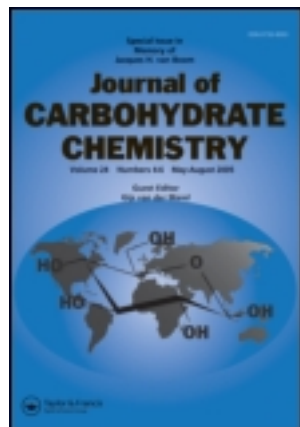


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# Improved Synthesis of 1-Benzenesulfinyl Piperidine and Analogs for the Activation of Thioglycosides in Conjunction with Trifluoromethanesulfonic Anhydride

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An improved protocol for the large-scale production of 1-benzenesulfinyl piperidine and other sulfinamides is described. It is demonstrated that 1-benzenesulfinyl pyrrolidine and *N,N*-diethyl benzenesulfinamide function analogously to 1-benzenesulfinyl piperidine in the trifluoromethanesulfonic anhydride-mediated activation of thioglycosides, and that their less crystalline nature enables them to be used at  $-78^{\circ}\text{C}$  as opposed to the  $-60^{\circ}\text{C}$  required to keep 1-benzenesulfinyl piperidine in solution. *N,N*-Dicyclohexyl benzenesulfinamide does not activate thioglycosides in combination with trifluoromethanesulfonic anhydride, which is attributed to its greater steric bulk.

**Keywords** Glycosylation, Sulfinamide, Thioglycoside, Trifluoromethanesulfonate

## INTRODUCTION

We recently introduced the combination of 1-benzenesulfinyl piperidine (**BSP, 1**) and trifluoromethanesulfonic anhydride ( $\text{Tf}_2\text{O}$ ) as a powerful means of activation of both armed and disarmed thioglycosides.<sup>[1,2]</sup> In designing the sulfinamide

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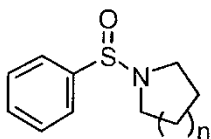
Dedicated with respect to the memory of Professor Jacques H. Van Boom.

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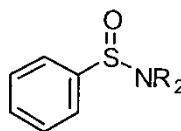
activating system we set several criteria, the most important of which was the need for a stable, crystalline reagent capable, in combination with triflic anhydride, of rapidly activating a wide range of armed and disarmed thioglycosides at low temperature.<sup>[1]</sup> **BSP** met these criteria admirably and activates most thioglycosides for coupling in a matter of minutes at  $-60^{\circ}\text{C}$  as demonstrated in a series of subsequent synthetic endeavors from this<sup>[3-7]</sup> and other laboratories.<sup>[8-13]</sup> The highly crystalline nature of **BSP**, however, limits its solubility below  $-60^{\circ}\text{C}$ , which explains the choice of this temperature for coupling reactions as opposed to the more convenient  $-78^{\circ}\text{C}$  achieved with dry ice/acetone cooling baths. This minor inconvenience and, more importantly, the recognition that liquid analogs of **BSP** might ultimately prove preferable in automated oligosaccharide synthesis applications requiring robotic dispensation prompted the synthesis and evaluation of other sulfinamides as described here.

We began with a refinement to our synthesis of **BSP**, the original version of which was based on the production of benzenesulfinyl chloride from diphenyl disulfide, sulfuryl chloride, and acetic anhydride,<sup>[1]</sup> which itself is a more convenient version of the protocol of Douglass and Norton employing sulfuryl chloride instead of chlorine gas.<sup>[14]</sup> Though adequate, this protocol requires careful control of temperature in the evaporation of the byproduct, acetyl chloride, if a high-quality sulfinyl chloride is to be obtained. The use of lower-quality sulfinyl chloride results in lower yields of **BSP** due to the formation of contaminants that hinder the direct isolation by crystallization. We have now found that a modification of a protocol described by Craig<sup>[15,16]</sup> affords higher-quality benzenesulfinyl chloride and thereby facilitates the isolation of the ensuing sulfinamides. In this method sodium benzenesulfinate is suspended in toluene in the presence of catalytic tetrabutylammonium bromide and is treated with thionyl chloride. Removal of the volatiles below  $25^{\circ}\text{C}$  then affords crude benzenesulfinyl chloride in admixture with sodium chloride. This mixture is used immediately in the derivatization of secondary amines. In this manner we were able to prepare high quality benzenesulfinyl piperidine in 86% yield on a 55 g scale. Similarly prepared on multigram scales were 1-benzenesulfinyl pyrrolidine (**2**),<sup>[17]</sup> *N,N*-diethyl benzenesulfinamide (**3**),<sup>[18]</sup> and *N,N*-dicyclohexyl benzenesulfinamide (**4**). Two of these sulfinamides (**2** and **4**) were crystalline, with melting points bracketing that of **BSP**, while a third (**3**) was a free-flowing, distillable liquid. All were found to be soluble in dichloromethane at  $-78^{\circ}\text{C}$ .



