A STEREOSELECTIVE TOTAL SYNTHESIS OF (9S)-9-DIHYDROERYTHRONOLIDE A FROM D-GLUCOSE¹

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<u>Summary</u> A rather facile stereoselective total synthesis of (9S)-9-dihydroerythronolide A starting from D-glucose was achieved via coupling of C1-C6 and C7-C15 segments and subsequent macrolactonization.

The well known macrolide antibiotic erythromycin A (1) has been an attractive synthetic target for modern synthetic chemists, because of its complex chemical structure as well as important biological activity, and many new synthetic methodologies have been developed.² As part of our synthetic studies of macrolide and polyether antibiotics by a common methodology,³ we report here a rather facile total synthesis of 9-dihydroerythronolide A (3)⁷ by use of some stereoselective reactions and suitable protecting groups. Two segments 4 (C7-C15)⁸ and 5 (C1-C6) were first chosen as synthetic intermediates in one of our various synthetic approaches to 1, 2, and 3.



The triol $(6)^9$ derived from D-glucose was first converted to a monobenzoate, and then the remaining diol was protected with a p-methoxyphenylethylidene group.¹⁰ After hydrolysis of the benzoate, Swern oxidation¹¹ readily gave the aldehyde (7), which was subjected to the Wittig reaction, followed by reduction to give the (E)-allyl alcohol (8). m-CPBA oxidation of 8 at -10 °C gave the epoxide (9) with 8 : 1 stereoselectivity. The regio- and stereoselective reductive opening of the epoxide ring was one of crucial steps in this total synthesis, and only the desired alcohol (10) was obtained in high yield by treatment with a large excess of NaBH₃CN in the presence of BF₃·OEt₂ in refluxing THF.^{12,13} Benzoyl protection of the primary alcohol of 10 followed by selective removal of the ketal protection gave the triol (11). When 11 was treated with 2-methoxypropene in the presence of a catalytic amount of PPTS at room temperature,^{2a} the expected kinetic product, 9,11-O-isopropylidene compound,¹⁶ was isolated



(A) 1) BzCl, Py, CH₂Cl₂, rt, 100%; 2) p-MeOC₆H₄CMe(OMe)₂, CSA, rt, 95%; 3) 1N-KOH, MeOH, rt, 98%; 4) (COCl)₂, DMSO, Et₃N, 100%. (B) 1) Ph₃P=CMeCO₂Et, EDC, reflux, 100%; 2) LiAlH₄, Et₂O, O°C, 94%. (C) m-CPBA, CH₂Cl₂, -15~-10°C, 95%. (D) NaBH₃CN (12 eq), BF₃'OEt₂ (6 eq), THF, reflux, 89%. (E) 1) BzCl, Py, CH₂Cl₂, rt, 84%; 2) 4N-HCl, THF, rt, 84%. (F) 1) CH₂=C(Me)OMe, PPTS, CH₂Cl₂, rt, 89%; 2) MMCl, i-Pr₂EtN, CH₂Cl₂, 45°C, 100%. (G) 1) 1N-NaOH, dioxane, 65~75°C, 100%; 2) TsCl, Et₃N, DMAP, CH₂Cl₂, reflux, 100%; 3) PhSNa, EtOH-DME, reflux, 90%; 4) NaIO₄, MeOH-H₂O, rt, 97%.

in high yield. MM protection of the remaining tertiary alcohol readily gave 12, which was converted to the C7-C15 segment (4) via treatment of a tosylate with PhSNa in excellent vield.^{17,18}

The diol $(13)^{19}$ derived also from D-glucose was protected as an acetonide, then removal of the Bn group followed by PCC oxidation gave the aldehyde, which was treated with the carbanion of 1,3-dithiane, and a stereoisomeric mixture (4.6 : 1) mainly consisting of the Cram adduct (14) was obtained. The acetonide group of 14 was removed, then the resulting primary alcohol was protected with a TBDPS group, and protection of the remaining diol gave 15, whose configuration was confirmed by observation of the NOE (8%) between an acetonide methyl group and each of Ha and Hb in the NMR spectrum. Finally, treatment of 15 with Mel²⁰ gave the C1-C6 segment (5) in high yield.²¹



(H) 1) Me₂C(OMe)₂, CSA, CH₂Cl₂, rt; 2) 10%Pd-C, H₂, EtOH, 96% (overall); 3) PCC, molecular sieves, CH₂Cl₂, rt, 82%; 4) 1.3-dithiane, n-BuLi, THF, -75~-70°C, 86%. (I) 1) TsOH H₂O, MeOH, rt, 92%; 2) TBDPSC1, imidazole, DMF, rt, 93%; 3) CH₂=C(Me)OMe, PPTS, rt, 95%. (J) MeI, NaHCO₃, 90%MeCN, 70°C, 88%.

