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Synthesis of a Novel Rhizobitoxine-Like Triazole-Containing Amino Acid

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Abstract The synthesis of the four stereoisomers of a new 1,2,3-triazole analogue of rhizobitoxine from serine is described. The key step is a Huisgen 1,3-dipolar cycloaddition on an ethynylglycine synthon.

Key word rhizobitoxine, triazole amino acid, Huisgen cycloaddition, ethynylglycine synthon, chiral HPLC

Rhizobitoxine **1** (Figure 1) is an unusual amino acid that belongs to the β , γ -enol ether family, a γ -substituted subclass of the naturally occurring vinylglycines.¹ It has been initially regarded as a phytotoxin because it induces chlorosis in soybeans.²⁻⁴



Rhizobitoxine is a metabolic product secreted by symbiotic bacteria such as *Rhizobium japonicum* (now *Bradyrhizobium elkanii*)⁵⁻⁷ or the plant pathogen *Pseudomonas andropogonis* (now *Burkholderia andropogonis*).⁸ Rhizobitoxine, which is a structural analogue of cystathionine, inhibits two pyridoxal phosphate (PLP) dependent enzymes: cystathionine β -lyase,^{5,9} involved in the methionine biosynthesis pathway and 1-aminocyclopropane-1-carboxylate (ACC) synthase¹⁰ involved in ethylene biosynthesis in plants. It inhibits the production of ethylene,¹¹ a gaseous stress phytohormone, and plays a positive role in establishing symbiosis between *B. elkanii* and its host legume by ethylene inhibition.¹² As plant growth regulators or inhibitors of sulfur assimilation, rhizobitoxine and analogues therefore have potential applications in agronomy and biotechnology.^{13,14}

Synthesis of such unusual amino acids is a challenge, especially because of the enol ether reactivity.^{1,15,16} Therefore, there is a need for readily accessible stable structural analogues. 1,2,3-Triazole derivatives have gained a recent interest in medicinal chemistry because they are pharmacophores with good stability and high aqueous solubility,¹⁷⁻²¹ particularly in the area of peptidomimetics²² and are readily accessible by the Huisgen 1,3-dipolar cycloaddition involving an alkyne and an azide.²³⁻²⁶

In this paper, we describe the synthesis of a new triazole-containing amino acid analogue of rhizobitoxine in protected form (compound (1*S*,2*S*)-**2**) from serine (Figure 2) where the central enol ether linkage in rhizobitoxine is replaced by the robust 1,2,3-triazole linker in such a way that there is no longer β , γ -unsaturation to the amino acid moiety. This analogue should be a stable analogue compared to unstable vinylglycine derivatives and, based on reported mechanisms of inhibition, such an unusual amino acid could be a potential inhibitor of PLP-dependent enzymes.^{27,28}



Figure 2 Protected triazole containing analogue of rhizobitoxine [(15,25)-2]

Retrosynthetic analysis shows that this compound should be accessible using a Huisgen 1,3-dipolar cycloaddition of azide (S)-**3** with alkyne (S)-**4** (Figure 3).

Alkyne (S)-**4** is an 'ethynylglycine synthon'.^{29,30} It is synthesized from D-serine in six steps using Garner aldehyde (R)-**5**^{31,32} as a key precursor (Scheme 1). Two principal methods have been described to synthesize alkyne **4** from



Figure 3 Precursors of the Huisgen 1,3-dipolar cycloaddition for triazole formation in (15,25)-**2**

(S)-4

(S)-3

aldehyde **5**: the Bestmann–Ohira and the the Corey–Fuchs strategies.²⁹ We decided here to use the Bestmann–Ohira strategy using diazophosphonate **6** for this aldehyde-to-alkyne transformation, as this was well-known in our laboratory.³³ In 2002, we described the one-pot synthesis of ethynylglycine synthon **4** in 70% in 72 hours.³⁴ This procedure involving in situ formation of diazophosphonate **6** is convenient on a small scale (0.95 mmol of aldehyde **5**) but we noticed a dramatic increase of the reaction time when performed on a larger scale. Therefore, we decided to return to the original strategy³⁵ with preparation of diazophosphonate **6** prior to the homologation. After flash chromatography, the ethynylglycine synthon **4** was obtained in 83% yield on 22 mmol scale (lit.³⁵ 80% on 11 mmol scale) (Scheme 1).



