



Pergamon

Bioorganic & Medicinal Chemistry Letters 12 (2002) 155–158

BIOORGANIC &
MEDICINAL
CHEMISTRY
LETTERS

Evaluation of Isotryptamine Derivatives at 5-HT₂ Serotonin Receptors

Jean Chang-Fong,^a James Addo,^a Małgorzata Dukat,^a Carol Smith,^b
Nicholas A. Mitchell,^b Katharine Herrick-Davis,^b Milt Teitler^b
and Richard A. Glennon^{a,*}

^aDepartment of Medicinal Chemistry, School of Pharmacy, Virginia Commonwealth University, Richmond, VA 23298, USA

^bCenter for Neuropharmacology and Neuroscience, Albany Medical College, Albany, NY 12208, USA

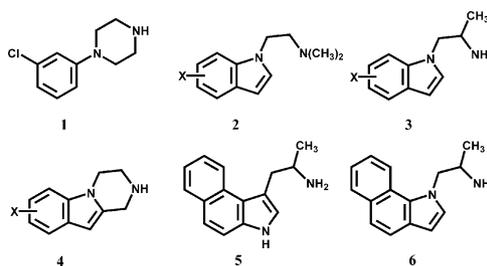
Received 26 September 2001; accepted 15 October 2001

Abstract—On the basis that *meta*-chlorophenylpiperazine (*m*CPP; **1**) is a nonselective 5-HT_{2C} agonist, that benz-fused tryptamines (e.g., **5**) display enhanced 5-HT₂ affinity, and that certain isotryptamines **3** reportedly bind with enhanced affinity and selectivity at 5-HT_{2C} receptors, we prepared and examined a series of isotryptamine-related analogues as potentially selective 5-HT_{2C} agonists. None of the compounds displayed selectivity for 5-HT_{2C} versus 5-HT_{2A} receptors. Detailed re-examination of a compound previously reported to display 100-fold 5-HT_{2C} selectivity [i.e., *S*(+)-5,6-difluoro- α -methylisotryptamine] revealed that its selectivity versus 5-HT_{2A} receptors was, at best, only 10-fold. © 2002 Elsevier Science Ltd. All rights reserved.

The identification of 5-HT_{2C}-selective serotonergic ligands—particularly 5-HT_{2C}-selective agonists—has been an elusive goal. 5-HT₂ receptors have been implicated in certain central disorders including depression, schizophrenia, anxiety, and eating disorders.^{1–3} However, agents with high affinity for 5-HT_{2C} receptors typically possess high affinity for 5-HT_{2A} receptors and rarely display >10-fold selectivity. Very few 5-HT_{2C}-selective agonists are known (e.g., refs 3 and 4). *meta*-Chlorophenylpiperazine (*m*CPP; **1**), although not necessarily selective for 5-HT₂ versus certain other populations of 5-HT receptors, is an agent that displays about 10-fold selectivity for 5-HT_{2C} receptors. Because *m*CPP behaves not as a 5-HT_{2C} agonist but as a 5-HT_{2A} antagonist,⁵ it might represent an attractive lead for the development of novel agents despite its low selectivity. In fact, others have already reported this strategy.⁶

There exists some remote structural similarity between *m*CPP (**1**) and the tryptamines.⁷ Tryptamine derivatives, though, lack selectivity for 5-HT_{2C} versus 5-HT_{2A} receptors.^{1,2} We have previously shown that isotryptamines (e.g., **2**) bind at 5-HT₂ receptors.⁸ Their subpopulation selectivity was not investigated at that time because 5-HT_{2C} receptors had not yet been identi-

fied. More recently, Bös et al.⁹ reported that certain isotryptamine derivatives (e.g., **3**) display upwards of 100-fold selectivity for 5-HT_{2C} versus 5-HT_{2A} receptors. Furthermore, certain pyrazino[1,2-*a*]indoles (e.g., **4**), which might be generally viewed as conformationally-restricted analogues of *m*CPP with an embedded isotryptamine moiety, have been identified as somewhat selective 5-HT_{2C} partial agonists.⁶



We have previously demonstrated that benz-fusion of tryptamines results in compounds with high affinity, although not necessarily enhanced selectivity, for 5-HT_{2C} receptors (e.g., **5**, 5-HT_{2C} K_i = 9 nM; 5-HT_{2A} K_i = 3 nM).¹⁰ Because it is claimed that certain isotryptamines **3** are 5-HT_{2C}-selective, the present study employed the strategy that if ring-fusion of tryptamines results in high-affinity 5-HT₂ ligands, and if the isotryptamine nucleus imparts 5-HT_{2C} selectivity, a ring-fused isotryptamine (i.e., **6**) might display enhanced 5-HT_{2C} selectivity. Also, given that pyrazino[1,2-*a*]indoles **4** possess an iso-

