

## Two new oleanane-type triterpene saponins from the leaves of *Schefflera sessiliflora* De P. V.

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### ABSTRACT

From the leaves of *Schefflera sessiliflora* De P. V., two new oleanane-type triterpene saponins, named scheffleraside A (**1**), scheffleraside B (**2**); together with two known saponins, chikusetsusaponin IVa (**3**), 3-O-[ $\alpha$ -L-rhamnopyranosyl-(1  $\rightarrow$  3)]- $\beta$ -D-glucuronopyranosyl hederagenin (**4**) were isolated by various chromatography methods. Its chemical structure was elucidated by IR, UV, HR-ESI-MS, NMR 1D and 2D experiments and comparison of their NMR data with previous reported data.

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## 1. Introduction

*Schefflera* is a large genus of the Araliaceae family with over 650 species (Hebbbar and Nalini, 2014), which 56 species are found in Vietnam (La et al., 2013). Some species in the *Schefflera* genus, such as *Schefflera actinophylla*, *Schefflera arboricola*, *Schefflera heptaphylla*, *Schefflera impressa*, *Schefflera leucantha*, and *Schefflerakwangsiensis* have been chemically investigated, which led to the isolation of triterpenoid (Wanas et al., 2010; Zhao et al., 2010; Wu et al., 2013; Srivastava, 1992; Pancharoen et al., 1994; Wang et al., 2014), sterol (Hansen and Boll, 1986), polyacetylene (Hansen and Boll, 1986), phenolic (Li et al., 2005), anthraquinone (Srivastava, 1992), trisaccharide (Tran et al., 1991). *Schefflera sessiliflora* De P. V. (Araliaceae) is a new species in Viet Nam, it were discovered and indentified in 2004 (Nguyen et al., 2004). Pharmacological of extracts from this species possessed antistress, enhanced physical strength (Nguyen et al., 2004; Huynh et al., 2005); antioxidant activities in DPPH and MDA test (Vo et al., 2008); the androgenic effects (Tran et al., 2012). However, only two sapogenins: oleanolic acid and hederagenin from the hydrolyzed product of total saponins were isolated (Vo et al., 2003, 2004). This paper reports

the isolation and structure elucidation of two new triterpene saponins (**1**, **2**) and 2 known saponins (**3**, **4**) from *S. sessiliflora* De P. V. growing in Vietnam.

## 2. Results and discussion

The 75% MeOH extract from the dried leaves of *S. sessiliflora* De P. V. was subjected to column chromatography over silica gel normal-phase and reversed-phase RP-18 to give two new triterpene saponins named scheffleraside A (3-O-[ $\alpha$ -L-rhamnopyranosyl-(1  $\rightarrow$  3)]- $\beta$ -D-glucuronopyranosyl oleanolic acid 28-O-[ $\alpha$ -L-rhamnopyranosyl-(1  $\rightarrow$  4)]- $\beta$ -D-glucopyranosyl ester) (**1**) and scheffleraside B (3-O-[ $\alpha$ -L-rhamnopyranosyl-(1  $\rightarrow$  3)]- $\beta$ -D-(6'-O-methyl)glucuronopyranosyl oleanolic acid 28-O-[ $\alpha$ -L-rhamnopyranosyl-(1  $\rightarrow$  4)]- $\beta$ -D-glucopyranosyl ester) (**2**), and two known saponins: 3-O- $\beta$ -D-glucuronopyranosyl oleanolic acid 28-O- $\beta$ -D-glucopyranosyl ester (chikusetsusaponin IVa) (**4**) (Mizui et al., 1990), 3-O-[ $\alpha$ -L-rhamnopyranosyl-(1  $\rightarrow$  3)]- $\beta$ -D-glucuronopyranosyl hederagenin (**5**) (El and Morta, 1998).

