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Piperazine sulfonamide BACE1 inhibitors: Design, synthesis, and in vivo characterization

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ABSTRACT

With collaboration between chemistry, X-ray crystallography, and molecular modeling, we designed and synthesized a series of novel piperazine sulfonamide BACE1 inhibitors. Iterative exploration of the non-prime side and S2' sub-pocket of the enzyme culminated in identification of an analog that potently lowers peripheral $A\beta_{40}$ in transgenic mice with a single subcutaneous dose.

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Alzheimer's disease (AD)¹ has become the sixth leading cause of death in the United States and is the most common form of dementia in the elderly. Despite significant research efforts in both industry and academia, there are currently no disease modifying therapies available to treat this illness. Significant evidence suggests that the pathology of AD is linked to generation of β -amyloid peptides (A $\beta_{40,42}$)² through proteolytic processing of amyloid precursor protein (APP), first by β -secretase (BACE1)³ and subsequently by γ -secretase. BACE1 in particular offers an attractive drug target,⁴ since BACE1 knockout mice are viable⁵ and produce a significant reduction in disease pathology when crossed with transgenic AD mice.⁶

As part of our peptidomimetic BACE1 inhibitor efforts, we have previously reported on a series of novel piperazinone structures **1** (Fig. 1).⁷ These cyclic amines achieved excellent BACE1 potency through a three point interaction in the active site: (1) requisite binding to the Asp32/Asp228 catalytic diad through the basic amine and hydroxyl group, (2) occupation of S2' with substituents

on the second ring nitrogen, and (3) hydrogen bonding to Thr72 NH of the flap from the ring carbonyl. X-ray crystal structures of the piperazinones bound to BACE1⁸ suggested that a similar hydrogen bond to the flap could be achieved if the ring carbonyl were



Figure 1. Initial design concepts of piperazine sulfonamide analogs 3 from earlier piperazinone series 1.

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Scheme 1. Synthesis of BACE1 inhibitor **3a** as a representative example for generation of piperazine sulfonamide analogs **3** (Ar = 3,5-difluorophenyl). Reagents and conditions: (a) LDA + **5** then **4**, $-78 \degree$ C, 41%; (b) H₂, Pd(OH)₂, HOAc, EtOH, \sim 90%; (c) BH₃·SMe₂, THF, reflux, 78%; (d) **9**, EDCI, DIEA 81%; (e) RSO₂Cl, DIEA, 60–90%; (f) TFA, DCM, \sim 80%.



Scheme 2. Synthesis of reference BACE1 inhibitors 12–14 from intermediates 10 and 11 (Ar = 3,5-difluorophenyl). Reagents: (a) TFA, ~80%; (b) PhC(O)Cl, DIEA.

deleted and an oxo moiety introduced on the substituent at the 4-position of the ring (e.g., structure **2**, Fig. 1). From this initial concept, with extensive input from molecular modeling,⁹ we designed piperazine sulfonamides **3**. The sulfonyl moiety in these analogs was intended to serve two functions. First, one of the sulfonyl oxygens would hydrogen bond with the Thr72 NH, and second, the sp³ nature of the sulfonyl itself would provide the correct trajectory for the pendant aryl group to occupy the S2' subsite.



Figure 2. Initial SAR scan¹⁴ of various piperazine core derivatives (Ar = 3,5-difluorophenyl), along with overlay of their X-ray crystal structures in BACE1. Only sulfonamide **3a** (green) simultaneously occupies S2′ and has a hydrogen bond to Thr72 NH of the flap (grey). *N*-Benzyl derivative **12** (yellow) only occupies S2′, and benzamide **13** (blue) only has the hydrogen bond to the flap.