*Corresponding author. Tel.: +1-804-828-8487; fax: +1-804-828-7404; e-mail: glennon@hsc.vcu.edu

tryptamine moiety and are structurally related to *m*CPP (**1**), we additionally explored several ring-substituted analogues of **4** incorporating substituents that might mimic the lipophilic or electronic effects of a fused ring. In particular, we prepared the 6-, 7-, and 8-chloro and the 6-, 7-, and 8-methoxy derivatives of **4** (e.g., analogues **7**).

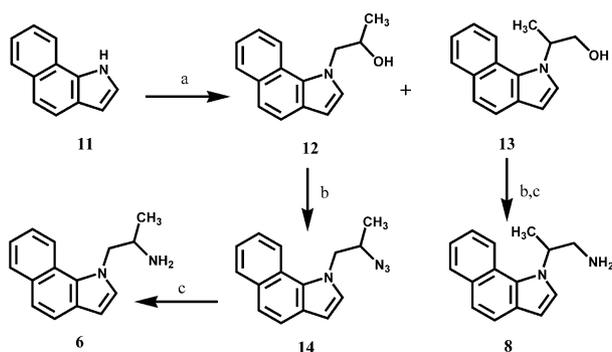
Chemistry

Ethyl benz[*g*]indole-2-carboxylate was prepared by the method of Rydon and Siddappa¹¹ except that hydrochloric acid was used in place of H₂SO₄ to effect cyclization; the ester was hydrolyzed to the acid, and the resulting acid was decarboxylated by heating at 210 °C to afford benz[*g*]indole (**11**).^{10,11}

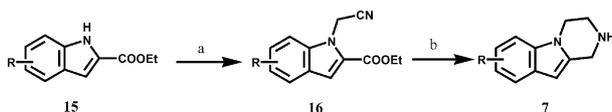
Compound **11** was allowed to react with propylene oxide to yield a 10:1 mixture of **12** and **13** which was separated by column chromatography (10% EtOAc in hexanes; silica gel) (Scheme 1). Reaction of **12** and **13** with diphenylphosphoryl azide (DPPA)/DBU gave the expected azides as oils. Catalytic reduction (10% Pd/C) of the azides afforded the target amines **6** (oxalate salt, mp 208–209 °C from MeOH/Et₂O) and **8** (0.75 oxalate, mp 123–124 °C from MeOH/Et₂O), respectively. Compound **14** was alternatively prepared in >90% yield by conversion of **12** to its mesylate, followed by reaction with sodium azide.

The pyrazino[1,2-*a*]indoles **7** were prepared by the general method of Rajur et al.¹² (Scheme 2). Compound **7h** has been reported by Bös et al.;⁶ certain of the other derivatives had also been reported¹² but their melting points were 60–100 °C lower than that found in the present study (see Table 1).

The synthesis of compound **7h** was initiated using the method of Bös et al.;⁶ however, once the necessary ethyl



Scheme 1. (a) Propylene oxide, THF, 0 °C → room temperature, 16 h; (b) DPPA/DBU, toluene, 0 °C → reflux 16 h; (c) (i) H₂, 10% Pd/C; (ii) oxalic acid.



Scheme 2. (a) (i) NaH; (ii) CICH₂CN, DMF; (b) (i) LiAlH₄/Et₂O; (ii) oxalic acid or HCl.

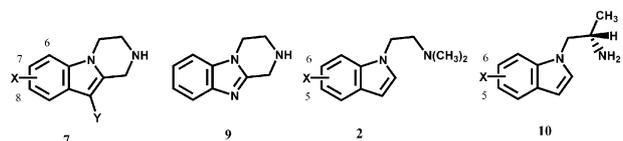
5-chloro-3-methoxyindole-2-carboxylate was obtained, the remainder of the synthesis followed the procedure shown in Scheme 1. Compounds **9**¹³ and **2a–2c**¹⁰ were prepared by literature methods. 5,6-Difluoroindole (mp 96–97 °C from hexanes), required for the synthesis of **2d** and **10**, was prepared by catalytic reduction of 3,4-difluoro-6-nitrophenylacetonitrile. The indole was elaborated to **2d** by treatment with NaH/HMPA and then 2-chloro-*N,N*-dimethylethylamine hydrochloride. Compound **10** was obtained from 5,6-difluoroindole as previously described.⁹