**1**:  $n = 2$ , **BSP**, mp  $83-84^{\circ}\text{C}$

**2**:  $n = 1$ , mp  $33-34^{\circ}\text{C}$

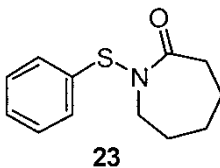


**3**:  $\text{R} = \text{Et}$ , liquid

**4**:  $\text{R} = \text{c-C}_6\text{H}_{11}$ , mp  $93^{\circ}\text{C}$

All three compounds (**2**–**4**) were assayed for their ability to activate to a standard mannosyl donor **6**,<sup>[19]</sup> in combination with triflic anhydride and the hindered base 2,4,6-tri-*tert*-butylpyrimidine (**TTBP**),<sup>[20]</sup> and to effect its coupling to 1,2;5,6-diacetone glucofuranose **5**. As reported in Table 1, with **2** and **3** the results were qualitatively the same as those previously obtained with **BSP** originally at  $-60^{\circ}\text{C}$ .<sup>[1]</sup> Reagent **4**, on the other hand, did not effect activation of **6** under these conditions. We attribute this failure to the greater steric bulk in the **4**- $\text{Tf}_2\text{O}$  adduct effectively preventing approach to the thioglycoside, and elected not to pursue this particular sulfinamide further. A number of other couplings were conducted by means of thioglycoside activation with **2** and/or **3** in combination with  $\text{Tf}_2\text{O}$ , each of which demonstrated comparable results to those obtained with **BSP** (Table 1).

Following the work of Gin on the activation of 1-hydroxy sugars (hemiacetals),<sup>[2,21]</sup> van Boom and coworkers introduced the combination of diphenyl sulfoxide and triflic anhydride for the activation of thioglycosides and found this reagent combination to be somewhat comparable to the **BSP**/ $\text{Tf}_2\text{O}$  couple.<sup>[2,10,22,23]</sup> While no actual comparisons between the **BSP** and  $\text{Ph}_2\text{SO}$  methods of thioglycoside activation have been carried out in this study, other work from our laboratory leads us to agree with the conclusion of van Boom and coworkers.<sup>[24–27]</sup> We anticipate that between **BSP**, its analogs introduced here, the recent modification of Wong (**23**),<sup>[28]</sup> and diphenyl sulfoxide, a reagent will be found to activate almost all classes of thioglycoside, in conjunction with trifluoromethanesulfonic anhydride, under milder conditions than have hitherto been possible.<sup>[29–31]</sup>

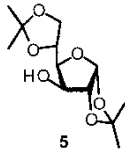
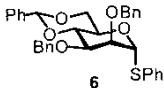
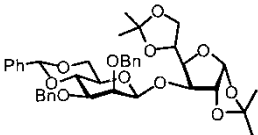
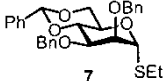
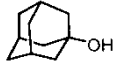
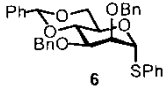
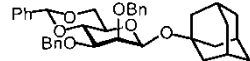
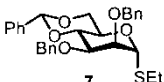
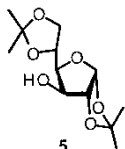
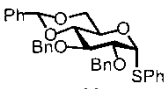
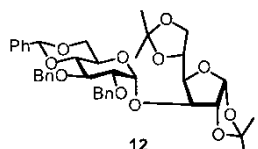


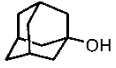
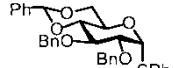
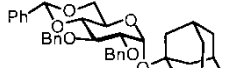
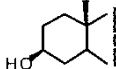
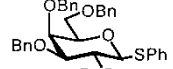
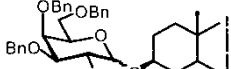
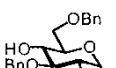
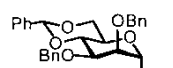
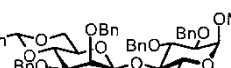
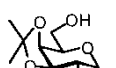
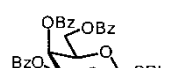
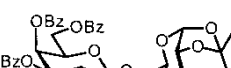
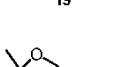
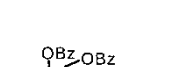
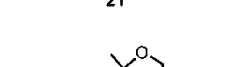
## EXPERIMENTAL