Compound (**1**) was obtained as a white amorphous powder. The molecular formula was established as C<sub>54</sub>H<sub>86</sub>O<sub>22</sub> by HR-ESI-MS data ([M-H]<sup>-</sup> *m/z*: 1085.5521, calcd. 1085.5527 and [M+Na]<sup>+</sup> *m/z* 1109.5533, calcd. 1109.5503). The IR spectrum of **1** showed absorptions of hydroxyl (3418 cm<sup>-1</sup>) and carbonyl (1726 cm<sup>-1</sup>) groups. The <sup>13</sup>C NMR and DEPT spectrum (Table 1), showed **1** has fifty four carbons including: one carbonyl carbon, two olefinic

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**Table 1**<sup>1</sup>H (500 Hz) and <sup>13</sup>C (125 Hz) NMR spectral data for compounds **1** and **2** in pyridine-*d*<sub>5</sub>.

| Aglycone | <b>1</b>         |                | <b>2</b>            |                | Sugar  | <b>1</b>       |                | <b>2</b>       |                |
|----------|------------------|----------------|---------------------|----------------|--------|----------------|----------------|----------------|----------------|
|          | δ <sub>H</sub>   | δ <sub>C</sub> | δ <sub>H</sub>      | δ <sub>C</sub> |        | δ <sub>H</sub> | δ <sub>C</sub> | δ <sub>H</sub> | δ <sub>C</sub> |
| 1        | 0.77, 1.30       | 38.6           | 0.78, 1.28          | 38.6           | GlcA   |                |                |                |                |
| 2        | 1.74, 2.24       | 26.4           | 1.72, 2.04          | 26.4           | 1'     | 4.78 d (6.5)   | 106.8          | 4.86 d (7.5)   | 107.0          |
| 3        | 3.26 br d (7.5)  | 89.1           | 3.26 dd (4.0, 11.5) | 89.3           | 2'     | 4.00           | 75.9           | 4.03           | 75.7           |
| 4        | –                | 39.4           | –                   | 39.5           | 3'     | 4.37           | 82.1           | 4.39           | 82.0           |
| 5        | 0.69             | 55.6           | 0.72                | 55.7           | 4'     | 4.28           | 72.0           | 4.39           | 71.4           |
| 6        | 1.21, 1.40       | 18.4           | 1.20, 1.41          | 18.4           | 5'     | 4.37           | 76.9           | 4.48           | 77.1           |
| 7        | 1.30, 1.40       | 33.1           | 1.28, 1.41          | 33.1           | 6'     |                | <sup>a</sup>   |                | 170.7          |
| 8        | –                | 39.8           | –                   | 39.9           | 6'-OMe |                |                | 3.76 s         | 52.1           |
| 9        | 1.53             | 47.9           | 1.56                | 48.0           | Rha I  |                |                |                |                |
| 10       | –                | 36.8           | –                   | 36.9           | 1''    | 6.28 br s      | 102.5          | 6.28 br s      | 102.9          |
| 11       | 1.81             | 23.7           | 1.83                | 23.7           | 2''    | 4.71 br s      | 72.5           | 4.73 d (1.5)   | 72.5           |
| 12       | 5.38 br s        | 122.9          | 5.38 br s           | 122.9          | 3''    | 4.56 d (8.0)   | 72.6           | 4.54           | 72.7           |
| 13       | –                | 144.1          | –                   | 144.1          | 4''    | 4.28           | 74.1           | 4.32           | 74.1           |
| 14       | –                | 42.0           | –                   | 42.1           | 5''    | 5.06           | 69.6           | 5.06           | 69.8           |
| 15       | 1.14, 1.30       | 28.2           | 1.14, 1.26          | 28.2           | 6''    | 1.68 d (5.0)   | 18.6           | 1.68 d (6.5)   | 18.6           |
| 16       | 1.89, 2.05       | 23.3           | 1.90, 2.04          | 23.3           | Glc-28 |                |                |                |                |
| 17       | –                | 47.0           | –                   | 47.0           | 1'''   | 6.18 d (8.0)   | 95.3           | 6.20 d (8.0)   | 95.4           |
| 18       | 3.14 br d (10.0) | 41.7           | 3.16 br d (13.0)    | 41.7           | 2'''   | 4.10           | 74.2           | 4.12           | 74.2           |
| 19       | 1.21, 1.74       | 46.1           | 1.20, 1.72          | 46.1           | 3'''   | 4.18           | 77.0           | 4.20           | 77.1           |
| 20       | –                | 30.7           | –                   | 30.7           | 4'''   | 4.47           | 77.8           | 4.52           | 77.9           |
| 21       | 1.03, 1.30       | 34.0           | 1.04, 1.28          | 34.0           | 5'''   | 3.76 d (9.5)   | 77.8           | 3.78 d (9.5)   | 77.9           |
| 22       | 1.68, 1.74       | 32.5           | 1.72, 1.72          | 32.5           | 6'''   | 4.04, 4.18     | 60.9           | 4.05, 4.20     | 61.0           |
| 23       | 1.21 s           | 28.1           | 1.21 s              | 28.0           | Rha II |                |                |                |                |
| 24       | 0.92 s           | 16.9           | 0.91 s              | 16.8           | 1'''   | 5.85 br s      | 102.6          | 5.87 br s      | 102.7          |
| 25       | 0.77 s           | 15.4           | 0.78 s              | 15.5           | 2'''   | 4.63 br s      | 72.5           | 4.64 d (1.5)   | 72.5           |
| 26       | 1.03 s           | 17.4           | 1.04 s              | 17.4           | 3'''   | 4.47           | 72.7           | 4.52           | 72.7           |
| 27       | 1.24 s           | 26.0           | 1.24 s              | 26.1           | 4'''   | 4.28           | 73.8           | 4.32           | 73.9           |
| 28       | –                | 176.4          | –                   | 176.4          | 5'''   | 4.86           | 70.3           | 4.90           | 70.4           |
| 29       | 0.89 s           | 33.1           | 0.90 s              | 33.1           | 6'''   | 1.65 d (6.0)   | 18.4           | 1.66 d (6.5)   | 18.5           |
| 30       | 0.86 s           | 23.6           | 0.87 s              | 23.6           |        |                |                |                |                |