Results and Discussion

Compound **6** was found to bind both at 5-HT_{2A} ($K_i = 38 \pm 9$ nM) and 5-HT_{2C} ($K_i = 42 \pm 4$ nM) receptors. Although 5-HT_{2A} affinity was high, and higher than nearly all of the isotryptamines already reported,⁹ selectivity was lacking. Interestingly, however, compound **8**, a positional isomer of **6**, was found to bind with greatly reduced affinity at both populations of receptors (5-HT_{2A} $K_i = 4300 \pm 1130$ nM; 5-HT_{2C} $K_i > 10,000$ nM). The low affinity of **8** relative to **6** suggests a possible steric interaction involving the β -methyl group that alters the conformation of the side chain. The individual optical isomers of **6** and **8** should be evaluated in future studies to determine the effect of stereochemistry on binding.

With few exceptions, the pyrazino[1,2-*a*]indoles **7** possessed low affinity at 5-HT_{2A} and 5-HT_{2C} receptors (Table 1). At 5-HT_{2A} receptors, only the 7-chloro derivative **7c** displayed appreciable affinity. Because 5-HT_{2A} agonists tend to show higher affinity for sites labeled by an agonist radioligand (e.g., [³H]DOB) than by an antagonist radioligand (e.g., [³H]ketanserin), 5-HT_{2A} sites were labeled with both radioligands (designated herein as 5-HT_{2A(D)} and 5-HT_{2A(K)}, respectively, in Table 1) in order to obtain a more realistic perspective of their affinity. Although somewhat higher affinity was observed in each case (Table 1), the pyrazino[1,2-*a*]indoles still bind only with modest affinity. The pyrazino[1,2-*a*]indoles also displayed relatively low affinity at 5-HT_{2C} receptors, and showed no selectivity for 5-HT_{2C} relative to 5-HT_{2A} receptors. Compound **7h** has been previously shown by Bös et al.⁹ to be a 5-HT_{2C} partial agonist with about 8-fold selectivity for 5-HT_{2C} receptors. They have commented that the presence of the 10-methoxy group contributes to affinity and 5-HT_{2C} selectivity; the present results (comparing **7h** with **7d**) confirm their prior claim that the methoxy group plays a role in binding.⁹ Data in Table 1 also show that replacement of the 10-methoxy group by a methyl group (i.e., **7i**) is tolerated, but that replacement of C₁₀ with a nitrogen atom (i.e., **9**) decreases affinity.

In an attempt to further determine why the pyrazino[1,2-*a*]indoles bind with low selectivity, we focused on the structurally simpler isotryptamines **2**. Although compounds **2a** and **2c** bind with modest affinity, they lack 5-HT_{2C} selectivity (Table 1). There are several possible explanations for the low affinity and selectivity

Table 1. 5-HT_{2A} and 5-HT_{2C} binding data for compounds examined in this investigation


	X	Y	Mp (°C) ^a	K _i (nM) (±SEM)			5-HT _{2A} selectivity ^c			
				5-HT _{2A(K)} ^b		5-HT _{2C}				
7a	H	H	233 ^d	1040	(30)	635	(75)	4780	(390)	7.5
7b	6-Cl	H	223–225	630	(40)	330	(40)	520	(40)	1.6
7c	7-Cl	H	221–222	58	(18)	80	(10)	560	(100)	7.0
7d	8-Cl	H	230 ^e	1670	(190)	320	(50)	1460	(250)	4.6
7e	6-OMe	H	225–227	1000	(60)	300	(20)	1990	(160)	6.6
7f	7-OMe	H	204–206	5190	(800)	1300	(120)	5160	(420)	4.0
7g	8-OMe	H	238 ^f	5830	(720)	930	(20)	9930	(200)	10.7
7h	8-Cl	OMe	—	130	(10)	60	(15)	330	(50)	5.5
7i	8-Cl	Me	—	1250	(250)	— ^g	—	450	(20)	—
9	—	—	—	> 10,000	—	> 10,000	—	> 10,000	—	—
2a	H	—	—	600	(80)	650	(170)	720	(80)	1.1
2b	5-OMe	—	—	> 10,000	—	> 10,000	—	> 10,000	—	—
2c	6-OMe	—	—	1040	(100)	— ^g	—	490	(100)	—
2d	5-F,6-F	—	206–207 ^h	640	(90)	230	(60)	300	(20)	1.3
10	5-F,6-F	—	—	320	(50)	15	(2)	85	(15)	5.7

^aAll new compounds (i.e., those for which melting points are provided), and **7a**, **7d**, and **7g** were submitted for microanalysis and determined values for C, H, and N were within 0.4% of theory. Analogues **7** were recrystallized from aqueous EtOH. All compounds were isolated as their oxalate salts except for **7h** (HCl), **9** (di HCl), and **10** (fumarate).