### General Methods

Unless otherwise stated  $^1\text{H}$  (300 MHz) and  $^{13}\text{C}$  (75 MHz) NMR spectra were recorded in  $\text{CDCl}_3$  solution. Optical rotations were recorded in  $\text{CHCl}_3$  solution, unless otherwise stated. All solvents were dried and distilled by standard protocols. All reactions were conducted under a blanket of dry nitrogen. All organic extracts were dried over sodium sulfate and concentrated under aspirator vacuum. Chromatographic purifications were carried out over silica gel. All glycosyl donors and acceptors were prepared by the literature methods or

**Table 1:** Glycosidic bond forming reactions.

Acceptor	Donor	Product	Activator					
			1 (BSP)		2		3	
			Yield (%)	$\alpha:\beta$ ratio	Yield (%)	$\alpha:\beta$ ratio	Yield (%)	$\alpha:\beta$ ratio
 5	 6	 8	77	1: > 9	90	1: > 9	87	1: > 9
	 7		—	—	91	1: > 9	89	1: > 9
 9	 6	 10	88	1: > 9	88	1: > 9	—	—
	 7		—	—	94	1: > 9	99	1: > 9
 5	 11	 12	74	>9:1	89	8.3:1	82	7.3:1

			72	>9:1	91 <sup>a</sup>	7.0:1	86 <sup>a</sup>	6.2:1
			78	1:1	87	1:3.0	87	1:3.9
			—	—	83	1: > 9	—	—
			78 <sup>b</sup>	1: > 9	85	1: > 9	80	1: > 9
			70 <sup>b</sup>	1: > 9	76 <sup>b</sup>	1: > 9	73 <sup>b</sup>	1: > 9

<sup>a</sup>Reaction was performed under dilute conditions (0.01 M donor in dichloromethane) as this led to enhanced  $\alpha$ : $\beta$  ratio.

<sup>b</sup>Reaction was conducted at  $-40^{\circ}\text{C}$  to achieve rapid activation of this disarmed donor.

were commercial samples. With the exception of the compounds reported below, all disaccharide physical characteristics and spectral data are consistent with the literature.<sup>[1]</sup>

**1-Benzenesulfinyl piperidine (1).** To an ice-cooled suspension of PhSO<sub>2</sub>Na (50.0 g, 0.31 mol) and Bu<sub>4</sub>NBr (4.90 g, 15.2 mmol) in dry toluene (200 mL) was added thionyl chloride (89.6 mL, 1.2 mol) dropwise over 30 min. After the addition, the reaction mixture was stirred at 0°C for 30 min, then warmed up to rt and stirred for a further 2 hr. The reaction mixture was then concentrated under reduced pressure keeping the temperature of the water bath below 25°C after which the residue was diluted with dry toluene (500 mL) and then treated with pyridine (24.6 mL, 0.31 mol) in one portion. The reaction mixture was cooled to 0°C, followed by dropwise addition of piperidine (60.8 mL, 0.61 mol) over 1 hr. After the addition, the reaction mixture was stirred in an ice bath for 2 hr, and then warmed up to rt and stirred for a further 1 hr. The reaction mixture was poured into a vigorously stirred mixture of ice and water (1 L) and NaHCO<sub>3</sub> (100 g, 1.2 mol). The organic layer was separated and washed with brine (500 mL), and then the aqueous layer was extracted with toluene (2 × 200 mL). The combined organic layer was dried and then concentrated. The residue was purified by recrystallization from ethanol to provide **1** as white crystals (55.0 g, 86%). mp: 83–84°C, lit. mp 83–84°C;<sup>[32]</sup> <sup>1</sup>H NMR δ: 7.62–7.65 (m, 2H), 7.43–7.50 (m, 3H), 3.05–3.12 (m, 2H), 2.88–2.95 (m, 2H), 1.47–1.62 (m, 6H); <sup>13</sup>C NMR δ: 143.3, 130.6, 128.7, 126.1, 46.9, 26.1, and 23.8.

**1-Benzenesulfinyl pyrrolidine (2).**<sup>[17]</sup> This compound was prepared analogously to **1** in 73% yield on a scale of 60 g. Colorless crystals were obtained by recrystallization from hexane and ethyl acetate. mp: 33–34°C, <sup>1</sup>H NMR δ: 7.66 (m, 2H), 7.49 (m, 3H), 3.33 (m, 2H), 3.00 (m, 2H), 1.83 (m, 4H). <sup>13</sup>C NMR δ: 130.6, 128.9, 125.9, 46.2, 26.1.

