Dalton Transactions

COMMUNICATION



View Article Online



Cite this: DOI: 10.1039/c5dt04088d

Received 19th October 2015, Accepted 23rd November 2015 DOI: 10.1039/c5dt04088d

www.rsc.org/dalton

New mechanistic insight into intramolecular arene hydroxylation initiated by $(\mu-1,2-peroxo)diiron(m)$ complexes with dinucleating ligands[†]

Mio Sekino,^a Hideki Furutachi,^{*a} Kyosuke Tasaki,^a Takanao Ishikawa,^a Shigeki Mori,^a Shuhei Fujinami,^a Shigehisa Akine,^a Yoko Sakata,^a Takashi Nomura,^b Takashi Ogura,^b Teizo Kitagawa^b and Masatatsu Suzuki^c

 $(\mu-1,2-\text{Peroxo})\text{diiron}(\mu)$ complexes (2-R) with dinucleating ligands (R-L) generated from the reaction of bis(μ -hydroxo)diiron(μ) complexes [Fe₂(R-L)(OH)₂]²⁺ (1-R) with dioxygen in acetone at -20 °C provide a diiron-centred electrophilic oxidant, presumably diiron(ν)-oxo species, which is involved in aromatic ligand hydroxylation.

The Fe/O2- and Fe2/O2-mediated arene hydroxylation is of current interest for understanding the reaction mechanism of dioxygen activating non-heme mononuclear and dinuclear iron enzymes such as tetrahydropterin-dependent aromatic amino acid hydroxylase¹ and toluene/o-xylene monooxygenase (ToMO).² In these enzymes, an iron(iv)-oxo and a (μ -peroxo)diiron(m) species have been spectroscopically identified,^{1,2} which are responsible for arene hydroxylation. To date, intramolecular aromatic ligand hydroxylation³⁻⁶ and intermolecular hydroxylation of aromatic compounds such as benzoic acid,⁷ benzene,8 and anthracene9 by synthetic mononuclear iron model complexes with various oxidants such as O_2 , H_2O_2 , m-CPBA, t-BuOOH, and PhIO have been extensively studied. In some of these, the iron(IV)-oxo species have been identified as the active oxidant in intramolecular aromatic ligand hydroxylation.^{5b,6b,9} Some synthetic ToMO model complexes have also been reported so far;¹⁰ yet only for one, a (µ-peroxo)diiron(III) complex $[Fe_2(L^{Ph4})(O_2)(Ph_3CCO_2)]^{2+}$ with a dinucleating ligand (L^{Ph4}) = N, N, N', N'-tetrakis(1-methyl-2-phenyl-4-imidazolyl)methyl-1,3-amino-2-propanolate) has been shown to be directly involved in intramolecular aromatic ligand hydroxylation,^{10d} although the electronic effect of substituents on the aromatic

ligand hydroxylation was not investigated. To gain further insights into the arene hydroxylation performed by (μ -peroxo)-diiron(III) species, further (μ -peroxo)diiron(III) model complexes are demanded, which exhibit the monooxygenase activity like ToMO.

In this study, we have applied the dinucleating ligands (R-L = 1.3-bis[bis(6-methyl-2-pyridylmethyl)aminomethyl]-5-Rbenzene; R = tBu, H, and NO₂) to the diiron complex system in order to examine the Fe2/O2-mediated arene hydroxylation (Scheme 1), since R-L provides crucial active oxygen species that are capable of hydroxylating the xylyl linker of R-L via an electrophilic aromatic substitution mechanism, as found for dicopper¹¹ and dinickel¹² complexes with R-L. Herein, we report intramolecular arene hydroxylation together with intermolecular acetone oxidation initiated by $(\mu-1,2-peroxo)$ diiron(m) complexes (2-R) with dinucleating ligands R-L (Scheme 1), generated from the reaction of $bis(\mu-hydroxo)diiron(\pi)$ complexes $[Fe_2(R-L)(OH)_2]^{2+}$ (1-R) with dioxygen in acetone at -20 °C. We found decisive mechanistic evidence that a diiron-centred electrophilic oxidant, presumably diiron(IV)oxo species, is involved in the aromatic ligand hydroxylation, whose species is capable of exchanging with exogenous water.