^b5-HT_{2A(K)} and 5-HT_{2A(D)} are 5-HT_{2A} sites labeled by [³H]ketanserin and [³H]DOB, respectively.

^cSelectivity = 5-HT_{2C} K_i value/5-HT_{2A(D)} K_i value.

^dLit.¹² mp 130 °C; also reported as HCl salt.⁷

^eLit.¹² mp 162 °C.

^fPresent compound is C₁₁H₁₁ClN₂·0.75C₂H₂O₄; lit.¹² mp 180 °C for oxalate salt.

^gBinding data not obtained.

^hCompound recrystallized from MeOH.

observed with these derivatives. One of the highest affinity and most 5-HT_{2C}-selective agents reported by Bös et al.⁹ is the 5,6-difluoro analogue of **3**; perhaps the fluoro groups are important contributors to selectivity. We examined the 5,6-difluoro derivative of **2a** (i.e., **2d**), but **2d** failed to display substantially improved affinity or selectivity over **2a**. Another explanation for low affinity and lack of selectivity is that compound **3** is an α -methylisotryptamine whereas compounds **2** are *N,N*-dimethylisotryptamines. Hence, we prepared compound **10**. Compound **10** has been reported to bind at 5-HT_{2A} sites with reduced affinity (K_i ca. 100 nM) relative to 5-HT_{2C} sites (K_i ca. 1 nM) resulting in a compound with 100-fold selectivity.⁹ We found that compound **10** binds with somewhat higher affinity at 5-HT_{2A} sites (K_i = 15 nM), and with lower affinity at 5-HT_{2C} sites (K_i = 85 nM) than previously reported.

Not only did the compounds in the present study lack selectivity for 5-HT_{2C} receptors, compound **10**, which was previously reported to be selective,⁹ also failed to exhibit the expected selectivity. Hence, the results cannot be solely attributed to structure; a remaining explanation for the apparent disparity might be the assay conditions. The earlier study used human 5-HT_{2A} and 5-HT_{2C} receptors whereas the present investigation used rat 5-HT_{2A} receptors and human 5-HT_{2C} receptors. The earlier study also used [³H]DOB to label 5-HT_{2A} recep-

tors and [³H]5-HT to label 5-HT_{2C} receptors, whereas, although [³H]DOB was used herein to label 5-HT_{2A} receptors, [³H]mesulergine was employed to label 5-HT_{2C} receptors. Compound **10** was re-examined using various assay conditions (Table 2). The results show that compound **10** binds with similar affinity at rat and human 5-HT_{2A} receptors when an agonist radioligand is employed, and that there is 20-fold increase in affinity when [³H]DOB is used in place of [³H]ketanserin. Compound **10** binds at human 5-HT_{2C} receptors with about 20-fold higher affinity when agonist radioligand is employed in place of [³H]mesulergine. Nevertheless, the selectivity of compound **10** for h5-HT_{2C} receptors over h5-HT_{2A} receptors is, at best, only about 10-fold using

Table 2. Binding of compound **10** at 5-HT_{2A} and 5-HT_{2C} receptors using different assay conditions

Receptor	Species	Radioligand	K _i , nM (SEM) ^a
5-HT _{2A}	Rat	[³ H]Ketanserin	320 (±50)
	Rat	[³ H]DOB	15 (±2)
	Human	[³ H]DOB	39 (±4)
	Human	[³ H]5-HT	12 (±1)
5-HT _{2C}	Human	[³ H]Mesulergine	85 (±15)
	Human	[³ H]DOB	3.9 (±0.8)
	Human	[³ H]5-HT	3.8 (±0.5)

^aK_i values, obtained at least in triplicate, are followed by SEM. Details of the assay procedures are provided.¹⁴

assay conditions that match the literature conditions as closely as possible. The h5-HT_{2A} binding results were confirmed using a second agonist radioligand ([³H]5-HT), as were the h5-HT_{2C} results [using ([³H]DOB)]. In fact, under the latter conditions, compound **10** displayed only 3-fold selectivity for 5-HT_{2C} receptors.

In conclusion, none of the examined pyrazinoindoles nor isotryptamines displayed substantial selectivity for 5-HT_{2C} versus 5-HT_{2A} receptors. As reported by Bös et al.,⁶ the presence of a 10-methoxy substituent on the pyrazinoindole nucleus might be necessary for enhanced 5-HT_{2C} affinity. In contrast, we were unable to confirm the selective nature of compound **10**. As a consequence, caution is advised when interpreting results of pharmacological studies using **10** as a selective 5-HT_{2C} agonist.

