

## Fijiolides A and B, Inhibitors of TNF- $\alpha$ -Induced NF $\kappa$ B Activation, from a Marine-Derived Sediment Bacterium of the Genus *Nocardiopsis*

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Fijiolide A, a potent inhibitor of TNF- $\alpha$ -induced NF $\kappa$ B activation, along with fijiolide B, were isolated from a marine-derived bacterium of the genus *Nocardiopsis*. The planar structures of fijiolides A (**1**) and B (**2**) were elucidated by interpretation of 2D NMR spectroscopic data, while the absolute configurations of these compounds were defined by interpretation of circular dichroism and 2D NMR data combined with application of the advanced Mosher's method. Fijiolides A and B are related to several recently isolated chloroaromatic compounds, which appear to be the Bergman cyclization products of enediyne precursors. Fijiolide A reduced TNF- $\alpha$ -induced NF $\kappa$ B activation by 70.3%, with an IC<sub>50</sub> value of 0.57  $\mu$ M. Fijiolide B demonstrated less inhibition, only 46.5%, without dose dependence. The same pattern was also observed with quinone reductase (QR) activity: fijiolide A was found to induce quinone reductase-1 (QR1) with an induction ratio of 3.5 at a concentration of 20  $\mu$ g/mL (28.4  $\mu$ M). The concentration required to double the activity was 1.8  $\mu$ M. Fijiolide B did not affect QR1 activity, indicating the importance of the nitrogen substitution pattern for biological activity. On the basis of these data, fijiolide A is viewed as a promising lead for more advanced anticancer testing.

Nature is an important resource for the discovery of anticancer drugs. The relevance of the marine environment as a source of novel anticancer compounds has been validated by the discovery of ca. 2500 new metabolites with antiproliferative activity.<sup>1</sup> Bacteria belonging to the order Actinomycetales, commonly known as actinomycetes, have provided nearly 50% of the bioactive microbial natural products discovered as of 2002.<sup>2</sup> Although most microbial small molecule discovery efforts have focused on actinomycetes from terrestrial environments, marine-derived actinomycetes are proving to be an important resource that has led to the discovery of biologically active molecules with unique chemical structures.<sup>3</sup> Excellent examples are the salinosporamides<sup>4a</sup> and marinomycins.<sup>4b</sup> Salinosporamide A, isolated from *Salinispora tropica*, is a potent 20S proteasome inhibitor that recently completed phase I clinical trials for treatment of solid tumors, lymphomas, and multiple myeloma. The marinomycins isolated from a "*Marinispora*" sp. showed significant antimicrobial activity against drug-resistant bacterial pathogens and selective cancer cell cytotoxicity against melanoma cell lines.<sup>4b</sup>

As part of a program to explore marine bacterial metabolites as inhibitors of tumor initiation and promotion, we have targeted nuclear factor kappa B (NF $\kappa$ B), a transcription factor that regulates the expression of more than 400 different genes. NF $\kappa$ B is an inducible, well-characterized protein, responsible for the regulation of complex phenomena, with a pivotal role in controlling cell signaling.<sup>5</sup> Many genes activated by NF $\kappa$ B encode proteins that are involved in tumorigenesis, including cyclooxygenase (COX)-2 and matrix metalloproteinase (MMP)-9, adhesion molecules, chemokines, inflammatory cytokines, inducible nitric oxide synthase (iNOS), and antiapoptosis proteins bcl-2 and bcl-xl.<sup>6</sup> Constitutive expression of NF $\kappa$ B is a frequent occurrence in tumor cells, and inhibitors of NF $\kappa$ B have the potential of suppressing carcinogenesis.<sup>7</sup> Since NF $\kappa$ B represents an important and attractive therapeutic target for drugs to treat many disorders, including arthritis, asthma, autoimmune diseases, and different types of cancer, it has become

a focal point for drug discovery from different sources, including the marine environment. Over the past decade, several compounds have been discovered that selectively interfere with the NF $\kappa$ B pathway. Examples include phenols and polyphenols (e.g., curcumin, resveratrol, caffeic acid phenethyl ester, quercetin, epigallocatechin-3-gallate), lignans, sesquiterpenes, diterpenes, and triterpenes.<sup>8</sup> The majority of these compounds are antioxidants, which act by blocking the protein kinase-mediated I $\kappa$ B degradation, thereby preventing NF $\kappa$ B activation.

An additional strategy for protecting cells from cancer initiation involves decreasing metabolic enzymes responsible for generating reactive species (phase I enzymes) while increasing phase II enzymes that can deactivate radicals and electrophiles known to intercede in normal cellular processes. Reduction of electrophilic quinones by quinone reductase (QR) is an important detoxification pathway, and induction of this enzyme often correlates with induction of other phase II enzymes such as glutathione S-transferase. Based on the evaluation of approximately 3800 marine organism extracts, the percentage characterized as "active" QR activators was established in the range of 17% of the total. About 60 QR inducers have been isolated from a diversity of sources, and substantial molecular diversity has been observed. These active compounds comprise ceramides, terpenoids, withanolides, flavonoids, chalcones, alkaloids, and diarylheptanoids.<sup>8</sup>

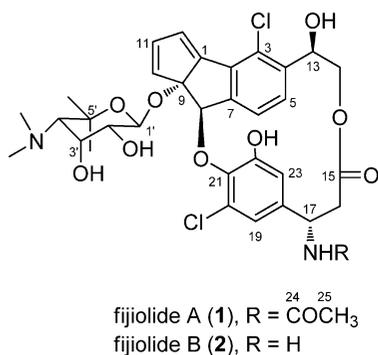
As part of our collaborative program to define new modulators of NF $\kappa$ B and QR, the marine-derived actinomycete strain CNS-653, isolated from a marine sediment sample collected from the Beqa Lagoon, Fiji, was evaluated. Analysis of the 16S rRNA gene sequence places this strain within the genus *Nocardiopsis* (GQ374440). Strain CNS-653 shares a nearly identical (99.8%) 16S sequence with another marine-derived strain, CNR-712 (EF392847), which produces the modified peptides lucentamycins A–D, and was isolated from a marine sediment sample collected in the Bahamas.<sup>9</sup> LC-MS analysis of a whole culture extract of this strain revealed peaks illustrating the characteristic MS isotopic patterns of dichlorinated secondary metabolites ([M + H]<sup>+</sup>: [M + H + 2]<sup>+</sup>: [M + H + 4]<sup>+</sup> = 10:6:1). Fractionation of a 30 L culture extract of this strain by standard silica column chromatography, followed by reversed-phase HPLC separation, yielded fijiolides A (**1**, 31.0

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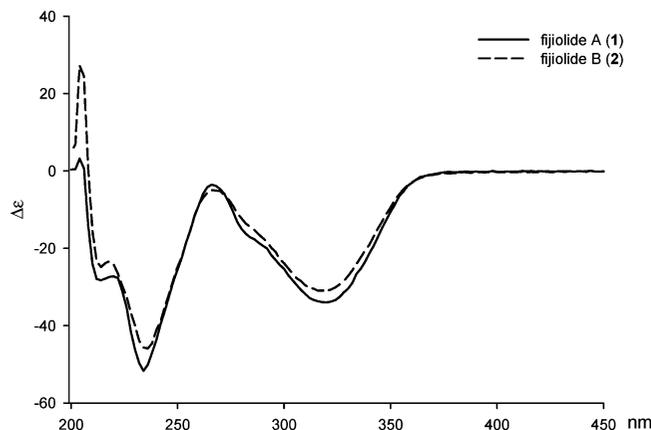
mg) and B (**2**, 3.0 mg) as colorless noncrystalline solids. Herein, we report details of the isolation and structure elucidation of fijiolides A and B and their effects on TNF- $\alpha$ -induced NF $\kappa$ B activity and QR induction.