***N,N*-Diethyl benzenesulfinamide (3).**<sup>[18]</sup> This compound was prepared analogously to **1** in 78% on a 55 g scale. The crude reaction mixture was purified by distillation under reduced pressure (110°C, 0.1 mm Hg) to yield the sulfinamide as colorless oil. <sup>1</sup>H NMR δ: 7.61 (dd, *J* = 8.1, 2.4 Hz, 2H), 7.42 (m, 3H), 3.07 (q, *J* = 7.5 Hz, 4H) 1.11 (t, *J* = 7.2 Hz, 6H); <sup>13</sup>C NMR δ: 130.7, 128.8, 126.4, 42.1, 14.5.

***N,N*-Dicyclohexyl benzenesulfinamide (4).** This compound was prepared analogously to **1** in 78% on a 16 g scale. Recrystallization from ethyl acetate and hexane gave the sulfonamide as colorless crystals. mp 93°C; <sup>1</sup>H NMR δ: 7.67–7.64 (m, 2H), 7.49–7.41 (m, 3H), 3.11–3.02 (m, 2H), 2.08–2.03 (m, 2H), 1.81–1.44 (m, 12H), 1.30–1.04 (m, 6H); <sup>13</sup>C NMR δ: 145.0, 130.2, 128.6, 126.7, 55.5, 34.9, 26.4, 25.5. Anal. calcd. for C<sub>18</sub>H<sub>27</sub>NOS: C, 70.77; H, 8.91. Found: C, 70.81; H, 8.98.

**Typical glycosylation protocol.** A stirred solution of substrate, **BSP** (1.1 equiv.), TTBP (2.0 equiv.) and 3 Å sieves in CH<sub>2</sub>Cl<sub>2</sub> (0.03 M in substrate) was kept at –78°C for 15 min. Then Tf<sub>2</sub>O (1.2 equiv.) was added and after 5 min. the acceptor (1.5 equiv.) in CH<sub>2</sub>Cl<sub>2</sub> (2.0 M) was added. Stirring was continued at –78°C for 0.5 hr and then the reaction mixture was allowed to warm to rt over a period of 2 hr before it was filtered, washed with a saturated solution of NaHCO<sub>3</sub> and brine, dried, and concentrated. Chromatographic purification (eluting with mixtures of ethyl acetate in hexane) afforded the coupled products.

**Methyl 2,3,6-tri-*O*-benzyl-4-*O*-(2,3-di-*O*-dibenzyl-β-D-mannopyranosyl)-α-D-glucopyranoside (18).** This compound was prepared by the standard protocol and had the following characteristics:  $[\alpha]_D^{20}$  –17.1 (c, 0.75); <sup>1</sup>H NMR (500 MHz) δ: 7.50–7.48 (m, 2H), 7.42–7.17 (m, 28H), 5.52 (s, 1H), 5.08–5.04 (d, *J* = 11.0 Hz, 1H), 4.86–4.79 (m, 3H), 4.77 (s, 1H), 4.73 (s, 1H), 4.66–4.56 (m, 4H), 4.36 (s, 1H), 4.30–4.26 (d, *J* = 12.0 Hz, 1H), 4.14–4.02 (m, 2H), 3.93–3.83 (m, 2H), 3.64–3.58 (m, 2H), 3.54–3.47 (m, 3H), 3.44–3.43 (d, *J* = 3.0 Hz, 1H), 3.41 (s, 3H), 3.45–3.31 (dd, *J* = 3.3, 9.6 Hz, 1H), 3.09–3.01 (m, 1H); <sup>13</sup>C NMR (125 MHz) δ: 139.9, 139.1, 139.0, 138.8, 138.1, 138.0, 129.3, 129.0, 128.8, 128.7, 128.6, 128.5, 128.2, 128.1, 128.0, 127.9, 127.8, 126.7, 101.9, 101.7, 98.8, 80.7, 79.4, 79.1, 78.7, 78.1, 75.7, 75.4, 74.1, 72.5, 70.0, 69.0, 68.8, 67.7, 55.8; ESI-HRMS calcd. for C<sub>55</sub>H<sub>58</sub>O<sub>11</sub>Na [M + Na]<sup>+</sup>: 917.3877. Found 917.3871.