The diiron(II) complex $[Fe_2(H-L)(OH)_2]^{2+}$ (1-H) was obtained by treatment of $[Fe(CH_3CN)_4(OTf)_2]$ with H-L in the presence



Scheme 1 Intramolecular arene hydroxylation and intermolecular acetone oxidation initiated by $(\mu$ -1,2-peroxo)diiron(III) complexes (2-R) with dinucleating ligands (R-L).

^aDepartment of Chemistry, Division of Material Sciences, Graduate School of Natural Science and Technology, Kanazawa University, Kakuma-machi, Kanazawa 920-1192, Japan. E-mail: h-furutachi@se.kanazawa-u.ac.jp

^bPicobiology Institute, Graduate School of Life Science, University of Hyogo, Ako-gun, Hyogo 678-1297, Japan

^cDepartment of Chemistry and Biochemistry, Graduate Engineering,

Kyushu University, 744 Moto-oka, Nishi-ku, Fukuoka 819-0395, Japan

[†]Electronic supplementary information (ESI) available: Experimental details of synthesis and structural, spectroscopic, and kinetic data. CCDC 1430633 and 1430634. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c5dt04088d

of triethylamine and H₂O in dry-THF under an N₂ atmosphere. **1**-H has a bis(μ -hydroxo)diiron(π) core bridged by exogenous hydroxo groups (Fig. 1 and S1[†]) as found for a closely related dicopper(π) complex [Cu₂(H-L)(OH)₂]²⁺.¹¹

Reaction of a pale green acetone solution of 1-H with O₂ at -20 °C generated a dark green peroxo species, 2-H, that exhibits a broad absorption band at 500-800 nm (Fig. 2(A), (a): 530 nm, $\varepsilon = \sim 710 \text{ M}^{-1} \text{ cm}^{-1}$; 700 nm, $\varepsilon = \sim 645 \text{ M}^{-1} \text{ cm}^{-1}$) that can be assigned to the CT transition from O_2^{2-} to the Fe(III) center, as found in related (µ-peroxo)diiron(III) complexes.^{13,14} Similar UV-vis spectral features were also observed for 2-tBu and 2-NO₂ (Fig. S3[†]). Unlike in acetone, no peroxo complexes 2-R were generated in acetonitrile and dichloromethane. The resonance Raman (rR) spectrum of 2-H prepared by ¹⁶O₂ showed a band at 845 cm⁻¹, which shifted to 800 cm⁻¹ when $^{18}O_2$ was used (Fig. 2(B) and S4[†]). The band at 845 cm⁻¹ can be assigned to ν (O–O) vibration and is similar to those of well characterized (µ-oxo)(µ-peroxo)diiron(m) complexes $(835-874 \text{ cm}^{-1})$,¹⁴ suggesting that 2-H has a similar (µ-oxo)- $(\mu$ -peroxo)diiron(III) core. In contrast to ν (O–O) vibration, ν (Fe–O) and ν (Fe–O–Fe) vibrations of 2-H were not observed (Fig. S4[†]), which might be due to their poor signal-to-noise ratios.



Fig. 1 An ORTEP view of $[Fe_2(OH)_2(H-L)]^{2+}$ (1-H). Hydrogen atoms are omitted for clarity.