<sup>a</sup> Not determined.

carbons, four anomeric carbons, seventeen oxygenated methine carbons, one oxygenated methylene carbon, six quaternary carbons, three methine carbons, ten methylene carbons, and nine methyl carbons. The presence of seven tertiary methyl groups, two olefinic carbons at δ<sub>C</sub> 144.0 (C-13) and 122.8 (C-12), and one carbonyl carbon 176.4 (C-28) which indicated an olean-12-en-28-oic acid as an aglycone (Tran et al., 1991). Thus **1** was an oleanane-type triterpenoid saponin bearing four sugar (six carbons) units. The <sup>1</sup>H NMR data (Table 1) also indicated aglycone of **1** was oleanolic acid with: one olefinic proton at δ<sub>H</sub> 5.38 (*br s*, H-12); one oxygenated methine proton at δ<sub>H</sub> 3.26 (*br d*, 7.5, H-3); one methine proton at δ<sub>H</sub> 3.14 (*br d*, 10.0, H-18) and seven tertiary methyl groups at δ<sub>H</sub> 0.76–1.24 (Maeda et al., 1994). Moreover, four anomeric protons at δ<sub>H</sub> 4.78 (*d*, 6.5, H-1'); 6.18 (*d*, 8.0, H-1''); 6.28 (*br s*, H-1'''); 5.85 (*br s* H-1''') to four anomeric carbons at δ<sub>C</sub> 106.8 (C-1'), 95.3 (C-1''), 102.5 (C-1'''), 102.6 (C-1''') were assigned to β-D-glucuronic (GlcA), β-D-glucose (Glc) and two α-L-rhamnose (Rha) units, respectively. It was completely appropriated with HR-ESI-MS data, a peak [M+2Na–H]<sup>+</sup> *m/z* 1131.5492 was indicated the presence of a sugar acid (Madl et al., 2006). The COSY, HSQC and NOESY spectrum, allowed analysis of their spin systems and assigned of their proton resonances to determine clearly every sugar unit. Beside, through acid hydrolysis followed by co-TLC in comparison with standard sugars the identification of the sugars was determined to be D for Glc, GlcA and L for Rha (see Section 3). The HMBC spectrum (Fig. 1), showed correlations between anomeric proton at δ<sub>H</sub> 4.78 (*d*, 6.5, H-1') of GlcA and carbons at δ<sub>C</sub> 89.1 (C-3) of aglycone; between anomeric proton at δ<sub>H</sub> 6.28 (*br s*, H-1'') of Rha I unit and carbons at δ<sub>C</sub> 82.1 (C-3') of GlcA; between oxygenated methine proton at δ<sub>H</sub> 4.37 (*m*, H-3') and carbon acetal at δ<sub>C</sub> 102.5 (C-1''). On the other hands, anomeric proton at δ<sub>H</sub> 6.18 (*d*, 8.0, H-1''') of Glc correlated with carbonyl carbon at δ<sub>C</sub> 176.4 (C-28); anomeric proton at δ<sub>H</sub> 5.85 (*br s*, H-1''') of Rha II correlated