Synthesis of these novel inhibitors began with a stereoselective Aldol addition of the conjugate base of piperazinone **5** to (*S*)- α -amino aldehyde **4**¹⁰ (Scheme 1). Adduct **6** was the major product of this reaction. The desired absolute and relative stereochemistry of the newly formed hydroxyl group was set under non-chelation control enforced by the *N*,*N*-dibenzyl groups on the amino aldehyde.^{11,12} Debenzylation of adduct **6** via palladium-mediated hydrogenation proceeded smoothly to give primary amine **7** that was then subjected to borane reduction to give piperazine **8**. The primary amine of intermediate **8** was coupled to 5-methyl-*N*,*N*-dipropylisophthalic acid (**9**)¹³ in the presence of EDCI and HOBt to give amide **10**. The piperazine *N*-benzyl group was removed with a second hydrogenation to give core **11**. Sulfonylation of this key intermediate followed by removal of the Boc group gave BACE1 inhibitors **3**, with phenylsulfonamide **3a** shown as a representative example.

In addition to sulfonamides, we also made analogous benzyl, benzamide, and N-unsubstituted analogs **12–14** (Scheme 2) for baseline comparison to sulfonamide **3a**. The BACE1 potencies¹⁴ of these four compounds, in conjunction with their X-ray crystal

structures (Fig. 2),¹⁵ clearly show the synergy of simultaneous contact with the S2' pocket and the flap. Sulfonamide **3a** is the only analog that can achieve both of these interactions and is by far the most active compound. It is 24-fold more potent than benzyl analog **12** that only occupies S2', albeit with excellent overlap. In addition, sulfonamide **3a** is 230-fold more potent than benzamide **13** that has a hydrogen bond to the flap but, due to the sp² nature of the amide linker, cannot occupy S2'.

With the piperazine sulfonamide core validated, we turned our attention to optimization of the prime and non-prime side substitution patterns of these inhibitors. On the prime side, there was a relatively steep SAR around substitution on or replacement of the S2' phenyl group, reflecting the relatively defined and rigid nature of that sub-site. In fact, most substitutions of the phenyl group caused a loss in BACE1 activity (Table 1) relative to unsubstituted phenyl **3a**. The exceptions to this general trend, with BACE1 K_i values <10 nM, were *ortho*-tolyl, *meta*-tolyl, and *meta*-chlorophenyl inhibitors, **3b**, **3c** and **3f**. The *meta*-tolyl compound **3c** in particular gave the best overall K_i and cell potency of these analogs.

Table 1

SAR of the piperazine sulfonamide S2' substituents with the N,N-dipropyl isophthalamide non-prime side binding motif (Ar = 3,5-difluorophenyl)¹⁴



Compd	R	BACE1 K _i (nM)	Cell IC ₅₀ (nM)	Compd	R	BACE1 K _i (nM)	Cell IC ₅₀ (nM)
3a	×	3	120	3m	N K	3	57
3b	×	5	64	3n	× ^N N	7	100
3c	Y CY	1	48	30	X-s	8	83
3d	Y C	60	1300	3р	X N	10	400
3e	X CI	400		3q	JON N	53	2900
3f	Y CI	7	240	3r	X S N	240	
3g	CI	42	1700	3s	N N	970	
3h	OMe	20	440	3t	N	1200	
3i	V OMe	120		3u	X N	5800	
3j	V CN	310		3v	Me	73	2300
3k	Y CI	73	1500	3w	Pr	95	1800
31	OMe OMe	45	1300				

All *para* substitution (examples **3d**, **3g**, **3i**, **3j**) produced a drop in potency of at least 10-fold, and addition of a *meta* substituent to these analogs to give 3,4-disubstitution (compounds **3k**, **3l**) did not significantly rescue the lost activity.

Further expanding the scope of S2' SAR, several heterocycles (Table 1, **3m**–**3p**) gave BACE1 K_i values ≤ 10 nM, though did not offer improvements in cell potency over *meta*-tolyl analog **3c** despite their increased polarity. A variety of other heterocycles were less well tolerated (**3q**–**3u**, BACE1 K_i >50 nM). Finally, small alkyl sulfonamides **3v** and **3w** that did not display significant lipophilicity to S2' showed only modest potency improvements (~5-fold) over unsubstituted piperazine **14**.