View Article Online Dalton Transactions

The yields of hydroxylated ligands (R-L-OH) were determined by ¹H NMR spectroscopy (Fig. S6–8, details in the ESI[†]). The hydroxylation yields of the xylyl linker depend on the electrondonating ability of the substituent (R-) of R-L and decreased as follows: tBu-L-OH (\sim 32%) > H-L-OH (\sim 26%) > NO₂-L-OH $(\sim 8\%)$ (Fig. S9(a)[†]). A similar observation was also made for a series of $(\mu-\eta^2;\eta^2-\text{peroxo})\text{dicopper}(\Pi)$ complexes $[Cu_2(R-L) (O_2)^{2+}$ (Cu-R)¹¹ and bis(µ-oxo)dinickel(III) complexes [Ni₂(R-L)- $(O)_2$ ²⁺ (Ni-R),¹² suggesting that aromatic hydroxylation of R-L in the present diiron system also proceeds via an electrophilic aromatic substitution mechanism. Thus, the substituentdependent intramolecular arene hydroxylation observed upon decay of (µ-peroxo)diiron(m) species 2-R is closely relevant to arene hydroxylation catalyzed by ToMO, where a (µ-peroxo)diiron(m) intermediate is directly involved in the arene hydroxylation.² However, the hydroxylation yield of 2-R is lower than those of Cu-R (98–72%), Ni-R (88–30%), and $[Fe_2(L^{Ph4}) (O_2)(Ph_3CCO_2)$ ²⁺ (~90%), suggesting that some other side reaction(s) takes place simultaneously in the present diiron system.

2-H is unstable at -20 °C and decomposes to give a red

solution that showed an absorption band at 550 nm (Fig. 2(A),

(b)). The red solution slowly converted into a purple solution

at ambient temperature that afforded purple crystals of

The ESI-TOF/MS of a red solution (Fig. 2(A) and (b)) obtained by decomposition of 2-H at -20 °C showed a signal at m/z 379.1 (Fig. S10(a)†), which can be assigned as [Fe₂(H-L-O)(O)(CH₃CO₂)]²⁺ having a hydroxylated ligand (H-L-O⁻) and an acetate. The signal at m/z 379.1 in acetone shifted to m/z 380.6 when 2-H decomposed in d₆-acetone (Fig. S10(b)†), indicating that acetone solvent is indeed the source of acetate in this species. The acetic acid was also identified by GC-MS analysis (Fig. S11†). Thus, these results clearly demonstrate intra-



Fig. 2 (A) Electronic spectra of **2**-H (a) and its decomposition species (b) in acetone at -20 °C. (B) Resonance Raman spectra of **2**-H generated from the reaction of **1**-H with ${}^{16}O_2$ (c) and ${}^{18}O_2$ (d) in d₆-acetone at -40 °C with a 647.1 nm laser excitation. The asterisk denotes the solvent band.



Fig. 3 An ORTEP view of $[Fe_2(OH)_2{H(H-L-O)}_2]^{4+}$ (3-H). Hydrogen atoms are omitted for clarity.

molecular arene hydroxylation and intermolecular acetone oxidation initiated by 2-H. Unlike the hydroxylation yields of R-L-H, the electron-donating ability of the substituent (R-) of R-L has only a small influence on the oxidation yields of acetone, where ~40% yield of acetic acid was obtained in each ligand system (Fig. S9(b), details in the ESI[†]). Moreover, the yields of R-L-OH and acetic acid observed upon decay of 2-R are not influenced by the reaction conditions under both O_2 and N_2 , indicating that there is no participation of radical species in the present reaction system. It has been reported that acetic acid is formed by decomposition of Fe- and Cu-HPP (HPP = 2-hydroxy-2-peroxy-propane) species generated by the reaction of mononuclear Fe and Cu complexes with H₂O₂ in acetone solvent.15,16 Recently, an electrophilic aromatic ligand hydroxylation by a mononuclear Cu-HPP complex has also been reported by Itoh et al., where an acetate species was not formed and only acetone is regenerated during the course of reaction.¹⁵ Thus, a diiron-HPP species, probably produced by a nucleophilic attack of 2-R to acetone, seems to be a candidate of the precursor to give acetic acid, but not to give rise to ligand hydroxylation in the present system (Scheme 2).