The molecular formula of **1** was deduced as C<sub>34</sub>H<sub>39</sub><sup>35</sup>Cl<sub>2</sub>N<sub>2</sub>O<sub>10</sub>, on the basis of analysis of HRESIMS data (a pseudomolecular ion peak at *m/z* 705.1997 [M + H]<sup>+</sup>) and interpretation of <sup>13</sup>C NMR data. The <sup>1</sup>H NMR spectrum of **1** displayed *ortho*-coupled aromatic protons [ $\delta_{\text{H}}$  7.08 (d, *J* = 8.0 Hz), 6.64 (d, *J* = 8.0 Hz)], *meta*-coupled aromatic protons [ $\delta_{\text{H}}$  6.52 (d, *J* = 2.0 Hz), 5.98 (d, *J* = 2.0 Hz)], three olefinic protons [ $\delta_{\text{H}}$  6.76 (d, *J* = 2.1 Hz), 6.65 (dd, *J* = 5.3, 2.1 Hz), 6.62 (d, *J* = 5.3 Hz)], and three downfield methine protons [ $\delta_{\text{H}}$  5.87 (s), 4.66 (dd, *J* = 11.0, 4.0 Hz), 4.61 (ddd, *J* = 13.4, 13.4, 3.8 Hz)]. The <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra also revealed two *N*-methyl groups [ $\delta_{\text{H}}$  2.79 ( $\delta_{\text{C}}$  44.4 and 42.9)], three methyl singlets [ $\delta_{\text{H}}$  1.46 ( $\delta_{\text{C}}$  20.6), 1.43 ( $\delta_{\text{C}}$  31.5), 1.70 ( $\delta_{\text{C}}$  22.6)], and six exchangeable protons [(4-OH, 2-NH);  $\delta_{\text{H}}$  9.09, 8.45, 8.41, 5.86, 5.73, 4.95]. Interpretation of 2D NMR spectroscopic data allowed three partial structures to be constructed: a benzodihydropentalene, a 3'-chloro-5'-hydroxy- $\beta$ -tyrosine, and an aminosugar unit. COSY NMR data, including cross-peaks from H-11 ( $\delta_{\text{H}}$  6.65, dd, *J* = 5.3, 2.1 Hz) to H-10 ( $\delta_{\text{H}}$  6.62, d, *J* = 5.3 Hz) and H-12 ( $\delta_{\text{H}}$  6.76, d, *J* = 2.1 Hz), along with the 1D proton coupling constants of these three protons, indicated that they belonged to a cyclopentadienyl spin system. The presence of *ortho*-coupled aromatic protons [H-5 ( $\delta_{\text{H}}$  6.64, d, *J* = 8.0 Hz) and H-6 ( $\delta_{\text{H}}$  7.08, d, *J* = 8.0 Hz)] indicated a 1,2,3,4-tetrasubstituted benzene ring. The connection between the cyclopentadienyl moiety and the 1,2,3,4-tetrasubstituted benzene ring was established by long-range HMBC correlations observed from H-12 ( $\delta_{\text{H}}$  6.76) to C-1 ( $\delta_{\text{C}}$  148.8), C-2 ( $\delta_{\text{C}}$  135.7), C-9 ( $\delta_{\text{C}}$  99.8), and C-10 ( $\delta_{\text{C}}$  135.5); from H-8 ( $\delta_{\text{H}}$  5.87) to C-1 ( $\delta_{\text{C}}$  148.8), C-2 ( $\delta_{\text{C}}$  135.7), C-6 ( $\delta_{\text{C}}$  125.3), C-7 ( $\delta_{\text{C}}$  147.7), C-9 ( $\delta_{\text{C}}$  99.8), and C-10 ( $\delta_{\text{C}}$  135.5); and from H-6 ( $\delta_{\text{H}}$  7.08) to C-2 ( $\delta_{\text{C}}$  135.7), C-7 ( $\delta_{\text{C}}$  147.7), and C-8 ( $\delta_{\text{C}}$  83.2). The oxygenated methine proton H-13 ( $\delta_{\text{H}}$  4.66), which was coupled to the methylene protons H<sub>2</sub>-14 ( $\delta_{\text{H}}$  4.23, and  $\delta_{\text{H}}$  3.64), also displayed long-range HMBC correlations to carbons C-3 ( $\delta_{\text{C}}$  126.7), C-4 ( $\delta_{\text{C}}$  139.3), and C-5 ( $\delta_{\text{C}}$  129.1). These data allowed construction of the benzodihydropentalene unit in **1** (C-1 to C-12).

The 3'-chloro-5'-hydroxy- $\beta$ -tyrosine amino acid component was established by the observation of COSY NMR cross-peaks between a downfield methine proton H-17 ( $\delta_{\text{H}}$  4.61) and the methylene protons H<sub>2</sub>-16 ( $\delta_{\text{H}}$  2.57, and  $\delta_{\text{H}}$  2.07). In addition, these assignments were also confirmed by long-range HMBC NMR correlations from H-17 to C-15 ( $\delta_{\text{C}}$  170.0), C-18 ( $\delta_{\text{C}}$  138.8), C-19 ( $\delta_{\text{C}}$  114.2), C-22 ( $\delta_{\text{C}}$  113.9), and C-24 ( $\delta_{\text{C}}$  168.4) and from the phenol proton 22-OH ( $\delta_{\text{H}}$  8.45) to C-21 ( $\delta_{\text{C}}$  138.3), C-22 ( $\delta_{\text{C}}$  151.0), and C-23 ( $\delta_{\text{C}}$  113.9). The connection between C-8 and C-21 was established by a three-bond HMBC correlation from H-8 ( $\delta_{\text{H}}$  5.87) to C-21 ( $\delta_{\text{C}}$  138.3).

The 4-deoxy-4-dimethylamino-5,5-dimethylribose sugar was constructed on the basis of analysis of COSY and HMBC NMR spectroscopic data. COSY NMR correlations permitted construction



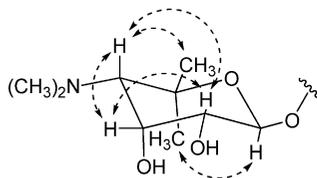
**Figure 1.** CD spectra for fijiolides A (**1**) and B (**2**) in MeOH ( $7.1 \times 10^{-4}$  M).

of the spin system from the anomeric proton H-1' through H-4' [H1' ( $\delta_{\text{H}}$  4.45, d, *J* = 8.0 Hz)—H-2' ( $\delta_{\text{H}}$  2.92, dd, *J* = 8.0, 3.4 Hz)—H-3' ( $\delta_{\text{H}}$  4.06, d, *J* = 8.0, 3.4 Hz)—H-4' ( $\delta_{\text{H}}$  2.98)] of the aminosugar. Long-range HMBC correlations from the *N*-dimethyl protons ( $\delta_{\text{H}}$  2.79) to C-4' ( $\delta_{\text{C}}$  69.6) and from the dimethyl protons ( $\delta_{\text{H}}$  1.46, 1.43) to C-4' ( $\delta_{\text{C}}$  69.6) and C-5' ( $\delta_{\text{C}}$  75.0) allowed placement of these protons at C-4' and C-5', respectively. A long-range HMBC correlation from the anomeric proton H-1' ( $\delta_{\text{H}}$  4.45) to C-9 ( $\delta_{\text{C}}$  99.8) provided evidence for placing the sugar at C-9. The three-bond HMBC correlations from the methylene protons H-14 ( $\delta_{\text{H}}$  4.23, 3.64) to the carbonyl carbon C-15 ( $\delta_{\text{C}}$  170.0) and from the methine proton H-17 ( $\delta_{\text{H}}$  4.61) to the carbonyl carbon C-24 ( $\delta_{\text{C}}$  168.4) permitted the ester linkage between C-14 and C-15 and the amide linkage between C-17 and C-24, respectively. Lastly, placement of the chlorine at C-3 completed the structure assignment of **1**.