Scheme 1 Reagents and conditions: (i) see ref.^{31,32}; (ii) NaH, toluene then 4-acetamidobenzenesulfonyl azide, THF, 72%, see ref.³⁶; (iii) K_2CO_3 , MeOH, 83%.

The diazophosphonate **6** was synthesized from phosphonate **7** with minor modifications of the procedure described by Pietruszka and Witt.³⁶ It was obtained in 72% yield (lit.³⁶ 77%) (Scheme 1).

Azide (*S*)-**3** was synthesized from protected L-serine (*S*)-**8**.³¹ We first envisaged synthesizing azide **3** by a nucleophilic substitution on reactive sulfonic ester derivatives of alcohol **8** (Scheme 2). The conversion of alcohol (*S*)-**8** into *p*-toluenesulfonate (*S*)-**9a** was performed using the conditions described by Jackson and Perez-Gonzales³⁷ to obtain (*S*)-**9a** in 68% yield, and the results were in agreement with the literature (lit.³⁷ 64–69%). Conversion of (*S*)-**8** into methanesulfonate (*S*)-**9b** was performed using the conditions of Shetty et al.³⁸ and allowed (*S*)-**9b** to be obtained in 64% yield after column chromatography purification (lit.³⁸ 81%, crude



Scheme 2 Reagents and conditions: (i) see ref.³¹; (ii) TsCl, Et₃N, 4-DMAP (cat.), Me₃NHCl (cat.), CH₂Cl₂, 0 °C, 2 h, 68%; (iii) MsCl, Et₃N, CH₂Cl₂, 0 °C, 0.5 h, 64%; (iv) NaN₃, DMF, see Table 1.

vield). The results of the transformation from (S)-9a and (S)-9b to (S)-3 are summarized in Table 1. The best reaction conditions in our hands for nucleophilic substitution were using sodium azide in DMF at 70 °C for a short reaction time (Table 1, entry 2). Under these reaction conditions, azide (S)-3 was obtained from p-toluenesulfonate (S)-9a in 39% vield, together with alkene **10** in 37% vield, resulting from an elimination reaction. Changing from *p*-toluenesulfonate 9a to methanesulfonate 9b, or lowering reaction temperature did not improve the vield for **3** (Table 1). We obtained enantiomer (R)-3 under the same conditions (Table 1, entry 2) through (*R*)-9a from D-serine with identical yields and opposite specific rotation. It is worth noting that Shelly et al.³⁸ reported the formation of compound (S)-**3** from methanesulfonate (S)-9b (NaN₃, DMF, 50 °C, 0.5 h; under the same conditions as Table 1, entry 4) in 56% yield but with a lower specific rotation. Moreover, Friscourt et al. reported the formation of the benzyl ester analogue of **3** (NaN₃, DMF, 40°C, 2 h) in only 18% yield.³⁹ All these observations show that this transformation is somewhat capricious.

(3) 30					
Entry	Starting material	T(°C)	Time (h)	Yield of (<i>S</i>)-3 (%)	Yield of 10 (%)
1	(S)- 9a	20	5	33	35
2	(S)- 9a	70	0.17	39	37
3	(S)- 9b	20	24	25	55
4	(S)- 9b	50	0.5	25	29

Table 1 Results of the NaN₃ Nucleophilic Substitution on (5)-9a and

^a See Scheme 2, reaction conditions (iv).

We then tried the direct formation of azide (S)-**3** using a Mitsunobu reaction as described by Stanley et al.⁴⁰ (Scheme 3).

(S)-**9h**a

В





Azide (*S*)-**3** was obtained in 41% yield from (*S*)-**8** (lit.⁴⁰ 69%) although with a cumbersome purification by column chromatography to isolate a somewhat impure material as observed on the NMR spectrum. We decided therefore to pursue the synthesis using pure compound **3** obtained following conditions described in Scheme 2,Table 1, entry 2.

