa\lambda = 0.71073 \text{ Å}.$  Finefocus sealed tube source with graphite monochromator. R =0.0514 (for 2576 reflection  $l > 2\sigma(l)$ ), wR = 0.1764 which was refined against |F2| and S = 0.941 for 281 parameters and 4739 unique reflections. The structure was obtained by direct methods using SHELXS-97. All non-hydrogen atoms were refined isotropically. The hydrogen atoms were fixed geometrically in the idealized position and refined in the fil cycle of refinement as riding over atoms to which they are bonded.  $\mu = 0.164 \text{ mm}^{-1}$ .

Crystal structure of compound 10b (CCDC 1525176): C32H30N2O2S; Compound 10b was crystallized from slow evaporation of CH<sub>2</sub>Cl<sub>2</sub>/hexane at room temperature. A colorless cubic shaped crystal with approximate dimensions 0.185 x 0.161 x 0.058 mm gave an Orthorhombic with space group P21/c; a = 8.0697(3) b = 32.4621(12) c = 9.8721(4) Å,  $\alpha =$  $90^{\circ} \beta = 90.5490^{\circ} \gamma = 90^{\circ}; V = 2585.97(17) \text{ Å}^{3}; T = 296 (2) \text{ K}; Z =$ 4;  $\rho_{calc} = 1.301 \text{ Mgm}^{-3}$ ;  $\vartheta_{max} = 68.384^{\circ}$ ;  $MoK\alpha\lambda = 0.71073 \text{ Å}$ . Fine-focus sealed tube source with graphite monochromator. R = 0.0434 (for 4143 reflection  $l > 2\sigma(l)$ ), wR = 0.1365 which was refined against |F2| and S = 1.101 for 335 parameters and 4726 unique reflections. The structure was obtained by direct methods using SHELXS-97. All non-hydrogen atoms were refined isotropically. The hydrogen atoms were fixed geometrically in the idealized position and refined in the final cycle of refinement as riding over atoms to which they are bonded.  $\mu$  = 1.364 mm<sup>-1</sup>.

**Crystal structure of compound 28 (CCDC 1525520):**  $C_{25}H_{28}N_2O_2S$ ; Compound **28** was crystallized from slow evaporation of CH<sub>2</sub>Cl<sub>2</sub>/hexane at room temperature. A colorless cubic shaped crystal with approximate dimensions 0.185 x 0.161 x 0.058 mm gave an Orthorhombic with space group  $P^{-1}$ ; a = 9.0423(4) b = 10.9846(5) c = 11.4374(5) Å,  $\alpha =$ 

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91.995° β = 100.164° γ = 103.818°; V = 2128.3(8) Å<sup>3</sup>; T = 0 (2) K; Z = 2;  $\rho_{calc}$  = 1.291 Mgm<sup>-3</sup>;  $\vartheta_{max}$  = 54.76°; *MoKaλ* = 0.71073 Å. Fine-focus sealed tube source with graphite monochromator. R = 0.0361 (for 4159 reflection *I*>2σ(*I*)), wR = 0.0820 which was refined against *IF2*I and S = 0.935 for 258 parameters and 4839 unique reflections. The structure was obtained by direct methods using SHELXS-97. All non-hydrogen atoms were refined isotropically. The hydrogen atoms were fixed geometrically in the idealized position and refined in the final cycle of refinement as riding over atoms to which they are bonded.  $\mu$  = 0.174 mm<sup>-1</sup>.

Crystal structure of compound 30 (CCDC 1525179): C38H34N2O2S; Compound 30 was crystallized from slow evaporation of CH<sub>2</sub>Cl<sub>2</sub>/hexane at room temperature. A colorless cubic shaped crystal with approximate dimensions 0.185 x 0.161 x 0.058 mm gave an Orthorhombic with space group *P21/c*; *a* = 12.375(4) *b* = 17.958(7) *c* = 14.179(6) Å, α =  $90^{\circ}$  β = 91.594° γ =  $90^{\circ}$ ; V = 3150(2) Å<sup>3</sup>; T = 296 (2) K; Z = 4;  $\rho_{calc}$ = 1.229 Mgm<sup>-3</sup>;  $\vartheta_{max}$ = 28.282°; *MoKαλ* = 0.71073 Å. Fine-focus sealed tube source with graphite monochromator. R = 0.0805(for 2133 reflection  $l>2\sigma(l)$ ), wR = 0.2471 which was refined against IF2I and S = 0.912 for 389 parameters and 5734 unique reflections. The structure was obtained by direct methods using SHELXS-97. All non-hydrogen atoms were refined isotropically. The hydrogen atoms were fixed geometrically in the idealized position and refined in the final cycle of refinement as riding over atoms to which they are bonded.  $\mu$  = 0.139 mm<sup>-1</sup>.

Crystal structure of compound 22b (CCDC 1525178): C33H29N3O2S; Compound 22b was crystallized from slow evaporation of CH<sub>2</sub>Cl<sub>2</sub>/hexane at room temperature. A colorless cubic shaped crystal with approximate dimensions 0.185 x 0.161 x 0.058 mm gave an Orthorhombic with space group  $P^{-1}$ ; a = 8.7353(11) b = 10.2521(13) c = 15.765(2) Å,  $\alpha =$  $101.925^{\circ} \beta = 98.258^{\circ} \gamma = 93.270^{\circ}; V = 1361.6(3) \text{ Å}^3; T = 296 (2)$ K; Z = 2;  $\rho_{calc}$  = 1.334 Mgm<sup>-3</sup>;  $\vartheta_{max}$  = 77.283°; *MoKαλ* = 0.71073 Fine-focus sealed tube source with graphite Å. monochromator. R = 0.0948 (for 4453 reflection  $l>2\sigma(l)$ ), wR =0.2636 which was refined against |F2| and S = 1.782 for 353 parameters and 5550 unique reflections. The structure was obtained by direct methods using SHELXS-97. All non-hydrogen atoms were refined isotropically. The hydrogen atoms were fixed geometrically in the idealized position and refined in the final cycle of refinement as riding over atoms to which they are bonded.  $\mu$  = 1.334 mm<sup>-1</sup>.

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