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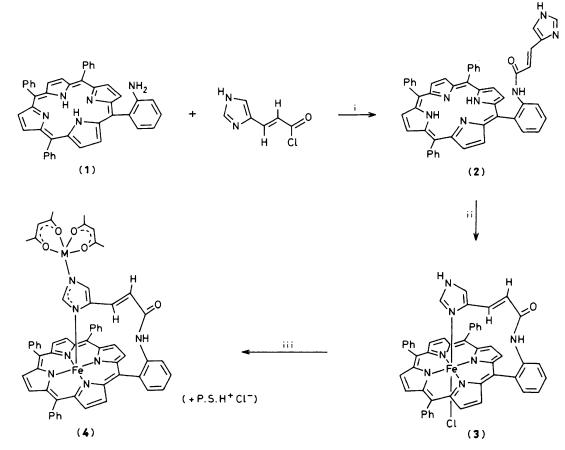
Cytochrome c Oxidase Models. A μ-Imidazolato Complex of Copper(") and Iron(") Derived from an Appended-tail Porphyrin

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An appended-tail Fe^{III} porphyrin complex derived from mono-NH₂TPPH₂ (TPPH₂=tetraphenylporphyrin) and urocanic acid chloride has been prepared and used as a precursor for synthesizing a μ -imidazolato binuclear species of Fe^{III} and Cu^{II} as a cytochrome c oxidase model compound.

Cytochrome c oxidase contains two heme and two copper sites and is the terminal enzyme in oxidative phosphorylation, catalysing a 4e⁻ reduction of O₂ to H₂O (O₂ + 4H⁺ + 4e⁻ \rightarrow 2H₂O) with the concomitant release of energy which is stored in the ATP-ADP cycle.¹ Much recent interest in the enzyme has focused on the nature and structure of the active site which appears as binuclear in copper (Cu_u) and heme (Cyt.a₃). Furthermore, in the fully oxidized or resting state, this binuclear [Cu²⁺_u/Cyt.a₃³⁺] centre exhibits magnetic and e.s.r. (silent) properties consistent with an S = 2 ground state, arising presumably from strong antiferromagnetic coupling $(-J \ge 200 \text{ cm}^{-1})$ between the S = 1/2 (Cu²⁺_u) and S = 5/2(Cyt.a₃³⁺) centres.^{2,3} Biochemically feasible bridges, *e.g.*, [Cu²⁺_u-(B)-Cyt.a₃³⁺], such as B = imidazolate (imid⁻) from histidine,⁴ oxo from O₂ or tyrosine,⁵⁻⁷ or mercapto from cysteine,^{8,9} have all been suggested as possible mediators of the 'strong' magnetic exchange interaction and several such $[Cu^{2+}-(B)-Fe^{3+}]$ model compounds have been reported by others¹⁰⁻¹⁵ and ourselves.^{6,16-18} In this communication, we report a B = imid⁻ model compound where a novel porphyrin ligand, UroTPPH₂ (TPPH₂ = tetraphenylporphyrin) [(2) in Scheme 1], has been synthesized from mono-NH₂TPPH₂ (1) and urocanic acid chloride [3-(1*H*-imidazol-4-yl)prop-2-enoyl chloride]. The UroTPPH₂ ligand is unusual among appendedtail porphyrins in possessing a site of unsaturation in the tail (lending rigidity) and a terminal imidazole moiety joined at C-4 rather than through N-1. The former property was selected to encourage S = 5/2 Fe³⁺ (short, rigid tail) to mimic the Cyt.a₃³⁺ spin state, and the latter to permit imid⁻ bridging. In general, both properties have been realized,



Scheme 1. Synthesis of (3) and (4). Reagents and conditions: i, DMF, 35 °C, pyridine, 24 h, N₂; ii, FeCl₂, DMF, reflux, 30 min; iii, Cu(acac)₂, Proton Sponge (P.S.), CH₂Cl₂, reflux, 6 h.

permitting a thoroughly characterized $[Fe_{hs}^{3+}(imid)Cu^{2+}]$ (hs = high spin) species to be isolated as an analytically pure material and studied for the first time in both the solid and solution states as a model compound for the oxidase active site.