To gain further insight into the oxidation mechanism, kinetic studies of 2-R were carried out by monitoring the absorbance change at 700 nm in acetone at -20 °C. Decomposition of 2-R under the conditions obeys first-order kinetics (Fig. S12[†]). It should be noted that the decomposition rates of 2-R are independent of the electron-donating ability of the substituent (R-) of R-L (Fig. S12(d)[†]). In contrast to those of substituent-dependent decay observed for the $(\mu - \eta^2; \eta^2 - \text{peroxo})$ dicopper(II) complex $[Cu_2(R-L)(O_2)]^{2+}$ (Cu-R)¹¹ and the bis- $(\mu$ -oxo)dinickel(III) complex $[Ni_2(R-L)(O)_2]^{2+}$ (Ni-R),¹² this observation indicates that the rate determining step in the decay of 2-R does not involve an electrophilic attack by the peroxo species 2-R to the xylyl linker, but involves unimolecular reaction such as conversion of 2-R into a diiron(IV)-oxo species. Activation parameters obtained from the temperature dependence of decay of 2-R are ΔH^{\ddagger} = 76–80 kJ mol⁻¹ and ΔS^{\ddagger} = –8 to –26 J $\rm K^{-1}\ mol^{-1}$ (Fig. S13, Tables S3 and S4†). The observed small negative activation entropy implies that a bimolecular reaction such as a nucleophilic attack of 2-R to acetone affording a diiron-HPP species (Scheme 2, step b) is also partially involved in the rate determining step of the decay of 2-R as well as O–O bond cleavage of 2-R (Scheme 2, step a), since a large negative activation entropy has been observed for the reaction system in which bimolecular reaction is dominantly included in the rate-limiting step.¹⁷

It should be noted that isotope labeling experiments by using ¹⁸O₂ and H₂¹⁸O strongly support involvement of the diiron(IV)-oxo species in the aromatic ligand hydroxylation. Surprisingly, decomposition of 2-H generated by ¹⁸O₂ in acetone resulted in only ~33% incorporation of ¹⁸O into H-L-OH, which is confirmed by ESI-TOF/MS (Fig. S14(b)[†]). This observation is in contrast to that of $[Fe_2(L^{Ph4})(O_2)]$ - (Ph_3CCO_2) ²⁺,^{10d} where incorporation of the ¹⁸O label mainly originates from ¹⁸O₂. The complementary experiment with 2-H generated by ${}^{16}O_2$ in the presence of $H_2{}^{18}O$ (1000 equiv.) corroborates this result (Fig. S14(c)[†]). Thus, incorporation of the ¹⁸O label from H₂¹⁸O into H-L-OH requires the participation of an oxidant capable of exchanging with exogenous water, such as a diiron(iv)-oxo species, that carries out the ligand hydroxylation (Scheme 2, step c). A similar labeling result has been reported for the formation of the DOPA208 residue from the F208Y mutant of the ribonucleotide reductase (RNR) R2 protein¹⁸ as well as a few synthetic iron model complexes.^{3a,4c,10c} An electrophilic character of this species is supported by the observed substituent-dependent hydroxylation yields of the xylyl linker. Thus, observation of ¹⁸O incorporation from H₂¹⁸O as well as substituent-dependent hydroxylation yields of the xylyl linker in this system are indirect but compelling evidence for high-valent diiron(IV)-oxo species bearing an electrophilic character, which is well-known for synthetic mononuclear iron(IV)-oxo model complexes.³⁻⁹

Based on the kinetics data together with the experimental results mentioned above, we thus propose a plausible mechanism for intramolecular arene hydroxylation and intermolecular acetone oxidation initiated by 2-R (Scheme 2), which involves rate determining O–O bond cleavage of 2-R into diiron(IV)-oxo species (step a) and nucleophilic attack of 2-R to acetone into diiron-HPP species (step b) prior to rapid decay. Conversion of (μ -peroxo)diiron(III) species to high-valent diiron(IV)-oxo species has also been reported for a few synthetic diiron model complexes.^{14a,d,f,19} The generated diiron(IV)-oxo species is the



Scheme 2 Proposed mechanism for intramolecular arene hydroxylation and intermolecular acetone oxidation initiated by 2-R.

electrophilic active oxidant capable of exchanging with exogenous water prior to an electrophilic attack on the aromatic ring, to provide the partially ¹⁸O labeled phenol (R-L-OH), whose hydroxylation yields are substituent-dependent (step c). It has been reported that a well-defined mononuclear non-heme iron(n)-oxo complex shows a poor nucleophilic oxidative reaction toward 2-phenylpropionaldehyde compared with (hydro)peroxo-iron(III) complexes.^{17a,20} Thus, the diiron(IV)-oxo species derived from 2-H seems not to give rise to acetone oxidation via the formation of diiron-HPP species (step e), although the formation of diiron-HPP species derived from the diiron(IV)oxo species may not be ruled out. Decay of diiron-HPP species is independent of the electron-donating ability of the substituent (R-) of R-L to give acetic acid and the yield of acetic acid is expected to be nearly constant in each ligand system (step d), which is consistent with the observed yield of acetic acid (~40%).

