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## The First Synthesis and Iron Binding Studies of the Natural Product, *myo*-Inositol 1,2,3-Trisphosphate.

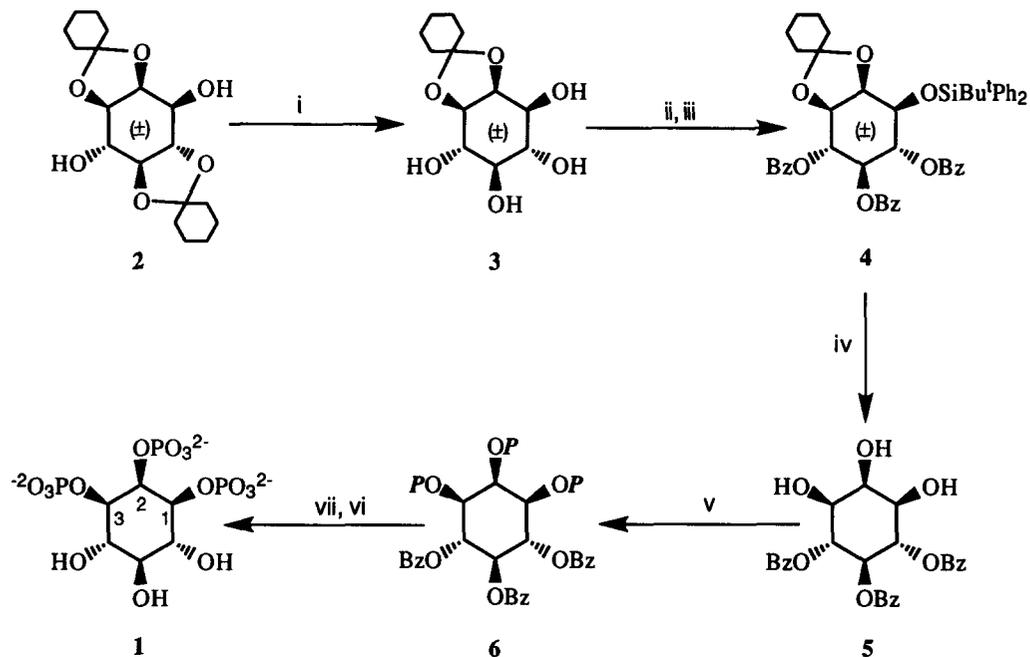
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**Abstract:** The natural product *myo*-inositol 1,2,3-trisphosphate **1** has been prepared and shown to inhibit Fe<sup>3+</sup>-catalysed hydroxyl radical formation.

*myo*-Inositol hexakisphosphate (phytic acid) is a widespread inositol phosphate, found in all eucaryotic cells, where it is present at concentrations 10-100 μM; however its biological role remains elusive. Phytic acid is an excellent chelator of Fe<sup>3+</sup> with an affinity<sup>1</sup> in the region of 10<sup>25</sup> and it also inhibits Fe<sup>3+</sup>-catalysed HO· production;<sup>2</sup> it has recently been suggested that it may act as both an intracellular, low molecular weight chelator of Fe<sup>3+</sup> and antioxidant. All *myo*-inositol pentakisphosphate isomers bind Fe<sup>3+</sup> with high affinity; however only those with the 1,2,3-trisphosphate groups are also antioxidant.<sup>3</sup> It has been proposed that *myo*-inositol 1,2,3-trisphosphate **1** represents the simplest structure able to bind Fe<sup>3+</sup> and function as an antioxidant, but this has never been tested. The synthesis of this recently discovered natural product<sup>4</sup> is discussed along with its Fe<sup>3+</sup> binding studies.

The synthesis of trisphosphate **1** is shown in Scheme 1. Diol **2** was synthesized from *myo*-inositol and 1,1-diethoxycyclohexane,<sup>5</sup> which was converted to **3** by the selective removal of the *trans*-cyclohexylidene ring.<sup>6</sup> The tetrol **3** was regioselectively silylated<sup>7</sup> at the 1-position, the product from which was then benzoylated to give the fully protected intermediate **4**. Both the *cis*-cyclohexylidene and silyl groups of **4** were removed by treatment with aqueous trifluoroacetic acid at 50°C to give **5**, which was crystallised from diethyl ether. The triol **5** was phosphorylated with dibenzyl *N,N*-diisopropylphosphoramidite<sup>8</sup> and the intermediate phosphite was oxidised<sup>9</sup> to yield the trisphosphate **6** as a crystalline compound, the structure of which was confirmed by X-ray diffraction analysis<sup>10</sup> (Figure 1). Deprotection of **6** was achieved in a quantitative yield by hydrogenolysis followed by base catalysed hydrolysis of the benzoyl esters to give the trisphosphate **1**, which was crystallised as its hexa(cyclohexylammonium) salt. All new compounds were fully characterised by <sup>1</sup>H, <sup>13</sup>C and <sup>31</sup>P NMR spectroscopy,<sup>11</sup> IR and mass spectrometry, and elemental analysis.

