Phenolic Glycosides from Roots of Adenophora tetraphylla Collected in Heilongjiang, China

Hai-Xue Kuang,^a Chun-Jie Shao,^b Ryoji Kasai,*,^c Kazuhiro Ohtani,^c Zhen-Kun Tian,^a Jing-Da Xu^b and Osamu Tanaka^d

Department of Chinese Pharmacy, Heilongjiang College of Traditional Chinese Medicine, 14 Ho-Ping Road, Harbin, China, Department of Organic Chemistry, Bethune University of Medical Sciences, 6 Xinmin Street, Changchun, China, Institute of Pharmaceutical Sciences, Hiroshima University School of Medicine, Kasumi, Minami-ku, Hiroshima 734, Japan and Suzugamine Women's College, 4-6-18 Inokuchi, Nishi-ku, Hiroshima 733, Japan. Received March 14, 1991

Adenophora tetraphylla (= A. triphylla, Campanulaceae) is a source of the traditional Chinese medicine "Shashen". From the roots of this plant, three new phenolic glycosides, called shashenosides I, II and III (2—4) were isolated together with siringinoside (1), β -sitosteryl- β -D-glucoside, linoleic acid and methyl stearate. The common aglycone of 2, 3 and 4 was identified as 3-methoxy-5-(2'-propenyl)-1,2-benzenediol [=1-O-methyl-5-(2'-propenyl)-pyrogallol=6-hydroxyeugenol, 6], and the structures of 2 and 3 were elucidated as 2,3-di-O- β -glucopyranoside and 3-O- α -arabinopyranosyl-(1 \rightarrow 6)- β -glucopyranoside of 6, respectively. Shashenoside III (4) was formulated as 2-O- β -glucopyranoside of 3. No saponin was found in the roots.

Keywords Adenophora tetraphylla; Campanulaceae; Shashen; shashenoside; Chinese traditional medicine; synapyl-glycoside; 3-methoxy-5-(2'-propenyl)-1,2-benzenediol; glycoside

The traditional Chinese medicine, Shashen (沙参), roots of Adenophora spp. (Campanulaceae) is known to be an antiinflammatory and antitussive drug used in the treatment of lung disease. From Adenophora species, Konno et al. reported the isolation of sterols and triterpenoids¹⁾ and Du et al. isolated triterpenoids and coumarins.²⁾ A. tetraphylla Fisch. ex Jackson (= A. triphylla (Thunb.) A.DC.,3) Chinese name: 輪葉沙参, 南参; Japanese name: saiyoushajin) is one of source plants of this crude drug. We have conducted studies on water soluble constituents of dangshen (党参), roots of Codonopsis spp. (Campanulaceae).4-6) Continuing our chemical studies on campanulaceous traditional Chinese medicines, the present paper reports the isolation and structural elucidation of four phenolic glycosides from roots of A. tetraphylla collected in Heilongjian province, China.

As in the case of Dangshen, a methanolic extract of the roots contained a large amount of saccharides, which was removed by chromatography on highly porous synthetic polymer (Diaion HP-20). The resulting crude glycoside fraction was separated by combination of a variety of chromatography as described in the Experimental section to give four glycosides, 1—4 together with β -sitosteryl- β -D-glucoside, linoleic acid and methyl stearate.

Compound 1, C₂₃H₃₄O₁₄ (from high resolution fast atom

bombardment mass spectrometry (HR-FAB-MS)) afforded glucose on acid hydrolysis. Inspection of the negative FAB-MS as well as the $^1\text{H-}$ and $^{13}\text{C-}$ nuclear magnetic resonance (NMR) spectrum (in $\text{C}_5\text{D}_5\text{N}$) led to the identification of 1 as syringinoside (4-O- β -gentiobiosyl-3,5-dimethoxy-4-hydroxy-trans-cinnamylalcohol) which was recently isolated from Wikstroemia sikokiana FRANCH. et SAV. (Thymelaeaceae).