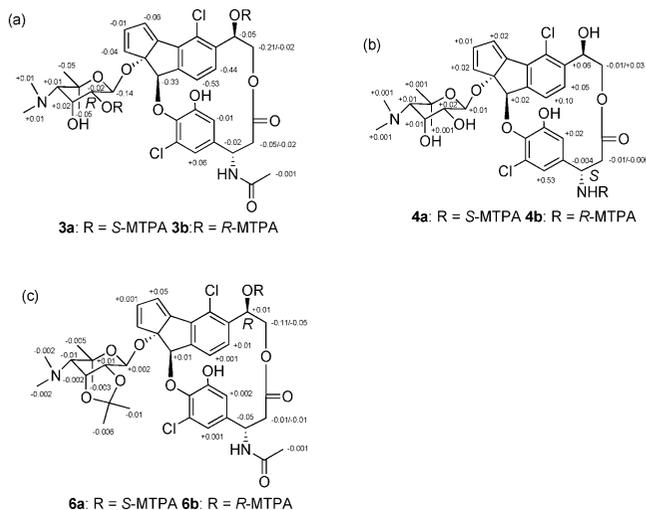
The molecular formula of **2** was assigned as C<sub>32</sub>H<sub>36</sub><sup>35</sup>Cl<sub>2</sub>N<sub>2</sub>O<sub>9</sub>, on the basis of the pseudomolecular ion peak at *m/z* 663.1871 [M + H]<sup>+</sup> observed in the HRESIMS spectrum. The <sup>1</sup>H NMR spectrum of **2** was almost identical to that of **1** except for an upfield shifted methine proton H-17 ( $\delta_{\text{H}}$  4.09) and the absence of one methyl singlet. This difference indicated **2** possessed a primary amine at C-17 instead of an acetamide group. Interpretation of 2D COSY, HSQC, and HMBC NMR spectroscopic data allowed the planar structure of **2** to be assigned as shown.

The relative and absolute configurations of **1** and **2** were determined by analysis of circular dichroism (CD) spectroscopic data, application of the advanced Mosher's method, and combined spectroscopic methods. Fijiolides A (**1**) and B (**2**) were assigned identical absolute configurations on the basis of the virtually identical CD curves observed for these two metabolites (Figure 1).

The relative configuration of the ribopyranose ring of **1** was determined by analysis of vicinal coupling constants and by analysis of NOESY spectroscopic data. The magnitude of the <sup>1</sup>*J*<sub>CH</sub> coupling constant for the anomeric proton (H-1', 162 Hz) indicated that this center possesses a  $\beta$ -configuration.<sup>10</sup> A large diaxial coupling constant (8.0 Hz) between H-1' and H-2' indicated an axial configuration for H-2', while a small vicinal coupling constant (3.4 Hz) between H-2' and H-3' and strong NOESY correlations between H-2' and H-3' and between H-3' and H-4' established that H-2' and H-3' are in the equatorial and axial positions, respectively (Figure 2). Determination of the absolute configuration of the ribopyranose of **1** was achieved using the advanced Mosher method involving NMR analysis of MTPA ester pairs produced by derivatization with *R*- and *S*-MTPA-Cl ( $\alpha$ -methoxy- $\alpha$ -(trifluoromethyl)phenylacetyl chloride).<sup>11</sup> Calculation of  $\Delta\delta_{S-R}$  values of bis-MTPA esters (**3a,b**) clearly established the absolute configuration of C-2' as *R* and allowed the absolute configuration of the ribopyranose to be assigned as 1'*S*, 2'*R*, 3'*R*, 4'*S* (Figure 3a).



**Figure 2.** Relative configuration of the ribopyranose based on key NOESY correlations (dashed arrows).

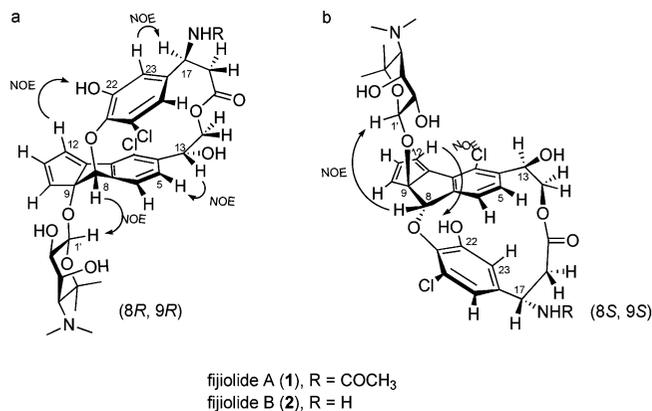
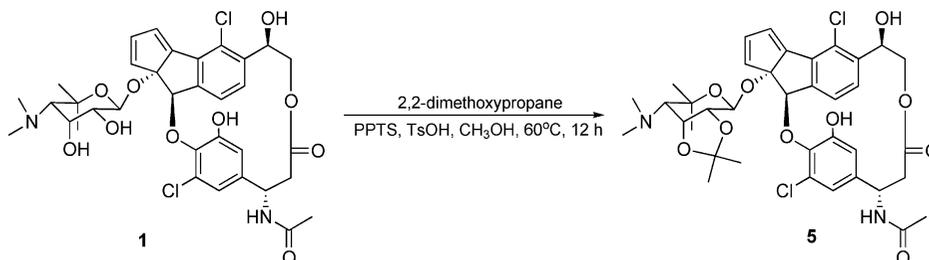


**Figure 3.**  $\Delta\delta_{S-R}$  of  $^1\text{H}$  for *S*- and *R*-bis-MTPA esters of **1** (a),  $\Delta\delta_{S-R}$  of  $^1\text{H}$  for *S*- and *R*-mono-MTPA amides of **2** (b),  $\Delta\delta_{S-R}$  of  $^1\text{H}$  for *S*- and *R*-mono-MTPA esters of **1** (c).

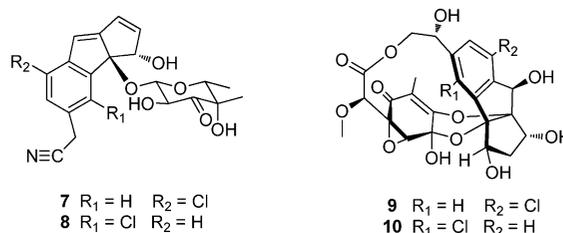
The absolute configurations at C-17 and C-13 in **1** and **2** were determined as *S* and *R*, respectively, also by the application of the advanced Mosher's method. Treatment of **2** in pyridine *R*(-)-MTPA-Cl and (*S*)(+)-MTPA-Cl yielded the MTPA amides **4a** and **4b**. Positive  $\Delta\delta_{S-R}$  values for H-19 and H-23 and negative  $\Delta\delta_{S-R}$  values for H-16 allowed the absolute configuration of C-17 to be assigned as *S* (Figure 3b). To assign the absolute configurations at C-13, the 1,2-diol groups of the ribopyranose in **1** were first protected by conversion to the acetonide **5** using 2,2-dimethoxypropane and pyridinium *p*-toluenesulfate in methanol (Scheme 1). Treatment of **5** in pyridine with *R*-MTPA-Cl and *S*-MTPA-Cl yielded the *S*-MTPA ester **6a** and *R*-MTPA ester **6b**, respectively. Calculation of  $\Delta\delta_{S-R}$  values of MTPA esters of **6a,b** established the absolute configuration of C-13 as *R* (Figure 3c).