To the best of our knowledge, there is only one precedent describing a Huisgen 1,3-dipolar cycloaddition using an ethynylglycine synthon as substrate. It is one example (with no further application of the product) in a methodology report of a click reaction between in situ generated β -azido styrenes from cinnamic acid using CAN/NaN₃ and alkynes to form *N*-styryl triazoles.⁴¹

The Huisgen 1,3-dipolar cycloaddition between ethynylglycine synthon (*S*)-**4** and azide (*S*)-**3** was performed using classical conditions⁴² (Scheme 4, path a): L-Ascorbate, CuSO₄ in the mixture of *tert*-butanol/water and yielded the desired 1,2,3-triazole (2*S*,4*S*)-**11** in 70% yield. Deprotection of the oxazolidine with APTS monohydrate in methanol⁴³ furnished the final compound (1*S*,2*S*)-**2** in 17% yield with 55% recovery of starting material. The overall yield from (*S*)-**4** was 12%.

In order to increase the global yield, inversion of the order of the two final steps was examined (Scheme 4, path b). Opening the oxazolidine ring in (*S*)-**4** using the same conditions as before yielded the protected amino alcohol (*S*)-**12** in 48% yield with recovery of starting material (*S*)-**4** in 32% yield. Subsequent Huisgen cycloaddition then led to the same compound (1*S*,2*S*)-**2** in 65% yield, with a 92:8 diastereomeric ratio and an enantiomeric excess higher than 99.5% (vide infra). Using this strategy, the overall yield from (*S*)-**4** increased to 31%.



Scheme 4 Reagents and conditions: (i) azide (S)-**3**, L-ascorbate (0.2 equiv), $CuSO_4$ (0.1 equiv), t-BuOH-H₂O (1:1), 70%, (path a), 65% (path b); (ii) PTSA-H₂O, MeOH, 20 °C, 2 h, 17% (path a), 48% (path b).

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The four stereoisomers of compound $\mathbf{2}$ were synthesized using the same strategy as described above from Dand L-serine with analogous results (see Supporting Information).⁴⁴

After a screening of several chiral stationary phases by HPLC, Lux-Cellulose-2, and Chiralpak AZ-H were found to be efficient for baseline separation of the mixture of the four stereoisomers of **11** and **2**, respectively, thus allowing the determination of the diastereomeric ratio and the enantiomeric excess of each isomer (Table 2). For all compounds, diastereomeric ratios were found to be greater than 90:10 and the enantiomeric excesses were higher than 96% (see Supporting Information)

Table 2 Diastereomeric Ratio and Enantiomeric Excess for Stereoisomers of 11 and 2

lsomer	dr	ee (%)	Isomer	dr	ee (%)
(2S,4R)- 11	10:1	99.5	(15,25)- 2	11:1	99.5
(2R,4S)- 11	16:1	96.2	(1 <i>R</i> ,2 <i>R</i>)- 2	10:1	99.5
(2 <i>S</i> ,4 <i>S</i>)- 11	14:1	99.5	(1 <i>S</i> ,2 <i>R</i>)- 2	14:1	99.5
(2 <i>S</i> ,4 <i>R</i>)- 11	9:1	99.5	(1 <i>R</i> ,2 <i>S</i>)- 2	9:1	97.2

^a Determined by chiral HPLC.

Deprotection of compounds **2** and biological evaluation of their activity on PLP-dependant enzymes are under investigation in our laboratory, and the results will be reported in due course.

Acknowledgment

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Supporting Information

Supporting information for this article is available online at http://dx.doi.org/10.1055/s-0036-1588300.

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- (44) General Synthetic Procedure for Click-Chemistry Reaction for the Synthesis of 2

(*S*)-Methyl 2-[(*tert*-Butoxycarbonyl)amino]-3-(4-{(*S*)-1-[(*tert*-butoxycarbonyl)amino]-2-hydroxyethyl}-1*H*-1,2,3triazol-1-yl)propanoate [(1*S*,2*S*)-2, (Scheme 4, Path a]

To a solution of (2*S*, 4*S*)-**11** (0.337 g, 0.72 mmol) in MeOH (5 mL) was added PTSA·H₂O (0.137 g, 0.72 mmol). The reaction mixture was stirred for 2 h at room temperature and sat. aq NaHCO₃ solution (40 mL) was poured into the solution. The aqueous solution was extracted with EtOAc (3 × 40 mL). The organic phases were combined, washed with sat. aq NaHCO₃ solution (40 mL), sat. aq NaCl solution (40 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by flash column chromatography (silica gel, EtOAc–PE = 0:100, increasing to 100:0, v/v) to give the desired compound (1*S*, 2*S*)-**2** (0.053 g, 17%) as a white solid and recovered starting material (2*S*, 4*S*)-**11** (0.184 g, 55%).