Coupling of 5 and 4 (2 eq) in the presence of LDA (2 eq) at -80~-70 °C proceeded quite smoothly to give a stereoisomeric mixture of 16 in high yield.²³ Desulfurization with Raney Ni and subsequent Swern oxidation¹¹ readily gave the 6-keto compound, which was treated with a large excess of MeLi in THF at -85 °C. The Cram addition (non-chelation) took place rather rapidly to give 17 in excellent yield (7.2 : 1 stereoselectivity).²⁴ Thus all chiral centers required for 3 were constructed. Four step conversion of 17 [MM protection of C6-OH group, desilylation with F⁻, Jones oxidation of the resulting primary alcohol, and final removal of the MPM protection by catalytic reduction or DDQ oxidation²⁵] gave the fully protected (except 1-C0₂H and 13-OH) seco-acid (18) in good yield. Macrolactonization of 18 was achieved only by Yamaguchi's method²⁶ at rather high concentration of DMAP. When a 2 mM toluene solution of a mixed anhydride, prepared from 18 and 2,4,6-trichlorobenzoyl chloride in the presence of Et₃N, was added dropwise to an equal volume of refluxing 50 mM toluene solution of DMAP (25 eq) over a period of 39 h, the expected lactone (19) was isolated in 27% yield.^{27,28} Removal of the protecting groups of 19 with 50% AcOH readily gave the title compound (3).²⁹ More efficient syntheses of 2 and 3 will be reported later.



(K) LDA, THF, -80~-70°C, 1.5h, 85%. (L) 1) Raney Ni (W-2), EtOH, rt, 83%; 2) (COC1)₂, DMSO, Et₃N, 92%; 3) MeLi (15 eq), THF, -85°C, 1h, 95%. (M) 1) MMC1, i-Pr₂EtN, CH₂C1₂, 50~55°C, 100%; 2) n-Bu₄NF, THF, 55~60°C, 94%; 3) 2.67M Jones reagent, Me₂CO, -17°C, 78%; 4) 10%Pd-C, H₂, EtOH, 55°C, 99%. (N) i) 2.4,6-C1₃C₆H₂COC1, Et₃N, THF, ii) DMAP (25 eq), toluene, reflux, 39h, 27%. (0) 50%AcOH, rt, 91%.

REFERENCES AND NOTES

- Chiral synthesis of polyketide-derived natural products. 18. For part 17, see: K. Horita, S. Nagato, Y. Oikawa, and O. Yonemitsu, <u>Tetrahedron Lett.</u>, 28, 0000 (1987).
- 2) Successful total synthetic studies directed toward 1, 2, and 3: a) E. J. Corey, P. B. Hopkins, S. Kim, S. Yoo, K. P. Nambiar, and J. R. Falk, <u>J. Am. Chem. Soc.</u>, 101, 7131 (1979); b) R. B. Woodward, et al., <u>J. Am. Chem. Soc.</u>, 103, 3210, 3213, 3215 (1981); c) B. Bernet, P. M. Bishop, M. Caron, T. Kawamata, B. L. Roy, L. Ruest, G. Sauve, P. Soucy, and