The ability of *myo*-inositol 1,2,3-trisphosphate **1** to inhibit Fe<sup>3+</sup>-catalysed hydroxyl radical formation was studied in a hypoxanthine/xanthine oxidase system. This generates HO·, which in turn generates formaldehyde from dimethylsulphoxide (present in the assay).<sup>3</sup> Both **1** and phytic acid bind to Fe<sup>3+</sup> with high affinities and both completely inhibited Fe<sup>3+</sup>-catalysed hydroxyl radical formation at >100 μM (Figure 2). We conclude that the 1,2,3 (equatorial-axial-equatorial) trisphosphate grouping in phytic acid is the orientation needed to inhibit Fe<sup>3+</sup>-catalysed hydroxyl radical formation. This may allow phytic acid to function as a 'safe' carrier of Fe<sup>3+</sup> in the cell.



Scheme 1: Synthesis of *myo*-inositol 1,2,3-trisphosphate **1**.

i. *p*-TsOH, toluene, hexane, EtOH; ii.  $\text{Bu}^t\text{Ph}_2\text{SiCl}$ , imidazole, pyridine (58%); iii. benzoyl chloride, DMAP, pyridine (73%); iv. aqueous  $\text{CF}_3\text{CO}_2\text{H}$  (49%); v.  $(\text{BnO})_2\text{PNPr}_2$ , 1*H*-tetrazole,  $\text{CH}_2\text{Cl}_2$ , then *m*-CPBA (56%); vi.  $\text{H}_2$ , Pd/C (10%), EtOH, room temperature, overnight; vii. NaOH (0.5M), quantitative. Abbreviations: P =  $(\text{BnO})_2\text{P}(\text{O})$ ; Bn = benzyl; Bz = benzoyl.

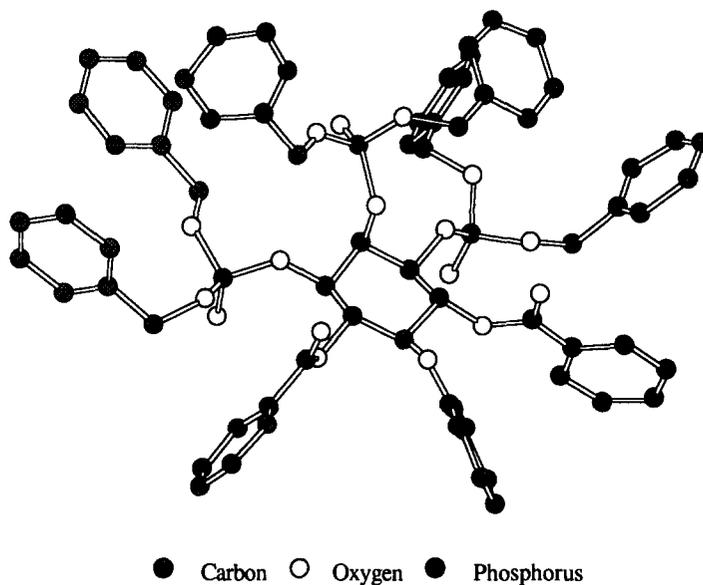


Figure 1: Crystal structure of 1,2,3-tris(dibenzylphosphoryl)-4,5,6-tribenzoyl *myo*-inositol **6**.<sup>11</sup>

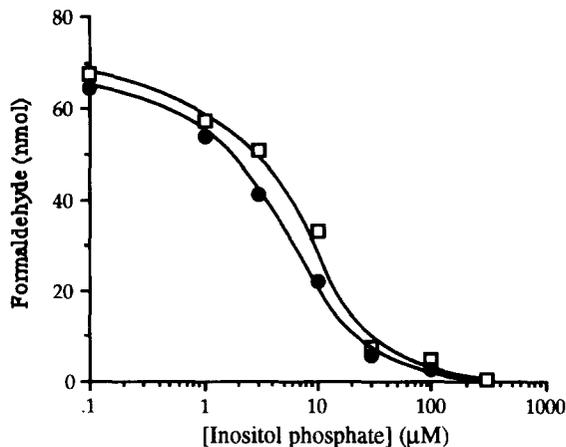


Figure 2: Effects of phytic acid (●) and *myo*-inositol 1,2,3-trisphosphate (□) on HO<sup>•</sup> generation. This result is typical of three independent experiments.

In summary, we report the first synthesis of the natural product *myo*-inositol 1,2,3-trisphosphate **1**. The crystal structure of the key intermediate 1,2,3-tris(dibenzylphosphoryl)-4,5,6-tribenzoyl *myo*-inositol **6** is presented. The 1,2,3 (equatorial-axial-equatorial) trisphosphate grouping present in both **1** and phytic acid inhibits Fe<sup>3+</sup>-catalysed hydroxyl radical formation. Further biological investigations and full experimental details for the synthesis of **1** will be reported in a forthcoming full paper.