The absolute configurations of the remaining components of **1** were determined by interpretation of NOESY NMR correlations. A NOESY correlation between H-8 ( $\delta_{\text{H}}$  5.87) and the anomeric proton H-1' ( $\delta_{\text{H}}$  4.45) of the ribopyranose suggested a *trans*-orientation of the C-9 glycoside and C-8 ether bonds.<sup>12</sup> There were only two possible stereoisomers, (8*R*, 9*R*) or (8*S*, 9*S*), for fijiolides, as shown in Figure 4. A NOESY correlation between H-12 and 22-OH revealed that ester and ether linkages between the two aromatic rings of **1** restrict free rotation of the  $\beta$ -tyrosine moiety in fijiolides (Figure 4a,b). The strong NOESY correlations between

**Scheme 1.** Production of Acetonide **5** from Fijiolide A (**1**)



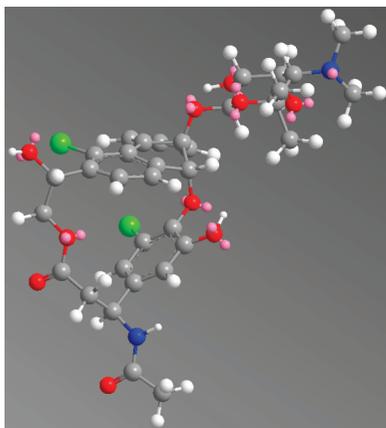
**Figure 4.** Two possible stereoisomers (a, b) and selected key NOESY correlations for determination of the configurations at C-8 and C-9 for fijiolides (a).



**Figure 5.** Chemical structures of cyanosporasides (**7** and **8**), sporolides (**9** and **10**), and aromatization products of C-1027 (**11** and **12**).

H-5 and H-13 and between H-17 and H-23 suggested that C-8 and C-9 could be assigned *R* and *R* configurations, respectively (Figure 4a).<sup>13</sup>

The chlorocyclopenta[*a*]indene carbon skeleton of the fijiolides resembles that of the cyanosporasides<sup>14a</sup> and sporolides,<sup>14b</sup> isolated from strains of the marine actinomycete genus *Salinispora* (Figure 5). Both cyanosporasides A and B (**7**, **8**) and sporolides A and B (**9**, **10**) are equally represented in the respective fermentation extracts of the producing strains. This is what would be expected for the cycloaromatization of the enediyne precursors to Bergman diradical intermediates followed by nonspecific quenching with hydrogen<sup>15a</sup> or chloride, a process that has been experimentally



**Figure 6.** Energy-minimized model of fijiolide A (**1**) (calculated by ChemBio 3D (ver. 11.0), red = oxygen, green = chlorine, white = hydrogen, blue = nitrogen, gray = carbon atom).

verified.<sup>15b</sup> Interestingly, chloride addition occurred primarily at the C-3 position in fijiolides A and B. Energy-minimized modeling of fijiolides revealed that C-6 in the benzene ring is in closer proximity to the chlorine atom of  $\beta$ -tyrosine than C-3 is to the 22-OH group of  $\beta$ -tyrosine (Figure 6). The ribopyranose is also located near C-6 in the fijiolides, further restricting chloride addition to that site.

With the exception of the chlorine substituents, the fijiolides are nearly identical to the aromatization product from the bacterial metabolite C-1027 chromophore (**11**), derived from an antitumor antibiotic enediyne, C-1027 (**12**), isolated in 1988 by Marunaka *et al.* from the culture broth of *Streptomyces globisporus*.<sup>16</sup> C-1027 is responsible for the activity of the complete C-1027 natural product, which consists of an apoprotein with a single polypeptide chain of 110 amino acid residues and a chromophore with the nine-membered enediyne and benzoxazine moieties. In 1995, Yu *et al.* demonstrated that loss of the benzoxazine moiety in the aromatization product of C-1027 resulted in a substantial decrease of DNA binding affinity (~400 fold).<sup>17</sup> Fijiolides A and B, which resemble the aromatization product of the C-1027 chromophore without its benzoxazine moiety, do not display significant activity against either HCT-116 or methicillin-resistant *Staphylococcus aureus* (MRSA) (data not shown).

NF $\kappa$ B plays a central role in a variety of disease states, including cancer. As a transcription factor, activated NF $\kappa$ B regulates the expression of numerous genes, which subsequently influence cell survival, proliferation, differentiation, and inflammation.<sup>18,19</sup> Numerous studies demonstrating the effects of different compounds on NF $\kappa$ B activity have been reported. However, the results of these studies are often inconsistent, which is likely due to the use of diverse cell types and assay methods. For assessment of NF $\kappa$ B activity, we used stably transfected 293 human embryonic kidney cells that demonstrated NF $\kappa$ B activation by treatment with 12-*O*-tetradecanoylphorbol-13-acetate, as well as TNF- $\alpha$ . We observed a 5- to 15-fold NF $\kappa$ B activation by treatment with 0.14 nM TNF- $\alpha$ , high values for luciferase luminescence, and good reproducibility.<sup>20</sup> With this assay, fijiolide A inhibited TNF- $\alpha$ -induced NF $\kappa$ B activity by 70.3%, and dose dependence demonstrated an IC<sub>50</sub> value of 0.57  $\mu$ M. Fijiolide B was less active, showing only 46.5% without a dose-dependent response.

An established mechanism of cancer chemoprevention involves the stimulation of metabolic detoxification activity. The detoxification enzyme quinone reductase 1 (QR1) is an important phase II enzyme (one that deactivates reactive and potentially carcinogenic species) that converts quinones to hydroquinones, reducing oxidative cycling.<sup>21</sup> The enzyme exhibits broad specificity, reducing a wide range of hydrophobic quinones of varying structure.<sup>22</sup> The induction of QR often coincides with induction of other phase II enzymes

and is therefore useful in the study of chemopreventive agents. Hepa 1c1c7 (mouse hepatoma) cells were used to assess quinone reductase activity via a color change as MTT is reduced to a blue formazan.<sup>23</sup> Fijiolide A (**1**) was found to enhance QR1 activity with an induction ratio of 3.5 at 20  $\mu$ g/mL (28.4  $\mu$ M). The concentration required to double induction (CDI) was 1.8  $\mu$ M (the CDI value of 4-bromoflavone, as a positive control, is 0.013  $\mu$ M). Fijiolide B did not demonstrate QR1 induction activity, indicating that substitutions on the nitrogen atom affect activity. Differential activity of this type was also observed with the NF $\kappa$ B assay.