Using the optimized *meta*-tolyl sulfonamide piperazine core, we next examined a number of isophthalamide variations that have found wide use in the literature as non-prime side binding motifs (Table 2). In particular, methoxymethyl pyrrolidine amide $3x^{16}$ and sultam $3z^{17}$ were both very potent against BACE1 ($K_i \sim 1$ nM) with exceptional cellular activity (IC₅₀ <10 nM). While some truncation of these moieties was tolerated in vitro (e.g., analogs **3y**, **3aa**,¹⁸

Table 2

SAR of non-prime side isophthalamide variations with the *meta*-tolyl sulfonamide core (Ar = 3,5-difluorophenyl)¹⁴



Compd	R	BACE1 K _i (nM)	Cell IC ₅₀ (nM)
3c	Pr ₂ N	1	48
3x		1	7
3у		3	270
3z		1	8
3aa	N ⁻ SO ₂	1	62
3bb		1	38
3cc	O BuN	14	1350
3dd	Me	365	4350

Table 3

SAR of the sulfonamide substituents occupying the S2' subsite with the methoxymethyl pyrrolidine non-prime side binding motif (Ar = 3,5-difluorophenyl)¹⁴



Compd	R	BACE1 K _i (nM)	Cell IC ₅₀ (nM)
3ee	×	1	13
3ff	X CI	1	22
3gg	X N	2	70
3hh	X-S-CI	1	68
3ii	× N N	2	53
3jj	\mathcal{N}	1	14
3kk	c-Pr	7	100
311	Me	13	220

3bb, ¹⁹ BACE $K_i \leq 3$ nM), these modifications all resulted in at least fourfold loss of cellular potency (IC₅₀ \geq 38 nM). Further reduction in the size of the non-prime side group, as exemplified by lactam **3cc** and simple acetamide **3dd**, resulted in larger losses of both in vitro and cellular potency.

Of the two potent lead sulfonamides **3x** and **3z** identified above, the former was chosen for additional SAR development based on its superior physico-chemical properties, specifically its lower molecular weight and fewer hydrogen bond donors and acceptors.²⁰ With this optimal methoxymethyl pyrrolidine isophthalamide group on the non-prime side, re-examination of the S2' pocket showed that a wide variety of aryl sulfonamide substituents **3ee–3jj** (Table 3) gave excellent BACE1 activity ($K_i \leq 3$ nM) and good cellular potencies (IC₅₀ <70 nM). Truncated alkysulfonamides **3kk** and **3ll** were also now fairly well tolerated, as opposed to the earlier S2' SAR (c.f. analog **3ll** in Table 3 vs analog **3v** in Table 1).

Table 4

In vitro, counterscreening, and permeability profile of piperazine sulfonamide **3x** (Ar = 3,5-difluorophenyl)



Assay	
BACE1 K _i (nM)	0.8
BACE2 K _i (nM)	5.4
Cell IC ₅₀ (nM)	7.0
Cathepsin D K _i (nM)	200
Cathepsin E K _i (nM)	86
Pepsin K _i (nM)	73
Caco-2 AP-BL (efflux ratio)	1 (177)



Figure 3. Acute dose–response of piperazinone sulfonamide **3x** in 6-week-old preplaque CRND8 mice along with average compound levels measured in brain and plasma for each dose (eight animals per dose, administered subcutaneously, $A\beta_{40}$ and compound levels measured at 3 h post-dose. Changes in plasma $A\beta_{40}$ are expressed as mean ± standard error as a percent of vehicle. For additional general experimental details, see Ref. 21).