Compound 2 called shashenoside I, C₂₂O₃₂H₁₃ (from HR-FAB-MS) afforded glucose and an aglycone (6), C₁₀H₁₂O₃ (from HR-FAB-MS) on acid hydrolysis. The ¹H-NMR spectrum of 6 (in CD₃OD) exhibited signals due to an allyl group at δ 3.27 (2H, dddd, J = 6.8, 1.6, 1.3, 0.6 Hz, $-CH_2-CH=CH_2$), 5.92 (1H, ddt, J=17.0, 10.1, 6.8 Hz, $-CH_2-CH_2-CH_2$, 5.05 (1H, ddt, J=10.1, 2.0, 1.3 Hz, $-CH_2-CH = C\underline{H}_2$ (cis)), 5.08 (1H, ddt, J = 17.0, 2.0, 1.6 Hz, $-CH_2-CH=CH_2$ (trans)). Presence of an allyl group in 6 was also supported by the ¹³C-NMR spectrum (in CD₃OD): signals at δ 40.1 (-CH₂-), 137.6 (-CH=), 115.7(=CH₂). The ¹³C-NMR (in CD₃OD) spectrum of **6** exhibited a methoxyl carbon signal at δ 56.2, four substituted aromatic carbon signals at δ 146.9, 143.9, 132.1, 130.7 and two unsubstituted aromatic carbon signals at δ 108.9, 103.5. The ¹H-NMR spectrum of $\bf 6$ showed two hydroxy proton signals at δ 5.28 and 5.34 (each 1H, s), a methoxyl proton signal at δ 3.86

© 1991 Pharmaceutical Society of Japan

(3H, s) and a couple of *meta*-located aromatic proton signals at δ 6.30 (1H, d, J=1.8 Hz) and 6.44 (1H, dt, J=1.8, 0.6 Hz, long range coupling with methylene protons). The nuclear Overhauser effect (NOE) difference spectrum of $\boldsymbol{6}$ revealed the presence of NOE between two aromatic proton signals and the allylic methylene signal as well as between one of the aromatic proton signals (at δ 6.30) and the methoxyl proton signal (see Chart 1). These results led to assignment of the structure of $\boldsymbol{6}$ as 3-methoxy-5-(2'-propenyl)-1,2-benzenediol [=1-O-methyl-5-(2'-propenyl)-pyrogallol=6-hydroxyeugenol]. Compound $\boldsymbol{6}$ was used as a starting material for syntheses of natural phenolic compounds such as (\pm)-eusiderin, ⁸⁾ but this is the first example of the occurrence of this compound in nature as an aglycone of glycosides.

The ¹H-NMR spectrum of **2** in CD₃OD exhibited two β -glucosyl anomeric proton signals at δ 4.79 (1H, d, J=7.9 Hz) and 5.80 (1H, d, J=7.9 Hz). The ¹³C-NMR spectrum of **2** in C₅D₅N exhibited signals assignable to two sets of terminal β -glucopyranosyl units together with signals due to the aglycone carbons. Accordingly, **2** can be formulated as 1,2-di-O- β -glucopyranoside of **6**.

Compound 3, called shashenoside II, C21H30O12 (from HR-FAB-MS), yielded glucose, arabinose and the aglycone (6) on acid hydrolysis. The ¹H-NMR spectrum of 3 in CD_3OD showed two anomeric proton signals at δ 4.75 (1H, d, J = 7.3 Hz) and 4.29 (1H, d, J = 6.8 Hz). The ¹³C-NMR spectrum of 3 in CD₃OD exhibited signals due to a terminal α -arabinopyranosyl unit and a 6-linked β -glucopyranosyl unit, indicating the presence of an α-arabinopyranosyl- $(1\rightarrow 6)$ - β -glucopyranosyl moiety. In order to elucidate the location of the glycosyl moiety, a methyl ether (7) was prepared from 3 by treatment with diazomethane. As reported, in the ¹H-NMR spectra of 5, ⁴⁾ signals due to two aromatic and two methoxy protons appear at the same position, respectively, showing the symmetrical nature of the aromatic ring moiety. In contrast, the ¹H-NMR spectrum of 7 in CD₃OD exhibited signals due to the two aromatic and two methoxy proton signals at dfferent positions: at δ 6.66, 6.55 (each 1H, d, J=1.8 Hz, C-2,6), 3.81, 3.79 (each 3H, s, OCH₃), leading to the structure of 3 as 1-O- α arabinopyranosyl- $(1\rightarrow 6)$ - β -glucopyranoside of **6**. Presence of NOE between the signals at δ 3.81 and 6.55 as well as absence of NOE of another aromatic signal at δ 6.66 with both the methoxyl proton signals supported this formulation.