QR1 is well established to undergo transcriptional activation through NF $\kappa$ B signaling pathways.<sup>24</sup> Although NF $\kappa$ B alone seems to account for only 20–30% of QR promoter activity, revealed by mutational analysis, NF $\kappa$ B is responsible for a proportion of the immediate response of the QR gene promoter.<sup>25</sup> Significant inhibition of luciferase NF $\kappa$ B activity and QR activation was detected with fijiolide A at reasonably low concentrations (IC<sub>50</sub> for NF $\kappa$ B, 0.57  $\mu$ M; CDI for QR1, 1.8  $\mu$ M). It is likely that QR activation is NF $\kappa$ B independent. In summary, fijiolide A modulates distinct and important mechanisms, and this would be worthy of more detailed investigation.

## Experimental Section

**General Experimental Procedures.** The optical rotations were measured using a Rudolph Research Autopol III polarimeter with a 10 cm cell. UV spectra were recorded in a Varian Cary UV–visible spectrophotometer with a path length of 1 cm, and IR spectra were recorded on a Perkin-Elmer 1600 FT-IR spectrometer. CD spectra were collected with an AVIV model 215 CD spectrometer with a 0.5 cm long cell. <sup>1</sup>H and 2D NMR spectral data were recorded at 500 or 600 MHz in DMSO-*d*<sub>6</sub>, CDCl<sub>3</sub>, or methanol-*d*<sub>4</sub> solution containing Me<sub>4</sub>Si as internal standard on Varian Inova spectrometers. <sup>13</sup>C NMR spectra were acquired at 150 MHz on a Varian Inova spectrometer. High-resolution ESI-TOF mass spectra were provided by The Scripps Research Institute, La Jolla, CA, or by the mass spectrometry facility at the Department of Chemistry and Biochemistry at the University of California, San Diego, La Jolla, CA. Low-resolution LC/MS data were measured using a Hewlett-Packard series 1100 LC/MS system with a reversed-phase C<sub>18</sub> column (Phenomenex Luna, 4.6 mm  $\times$  100 mm, 5  $\mu$ m) at a flow rate of 0.7 mL/min.

**Collection and Phylogenetic Analysis of Strain CNS-653.** The marine-derived actinomycete, strain CNS-653, was isolated from a surf zone sediment sample collected near Beqa Island in Beqa Lagoon in Fiji, July 2006. NCBI BLAST analysis of the partial 16S rDNA sequence of CNQ-653 (deposited with GenBank as accession number GQ374440) indicates that this strain is affiliated with the genus *Nocardioopsis*.

**Cultivation and Extraction.** The bacterium (strain CNS-653) was cultured in 30 2.8 L Fernbach flasks each containing 1 L of a seawater-based medium (10 g starch, 4 g yeast extract, 2 g peptone, 1 g CaCO<sub>3</sub>, 40 mg Fe<sub>2</sub>(SO<sub>4</sub>)<sub>3</sub>·4H<sub>2</sub>O, 100 mg KBr) and shaken at 230 rpm at 27 °C. After seven days of cultivation, sterilized XAD-16 resin (20 g/L) was added to adsorb the organic products, and the culture and resin were shaken at 215 rpm for 2 h. The resin was filtered through cheesecloth, washed with deionized water, and eluted with acetone. The acetone was removed under reduced pressure, and the resulting aqueous layer was extracted with EtOAc (3  $\times$  500 mL). The EtOAc-soluble fraction was dried *in vacuo* to yield 3.5 g of extract.

**Isolation of Fijiolides A and B.** The extract (3.5 g) was fractionated by open column chromatography on silica gel (25 g), eluted with a step gradient of CH<sub>2</sub>Cl<sub>2</sub> and MeOH. The CH<sub>2</sub>Cl<sub>2</sub>/MeOH (5:1) fraction contained a mixture of both fijiolides, which were purified by reversed-phase HPLC (Phenomenex Ultracarb C30, 250  $\times$  100 mm, 2.0 mL/min, 5  $\mu$ m, 100 Å, UV = 210 nm) using a gradient solvent system from 5% to 100% CH<sub>3</sub>CN (0.05% TFA) over 60 min to afford fijiolides A (**1**, 30.0 mg) and B (**2**, 3.0 mg), as pale yellow oils.

**Fijiolide A (1):** pale yellow oil; [ $\alpha$ ]<sub>D</sub><sup>21</sup> –440 (*c* 0.5, MeOH); UV (MeOH)  $\lambda$ <sub>max</sub> (log  $\epsilon$ ) 213 (4.4), 285 (3.7), 330 (2.7) nm; IR (KBr)  $\nu$ <sub>max</sub> 3380, 1725, 1677, 1512, 1190, 1128 cm<sup>-1</sup>; <sup>1</sup>H and 2D-NMR (600 MHz, DMSO-*d*<sub>6</sub>), see Table 1; HRESIMS [M + H]<sup>+</sup> *m/z* 705.1997 (calcd for C<sub>34</sub>H<sub>39</sub><sup>35</sup>Cl<sub>2</sub>N<sub>2</sub>O<sub>10</sub>, 705.1976).

**Table 1.** NMR Spectroscopic Data for Fijiolide A (**1**) (DMSO- $d_6$ )<sup>a</sup>

no.	$\delta_C$ , mult. <sup>b</sup>	$\delta_H$ (J in Hz)	COSY	HMBC <sup>c</sup>
1	148.8, C			
2	135.7, C			
3	126.7, C			
4	139.3, C			
5	129.1, CH	6.64, d (8.0)	6	3, 4, 6, 7, 13
6	125.3, CH	7.08, d (8.0)	5	2, 4, 5, 7, 8
7	147.7, C			
8	83.2, CH	5.87, s		1, 2, 6, 7, 9, 10, 21
9	99.8, C			
10	135.5, CH	6.62, d (5.3)	11	1, 8, 9, 11, 12
11	138.0, CH	6.65, dd (5.3, 2.1)	10, 12	1, 9, 10, 12
12	129.4, CH	6.76, d (2.1)	11	1, 2, 9, 10, 11
13	73.1, CH	4.66, dd (11.0, 4.0)	14, 13-OH	3, 4, 5, 14
14	65.9, CH <sub>2</sub>	4.23, dd (11.0, 4.0), 3.64, dd (11.0, 4.0)	13	4, 13, 15
15	170.0, C			
16	42.9, CH <sub>2</sub>	2.57, dd (13.4, 3.8), 2.07, t (13.4)	17	15, 17, 18
17	49.0, CH	4.61, ddd (13.4, 13.4, 3.8)	16, 17-NH	15, 16, 18, 19, 23, 24
18	138.8, C			
19	114.2, CH	6.52, d (2.0)	23	17, 18, 20, 21, 23
20	129.2, C			
21	138.3, C			
22	151.0, C			
23	113.9, CH	5.98, d (2.0)	19	17, 18, 21, 22
24	168.4, C			
25	22.6, CH <sub>3</sub>	1.70, s		24
1'	93.1, CH	4.45, d (8.0)	2'	9, 2', 3', 5'
2'	69.6, CH	2.92, dd (8.0, 3.4)	1', 3'	1', 4'
3'	67.5, CH	4.06, dd (8.0, 3.4)	2', 4'	1', 5'
4'	69.6, CH	2.98 <sup>d</sup>	3'	
5'	75.0, C			
4'-NMe <sub>2</sub>	42.9, CH <sub>3</sub>	2.79, br s		4'
4'-NMe <sub>2</sub>	44.4, CH <sub>3</sub>	2.79, br s		4'
6'-Me $\alpha$	31.5, CH <sub>3</sub>	1.43, s		4', 5', 6'-Me $\beta$
6'-Me $\beta$	20.6, CH <sub>3</sub>	1.46, s		4', 5', 6'-Me $\alpha$
13-OH		5.86	13	
17-NH		8.41, d (8.0)		16, 17, 24
22-OH		8.45, s		21, 22, 23
2'-OH		4.95, br s	2'	
3'-OH		5.73, br s	3'	
4'-NH <sup>+</sup>		9.09, br s	4'-NMe <sub>2</sub>	

<sup>a</sup> 600 MHz for <sup>1</sup>H NMR and 150 MHz for <sup>13</sup>C NMR. <sup>b</sup> Numbers of attached protons were determined by analysis of 2D spectra. <sup>c</sup> HMBC correlations are from the stated protons to the indicated carbons. <sup>d</sup> Coupling constant could not be determined because of overlapping signals.