Even with this subsequent exploration of the S2' subsite, however, sulfonamide 3x (in vitro profile in Table 4) still represented the best lead in this series and was evaluated in vivo in 6-weekold, pre-plaque CRND8 mice.²¹ This potent compound reduced plasma $A\beta_{40}$ levels to background with single subcutaneous²² doses at or above 30 mg/kg (Fig. 3). Interestingly, these single doses produced an effect that was not only robust at the 3 h time point, but also persistent. The effect lasted 6 h for the 30 mg/kg dose, at least 24 h for the 100 mg/kg dose, and the 300 mg/kg dose still showed more than 90% reduction of plasma $A\beta_{40}$ at 12 h (Fig. 4). Unfortunately, there was no effect on $A\beta_{40}$ levels in the cortex of the mice at any of these doses or time points. Like many hydroxyethylamine-based peptidomimetics.^{16b,23,24} sulfonamide **3x** is a substrate for P-gp (Caco-2 efflux ratio >100). Despite this high ratio, subcutaneous dosing generated modest levels of compound in the brain (Fig. 3). With mouse plasma protein binding of approximately 95%, however, the fraction of compound unbound in the brain is only at or slightly above the cellular IC_{50} , likely accounting for lack of efficacy in that compartment.



Figure 4. Time course of changes in plasma $A\beta_{40}$ following a single dose of piperazinone sulfonamide **3x** in 6-week-old pre-plaque CRND8 mice (three to eight animals per dose, compound administered subcutaneously, $A\beta_{40}$ levels for each dose were measured at time points shown. Changes in $A\beta_{40}$ levels are normalized relative to vehicle (100%) and are expressed as mean ± standard error. For additional general experimental details, see Ref. 21).

In conclusion, using an iterative structure-based approach, we have designed a series of novel peptidomimetic piperazine sulfonamide BACE1 inhibitors. The sulfonamide portion of these structures achieves both occupancy of S2' and capture of a key hydrogen bond to the flap of the active site. These interactions, in conjunction with optimal non-prime side binding motifs, gave compounds with excellent BACE1 potency, both in vitro and in cells. In addition, analog **3x** produced a robust and persistent lowering of peripheral A β in a transgenic mouse model following a single subcutaneous dose. However, these compounds, as with many peptidomimetics, are substrates for P-pg, and this liability limits their oral bioavailability, brain penetration, and ultimately their central efficacy. Based on these observations, we have directed significant effort toward identification of non-peptidic BACE1 inhibitors, and results will be reported in due course.²⁵