- K. Tomooka, and M. Nakata, Tetrahedron Lett., 27, 1814, 1815 (1986); e) G. Stork and S.
- D. Rychnovsky, <u>J. Am. Chem. Soc.</u>, 109, 1565 (1987); f) T. Nakata, M. Fukui, and T. Oishi, to be published.
- 3) Recently we reported highly storeoselective syntheses of macrolide aglycons such as methynolide,⁴ pikronolide,⁵ and tylonolide.⁶
 4) Y. Oikawa, T. Tanaka, and O. Yonemitsu, <u>Tetrahedron Lett</u>., 27, 3647 (1986).
- 5) N. Nakajima, T. Hamada, T. Tanaka, Y. Oikawa, and O. Yonemitsu, J. Am. Chem. Soc., 108, 4645 (1986).
- 6) T. Tanaka, Y. Oikawa, T. Hamada, and O. Yonemitsu, Tetrahedron Lett., 27, 3651 (1986).
- 7) ${\bf 3}$ was already converted to ${\bf 1}^{2b}$ and ${\bf 2.}^{2d}$
- 8) Unless otherwise noted, the numberings are based on that of 3.
- 9) Y. Oikawa, T. Nishi, and O. Yonemitsu, J. Chem. Soc. Perkin 1, 7 (1985), and alternative syntheses of 6 from propionaldehyde will be reported soon.
- 10) B. H. Lipshutz and M. C. Morey, J. Org. Chem., 46, 2419 (1981).
- 11) A. J. Mancuso, S.-L. Huang, and D. Swern, <u>J. Org. Chem.</u>, 43, 248 (1978).
- 12) cf. R. O. Hutchins, I. M. Taffer, and W. Burgoyne, <u>J. Org. Chem.</u>, 46, 5214 (1981).
- 13) Use of more excess NaBH₃CN than $BF_3^{\bullet}OEt_2$ was very important. Other reduction methods (with Red-al in THF, ¹⁴ AICl₃ and LiAlH₄ in ether, ¹⁵ etc.) gave only poor results.
- 14) S. M. Viti, Tetrahedron Lett., 23, 4541 (1982); P. Ma, V. S. Martin, S. Masamune, K. B. Sharpless, and S. M. Viti, J. Org. Chem., 47, 1378 (1982); M. Honda, T. Katsuki, and M. Yamaguchi, Tetrahedron Lett., 25, 3857 (1984).
- 15) N. M. Yoon and C. H. Brown, J. Am. Chem. Soc., 90, 2927 (1968).
- 16) Protection of 9,11-diol, not of 11,12-diol, as a cyclic acetal is extremely important for the macrolactonization.^{2b,d,e}
- 17) cf. a) G. Stork, I. Paterson, and F. K. C. Lee, <u>J. Am. Chem. Soc.</u>, 104, 4686 (1982); b) 1. Paterson, <u>Tetrahedron Lett.</u>, 24, 1311 (1983).
- 18) 4 is a l : 1 stereoisomeric mixture with regard to the SO position.
- 19) Y. Oikawa, T. Nishi, and O. Yonemitsu, <u>J. Chem. Soc. Perkin 1</u>, 19 (1985), and an alternative synthesis of 13 from methally1 alcohol via the Sharpless asymmetric epoxidation will be reported soon.
- 20) M. Fetizon and M. Jurion, <u>J. Chem. Soc. Chem. Comm.</u>, 382 (1972).
- 21) Corey's method²² (NBS, 2,6-lutidine, 80% MeCN, rt) also gave 5 in 71% yield.
- 22) E. J. Corey and B. W. Erickson, <u>J. Org. Chem.</u>, 36, 3553 (1971).
- 23) Most of excess 4 was recovered (86%). Coupling between 4 and the 6-methylketone derived from 5^{17a} gave only poor results in <20% yield.
- 24) Treatment of the ketone with MeLi (10 eq) in the presence of HMPA (40 eq) in ether at -86 $^\circ \mathrm{C}$ for 1 h also gave 17 in 95% yield with 5.6 : 1 stereoselectivity. On the other hand, in the absence of HMPA, the chelation controlled addition of MeLi in ether occurred quite smoothly to give the C6 epimer of 17 in quantitative yield with 5: 1 stereoselctivity. When the ketone was treated with MeMgI in ether or with MeCeCl $_2$ in THF, no reaction occurred.
- 25) a) Y. Oikawa, T. Yoshioka, and O. Yonemitsu, Tetrahedron Lett., 23, 885 (1982); b) K. Horita, T. Yoshioka, T. Tanaka, Y. Oikawa, and O. Yonemitsu, Tetrahedron, 42, 3021 (1986).
- 26) J. Inanaga, K. Hirata, H. Saeki, T. Katsuki, and M. Yamaguchi, Bull. Chem. Soc. Jpn., 52, 1989 (1979).
- 27) In this macrolactonization, concentration of DMAP was very important. Yields of 19 in various concentrations of DMAP are as follws [final mM of DMAP (% yield of 19)]: 25 mM (27%); 9.5 mM (14%); 6 mM (13%); 3 mM (0%).
- 28) As pointed out by Stork,^{2e} an axial methyl group of 9,11-acetonide protection hindered this lactonization. Actually we could not obtain 19 by the modified Corey method. 2b and also 20 gave only a trace of lactonization product, which was, however, obtained in 63% yield by highly reactive Yamaguchi's method 26 at high concentration of DMAP.
- 29) 3, 18, 19, and a 6-0-MM derivative of 21 were identical with the respective authentic samples derived from natural erythromycin A in terms of JR, NMR, and MS spectra.

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