**Fijiolide B (2):** pale yellow oil; [ $\alpha$ ]<sub>D</sub><sup>21</sup> -360 (c 0.2, MeOH); UV (MeOH)  $\lambda_{\max}$  (log  $\epsilon$ ) 213 (4.3), 285 (3.8), 330 (2.7) nm; IR (KBr)  $\nu_{\max}$  3383, 1678, 1512, 1201, 1134, 1048 cm<sup>-1</sup>; <sup>1</sup>H and 2D-NMR (600 MHz, DMSO- $d_6$ ), see Table 2; HRESITOFMS [M + H]<sup>+</sup>  $m/z$  663.1871 (calcd for C<sub>32</sub>H<sub>37</sub><sup>35</sup>Cl<sub>2</sub>N<sub>2</sub>O<sub>9</sub>, 663.1871).

**Bis-MTPA Esters of 1 (3a, 3b).** Fijiolide A (7.0 mg) was dissolved in freshly distilled dry pyridine (2 mL), and several dry crystals of dimethylaminopyridine were added. The mixture was stirred for 15 min at RT. Treatment with *R*-(-)-MTPA-Cl yielded the bis-*S*-MTPA ester at 60 °C after 12 h. The reaction was quenched by 1 mL of MeOH. After removal of solvent under vacuum, the residue was purified by reversed-phase HPLC (Phenomenex Luna 5u C18 (2) 100 Å, 250 × 100 mm, 2.0 mL/min, UV detection at 210 nm) using a gradient solvent system from 5% to 100% CH<sub>3</sub>CN (0.05% TFA) over 50 min. The bis-*S*-MTPA ester was obtained at 36 min. The bis-*R*-MTPA ester was prepared with *S*-MTPA-Cl in the same manner.  $\Delta\delta_{S-R}$  values for the *S*- and *R*-MTPA esters of **1** were recorded in ppm in methanol- $d_4$ .

**Bis-*S*-MTPA ester of fijiolide A (3a):** <sup>1</sup>H NMR (600 MHz, methanol- $d_4$ )  $\delta$  7.55 (m, 2H), 7.45–7.30 (m, 8H), 6.89 (d, 1H,  $J$  = 1.5 Hz), 6.63 (d, 1H,  $J$  = 5.9 Hz), 6.589 (dd, 1H,  $J$  = 5.9, 3.0 Hz), 6.484 (d, 1H,  $J$  = 8.0 Hz), 6.42 (d, 1H,  $J$  = 3.0 Hz), 6.292 (d, 1H,  $J$  = 8.0 Hz), 5.802 (dd, 1H,  $J$  = 11.0, 4.0 Hz), 5.54 (s, 1H), 4.717 (d, 1H,  $J$  = 11.0 Hz), 4.69 (dd, 1H,  $J$  = 11.0, 4.0 Hz), 4.678 (dd, 1H,  $J$  = 11.0, 4.0 Hz), 4.465 (t, 1H,  $J$  = 11.0 Hz), 4.39 (t, 1H,  $J$  = 1.2 Hz), 3.71 (s, 1H), 3.646 (dd, 1H,  $J$  = 11.0, 4.0 Hz), 3.37 (s, 3H), 3.270 (d, 1H,  $J$  = 2.0 Hz), 2.910 (s, 6H), 2.60 (t, 1H,  $J$  = 11.0 Hz), 2.55 (s, 3H), 2.14 (t, 1H,  $J$  = 11.0 Hz), 1.822 (s, 3H), 1.449 (s, 3H), 1.443 (s, 3H); ESIMS  $m/z$  1137 [M + H]<sup>+</sup>, 1159 [M + Na]<sup>+</sup>.

**Bis-*R*-MTPA ester of fijiolide A (3b):** <sup>1</sup>H NMR (600 MHz, methanol- $d_4$ )  $\delta$  7.74–7.30 (m, 12H), 7.021 (d, 1H,  $J$  = 8.0 Hz), 6.83 (d, 1H,  $J$  = 1.5 Hz), 6.736 (d, 1H,  $J$  = 8.0 Hz), 6.67 (d, 1H,  $J$  = 5.9

Hz), 6.59 (dd, 1H,  $J$  = 5.9, 3.0 Hz), 5.86 (s, 1H), 5.856 (dd, 1H,  $J$  = 11.0, 4.0 Hz), 4.857 (d, 1H,  $J$  = 11.0 Hz), 4.72 (dd, 1H,  $J$  = 11.0, 4.0 Hz), 4.680 (dd, 1H,  $J$  = 11.0, 4.0 Hz), 4.492 (t, 1H,  $J$  = 11.0 Hz), 4.376 (t, 1H,  $J$  = 1.2 Hz), 3.864 (dd, 1H,  $J$  = 11.0, 4.0 Hz), 3.52 (s, 3H), 3.263 (d, 1H,  $J$  = 1.2 Hz), 2.95 (s, 3H), 2.899 (s, 6H), 2.62 (t, 1H,  $J$  = 11.0 Hz), 2.20 (t, 1H,  $J$  = 11.0 Hz), 1.822 (s, 3H), 1.499 (s, 3H), 1.493 (s, 3H); ESIMS  $m/z$  1137 [M + H]<sup>+</sup>, 1159 [M + Na]<sup>+</sup>.

**MTPA Amides of 2 (4a, 4b).** Fijiolide B (1.3 mg) was dissolved in freshly distilled dry pyridine (2 mL), several dry crystals of dimethylaminopyridine were added, and the mixture was stirred for 15 min at RT. Treatment with *R*-(-)-MTPA-Cl yielded the *S*-MTPA amide at room temperature after 11 h. The reaction was quenched by 1 mL of MeOH. After removal of solvent under vacuum, the residue was purified by reversed-phase HPLC (Phenomenex Luna 5u C18 (2) 100 Å, 250 × 100 mm, 2.0 mL/min, UV detection at 210 nm) using a gradient solvent system from 5% to 100% CH<sub>3</sub>CN (0.05% TFA) over 60 min. The *S*-MTPA amide was obtained at 41 min. The *R*-MTPA amide was prepared with *S*-MTPA-Cl in the same manner.  $\Delta\delta_{S-R}$  values for the *S*- and *R*-MTPA amides of **2** were recorded in ppm in DMSO- $d_6$ .