Analytical Data

$$\begin{split} R_f &= 0.26 \; (\text{EtOAc}); \; \text{mp } 55-57 \; ^\circ\text{C}. \; ^1\text{H NMR } (300 \; \text{MHz}, \text{CDCI}_3); \; \delta = 1.42 \; [\text{s}, 18 \; \text{H}, \text{C(CH}_{3})_3], 2.89 \; (\text{br s}, 1 \; \text{H}, \text{OH}), 3.79 \; (\text{s}, 3 \; \text{H}, \text{CO}_2\text{CH}_3), 3.84-3.87, 4.10-4.12 \; (2 \; \text{m}, 2 \; \text{H}, \text{CH}_2\text{O}), 4.70-4.86 \; (\text{m}, 4 \; \text{H}, 2 \; \text{CH}, \text{CH}_2\text{N}), 5.43 \; (\text{br s}, 1 \; \text{H}, \text{NH}), 5.59 \; (\text{br s}, 1 \; \text{H}, \text{NH}), 7.57 \; (\text{s}, 1 \; \text{H}, \text{CH}_{\text{triazole}}), \; ^{13}\text{C} \; \text{NMR } \; (75 \; \text{MHz}, \text{CDCI}_3); \; \delta = 28.4, \; 28.5 \; [2 \; \text{s}, 18 \; \text{H}, \text{C(CH}_3)_3], 48.2 \; (\text{CH}), 51.5 \; (\text{CH}_2\text{N}), 53.4 \; (\text{CO}_2\text{CH}_3), 53.9 \; (\text{CH}), 65.0 \; (\text{CH}_2\text{O}), \; 80.1 \; [\text{C(CH}_3)_3], 81.0 \; [\text{C(CH}_3)_3], 123.9 \; (\text{CH}_{\text{triazole}}), 147.1 \; (\text{C}_{\text{triazole}}), 155.2 \; (\text{NCO}_2), 155.8 \; (\text{NCO}_2), 169.5 \; (\text{CO}_2\text{CH}_3). \; [\alpha]_D^{20} \; +49.9 \; (c \; 0.91, \; \text{CHCI}_3). \; \text{HRMS } \; (\text{ES}^+): \; m/z \; [\text{M} \; + \; \text{H}]^+ \; \text{calcd for} \; \text{C}_{18}\text{H}_{32}\text{N}_5\text{O}_7: \; 430.2302; \; \text{found: } 430.2303. \; \text{HPLC: purity } = 99.6\%, \; t_{R} = 12.43 \; \text{min. IR: } 3358, 2362, 2338, 1742, 1683 \; \text{cm}^{-1}. \end{split}$$

Nitrogen inversion in the oxazolidine ring or slow interconversion of both amide or carbamate conformers of compounds **4**, **11**, and **2** causes considerable line broadening and duplication of signals in the ¹H NMR and ¹³C NMR spectra (see Supporting Information).

General Synthetic Procedure for Click-Chemistry Reaction for the Synthesis of 2

(*S*)-Methyl 2-[(*tert*-Butoxycarbonyl)amino]-3-(4-{(*S*)-1-[(*tert*-butoxycarbonyl)amino]-2-hydroxyethyl}-1*H*-1,2,3-triazol-1-yl)propanoate [(1*S*,2*S*)-2, Scheme 4, Path b)

Azide **3** (0.420 g, 1.72 mmol) and alkyne **12** (0.318 g, 1.72 mmol) were dissolved in a mixture of *t*-BuOH–H₂O (10 mL, 1:1, v/v). Sodium L-asborbate (0.068 g, 20 mol%) and CuSO₄·5H₂O (0.041 g, 10 mol%) were added. The reaction mixture was stirred at room temperature for 24 h, the solution was concentrated under vacuum and diluted with H₂O (70 mL). The aqueous phase was extracted with EtOAc (3×50 mL). The organic phases were combined, dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by flash chromatography (silica gel, EtOAc–PE = 0:100, increasing to 100:0, v/v) to give the desired compound (15,25)-**2** as a white solid (0.480 g, 65% yield). The compound exhibited the same analytical properties as described above.

See Supporting Information for the characterization data of other products.