Scheme 1 illustrates the synthesis of the metal-free ligand, UroTPPH₂ (2), the precursor iron(III) complex, $[Fe^{III}]$ -(UroTPP)Cl] (3), and the subsequent reaction with a copper centre [in this case the bis(acetylacetonato) (accac) complex] to form the binuclear complex (4). Unlike many reactions of acid chlorides with primary amines, reaction between urocanic acid chloride and (1) does not proceed readily at room temperature, but rather requires prolonged stirring under dry N₂ (35 °C for 24 h) to afford complete reaction. In addition, porphyrins are known to photosensitize the production of singlet dioxygen,¹⁹ and reactions involving the attached tail-base were therefore performed in the dark or under dioxygen-free conditions since imidazole moieties are degraded by ${}^{1}O_{2}$.²⁰ Thus, (2) is susceptible to photochemical decomposition while dispersed on a silica gel column, and all column work was also performed excluding light. A typical preparatory reaction for (2) involved stirring 1 mol. equiv. of the amine with 5 mol. equiv. of urocanic acid chloride (from the acid and SOCl₂ in dimethylformamide, DMF) in the presence of excess of pyridine. UroTPPH₂ was then separated from the reaction mixture by chromatography on a silica gel column. Refluxing (2) in DMF with anhydrous $FeCl_2$ gave (3). Although the metallated complex is less photosensitive, all manipulations were nevertheless also performed in the dark. Purification by silica gel column chromatography and recrystallization from toluene-hexane afforded (3) with an S = 5/2

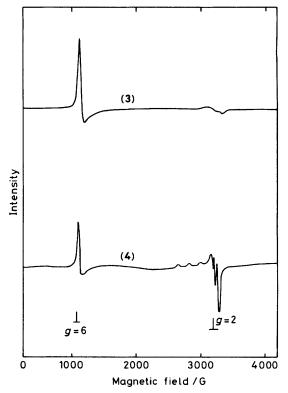


Figure 1. E.s.r. spectra of (3) and (4) at 10 K in CH_2Cl_2 glasses (1 G = 10^{-4} T).

Fe^{III} centre (*vide infra*). Finally, addition of 1 mol. equiv. of [1,2-bis(dimethylamino)naphthalene] (Proton Sponge) or Bu'O⁻K⁺ and a 1.1 molar excess of [Cu(acac)₂] to (**3**) in CH₂Cl₂ under reflux gave the appropriate imidazolate binuclear complex (**4**). In a typical preparation of (**4**), a solution of (**3**) (0.18 mmol), [Cu(acac)₂] (0.20 mmol), and Proton Sponge (0.18 mmol) in CH₂Cl₂ (40 ml) was refluxed for 6 h. The solvent was then removed under reduced pressure and the resulting solid washed thoroughly with benzene to remove any unreacted [Cu(acac)₂]. The solid was then recrystallized from CH₂Cl₂-heptane, filtered off, and dried over P₂O₅ at room temperature for 12 h. Elemental analyses (C, H, N, Cl, metal) were consistent with the molecular formulations for (**3**) and (**4**) shown in Scheme 1.

With μ_{eff} (298 K, solid) = 5.4 μ_B and the frozen-solution e.s.r. spectrum shown in Figure 1, (3) displays properties consistent with an Fe^{III} S = 5/2 species in both the solid and solution states. Whether the imidazole terminus in (3) is co-ordinated to Fe^{III}, as shown in Scheme 1, is not indicated by these results, but space-filling molecular models indicate no special problem in accommodating the porphyrin tail in an axial position when the ethylenic hydrogens are in their *trans* configuration. The presence of a binuclear [Fe^{III}(imid)Cu^{II}] centre in (4) is more certain since base is required for adduct formation of (3) with [Cu(acac)₂], conductivity data in toluene or CH₂Cl₂ have established the non-electrolyte behaviour of (4), and, finally, a molecular weight determination of (4) by osmometry in CH₂Cl₂ yielded a value of *ca.* 1030 (1066 calc.).

Compound (4) exhibits nearly Curie-Weiss magnetic susceptibility behaviour between 20 and 300 K, with $\theta = ca. - 2.0$ K. Over this temperature range, μ_{eff} varies only between 5.4 μ_B (20 K) and 5.67 μ_B (298 K). Using a value of $\mu_{eff.}(298~K)$ = 5.4 μ_B for the Fe^{III} centre, as found for (3), and assuming μ_{eff} . = 2.0 μ_B for Cu^{II}, a room temperature value of *ca*. 5.8 μ_B can be calculated for an [Fe^{III}(imid)Cu^{II}] centre in (4), assuming magnetically non-interacting Fe^{III} and Cu^{II} ion centres. This value compares well with the experimentally determined value of 5.67 μ_B (298 K). Moreover, the e.s.r. spectrum of (4) in a frozen CH₂Cl₂ glass at 10 K displays spectral characteristics consistent with the presence of magnetically dilute S =5/2 Fe^{III} centres [g = 6, 2 (weak)] and S = 1/2 Cu^{II} centres (g ca. 2) in a 1:1 ratio. The well resolved four-line hyperfine pattern on the g ca. 2 signal establishes that the signal arises from Cu^{II}, and the narrow line width of the signal is consistent with a magnetically isolated Cu^{II} centre. Thus, the magnetic and e.s.r. properties of this appended-tail µ-imidazolato porphyrin species, with essentially non-interacting Fe^{III} and Cu^{II} centres, do not resemble those of the strongly coupled $[Cu_n^{2+}/Cyt.a_3^{3+}]$ pair in oxidase. This conclusion is similar to other reports for [Fe^{III}_{hs}(2-Meimid)Cu^{II}],¹⁰ [F^{III}_{hs}(imid)Cu^{II}]¹⁶ (ls = low spin), and [Fe^{III}_{hs}(imid)Co^{II}_{ls}],²¹ species where -J is \ll