with carbon at δ<sub>C</sub> 77.8 (C-4'') of Glc; between oxygenated methine proton at δ<sub>H</sub> 4.47 (*m*, H-4'') and carbon acetal at δ<sub>C</sub> 102.6 (C-1'''). These connectivities were also confirmed by correlations observed in the NOESY spectrum between H-3 (δ<sub>H</sub> 3.26) of aglycone and H-1' (δ<sub>H</sub> 4.78) of GlcA; between H-1'' (δ<sub>H</sub> 6.28) of RhaI and H-3' (δ<sub>H</sub> 4.37) of GlcA. Based on data of HR-ESI-MS, 1D, 2D-NMR and compared with previous published data (Tapondjou et al., 2006; Wang et al., 2014), the structure of **1** was identified as 3-O-[α-L-rhamnopyranosyl-(1 → 3)]-β-D-glucuronopyranosyl oleanolic acid 28-O-[α-L-rhamnopyranosyl-(1 → 4)]-β-D-glucopyranosyl ester, and named scheffleraside A.

Compound (**2**) was obtained as a white amorphous powder. The molecular formula was established as C<sub>55</sub>H<sub>88</sub>O<sub>22</sub> by HR-ESI-MS data ([M+Na]<sup>+</sup> *m/z* 1123.5633, calcd. 1123.5659). The <sup>1</sup>H and <sup>13</sup>C NMR data (Table 1) demonstrated that **2** has the same aglycone and sugar chains as **1**, except for presence of an oxygenated methyl group [δ<sub>H</sub> 3.76 (*s*); δ<sub>C</sub> 52.1]. Further, the HMBC spectrum (Fig. 1) showed correlation between this oxygenated methyl at δ<sub>H</sub> 3.76(*s*) and carbon carbonyl of glucuronic unit at δ<sub>C</sub> 170.7. Based on data of HR-ESI-MS, 1D, 2D-NMR and compared with previous published data (Tapondjou et al., 2006; Wang et al., 2014), the structure of **2** was identified as 3-O-[α-L-rhamnopyranosyl-(1 → 3)]-β-D-(6'-O-methyl)glucuronopyranosyl oleanolic acid 28-O-[α-L-rhamnopyranosyl-(1 → 4)]-β-D-glucopyranosyl ester, and named scheffleraside B.

Some species in the *Schefflera* genus, such as *S. actinophylla* (Wanas et al., 2010), *S. arboricola* (Zhao et al., 2010), *S. impressa* (Srivastava, 1992), *S. leucantha* (Pancharoen et al., 1994) are reported as a rich source of saponins which possessed lupane-type and/or ursane-type aglycones. Whereas, the genins of *S. heptaphylla* (Maeda et al., 1994), *S. kwangsiensis* (Wang et al., 2014) as well as *S. sessiliflora* were oleanolic acid and hederagenin. However, all compounds were isolated for the first time from the genus *Schefflera*.