Compound **4**, called shashenoside III, $C_{27}H_{40}O_{17}$ (from HR-FAB-MS) gave glucose, arabinose and **6** on acid hydrolysis. The ¹H-NMR spectrum of **4** in CD₃OD exhibited three anomeric proton signals at δ 4.94 (1H, d, J=7.3 Hz), 4.89 (1H, d, J=6.3 Hz), 4.29 (1H, d, J=6.8 Hz). Comparison of the ¹³C-NMR of **4** in CD₃OD with that of **3** revealed the presence of a terminal β -glucopyranosyl unit together with a α -arabinopyranosyl-(1 \rightarrow 6)- β -glucopyranosyl moiety in **4**. On mild acid hydrolysis, **4** afforded **3**. It follows that **4** can be formulated as 1-O- β -glucopyranoside of **3**.

It has been mentioned that Shashen contains some saponins, and can be used as a substitute of Ginseng. However, in the present study, no saponin was isolated from the roots.

Experimental

General Procedures ¹H- (400 MHz) and ¹³C- (100 MHz) NMR spectra were recorded on a JEOL JNM GX-400 NMR spectrometer. FAB-MS was taken on a JEOL JMS SX-102 mass spectrometer by the direct inlet method. Preparative high performance liquid chromatography (HPLC) was carried out on a column of TSK-gel ODS-120T (21.5 mm i.d. × 30 cm): detection, ultraviolet (UV) at 254 nm; flow rate, 6 ml/min. Acid hydrolysis of glycosides and identification of the resulting monosaccharides by gas-liquid chromatography (GLC) were conducted in the usual manner.

Extraction and Separation Dried roots (8 kg) collected at Xiaoxinganling, Heilongjiang Province of China, were extracted with hot MeOH to give MeOH-extract (700 g). A suspension of the MeOH-extract (350 g) in $\rm H_2O$ was chromatographed on a column of Diaion HP-20 (Mitsubishi Kasei Co., Ltd.) with $\rm H_2O$ and then with MeOH. The $\rm H_2O$ -eluate was composed of a large amount of saccharides and other highly water soluble substances. Twenty grams of the MeOH-eluate (total: 40 g) was subjected to chromatography on silica gel with CHCl₃–MeOH- $\rm H_2O$ (30:10:1 then 14:6:1, homogenous) to give six fractions tentatively designated as fr. I—VI in their order of elution. Further chromatography of fr. I on silica gel with CHCl₃–MeOH (20:1) afforded β-sitosteryl 3- $\rm O$ -β-glucoside (150 mg) and linoleic acid (10 mg). Fraction II was chromatograhed on silica gel with CHCl₃–MeOH (6:1), affording methyl stearate. All of these compounds were identified by comparison of thin layer chromatography (TLC) and the $^{\rm 1}$ H- and $^{\rm 13}$ C-NMR spectra with corresponding authentic specimens.

Fractions II and III were respectively separated by HPLC on an octadecyl silica (ODS) column with 30% MeOH to give 3 (90 mg) from fr. III and 2 (228 mg) from fr. IV. Fractions V and VI were respectively subjected to chromatography on LiChroprep RP-18 (40—63 m μ , Merck) with 25% MeOH followed by HPLC on an ODS column with 24% MeOH, affording 1 (30 mg) from fr. V and 4 (40 mg) from fr. VI.

Syringinoside (1): A white powder, $[\alpha]_D^{18} - 22.6^{\circ}$ (c = 0.31, MeOH), HR-FAB-MS (positive) m/z: Calcd for $[C_{23}H_{34}O_{14}+Na]^+$ 557.1846. Found 557.1884. FAB-MS m/z (negative): 533 (M-H)⁻, 371 (M-Glc-H)⁻, 209 (M-Glc₂-H)⁻. ¹H-NMR in C_5D_5N : δ 6.89 (2H, s), 6.64 (1H, dt, J = 16.1, 5.2 Hz), 6.86 (1H, d, J = 16.1 Hz), 5.71 (1H, d, J = 6.8 Hz), 4.93 (1H, d, J = 7.8 Hz), 4.57 (2H, d, J = 5.2 Hz), 3.83 (6H, s). ¹³C-NMR in CD₃OD: aglycone carbons, δ 57.0 (2C), 63.6, 105.5, 130.0 (2C), 131.0, 131.3, 135.3, 154.3 (2C); sugar carbons, δ 62.8, 69.3, 71.4, 71.7, 75.2, 75.7, 77.9 (4C), 104.5, 105.1.