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References and notes

- (a) An, Y.; Zhang, C.; He, S.; Yao, C.; Zhang, L.; Zhang, Q. Life Sci. J. 2008, 5, 1; (b) Herz, J. Neuron 2007, 53, 477; (c) Nguyen, J.; Yamani, A.; Kiso, Y. Curr. Pharm. Des. 2006, 12, 4295; (d) Zimmermann, M.; Gardoni, F.; Di Luca, M. Drugs Aging 2005, 22, 27; (e) Citron, M. Nat. Rev. Neurosci. 2004, 5, 677; (f) For excellent online references to general information and statistics about Alzheimer's disease, as well as ongoing research in the field, see www.alz.org and www.ahaf.org.
- 2 For examples of genetic, biochemical, and clinical evidence linking Aβ and AD, see: (a) Klyubin, I.; Betts, V.; Welzel, A. T.; Blennow, K.; Zetterberg, H.; Wallin, A.; Lemere, C. A.; Cullen, W. K.; Peng, Y.; Wisniewski, T.; Selkoe, D. J.; Anwyl, R.; Walsh, D. M.; Rowan, M. J. J. Neurosci. 2008, 28, 4231; (b) George-Hyslop, P. H., St. Biol. Psychiatry 2000, 47, 183; (c) Patterson, D.; Gardiner, K.; Kao, F. T.; Tanzi, R.; Watkins, P.; Gusella, J. F. Proc. Natl. Acad. Sci. U.S.A. 1988, 85, 8266; (d) Chishti, M. A.; Yang, D. S.; Janus, C.; Phinney, A. L.; Horne, P.; Pearson, J.; Strome, R.; Zuke, N.; Loukides, J.; French, J.; Turner, S.; Lozza, G.; Grilli, M.; Kunicki, S.; Morissette, C.; Paquette, J.; Gervais, F.; Bergeron, C.; Fraser, P.; Carlson, G.; George-Hyslop, P., St.; Westaway, D. J. Biol. Chem. 2001, 276, 21562; (e) Hock, C.; Konietzko, U.; Streffer, J. R.; Tracy, J.; Signorell, A.; Muller-Tillmanns, B.; Lemke, U.; Henke, K.; Moritz, E.; Garcia, E.; Wollmer, M. A.; Umbricht, D.; de Quervain, D. J.; Hofmann, M.; Maddalena, A.; Papassotiropoulos, A.; Nitsch, R. M. Neuron 2003, 38, 547; (f) Glenner, G. G.; Wong, C. W. Biochem. Biophys. Res. Commun. 1984, 120, 885
- For an overviews of BACE1 biology, see: (a) Vassar, R. J. Mol. Neurosci. 2004, 23, 105; (b) Cummings, J. L. N. Eng. J. Med. 2004, 351, 56.
- For reviews of BACE1 inhibitor research, see: (a) Hamada, Y.; Kiso, Y. Expert Opin. Drug Discovery 2009, 4, 391; (b) Huang, W.-H.; Sheng, R.; Hu, Y.-Z. Curr. Med. Chem. 2009, 16, 1806; (c) Ghosh, A. K.; Bilcer, G.; Hong, L.; Koelsch, G.; Tang, J. Curr. Alzheimer Res. 2007, 4, 418; (d) Durham, T. B.; Shepherd, T. A. Curr. Opin. Drug Discovery Dev. 2006, 9, 776; (e) Guo, T.; Hobbs, D. W. Curr. Med. Chem. 2006, 13, 1811; (f) Thompson, L. A.; Bronson, J. J.; Zusi, F. C. Curr. Pharm. Des. 2005, 11, 3383; (g) Cumming, J. N.; Iserloh, U.; Kennedy, M. E. Curr. Opin. Drug Discovery Dev. 2004, 7, 536.
- Modest developmental phenotypes in BACE1 knockout mice have been identified: (a) Willem, M.; Garratt, A. N.; Novak, B.; Citron, M.; Kaufmann, S.; Rittger, A.; DeStrooper, B.; Saftig, P.; Birchmeier, C.; Haass, C. Science 2006, 314, 664; (b) Dominguez, D.; Tournoy, J.; Hartmann, D.; Huth, T.; Cryns, K.; Deforce, S.; Serneels, L.; Camacho, I. E.; Marjaux, E.; Craessaerts, K.; Roebroek, A. J. M.; Schwake, M.; D'Hooge, R.; Bach, P.; Kalinke, U.; Moechars, D.; Alzheimer, C.; Reiss, K.; Saftig, P.; De Strooper, B. J. Biol. Chem. 2005, 280, 30797.
- McConlogue, L.; Buttini, M.; Anderson, J. P.; Brigham, E. F.; Chen, K. S.; Freedman, S. B.; Games, D.; Johnson-Wood, K.; Lee, M.; Zeller, M.; Liu, W.; Motter, R.; Sinha, S. J. Biol. Chem. 2007, 282, 26326.
- Cumming, J. N.; Le, T. X.; Babu, S.; Carroll, C.; Chen, X.; Favreau, L.; Gaspari, P.; Guo, T.; Hobbs, D. W.; Huang, Y.; Iserloh, U.; Kennedy, M. E.; Kuvelkar, R.; Li, G.; Lowrie, J.; McHugh, N. A.; Ozgur, L.; Pan, J.; Parker, E. M.; Saionz, K.; Stamford, A. W.; Strickland, C.; Tadesse, D.; Voigt, J.; Wang, L.; Wu, Y.; Zhang, L.; Zhang, Q. Bioorg. Med. Chem. Lett. 2008, 18, 3236.
- As a representative example, coordinates for the X-ray structure of compound 1 complexed with BACE1 have been deposited in the Protein Data Bank (www.rcsb.org), and can be accessed under PDB ID 2QP8.