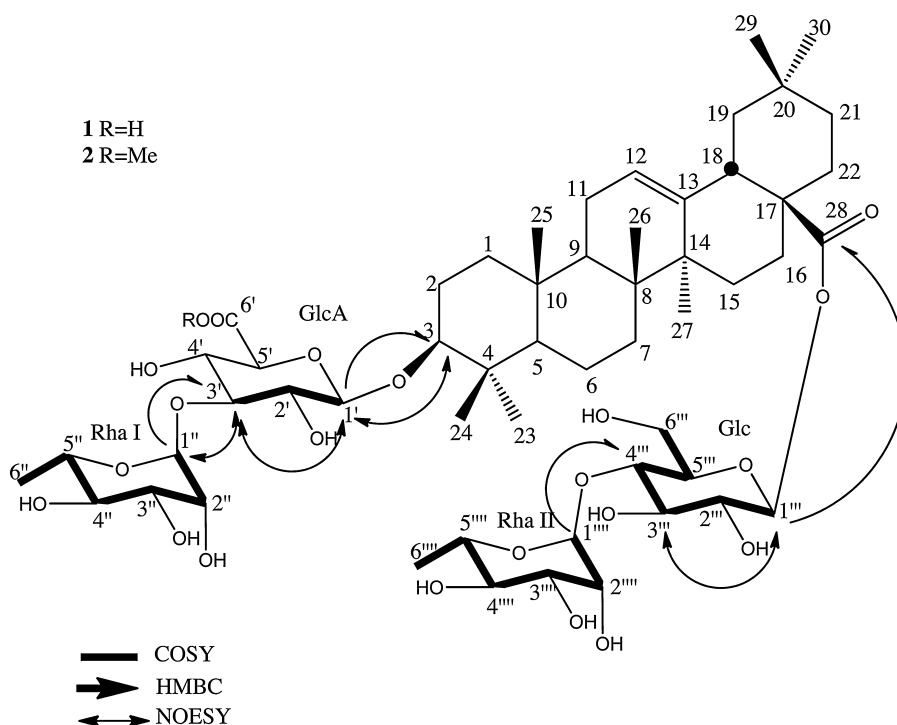


Fig. 1. Chemical structures and selected HMBC, COSY and NOESY correlations of **1** and **2**.

### 3. Experimental

#### 3.1. General experimental procedures

The high resolution electrospray ionization mass spectroscopy (HR-ESI-MS) was recorded on a Bruker MicrOTOF-QII spectrometer. The  $^1\text{H}$  NMR (500 MHz),  $^{13}\text{C}$  NMR (125 MHz), DEPT, COSY, HSQC and HMBC spectra were recorded on a Bruker AM500 FT-NMR spectrometer using tetramethylsilane (TMS) as internal standard. Column chromatography was carried out using Merck Silica gel normal-phase (230–240 mesh) and reversed-phase C<sub>18</sub> (Merck). Analytical TLC was carried out in silica gel plates (Merck DC-Alufolien 60 F<sub>254</sub>). Compounds were visualized by spraying with aqueous 10% H<sub>2</sub>SO<sub>4</sub> and heating for 3–5 min.

#### 3.2. Plant material

The leaves of *S. sessiliflora* De P. V. were provided by Center Cultivation and Processing of Medicinal Plants Da Lat, No. 18 Hoang Van Thu Street, Da Lat City, Viet Nam and identified by Dr. Phan Van De – Department of Medicinal Resources, The Research Center of Ginseng and Medicinal Materials, Ho Chi Minh City, Viet Nam. A voucher specimen was deposited in Bioactive Compounds Laboratory, Institute of Chemical Technology, Vietnam Academy of Science and Technology, Viet Nam.

The leaves were collected, washed, dried, and powdered.

#### 3.3. Extraction and isolation

Powder of leaves *S. sessiliflora* De P. V. (5 kg) were extracted with 96% EtOH (v/v), filtered residue, removed solvents under low pressure, obtained crude extract (890 g). Then, crude extract was added water and extracted with *n*-hexane, ethyl acetate, obtained extracts respectively. The aqueous portion was eluted by Diaion HP-20 column with: H<sub>2</sub>O, 25% MeOH, 50% MeOH, 75% MeOH and 100% MeOH. Then distilled solvents, gave five major fractions (I–V), respectively. Fraction IV (80 g) was subjected by column chromatography with mobile phase (EtOAc–MeOH) gradient (0–100%) gave

seven subfractions (IV.1–IV.7). Fraction IV.2 (15 g), was eluted with solvent system CHCl<sub>3</sub>–MeOH–H<sub>2</sub>O (85:15:0.1 → 75:25:0.3, v/v), then, separated by RP-18 using gradient mixtures of MeOH–H<sub>2</sub>O gave **3** (15 mg) and **4** (8 mg). Similar, Fraction IV.3 (20 g) was eluted with solvent system CHCl<sub>3</sub>–MeOH–H<sub>2</sub>O (80:20:0.2 → 70:30:0.5, v/v), then, separated by RP-18 using gradient mixtures of MeOH–H<sub>2</sub>O gave **1** (12 mg) and **2** (16 mg).