Acknowledgements

This work was supported in part by DA 01642.

References and Notes

1. Glennon, R. A.; Dukat, M.; Westkaemper, R. B. Serotonin Receptor Subtypes and Ligands. In *Psychopharmacology, a Generation of Progress*. CD ROM Version, 1999.
2. *Serotonin Receptors and their Ligands*, Olivier, B., van Wijngaarden, I., Soudin, W., Eds. Elsevier: Amsterdam, 1997.
3. Issac, M. *Drugs Future* **2001**, *26*, 383.
4. Isaac, M.; Slassi, A.; O'Brien, A.; Edwards, L.; MacLean, N.; Bueschkens, D.; Lee, D. K. H.; McCallum, K.; De Lanoy, I.; Demchyshyn, L.; Kamboj, R. *Bioorg. Med. Chem. Lett.* **2000**, *10*, 919.
5. Kennett, G. *Curr. Opin. Invest. Drugs* **1993**, *2*, 317.
6. Bös, M.; Jenck, F.; Martin, J. R.; Moreau, J. L.; Mutel, V.; Sleight, A. J.; Widmer, U. *Eur. J. Med. Chem.* **1997**, *32*, 253.
7. Mokrosz, J.; Boksa, J.; Bojarski, A. J.; Charakchieva-Minol, S. *Med. Chem. Res.* **1993**, *3*, 240.
8. Glennon, R. A.; Jacyno, J. M.; Young, R.; McKenney, J. D.; Nelson, D. *J. Med. Chem.* **1984**, *27*, 41.
9. Bös, M.; Jenck, F.; Martin, J. R.; Moreau, J. L.; Sleight, A. J.; Wichmann, J.; Widmer, U. *J. Med. Chem.* **1997**, *40*, 2762.
10. Glennon, R. A.; Bartyzel, P.; Teitler, M. *Med. Chem. Res.* **1991**, *1*, 201.
11. Rydon, H. N.; Siddappa, S. *J. Chem. Soc.* **1951**, 2462.
12. Rajur, S. B.; Merwade, A. Y.; Hendi, S. B.; Basanagoudar, L. D. *Indian J. Chem.* **1989**, *28B*, 1065.
13. von Schmutz, J.; Kunzle, F. *Helv. Chim. Acta* **1956**, *39*, 1144.
14. **Cell culture and transfection (for human 5HT_{2A} assays):** COS-7 cells were grown in DMEM (Life Technologies) with 10% fetal bovine serum (Life Technologies) in 5% CO₂ at 37 °C and subcultured 1:12 twice a week. Twenty-four hours before transfection, cells were seeded at 80% confluence in 100-mm dishes. Cells were transfected with human 5-HT_{2A} DNA by Lipofectamine (Life Technologies). This was accomplished by combining 20 µL Lipofectamine with 5 µg of plasmid per dish. Transfections were performed in serum-free DMEM for 4.5 h at 37 °C. **Radioligand binding:** Thirty-six hours after transfection, membranes were prepared from COS-7 cells (for human 5-HT_{2A}) by scraping in 50 mM Tris-HCl, 0.5 mM EDTA, 5 mM MgCl₂, pH 7.4 (tissue buffer) and centrifugation at 10,000g for 30 min. Membranes were re-suspended in tissue buffer, homogenized and centrifuged again. For human 5-HT_{2C}, NIH3T3 cells stably transfected with human 5-HT_{2C} INI receptors were grown to confluence and prepared in the same manner as the COS-7 cells. After re-suspension in assay buffer (tissue buffer + 0.1% ascorbate), 0.5-mL membrane aliquots were added to each assay tube containing either 2 nM [³H]DOB or 0.4 nM [³H]ketanserin for 5-HT_{2A} or 2 nM [³H]5-HT or 1 nM [³H]mesulergine for 5-HT_{2C} and varying concentrations of competing drug in a final volume of 1 mL mianserin (10 µM) was used to define non-specific binding. Samples were incubated at room temperature for 30 min (5-HT_{2A}) or at 37 °C for 30 min (5-HT_{2C}), filtered through glass fiber filters (presoaked in 0.3% polyethyleneimine) on a Brandel cell harvester, and counted in Ecoscint cocktail in a liquid scintillation counter (Beckman, Berkeley, CA, USA) at 40% efficiency. **Data analyses:** Data analyses were performed using Prism software (GraphPad, San Diego, CA, USA). The Cheng-Prusoff equation was used to calculate K_i values from the IC₅₀ values.