Shashenoside I (2): A pale yellow powder, $[\alpha]_D^{18} - 47.6^{\circ}$ (c = 0.42, H_2O). HR-FAB-MS (positive) m/z: Calcd for $[C_{22}H_{32}O_{13}+Na]^+$ 527.1741. Found: 527.1780. FAB-MS (negative) m/z: 503 (M – H) –, 341 (M – Glc – H) –, 179 (M – Glc $_2$ – H) – . ¹H-NMR in CD $_3$ OD: δ 3.84 (3H, s, OCH $_3$), 6.81, 6.60 (each 1H, d, J = 1.8 Hz, a pair of meta-located aromatic protons) and allyl group signals at δ 3.29 (2H, ddd, J = 6.4, 1.6, 1.3 Hz), 5.95 (1H, ddt, J = 17.0, 10.1, 6.8 Hz), 5.05 (1H, ddt, J = 10.1, 2.0, 1.3 Hz), 5.06 (1H, ddt, J = 17.0, 2.0, 1.6 Hz). ¹³C-NMR in CD $_3$ OD: aglycone carbons, δ 41.2 (C- α), 57.0 (OCH $_3$), 109.3, 112.0 (C- α), 116.3 (C- γ), 135.5 (C-4), 138.5 (C- β), 138.7 (C-1), 152.4, 154.3 (C-3,5); sugar carbons, δ 62.5, 62.6 (C- α), 71.3, 71.4 (C-4), 75.1, 75.7 (C-2), 77.7, 78.0, 78.4 (2C) (C-3,5), 103.8, 105.5 (C-1).

A solution of **2** (45 mg) in a mixture of 2% HCl and 50% dioxan for 2 h at 75 °C. The reaction mixture was extracted with C_6H_6 and the benzene extract was chromatographed on silica gel with $CHCl_3$ –MeOH (20:1) to give **6**: A yellow powder. HR-EI-MS m/z: Calcd for $C_{10}H_{12}O_3^+$ 180.0786. Found: 180.0766.

Shashenoside II (3): A yellow powder, $[\alpha]_D^{18} - 51.4^{\circ}$ (c = 0.68, MeOH). HR-FAB-MS (positive) m/z: Calcd for $[C_{21}H_{30}O_{12} + Na]^+$ 497.1635. Found: 497.1651. ¹H-NMR in CD₃OD: δ 6.66, 6.54 (each 1H, d, J = 1.8 Hz, C-2,4), 5.94 (1H, ddt, J = 16.8, 10.1, 6.8 Hz, C- β), 5.06 (1H, dd, J = 16.8, 1.8 Hz, C- γ trans), 5.03 (1H, ddt, J = 10.1, 1.8, 1.0 Hz, C- γ cis), 4.75 (1H, d, J=7.3 Hz, anomeric H), 4.29 (1H, d, J=6.6 Hz, anomeric H), 3.82 (3H, s, OCH₃), 3.29 (2H, dd, J = 6.8, 1.0 Hz, C- α). ¹³C-NMR in CD₃OD: aglycone carbons, δ 40.9 (C- α), 56.8 (OCH₃), 108.9, 112.3 (C-6,2), 115.9 $(C-\gamma)$, 132.5 (C-1), 136.0 (C-4), 139.1 $(C-\beta)$, 146.7, 149.6 (C-5,3); anomeric carbons δ 104.3, 104.8; α -arabinoside carbons, δ 66.6 (C-5), 69.2 (C-4), 72.3 (C-2), 74.1 (C-3); β -glucoside carbons, δ 69.3 (C-6), 71.3 (C-4), 74.8 (C-2), 77.0 (C-5), 77.4 (C-3). On acid hydrolysis in the same way as 2, 3 afforded glucose, arabinose and 6. Shashenoside III (3) (15 mg) was treated with CH2N2 in Et2O-MeOH and the product was purified by HPLC on an ODS column with 35% MeOH to give a methyl ether 7: A yellow powder. HR-FAB-MS (positive) m/z: Calcd for $[C_{22}H_{32}O_{12} + Na]$ 511.1791. Found: 511.1820. ¹H-NMR in CD₃OD: δ 6.66 (1H, d, J=1.8

Hz, C-2), 6.55 (1H, d, J=1.8 Hz, C-6), 5.96 (1H, m, C- β), 5.10 (1H, dd, J=16.7, 2.0 Hz, C- γ trans), 5.04 (1H, dd, J=10.0, 2.0 Hz, C- γ cis), 4.94 (1H, d, J=7.0 Hz, anomeric H), 4.26 (1H, d, J=6.7 Hz, anomeric H), 3.81 (3H, s, 5-OCH₃), 3.79 (3H, s, 4-OCH₃), 3.29 (2H, d, J=7.3 Hz, C- α).