##### 3.3.1. Scheffleraside A (**1**)

Amorphous powder (MeOH);  $[\alpha]_{\text{D}}^{25}$  –17.0 (c 0.10, MeOH); IR (KBr)  $\nu_{\text{max}}$ : 3418, 2927, 1726, 1615, 1075, 1030 cm<sup>–1</sup>; UV (MeOH)  $\lambda_{\text{max}}$ : 206 and 220 nm; HR-ESI-MS  $m/z$  1085.5521 [M–H]<sup>–</sup> (calcd for C<sub>54</sub>H<sub>85</sub>O<sub>22</sub>, 1085.5527) and  $m/z$  1109.5533 [M+Na]<sup>+</sup> (calcd for C<sub>54</sub>H<sub>86</sub>O<sub>22</sub>Na, 1109.5503);  $^1\text{H}$  and  $^{13}\text{C}$  NMR data (pyridine-*d*<sub>5</sub>), see Table 1.

##### 3.3.2. Scheffleraside B (**2**)

Amorphous powder (MeOH);  $[\alpha]_{\text{D}}^{25}$  –13.5 (c 0.10, MeOH); IR (KBr)  $\nu_{\text{max}}$ : 3423, 2924, 1741, 1635, 1067, 1039 cm<sup>–1</sup>; UV (MeOH)  $\lambda_{\text{max}}$ : 240 nm; HR-ESI-MS  $m/z$  1123.5633 [M+Na]<sup>+</sup>, calcd for C<sub>58</sub>H<sub>88</sub>O<sub>22</sub>Na, 1123.5659;  $^1\text{H}$  and  $^{13}\text{C}$  NMR data (pyridine-*d*<sub>5</sub>), see Table 1.

#### 3.4. Acid hydrolysis

Each saponin (2 mg) was refluxed with 2 N aq. CH<sub>3</sub>COOH (5 ml) for 2 h at 100 °C. After extraction with CH<sub>2</sub>Cl<sub>2</sub> (3 × 5 ml), the aqueous layer was repeatedly evaporated to dryness with MeOH until neutral, and then analyzed by TLC over silica gel (MeCOEt–isoPrOH–Me<sub>2</sub>CO–H<sub>2</sub>O 20:10:7:6) by comparison with authentic samples (l-rhamnose *R<sub>f</sub>* 0.65; D-glucose *R<sub>f</sub>* 0.40; D-glucuronic acid *R<sub>f</sub>* 0.05) (Haddad et al., 2013; Voutquenne-Nazabadioko et al., 2013).

### References

- Haddad, M., Lelamer, A.C., Banuls, L.M.Y., Vazquez, P., Carraz, M., Vaisberg, A., Castillo, D., Sauvain, M., Rojas, R., Kiss, R., 2013. *In vitro* growth inhibitory effects of 13,28-epoxyoleanane triterpene saponins in cancer cells. *Phytochem. Lett.* 6, 128–134.