**1,2:3,4-Di-*O*-isopropylidene-3-*O*-(2,3,4,6-tetra-*O*-benzoyl-β-D-β-galactopyranosyl)-α-D-galactopyranose (21).** This compound was prepared by the standard protocol and had the following characteristics:  $[\alpha]_D^{26}$  +34.7° (c, 3.0), lit.<sup>[33]</sup>  $[\alpha]_D$  = +45° (CHCl<sub>3</sub>); <sup>1</sup>H NMR (500 MHz) δ: 8.09 (d, *J* = 7.6 Hz, 2H), 8.03 (d, *J* = 7.7 Hz, 2H), 7.98 (d, *J* = 7.7 Hz, 2H), 7.79 (d, *J* = 7.7 Hz, 2H), 7.23–7.49 (m, 12H), 6.00 (d, *J* = 2.8 Hz, 1H), 5.82 (t, *J* = 10.0 Hz, 1H), 5.62 (dd, *J* = 3.2, 10.4 Hz, 1H), 5.42 (d, *J* = 4.9 Hz, 1H), 5.03 (d, *J* = 8.0 Hz, 1H), 4.68 (dd, *J* = 6.6, 11.2 Hz, 1H), 4.41–4.45 (m, 2H), 4.35 (t, *J* = 6.5 Hz, 1H), 4.22 (d, *J* = 2.7 Hz, 1H), 4.05–4.12 (m, 2H), 3.90–3.93 (m, 2H), 1.40 (s, 3H), 1.24 (s, 3H), 1.22 (s, 3H), 1.20 (s, 3H); <sup>13</sup>C NMR (125 MHz) δ: 166.0, 165.6, 165.3, 133.6, 133.2, 133.0, 130.02, 129.98, 129.79, 129.4, 129.3, 129.0, 128.8, 128.6, 128.4, 128.3, 128.2, 124.8, 109.3, 108.4, 101.7, 96.2, 71.8, 71.3, 71.0, 70.5, 70.3, 69.7, 68.4, 68.2, 67.4, 62.4, 62.0, 25.9, 25.7, 24.8, 24.2.

**1,2:5,6-Di-*O*-isopropylidene-3-*O*-(2,3,4,6-tetra-*O*-benzoyl-β-D-β-galactopyranosyl)-α-D-glucofuranose (22).**<sup>[34]</sup> This compound was prepared by the standard protocol and had the following characteristics:  $[\alpha]_D^{24}$  +46.1 (c, 1.8); <sup>1</sup>H NMR (500 MHz) δ: 8.10–8.08 (d, *J* = 7.5 Hz, 2H), 8.04–8.02 (d, *J* = 8.0 Hz, 2H), 8.00–7.94 (d, *J* = 8.0 Hz, 2H), 7.79–7.77 (d, *J* = 7.5 Hz, 2H), 7.63–7.60 (t, *J* = 7.5 Hz, 1H), 7.58–7.55 (t, *J* = 7.5 Hz, 1H), 7.51–7.41 (m, 6H),



7.38–7.35 (t,  $J = 7.5$  Hz, 2H), 7.26–7.22 (t,  $J = 8.0$  Hz, 2H), 5.99 (d,  $J = 2.5$  Hz, 1H), 5.90 (d,  $J = 3.5$  Hz, 1H), 5.83–5.80 (t,  $J = 8.0$  Hz, 1H), 5.61–5.59 (dd,  $J = 3.5$ , 10.5 Hz, 1H), 4.93–4.92 (d,  $J = 8.0$  Hz, 1H), 4.69–4.65 (dd,  $J = 6.5$ , 11.0 Hz, 1H), 4.50–4.49 (d,  $J = 3.5$  Hz, 1H), 4.44–4.41 (dd,  $J = 6.5$ , 11.0 Hz, 1H), 4.34–4.31 (t,  $J = 6.5$  Hz, 1H), 4.17–4.14 (m, 2H), 4.08–4.07 (d,  $J = 3.5$  Hz, 1H), 3.76–3.71 (m, 2H), 1.40 (s, 3H), 1.29 (s, 3H), 1.13 (s, 3H), 1.09 (s, 3H);  $^{13}\text{C}$  NMR (125 MHz)  $\delta$ : 166.5, 166.0, 165.9, 165.5, 134.0, 133.7, 133.6, 130.5, 130.3, 130.2, 129.85, 129.81, 129.4, 129.2, 129.0, 128.9, 128.7, 128.6, 112.6, 106.8, 102.6, 101.3, 84.4, 79.6, 75.3, 72.0, 71.7, 70.9, 70.1, 68.5, 62.4, 27.5, 26.9, 24.1, 23.1; ESI-HRMS calcd. for  $\text{C}_{46}\text{H}_{46}\text{O}_{15}\text{Na}$   $[\text{M} + \text{Na}]^+$ : 861.2734. Found 861.2704.

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