- We employed both the de novo design method Allegrow (v. 020802, Boston De Novo, Boston, MA) as well as minimization using the Polak–Ribiere conjugated gradient algorithm until convergence of 0.001, CFF91 force field, Insight 2000/ CDISCOVER (Accelrys Inc., San Diego, CA).
- Prepared from commercially available Boc-3,5-difluorophenylalanine: Reetz, M. T.; Drewes, M. W.; Schwickardi, R. Org. Synth. 1999, 76, 110.
- (a) Reetz, M. T.; Drewes, M. W.; Schmitz, A. Angew. Chem., Int. Ed. Engl. 1987, 26, 1141; For an extensive review of this chemistry, see: (b) Reetz, M. T. Chem. Rev. 1999, 99, 1121.
- 12. Diastereomeric ratios at the stereocenter formed on the piperazinone ring typically ranged from 5:4 to 3:2, always in favor of the desired isomer. The minor component was easily separable by silica gel chromatography to cleanly afford adduct **6**.
- Maillaird, M.; Hom, R.; Gailunas, A.; Jagodzinska, B.; Fang, L. Y.; John, V.; Freskos, J. N.; Pulley, S. R.; Beck, J. P.; Tenbrink, R. E. PCT Int. Appl. WO 2002002512.
- 14. Inhibition of BACE1 in vitro was determined using an APP-derived peptide containing the Swedish mutant: (a) Kennedy, M. E.; Wang, W.; Song, L.; Lee, J.; Zhang, L.; Wong, G.; Wang, L.; Parker, E. *Anal. Biochem.* **2003**, 319, 49; Cellular IC₅₀ values for inhibition of A β_{40} production were determined by incubating HEK293 cells, stably transfected with the human APP cDNA containing both Swedish and London FAD mutations, with increasing concentrations of BACE inhibitors. A β_{40} levels were measured in the cell culture media using an A β_{1-40} specific ELISA assay: (b) Zhang, L; Song, L; Terracina, G.; Liu, Y.; Pramanik, B.; Parker, E. *Biochemistry* **2001**, *40*, 5049.
- Coordinates for the X-ray structures of compounds 3a, 12, and 13 complexed with BACE1 have been deposited in the Protein Data Bank (www.rcsb.org), and can be accessed under PDB ID 3LPI, 3LPJ, and 3LNK, respectively.
- (a) Varghese, J.; Maillard, M.; Jagodzinska, B.; Beck, J. P.; Gailunas, A.; Fang, L.; Sealy, J.; Tenbrink, R.; Freskos, J.; Mickelson, J.; Samala, L.; Hom, R. PCT Int. Appl. WO 2003040096.; (b) Iserloh, U.; Pan, J.; Stamford, A. W.; Kennedy, M. E.; Zhang, Q.; Zhang, L.; Parker, E. M.; McHugh, N. A.; Favreau, L.; Strickland, C.; Voigt, J. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 418; (c) Thompson, L. A.; Boy, K., M.; Shi, J.; Macor, J. E. PCT Int. Appl. WO 2006099352.
- (a) Barrow, J. C.; Coburn, C. A.; Nantermet, P. G.; Selnick, H. G.; Stachel, S. J.; Stanton, M. G.; Stauffer, S. R.; Zhuang, L.; Davis, J. R. PCT Int. Appl. WO 2005065195.; (b) Eickmeier, C.; Fuchs, K.; Peters, S.; Dorner-Ciossek, C.; Heine, N.; Handschuh, S.; Klinder, K.; Kostka, M. PCT Int. Appl. WO 2006103038.; (c) Chirapu, S. R.; Pachaiyappan, B.; Nural, H. F.; Cheng, X.; Yuan, H.; Lankin, D. C.; Abdul-Hay, S. O.; Thatcher, G. R. J.; Shen, Y.; Kozikowski, A. P.; Petukhov, P. A. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 264.
- (a) Beswick, P.; Charrier, N.; Clarke, B.; Demont, E.; Dingwall, C.; Dunsdon, R.; Faller, A.; Gleave, R.; Hawkins, J.; Hussain, I.; Johnson, C. N.; MacPherson, D.; Maile, G.; Matico, R.; Milner, P.; Mosley, J.; Naylor, A.; O'Brien, A.; Redshaw, S.; Riddell, D.; Rowland, P.; Skidmore, J.; Soleil, V.; Smith, K. J.; Stanway, S.; Stemp, G.; Stuart, A.; Sweitze, S.; Theobald, P.; Vesey, D.; Walter, D. S.; Ward, J.; Wayne, G. Bioorg. Med. Chem. Lett. **2008**, *18*, 1022; (b) Clarke, B.; Demont, E.; Dingwall, C.; Dunsdon, R.; Faller, A.; Hawkins, J.; Hussain, I.; MacPherson, D.; Maile, G.; Matico, R.; Milner, P.; Mosley, J.; Naylor, A.; O'Brien, A.; Redshaw, S.; Riddell, D.; Rowland, P.; Soleil, V.; Smith, K. J.; Stanway, S.; Stemp, G.; Sweitzer, S.; Theobald, P.; Vesey, D.; Walter, D. S.; Ward, J.; Wayne, G. Bioorg. Med. Chem. Lett. **2008**, *18*, 1017; (c) Hussain, I.; Hawkins, J.; Harisson, D.; Hille, C.; Wayne,