In summary, we have succeeded in intramolecular arene hydroxylation and intermolecular acetone oxidation initiated by the (μ -1,2-peroxo)diiron(III) complexes (2-R) with the dinucleating ligands (R-L) for the first time. The former reaction mimics the function of ToMO² and involves the diiron(IV)-oxo species as the electrophilic active oxidant capable of exchanging with exogenous water, whereas the latter one is closely relevant to a nucleophilic attack of peroxo species to the C=O group proposed for aldehyde deformylating oxygenase (ADO).^{17*a*,21} The observed oxidation reactions initiated by 2-R provide a chemical insight into the nature of (μ -1,2-peroxo)diiron(III) species for oxidation reactivity and O–O bond activation, which are found for dioxygen-activating non-heme diiron proteins, although further comprehensive functional model studies are needed.

This work was supported by Grants-in-Aid for Scientific Research from the Ministry of Education, Culture, Sports, Science and Technology, Japan and Kanazawa University SAKI-GAKE Project.

Notes and references

- (a) P. F. Fitzpatrick, *Biochemistry*, 2003, 42, 14083;
 (b) M. Costas, M. P. Mehn, M. P. Jensen and L. Que Jr., *Chem. Rev.*, 2004, 104, 939; (c) A. R. McDonald and L. Que Jr., *Coord. Chem. Rev.*, 2013, 257, 414; (d) M. Puri and L. Que Jr., *Acc. Chem. Res.*, 2015, 48, 2443.
- 2 (a) M. H. Sazinsky and S. J. Lippard, Acc. Chem. Res., 2006,
 39, 558; (b) L. J. Murray, S. G. Naik, D. O. Ortillo, R. García-Serres, J. K. Lee, B. H. Huynh and S. J. Lippard, J. Am. Chem. Soc., 2007, 129, 14500; (c) L. J. Bailey and B. G. Fox, Biochemistry, 2009, 48, 8932.
- 3 (a) S. J. Lange, H. Miyake and L. Que Jr., *J. Am. Chem. Soc.*, 1999, **121**, 6330; (b) M. P. Jensene, S. J. Lange, M. P. Mehn, E. L. Que and L. Que Jr., *J. Am. Chem. Soc.*, 2003, **125**, 2113.
- 4 (a) E. L. Hegg, R. Y. N. Ho and L. Que Jr., *J. Am. Chem. Soc.*, 1999, **121**, 1972; (b) M. P. Mehn, K. Fujisawa, E. L. Hegg and L. Que Jr., *J. Am. Chem. Soc.*, 2003, **125**, 7828;

(c) S. Chatterjee and T. K. Paine, *Angew. Chem., Int. Ed.*, 2015, 54, 9338.