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 200 cm^{-1} across imidazolate, but contrary to conclusions for at least one other [Fe^{III}(imid)Cu^{II}] trifluoromethane sulphonate complex¹⁶ that could only be studied in the solid state. Nevertheless, accumulating evidence suggests that imidazolate cannot, in general, foster as strong a magnetic interaction as apparently observed for the active site of resting cytochrome c oxidase.

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References

- 1 A. L. Lehninger, 'The Mitrochondrion,' Benjamin, New York, 1973.
- 2 M. F. Tweedle, L. J. Wilson, L. Garcia-Iniguez, G. T. Babcock, and G. Palmer, J. Biol. Chem., 1978, 253, 8065.
- 3 T. H. Moss, E. Shapiro, T. E. King, H. Beinert, and C. R. Hartzell, J. Biol. Chem., 1978, 253, 8072.
- 4 G. Palmer, G. T. Babcock, and L. E. Vickery, *Proc. Natl. Acad. Sci. USA*, 1973, **73**, 2006.
- 5 C. A. Reed and J. T. Landrum, FEBS Lett., 1979, 106, 265.
- 6 R. H. Petty, B. R. Welch, L. J. Wilson, L. A. Bottomley, and K. M. Kadish, J. Am. Chem. Soc., 1980, 102, 611.
- 7 R. W. Shaw, J. E. Rif, M. H. O'Leary, and H. Beinert, J. Biol. Chem., 1980, 256, 1105.
- 8 B. Chance and L. Powers, Biophys. J., 1981, 33, 95.
- 9 L. Powers, B. Chance, Y. Ching, and P. Angiolillo, *Biophys. J.*, 1981, **33**, 95a.
- 10 T. Prosperi and A. A. G. Tomlinson, J. Chem. Soc., Chem. Commun., 1979, 196.
- 11 D. Kovacs and R. E. Shepherd, J. Inorg. Biochem., 1979, 10, 67.
- 12 M. J. Gunter, L. N. Mander, G. M. McLaughlin, K. S. Murry, K. J. Berry, P. E. Clark, and D. A. Buckingham, J. Am. Chem. Soc., 1980, 102, 1470; M. J. Gunter, L. N. Mander, K. S. Murry, and P. E. Clark, *ibid.*, 1981, 103, 6784.
- 13 C. K. Chang, M. S. Koo, and B. Ward, J. Chem. Soc., Chem. Commun., 1982, 716.
- 14 M. Okawa, W. Kanda, and S. Kida, Chem. Lett., 1980, 1281; J. Jaud, Y. Journaux, J. Galy, and O. Kahn, Nouveau J. Chim., 1980, 4, 629.
- 15 M. C. Elliott and K. Akabori, J. Am. Chem. Soc., 1982, 104, 2671.
- 16 S. E. Dessens, C. L. Merrill, R. J. Saxton, R. L. Ilaria, Jr., J. W. Lindsey, and L. J. Wilson, J. Am. Chem. Soc., 1982, 104, 4357.
- 17 R. J. Saxton, L. W. Olsen, and L. J. Wilson, J. Chem. Soc., Chem. Commun., 1982, 984.
- 18 L. J. Wilson, V. Chunplang, B. K. Lemke, C. L. Merrill, R. J. Saxton, and M. L. Watson, *Inorg. Chem. Acta Bioinorg.*, 1983, 79, 107.
- 19 M. D. Maines and A. Kappos, J. Biol. Chem., 1975, 250, 2363, and references therein.
- 20 H. H. Wasserman, K. Stiller, and M. B. Floyd, *Tetrahedron Lett.*, 1968, 3277.
- 21 J. T. Landrum, C. A. Reed, K. Hatano, and W. R. Scheidt, J. Am. Chem. Soc., 1978, 100, 3232.