***S*-MTPA amide of fijiolide B (4a):** <sup>1</sup>H NMR (600 MHz, DMSO- $d_6$ )  $\delta$  9.96 (d, 1H,  $J$  = 7.4 Hz), 8.98 (br s, 1H), 7.36–7.25 (m, 5H), 7.08 (d, 2H,  $J$  = 8.0 Hz), 6.738 (m, 1H), 6.64–6.59 (m, 4H), 6.040 (m, 1H), 6.03 (m, 1H), 5.89 (m, 1H), 5.80 (m, 1H), 4.99 (m, 1H), 4.73 (m, 1H), 4.640 (m, 1H), 4.48 (d, 1H,  $J$  = 10.0 Hz), 4.27 (dd, 1H,  $J$  = 10.4, 10.4 Hz), 4.06 (m, 1H), 3.78 (s, 3H), 3.62 (dd, 1H,  $J$  = 11.0, 6.0 Hz), 3.00 (s, 6H), 2.92 (br s, 1H), 2.91 (m, 1H), 2.840 (m, 1H), 2.110 (dd, 1H,  $J$  = 11.0, 2.0 Hz), 1.37 (br s, 6H); ESIMS  $m/z$  879 [M + H]<sup>+</sup>, 901 [M + Na]<sup>+</sup>.

***R*-MTPA amide of fijiolide B (4b):** <sup>1</sup>H NMR (600 MHz, DMSO- $d_6$ )  $\delta$  8.98 (br s, 1H), 8.91 (d, 1H,  $J$  = 7.4 Hz), 7.36–7.25 (m, 5H), 7.17 (d, 2H,  $J$  = 8.0 Hz), 6.718 (m, 1H), 6.711 (d, 1H,  $J$  = 2.0),

**Table 2.** NMR Spectroscopic Data for Fijiolide B (2) (DMSO- $d_6$ )<sup>a</sup>

no.	$\delta_C$ , mult. <sup>b</sup>	$\delta_H$ (J in Hz)	COSY	HMBC <sup>c</sup>
1	148.2, C			
2	135.7, C			
3	126.9, C			
4	139.8, C			
5	129.5, CH	6.66, d (8.0)	6	3, 4, 6, 7, 13
6	125.5, CH	7.16, d (8.0)	5	2, 4, 5, 7, 8
7	147.8, C			
8	83.9, CH	5.88, s		1, 2, 6, 7, 9, 10, 21
9	100.4, C			
10	135.8, CH	6.62, d (5.3)	11	1, 8, 9, 11, 12
11	138.5, CH	6.65, dd (5.3, 2.1)	10, 12	1, 9, 10, 12,
12	129.9, CH	6.75, d (2.1)	11	1, 2, 9, 10, 11
13	72.7, CH	4.66, dd (11.0, 4.0)	14, 13-OH	3, 4, 5, 14
14	65.5, CH <sub>2</sub>	4.28, dd (11.0, 4.0), 3.68, dd (11.0, 4.0)	13	4, 13, 15
15	168.4, C			
16	41.7, CH <sub>2</sub>	2.85, dd (13.4, 1.8), 2.20, t (13.4)	17	15, 17, 18
17	51.2, CH	4.09, dd (13.4, 1.8)	16	15, 16, 18, 19, 23
18	133.1, C			
19	114.4, CH	6.76, d (2.0)	23	17, 18, 20, 21, 23
20	130.1, C			
21	139.9, C			
22	151.7, C			
23	115.2, CH	6.06, d (2.0)	19	17, 18, 21, 22
1'	93.4, CH	4.44, d (8.0)	2'	9, 2', 3', 5'
2'	69.6, CH	2.91, br s	1', 3'	1', 4'
3'	67.8, CH	4.05, br s	2', 4'	1', 5'
4'	69.6, CH	2.99, br s	3'	4'-NMe <sub>2</sub>
5'	75.3, C			
4'-NMe <sub>2</sub>	42.9, CH <sub>3</sub>	2.79, br s		4'
4'-NMe <sub>2</sub>	44.6, CH <sub>3</sub>	2.79, br s		4'
6'-Me $\alpha$	31.9, CH <sub>3</sub>	1.37, s		4', 5', 6'-Me $\beta$
6'-Me $\beta$	21.2, CH <sub>3</sub>	1.41, s		4', 5', 6'-Me $\alpha$
13-OH		5.86	13	
17-NH <sub>2</sub>		8.58		
22-OH		8.84, s		21, 22, 23
2'-OH		4.97, br s	2'	
3'-OH		5.72, br s	3'	
4'-NH <sup>+</sup>		9.13, br s	4'-NMe <sub>2</sub>	

<sup>a</sup> 600 MHz for <sup>1</sup>H NMR. <sup>b</sup> Numbers of attached protons were determined by analysis of 2D spectra. <sup>c</sup> HMBC correlations are from the stated protons to the indicated carbons.

6.67–6.62 (m, 3H), 6.10 (d, 1H,  $J = 2.0$ ), 6.020 (m, 1H), 5.87 (m, 1H), 5.77 (m, 1H), 4.99 (m, 1H), 4.68 (m, 1H), 4.644 (m, 1H), 4.46 (d, 1H,  $J = 10.0$  Hz), 4.24 (dd, 1H,  $J = 10.4, 10.4$  Hz), 4.06 (m, 1H), 3.82 (s, 3H), 3.64 (dd, 1H,  $J = 11.0, 6.0$  Hz), 2.92 (br s, 1H), 2.91 (m, 1H), 2.79 (s, 6H), 2.841 (m, 1H), 2.116 (dd, 1H,  $J = 11.0, 2.0$  Hz), 1.40 (br s, 6H); ESIMS  $m/z$  879 [M + H]<sup>+</sup>, 901 [M + Na]<sup>+</sup>.

**Acetonide 5.** Fijiolide A (1, 5.0 mg) was dissolved in 4 mL of dry MeOH solution, along with 5 mg of pyridinium-*p*-toluenesulfonate and 5 mg of *p*-toluenesulfonic acid. The solution was cooled in an ice water bath, and 2 mL of 2,2-dimethoxypropane was added. The solution was stirred at 60 °C for 12 h. The reaction was quenched with 5% aqueous NaHCO<sub>3</sub>, and the product was extracted with EtOAc. The EtOAc was removed *in vacuo*, and the residual material was purified by reversed-phase HPLC (Phenomenex Ultracarb C30 100 Å, 250 × 100 mm, 2.0 mL/min, UV detection at 210 nm) using a gradient solvent system from 5% to 100% CH<sub>3</sub>CN (0.05% TFA) over 60 min. The acetonide **5** (4.8 mg) was isolated with a retention time of 31 min.

**Acetonide 5:** <sup>1</sup>H NMR (500 MHz, methanol- $d_4$ )  $\delta$  7.35 (d, 1H,  $J = 8.0$  Hz), 6.99 (d, 1H,  $J = 1.5$  Hz), 6.82 (dd, 1H,  $J = 5.8, 3.0$  Hz), 6.78 (d, 1H,  $J = 5.8$  Hz), 6.59 (d, 1H,  $J = 3.0$  Hz), 6.15 (d, 1H,  $J = 1.5$  Hz), 5.94 (s, 1H), 4.83 (m, 2H), 4.65 (m, 1H), 4.55 (dd, 1H,  $J = 10.4, 10.4$  Hz), 4.31 (d, 1H,  $J = 11.0$  Hz), 3.79 (dd, 1H,  $J = 10.4, 3.5$  Hz), 3.62 (t, 1H,  $J = 3.5$  Hz), 3.55 (m, 1H), 3.44 (m, 1H), 3.04 (br s, 6H), 2.72 (dd, 1H,  $J = 12.4, 2.8$  Hz), 2.26 (t, 1H,  $J = 12.4$  Hz), 1.93 (s, 3H), 1.60 (s, 3H), 1.54 (s, 3H), 1.35 (s, 3H), 1.21 (s, 3H); ESIMS  $m/z$  745 [M + H]<sup>+</sup>, 767 [M + Na]<sup>+</sup>.