- 5 (a) S. Sahu, L. R. Widger, M. G. Quesne, S. P. de Visser, H. Matsumura, P. Moënne-Loccoz, M. A. Siegle and D. P. Goldberg, J. Am. Chem. Soc., 2013, 135, 10590; (b) S. Sahu, M. G. Quesne, C. G. Davies, M. Dürr, I. Ivanovíc-Burmazovíc, M. A. Siegle, G. N. L. Jameson, S. P. de Visser and D. P. Goldberg, J. Am. Chem. Soc., 2014, 136, 13542.
- 6 (a) W. H. Harman and C. J. Chang, J. Am. Chem. Soc., 2007, 129, 15128; (b) J. P. Bigi, W. H. Harman, B. Lassalle-Kaiser, D. M. Robles, T. A. Stich, J. Yano, R. D. Britt and C. J. Chang, J. Am. Chem. Soc., 2012, 134, 1536.
- 7 (a) S. Taktak, M. Flook, B. M. Foxman, L. Que Jr. and E. V. Rybak-Akimova, *Chem. Commun.*, 2005, 5301;
 (b) N. Y. Oh, M. S. Seo, M. H. Lim, M. B. Consugar, M. J. Oark, J.-U. Rohde, J. Han, K. M. Kim, J. Kim, L. Que Jr. and W. Nam, *Chem. Commun.*, 2005, 5644;
 (c) O. V. Makhlynets, P. Das, S. Taktak, M. Flook, R. Mas-Ballesté, E. V. Rybak-Akimova and L. Que Jr., *Chem. Eur. J.*, 2009, **15**, 13171; (d) O. V. Makhlynets and E. V. Rybak-Akimova, *Chem. Eur. J.*, 2010, **16**, 13995.
- 8 (a) V. Balland, D. Mathieu, Y. M. N. Pons, J. F. Bartoli, F. Banse, P. Battioni, J. J. Girerd and D. Mansuy, *J. Mol. Catal. A: Chem.*, 2004, 215, 81; (b) A. Thibon, J.-F. Bartoli, R. Guillot, J. Sainton, M. Martinho, D. Mansuy and F. Banse, *J. Mol. Catal. A: Chem.*, 2008, 287, 115; (c) A. Thibon, V. Jollet, C. Ribal, K. Sénéchal-David, L. Billon, A. B. Sorokin and F. Banse, *Chem. – Eur. J.*, 2012, 18, 2715.
- 9 S. P. de Visser, K. Oh, A.-R. Han and W. Nam, *Inorg. Chem.*, 2007, **46**, 4632.
- 10 (a) S. Ménage, J.-B. Galey, J. Dumats, G. Hussler, M. Seité, I. G. Luneau, G. Chottard and M. Fontecave, J. Am. Chem. Soc., 1998, 120, 13370; (b) H. Furutachi, M. Murayama, A. Shiohara, S. Yamazaki, S. Fujinami, A. Uehara, M. Suzuki, S. Ogo, Y. Watanabe and Y. Maeda, Chem. Commun., 2003, 1900; (c) F. Avenier, L. Dubois and J.-M. Latour, New J. Chem., 2004, 28, 782; (d) M. Yamashita, H. Furutachi, T. Tosha, S. Fujinami, W. Saito, Y. Maeda, K. Takahashi, K. Tanaka, T. Kitagawa and M. Suzuki, J. Am. Chem. Soc., 2007, 129, 2.
- 11 T. Matsumoto, H. Furutachi, M. Kobino, M. Tomii, S. Nagatomo, T. Tosha, T. Osako, S. Fujinami, S. Itoh, T. Kitagawa and M. Suzuki, *J. Am. Chem. Soc.*, 2006, **128**, 3874.
- 12 K. Honda, J. Cho, T. Matsumoto, J. Roh, H. Furutachi, T. Tosha, M. Kubo, S. Fujinami, T. Ogura, T. Kitagawa and M. Suzuki, *Angew. Chem., Int. Ed.*, 2009, 48, 3304.
- 13 (a) L. Que Jr., J. Chem. Soc., Dalton Trans., 1997, 3933;
 (b) M. Suzuki, H. Furutachi and H. Okawa, Coord. Chem. Rev., 2000, 200-202, 105; (c) E. Y. Tshuva and S. J. Lippard, Chem. Rev., 2004, 104, 987.
- 14 (a) Y. H. Dong, Y. Zhang, L. J. Shu, E. C. Wilkinson, L. Que Jr., K. Kauffmann and E. Münck, *J. Am. Chem. Soc.*, 1997, 119, 12683; (b) X. Zhang, H. Furutachi, S. Fujinami,