Shashenoside III (4): A yellow powder, $[\alpha]_D^{18} - 42.2^{\circ}$ (c = 0.45, MeOH). HR-FAB-MS (positive) m/z: Calcd for $[C_{27}H_{40}O_{17} + Na]^+659.2163$. Found: 659.2184. Calcd for $[C_{27}H_{40}O_{17}+K]^+$ 675.1903. Found: 675.1940. FAB-MS (negative) m/z: 635 (M-H)⁻, 473 (M-Glc-H)⁻, 341 $(M-Glc-Ara-H)^{-}$, 179 $(M-Glc_2-Ara-H)^{-}$. ¹H-NMR in CD₃OD: δ 6.77, 6.66 (each 1H, d, J = 1.8 Hz, C-2,6), 5.96 (1H, m, C- β), 5.07 (2H, m, C- γ), 4.94 (1H, d, J=7.3 Hz, anomeric H), 4.89 (1H, d, J = 6.3 Hz, anomeric H), 4.29 (1H, d, J = 6.8 Hz, anomeric H), 3.82 (3H, s, OCH₃), 3.31 (2H, d, J = 6.6 Hz, C- α). ¹³C-NMR in CD₃OD: aglycone carbons, δ 41.2 (C-α), 57.2 (OCH₃), 109.4, 112.1 (C-2,6), 116.4 (C-γ), 135.6 (C-4), 138.6 (C- β), 138.7 (C-1), 152.0, 154.4 (C-3,5); anomeric carbons, δ 103.7, 105.0, 105.4; α -arabinoside carbons, δ 66.6 (C-5), 69.3 (C-4), 72.4 (C-2), 74.2 (C-3); β -glucoside carbons, δ 62.5 (C-6), 69.4 (C-6), 71.2 (C-4), 71.3 (C-4), 75.0 (C-2), 75.8 (C-2), 77.1, 77.4, 77.9, 78.3 (C-3,5). On acid hydrolysis in the same way as 2, 4 gave glucose, arabinose and 6.

A solution of 4 (15 mg) in 1% H₂SO₄ (20 ml) was heated at 70 °C for 3 h. The reaction mixture was neutralized with diluted NaOH solution and then extracted with 1-BuOH saturated with H₂O. The BuOH extract was chromatographed on silica gel with CHCl₃-MeOH-H₂O (30:10:1,

homogeneous) to give 3 which was identified by comparison of the ¹H- and ¹³C-NMR spectra, optical rotation and TLC with those of an authentic sample.

Acknowledgements We are grateful to the Japan Society for the Promotion of Science, Tokyo for the fellowship for C. H. Shao's study in Hiroshima University. Thanks are also due to Professor M. Kozawa, Osaka University of Pharmaceutical Sciences for his valuable advice.

References

- 1) C. Konno, T. Saito, Y. Oshima, H. Hikino and C. Kabuto, *Planta Medica*, 42, 268 (1981).
- 2) S. J. Du, P. Gariboldi and G. Jommi, Planta Medica, 1986, 317.
- 3) J. Ohwi, "Flora of Japan," Shibundo, Tokyo, 1965, pp. 1280—1281.
- 4) K. Mizutani, M. Yuda, O. Tanaka, Y. Saruwatari, T. Fuwa, M. R. Jia, Y. R. Ling and X. F. Pu, *Chem. Pharm. Bull.*, **36**, 2689 (1988).
- K. Mizutani, M. Yuda, O. Tanaka, Y. Saruwatari, M. R. Jia, Y. R. Ling and Y. F. Pu, Chem. Pharm. Bull., 36, 2726 (1988).
- M. Yuda, K. Ohtani, K. Mizutani, R. Kasai, O. Tanaka, M. R. Jia, Y. R. Ling, X. F. Pu and Y. Saruwatari, *Phytochemistry*, 29, 1989 (1990).
- M. Niwa, Y. Iwadare, Y. Wu and Y. Hirata, Chem. Pharm. Bull., 36, 1158 (1988).
- 8) L. Marlini and A. Zanarotti, Tetrahedron Lett., 1975, 3621.