- Hansen, L., Boll, P.M., 1986. The polyacetylenic falcarinol as the major allergen in *Schefflera arboricola*. *Phytochemistry* 25, 529–530.
- Hebbbar, D.R., Nalini, M.S., 2014. Evaluation of phytochemicals, total phenolics and antioxidant activities of *Schefflera* spp. (Araliaceae) from southern India. *J. Pharmacogn. Phytochem.* 2, 10–14.
- Huynh, N.T., Phan, K.L., Nguyen, P.D., Tran, C.L., 2005. Studies on the effects of enhancing physical strength and antistress from the leaves of three *Schefflera* species as well as their combinative effects with red ginseng on mice. *J. Med. Hochiminh City* 9, 91–95.
- La, D.M., Chau, V.M., Tran, V.S., Pham, Q.L., Phan, V.K., Tran, H.T., Tran, M.H., Ninh, K.B., Le, M.H., 2013. Prospects of natural bioactive products from Araliaceae Juss. Family in VietNam. In: The 5-th National Conference on Ecology and Biological Resources. pp. 1152–1158.
- Li, Y.L., But, P.P.H., Ooi, V.E.C., 2005. Antiviral activity and mode of action of caffeoyl-quinic acids from *Schefflera heptaphylla* (L.) Frodin. *Antivir. Res.* 68, 1–9.
- Madl, T., Sterk, H., Mittelbach, M., 2006. Tandem mass spectrometric analysis of a complex triterpene saponin mixture of *Chenopodium quinoa*. *J. Am. Soc. Mass Spectrom.* 17, 795–806.
- Maeda, C., Ohtani, K., Kasai, R., Yamasaki, K., Nguyen, M.D., Nguyen, T.N., Nguyen, K.Q.C., 1994. Oleanane and ursane glycosides from *Schefflera octophylla*. *Phytochemistry* 37, 1131–1137.
- Mizui, F., Kasai, R., Ohtani, K., Tanaka, O., 1990. Saponins from bran of quinoa, *Chenopodium quinoa* Willd. II. *Chem. Pharm. Bull.* 38, 375–377.
- El, S., Morta, M., 1998. Study of the saponin content of *Atriplex stylosa* VIV, and its molluscicidal effect. *Bull. Pharm. Sci. Assiut Univ.* 21, 237–243.
- Nguyen, T.T.H., Nguyen, T.C., Do, M.A., Tran, C.L., Nguyen, P.D., 2004. Studies on the combinative effects of the leaves of three *Schefflera* species (Araliaceae) with red ginseng on the effects of enhancing physical strength and antistress. *J. Med. Hochiminh City* 8, 151–155.
- Pancharoen, O., Tuniwachuttikul, P., Taylor, W.C., Picker, K., 1994. Triterpenoid glycosides from *Schefflera leucantha*. *Phytochemistry* 35, 987–992.
- Srivastava, S.K., 1992. A new triterpenic acid from *Schefflera impressa*. *J. Nat. Prod.* 55, 298–302.
- Tapondjou, A.L., Miyamoto, T., Lacaille-Dubois, M.A., 2006. Glucuronide triterpene saponins from *Bersama engleriana*. *Phytochemistry* 67, 2126–2132.
- Tran, V.S., Peter-Katalinic, J., Günter, A., 1991. A bidesmosidic triterpenoid saponin from *Schefflera octaphylla*. *Phytochemistry* 30, 3717–3720.
- Tran, M.T., Dang, T.T.N., Tran, C.L., Nguyen, T.T.H., 2012. Study on androgenic effect of *Schefflera* sp3. *J. Med. Mater.: Hanoi* 17, 17–23.
- Vo, D.H., Tran, C.L., Duong, H.T.Q., 2003. Survey characteristics and preliminary study of the chemical composition of leaves, stems and roots of *Schefflera* sp3. *J. Med. Mater.: Hanoi* 6, 161–167.
- Vo, D.H., Tran, C.L., Duong, H.T.Q., 2004. Studies on saponin composition of *Schefflera* sp3. *J. Med. Mater.: Hanoi* 9, 46–50.
- Vo, D.H., Nguyen, T.T.H., Tran, C.L., Huynh, T.C.H., 2008. Study on the chemistry and *in vitro* antioxidant activity of saponin from *Schefflera* sp3. *J. Med. Mater.: Hanoi* 13, 17–21.
- Voutquenne-Nazabadioko, L., Gevrenova, R., Borie, N., Harakat, D., Sayagh, C., Weng, A., Thakur, M., Zaharieva, M., Henry, M., 2013. Triterpenoid saponins from the roots of *Gypsophila trichotoma* Wender. *Phytochemistry* 90, 114–127.
- Wanas, A.S., Matsunami, K., Otsuka, H., Desoukey, S.Y., Fouad, M.A., Kamel, M.S., 2010. Triterpene glycosides and glucosyl esters, and a triterpene from the leaves of *Schefflera actinophylla*. *Chem. Pharm. Bull.* 58, 1596–1601.
- Wang, Y., Wang, L., Wang, W.J., Zhang, X.Q., Tian, H.Y., Zhang, Q.W., Li, Y.L., Ye, W.C., 2014. New triterpenoid saponins from the aerial parts of *Schefflera kwangsiensis*. *Carbohydr. Res.* 385, 65–71.
- Wu, C., Duan, Y.H., Li, M.M., Tang, W., Wu, X., Wang, G.C., Ye, W.C., Zhou, G.X., Li, Y.L., 2013. Triterpenoid saponins from the stem barks of *Schefflera heptaphylla*. *Planta Med.* 79, 1348–1355.
- Zhao, Z., Matsunami, K., Otsuka, H., Shinzato, T., Takeda, Y., Kawahata, M., Yamaguchi, K., 2010. Schefflerins A–G, new triterpene glucosides from the leaves of *Schefflera arboricola*. *Chem. Pharm. Bull.* 58, 1343–1348.