S. Nagatomo, Y. Maeda, Y. Watanabe, T. Kitagawa and M. Suzuki, J. Am. Chem. Soc., 2005, **127**, 826; (c) S. V. Kryatov, S. Taktak, I. V. Korendovych, E. V. Rybak-Akimova, J. Kaizer, S. Torelli, X. Shan, S. Mandal, V. L. MacMurdo, A. M. i. Payeras and L. Que Jr., *Inorg. Chem.*, 2005, **44**, 85; (d) M. A. Cranswick, K. K. Meier, X. Shan, A. Stubna, J. Kaizer, M. P. Mehn, E. Münck and L. Que Jr., *Inorg. Chem.*, 2012, **51**, 10417; (e) J. S. Pap, M. A. Cranswick, É. Balogh-Hergovich, G. Baráth, M. Giorgi, G. T. Rohde, J. Kaizer, G. Speier and L. Que Jr., *Eur. J. Inorg. Chem.*, 2013, 3858; (f) M. Kodera, T. Tsuji, T. Yasunaga, Y. Kawahara, T. Hirano, Y. Hitomi, T. Nomura, T. Ogura, Y. Kobayashi, P. K. Sajith, Y. Shiota and K. Yoshizawa, *Chem. Sci.*, 2014, **5**, 2282.

- 15 (a) A. Kunishita, J. D. Scanlon, H. Ishimaru, K. Honda, T. Ogura, M. Suzuki, C. J. Cramer and S. Itoh, *Inorg. Chem.*, 2008, 47, 8222; (b) A. Kunishita, J. Teraoka, J. D. Scanlon, T. Matsumoto, M. Suzuki, C. J. Cramer and S. Itoh, *J. Am. Chem. Soc.*, 2007, **129**, 7248.
- 16 A. Mairata -i Payeras, R. Y. N. Ho, M. Fujita and L. Que Jr., *Chem. –Eur. J.*, 2004, **10**, 4944.
- 17 (a) J. Annaraj, Y. Suh, M. S. Seo, S. O. Kim and W. Nam, *Chem. Commun.*, 2005, 4529; (b) M. S. Seo, J. Y. Kim, J. Annaraji, Y. Kim, Y.-M. Lee, S.-J. Kim, J. Kim and W. Nam, *Angew. Chem., Int. Ed.*, 2007, 46, 377; (c) Y. Jo,

J. Annaraji, M. S. Seo, Y.-M. Lee, S. Y. Kim, J. Cho and W. Nam, *J. Inorg. Biochem.*, 2008, **102**, 2155; (*d*) J. Cho, R. Sarangi, H. Y. Kang, J. Y. Lee, M. Kubo, T. Ogura, E. I. Solomon and W. Nam, *J. Am. Chem. Soc.*, 2010, **132**, 16977; (*e*) J. Nakazawa, S. Terada, M. Yamada and S. Hikichi, *J. Am. Chem. Soc.*, 2013, **135**, 6010; (*f*) S. Kundu, J. V. K. Thompson, L. Q. Shen, M. R. Mills, E. L. Bominaar, A. D. Ryabov and T. J. Collins, *Chem. – Eur. J.*, 2015, **21**, 1803.

- 18 J. Ling, M. Shalin, B.-M. Siöberg, T. M. Loeher and J. Sanders-Loehr, J. Biol. Chem., 1994, 269, 5595.
- (*a*) G. Xue, A. T. Fiedler, M. Martinho, E. Münck and L. Que Jr., *Proc. Natl. Acad. Sci. U. S. A.*, 2008, **105**, 20615;
 (*b*) M. Kodera, Y. Kawahara, Y. Hitomi, T. Nomura, T. Ogura and Y. Kobayashi, *J. Am. Chem. Soc.*, 2012, **134**, 13236.
- 20 J. Cho, S. Jeon, S. A. Wilson, L. V. Liu, E. A. Kang, J. J. Braymer, M. H. Lim, B. Hedman, K. O. Hodgson, J. S. Valentine, E. I. Solomon and W. Nam, *Nature*, 2011, 478, 502.
- 21 (a) A. Shokri and L. Que Jr., J. Am. Chem. Soc., 2015, 137, 7686; (b) M. E. Pandelia, N. Li, H. Nørgaard, D. M. Warui, L. J. Rajakovich, W.-C. Chang, S. J. Booker, C. Krebs and J. M. Bollinger Jr., J. Am. Chem. Soc., 2013, 135, 15801; (c) E. N. G. Marsh and M. W. Waugh, ACS Catal., 2013, 3, 2515.