G.; Cutler, L.; Buck, T.; Walter, D.; Demont, E.; Howes, C.; Naylor, A.; Jeffrey, P.; Gonzalez, M. I.; Dingwall, C.; Michel, A.; Redshaw, S.; Davis, J. B. *J. Neurochem.* **2007**, *100*, 802; (d) Demont, E. H.; Faller, A.; MacPherson, D. T.; Milner, P. H.; Naylor, A.; Redshaw, S.; Stanway, S. J.; Vesey, D. R.; Walter, D. S PCT Int. Appl. WO 2004050619.

- (a) Coburn, C. A.; Stachel, S. J.; Li, Y.-M.; Rush, D. M.; Steele, T. G.; Chen-Dodson, E.; Holloway, M. K.; Xu, M.; Huang, Q.; Lai, M.-T.; DiMuzio, J.; Crouthamel, M.-C.; Shi, X.-P.; Sardana, V.; Chen, Z.; Munshi, S.; Kuo, L.; Makara, G. M.; Annis, D. A.; Tadikonda, P. K.; Nash, H. M.; Vacca, J. P. *J. Med. Chem.* **2004**, *47*, 6117; (b) Stachel, S. J.; Coburn, C. A.; Steele, T. G.; Jones, K. G.; Loutzenhiser, E. F.; Gregro, A. R.; Rajapakse, H. A.; Lai, M.-T.; Crouthamel, M.-C.; Xu, M.; Tugusheva, K.; Lineberger, J. E.; Pietrak, B. L.; Espeseth, A. S.; Shi, X.-P.; Chen-Dodson, E.; Holloway, M. K.; Munshi, S.; Simon, A. J.; Kuo, L.; Vacca, J. P. *J. Med. Chem.* **2004**, *47*, 6447.
- Coordinates for the X-ray structure of compound 3x complexed with BACE1 have been deposited in the Protein Data Bank (www.rcsb.org), and can be accessed under PDB ID 3LPK.
- 21. CRND8-APP mice are models for early onset (familial) AD that express human APP containing both Swedish and London mutations that enhance the rate of APP cleavage by BACE1 and favor production of A β_{42} over A β_{40} in the γ -secretase cleavage step: Hyde, L. A.; Kazdoba, T. M.; Grilli, M.; Lozza, G.; Brussa, R.; Zhang, Q.; Wong, G. T.; McCool, M. F.; Zhang, L.; Parker, E. M.; Higgins, G. A. Behav. Brain Res. **2005**, *160*, 344.
- 22. Subceutaneous dosing was chosen due to the poor oral bioavailability of lead compound 3x: rapid rat AUC_{0-6h} (10 mg/kg, PO) = 0 nM h.