**MTPA Esters of 5 (6a, 6b).** The acetonide **5** (2.4 mg) was dissolved in freshly distilled dry pyridine (2 mL), and several dry crystals of dimethylaminopyridine were added. The mixture was stirred for 15 min at RT. Treatment with *R*-(-)-MTPA-Cl yielded the *S*-MTPA ester at room temperature after 11 h. The reaction was quenched by 1 mL

of methanol. After removal of solvent under vacuum, the residue was purified by reversed-phase HPLC (Phenomenex Luna 5u C18 (2) 100 Å, 250 × 100 mm, 2.0 mL/min, UV detection at 210 nm) using a gradient solvent system from 5% to 100% CH<sub>3</sub>CN (0.05% TFA) over 40 min. The *S*-MTPA ester was eluted at 30 min. The *R*-MTPA ester was prepared with *S*-MTPA-Cl in the same manner.  $\Delta\delta_{S-R}$  values for the *S*- and *R*-MTPA esters of acetonides were recorded in ppm in CDCl<sub>3</sub>.

**S-MTPA ester of acetonide 5 (6a):** <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.428 (d, 1H,  $J = 8.0$  Hz), 7.390 (d, 1H,  $J = 8.0$  Hz), 7.34–7.28 (m, 5H), 6.93 (s, 1H), 6.85 (d, 1H,  $J = 8.0$  Hz), 6.79 (s, 1H), 6.40 (s, 1H), 6.20 (d, 1H,  $J = 2.0$  Hz) 5.855 (dd, 1H,  $J = 12.1, 4.0$  Hz), 5.73 (m, 1H), 4.85 (m, 1H), 4.605 (t, 1H,  $J = 12.1$  Hz), 4.48 (d, 1H,  $J = 2.0$  Hz), 4.16 (d, 1H,  $J = 11.0$  Hz), 3.774 (dd, 1H,  $J = 11.0, 4.0$  Hz), 3.47 (m, 1H), 3.43 (s, 3H), 3.27 (m, 1H), 2.90 (m, 2H), 2.85 (s, 6H), 2.11 (t, 1H,  $J = 11.0$  Hz), 1.91 (s, 3H), 1.56 (s, 3H), 1.51 (s, 3H), 1.18 (s, 3H), 1.07 (s, 3H); ESIMS  $m/z$  961 [M + H]<sup>+</sup>, 983 [M + Na]<sup>+</sup>.

**R-MTPA ester of acetonide 5 (6b):** <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.428 (d, 1H,  $J = 8.0$  Hz), 7.390 (d, 1H,  $J = 8.0$  Hz), 7.34–7.28 (m, 5H), 6.90 (s, 1H), 6.84 (d, 1H,  $J = 8.0$  Hz), 6.79 (s, 1H), 6.40 (s, 1H), 6.20 (d, 1H,  $J = 2.0$  Hz), 5.845 (dd, 1H,  $J = 12.1, 4.0$  Hz), 5.70 (m, 1H), 4.85 (m, 1H), 4.660 (t, 1H,  $J = 12.1$  Hz), 4.46 (d, 1H,  $J = 2.0$  Hz), 4.15 (d, 1H,  $J = 11.0$  Hz), 3.884 (dd, 1H,  $J = 11.0, 4.0$  Hz), 3.47 (m, 1H), 3.44 (s, 3H), 3.27 (m, 1H), 2.90 (m, 2H), 2.87 (s, 6H), 2.12 (t, 1H,  $J = 11.0$  Hz), 1.91 (s, 3H), 1.61 (s, 3H), 1.51 (s, 3H), 1.18 (s, 3H), 1.07 (s, 3H); ESIMS  $m/z$  961 [M + H]<sup>+</sup>, 983 [M + Na]<sup>+</sup>.

**NF $\kappa$ B Assay.** Panomics Inc. (Fremont, CA) has established NF $\kappa$ B reporter stable cells in a number of popular cell lines suitable for specific applications in the study of NF $\kappa$ B. We employed human embryonic kidney cells 293 (Panomics Inc., Fremont, CA) for monitoring any changes occurring along the NF $\kappa$ B pathway. Stably transfected cells

were seeded into sterile 96-well plates at a density of  $20 \times 10^3$  cells per well. Cells were maintained in Dulbecco's modified Eagle's medium (Invitrogen, Carlsbad, CA), supplemented with 10% FBS, 100 units/mL penicillin, 100  $\mu\text{g}/\text{mL}$  streptomycin, and 2 mM L-glutamine. After an incubation period of 48 h, the medium was replaced and various concentrations of test chemicals (dissolved in PBS) were added. Human recombinant TNF- $\alpha$  (Calbiochem, Gibbstown, NJ) was used as an activator (2 ng/mL). Following a 6 h incubation period, the medium was discarded and the cells were washed once with PBS. The cells were then lysed by adding 50  $\mu\text{L}/\text{well}$  (diluted with water five times) of Reporter Lysis Buffer (Promega, Madison, WI) and incubating for 5 min with shaking. The plates were then stored at  $-80^\circ\text{C}$  until the luciferase assay was performed using the Luc assay system from Promega.<sup>20</sup> The gene product, luciferase enzyme, reacts with luciferase substrate, emitting light, which was detected in a luminometer (LUMistar Galaxy BMG). Data are expressed as  $\text{IC}_{50}$  values (i.e., concentration required to inhibit TNF- $\alpha$ -activated NF $\kappa$ B activity by 50%).

**Quinone Reductase 1 Assay.** QR1 activity was assessed in 96-well plates using Hepa 1c1c7 murine hepatoma cells as previously described.<sup>26</sup> Briefly, cells were grown to a density of  $2 \times 10^4$  cells/mL in 200  $\mu\text{L}$  of MEM- $\alpha$  containing 5% antibiotic-antimycotic (Gibco) and 10% fetal bovine serum at  $37^\circ\text{C}$  in a 5%  $\text{CO}_2$  atmosphere. After preincubation for 24 h, the media was changed and test compounds were added to a final concentration of 20  $\mu\text{g}/\text{mL}$ . For dose dependence, samples were added in five serial 3-fold dilutions, starting at 20  $\mu\text{g}/\text{mL}$ . The cells were incubated with test samples for an additional 48 h. Quinone reductase activity was measured as a function of the NADPH-dependent menadiol-mediated reduction of 3-(4,5-dimethylthiazo-2-yl)-2,5-diphenyltetrazolium bromide (MTT) to a blue formazan.<sup>23</sup> Cytotoxicity was determined via crystal violet staining of identical plates. Specific activity is defined as nmol of formazan formed per mg protein per min.<sup>27</sup> The induction ratio of QR activity represents the specific enzyme activity of agent-treated cells compared with a DMSO-treated control. The concentration to double activity (CDI) was determined through a dose-response assay for active substances (induction ratio  $> 2$ ).

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**Supporting Information Available:**  $^1\text{H}$ ,  $^{13}\text{C}$ , and 2D NMR spectra of fijiolide A (1) and  $^1\text{H}$  spectra of fijiolide B (2), MTPA esters 3a, 3b, 4a, 4b, 6a, 6b, and acetone 5 are available free of charge via the Internet at <http://pubs.acs.org>.

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