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#### References and notes

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10. Crystallographic data for **6**: Monoclinic  $P2_1$ ,  $a = 15.564(3)$ ,  $b = 18.835(2)$ ,  $c = 22.738(6)$  Å,  $\beta = 95.04(2)^\circ$ ,  $V = 6640(2)$  Å<sup>3</sup>,  $Z = 4$ ,  $\rho_{\text{calc}} = 1.292$  g/cm<sup>3</sup>,  $\mu = 0.162$  mm<sup>-1</sup>,  $R = 0.0566$ ,  $wR2 = 0.1429$  for 7032 observed data [ $F_o \geq 4.0\sigma(F_o)$ ] from 12807 collected data. The diffraction data were collected on an Enraf-Nonius CAD4 diffractometer at 293(2) K in the  $\omega$ - $2\theta$  scan mode using Mo-K $\alpha$  radiation to a maximum  $2\theta$  value of 50°. The intensity data were corrected for Lorentz-polarization and intensity decay using DATRED (Brookhaven Natl. Lab. & University of Birmingham). The structure was solved by direct methods (SHELXS-86, Sheldrick 1990) and refined by the full-matrix least-squares method (SHELXL-93, Sheldrick 1993). All non-hydrogen positions were found including two molecules of water. The atomic coordinates and data for the X-ray structure are available from the Cambridge Crystallographic Data Centre, Lensfield Road, Cambridge, CB2 1EU, United Kingdom.
11. Melting points and selected NMR data:
4. M.p. 173.5 - 175 °C; <sup>1</sup>H-NMR (250.1 MHz, CDCl<sub>3</sub>):  $\delta$  0.95 (s, 9H, But), 1.19-2.06 (m, 10H, CH<sub>2</sub> of cyclohexylidene), 4.09-4.16 (m, 2H), 4.30 (dd, 1H,  $J_{\text{HH}} = 3.9$  Hz,  $J_{\text{HH}} = 9.3$  Hz), 5.36 (t, 1H,  $J_{\text{HH}} = 9.8$  Hz), 5.79 (dd, 1H,  $J_{\text{HH}} = 10.3$  Hz,  $J_{\text{HH}} = 7.0$  Hz), 6.03 (t, 1H,  $J_{\text{HH}} = 9.4$  Hz), 7.16-7.86 (m, 25H, Ph).
5. M.p. 100.5 - 103 °C; <sup>1</sup>H-NMR (250.1 MHz, CD<sub>3</sub>OD):  $\delta$  4.02 (dd, 2H, H-1/3,  $J_{\text{HH}} = 2.7$  Hz,  $J_{\text{HH}} = 9.7$  Hz), 4.19 (t, 1H, H-2,  $J_{\text{HH}} = 2.7$  Hz), 5.67 (t, 1H, H-5,  $J_{\text{HH}} = 9.9$  Hz), 5.85 (t, 2H, H-4/6,  $J_{\text{HH}} = 9.9$  Hz), 7.19-7.50 (m, 9H, Ph), 7.69-7.72 (m, 2H, Ph), 7.88-7.92 (m, 4H, Ph). <sup>1</sup>H-NMR (250.1 MHz, DMSO-*d*<sub>6</sub>): peaks include  $\delta$  5.27 (d, OH-1/3,  $J_{\text{HH}} = 6.1$  Hz), 5.50 (d, OH-2,  $J_{\text{HH}} = 2.9$  Hz), exchangeable in D<sub>2</sub>O.
6. M.p. 131.5 - 132.5 °C.
1. <sup>31</sup>P-NMR (101.3 MHz, D<sub>2</sub>O, <sup>1</sup>H decoupled, referenced to 85% H<sub>3</sub>PO<sub>4</sub>):  $\delta$  1.78 (s, P-2), 4.08 (s, P-1/3). <sup>1</sup>H-NMR (250.1 MHz, D<sub>2</sub>O, referenced to acetone at 2.05 ppm):  $\delta$  [0.97-1.25 (m, 30H), 1.47 (br d, 6H,  $J_{\text{HH}} = 11.9$  Hz), 1.62 (br d, 12H,  $J_{\text{HH}} = 3.8$  Hz), 1.80 (br s, 12H), 2.85-3.07 (m, 6H), (6 x cyclohexylammonium)], 3.22 (t, 1H, H-5,  $J_{\text{HH}} = 9.2$  Hz), 3.66 (t, 2H, H-4/6,  $J_{\text{HH}} = 9.5$  Hz), 3.81 (br t, 2H, H-1/3,  $J_{\text{HH}} = J_{\text{PH}} \approx 8.9$  Hz), 4.55 (br d, 1H, H-2,  $J_{\text{PH}} = 9.8$  Hz).

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