- (a) Meredith, J. E., Jr.; Thompson, L. A.; Toyn, J. H.; Marcin, L.; Barten, D. M.; Marcinkeviciene, J.; Kopcho, L.; Kim, Y.; Lin, A.; Guss, V.; Burton, C.; Iben, L.; Polson, C.; Cantone, J.; Ford, M.; Drexler, D.; Fiedler, T.; Lentz, K. A.; Grace, J. E., Jr.; Kolb, J.; Corsa, J.; Pierdomenico, M.; Jones, K.; Olson, R. E.; Macor, J. E.; Albright, C. F. *J. Pharmacol. Exp. Ther.* **2008**, *326*, 502; (b) Stanton, M. G.; Stauffer, S. R.; Gregro, A. R.; Steinbeiser, M.; Nantermet, P.; Sankaranarayanan, S.; Price, E. A.; Wu, G.; Crouthamel, M.; Ellis, J.; Lai, M.; Espeseth, A. S.; Shi, X.; Jin, L.; Colussi, D.; Pietrak, B.; Huang, Q.; Xu, M.; Simon, A. J.; Graham, S. L.; Vacca, J. P.; Selnick, H. *J. Med. Chem.* **2007**, *50*, 3431.
- 24. PK/PD data for BACE1 inhibition in a rhesus monkey model were recently reported for a fully de-peptidized transition state isostere with low susceptibility to P-gp transport: Sankaranarayanan, S.; Holahan, M. A.; Colussi, D.; Crouthamel, M.-C.; Devanarayan, V.; Ellis, J.; Espeseth, A.; Gates, A. T.; Graham, S. L.; Gregro, A. R.; Hazuda, D.; Hochman, J. H.; Holloway, K.; Jin, L.; Kahana, J.; Lai, M.-t.; Lineberger, J.; McGaughey, G.; Moore, K. P.; Nantermet, P.; Pietrak, B.; Price, E. A.; Rajapakse, H.; Stauffer, S.; Steinbeiser, M. A.; Seabrook, G.; Selnick, H. G.; Shi, X.-P.; Stanton, M. G.; Swestock, J.; Tugusheva, K.; Tyler, K. X.; Vacca, J. P.; Wong, J.; Wu, G.; Xu, M.; Cook, J. J.; Simon, A. J. J. Pharmacol. Exp. Ther. 2009, 328, 131.
- (a) Zhu, Z.; Sun, Z.-Y.; Ye, Y.; Voigt, J.; Strickland, C.; Smith, E. M.; Cumming, J.; Wang, L.; Wong, J.; Wang, Y.-S.; Wyss, D. F.; Chen, X.; Kuvelkar, R.; Kennedy, M. E.; Favreau, L.; Parker, E.; McKittrick, B. A.; Stamford, A.; Czarniecki, M.; Greenlee, W.; Hunter, J. C. J. Med. Chem. 2010, 53, 951; (b) Wang, Y.-S.; Strickland, C.; Voigt, J.; Kennedy, M. E.; Beyer, B. M.; Senior, M. M.; Smith, E. M.; Nechuta, T.; Madison, V.; Czarniecki, M.; McKittrick, B. A.; Stamford, A.; Parker, E.; Hunter, J.; Greenlee, W.; Wyss, D. F. J. Med. Chem. 2010, 53, 942.