

The Total Synthesis of AB3217-A, a Novel Anti-mite Substance, via Intermolecular Etherification and Intramolecular Glycosylation

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The first total synthesis of AB3217-A has been achieved. The deacetylanisomycin unit, (3*S*,4*S*,5*R*)-3-benzyloxy-1-(benzyloxycarbonyl)-5-[(1*R*)-1-hydroxy-1-(4-methoxyphenyl)methyl]-4-(2-tetrahydropyranyloxy)-pyrrolidine (**18**), was prepared from dimethyl L-tartrate by a stereoselective pyrrolidine-ring formation and a stereoselective reduction of phenyl ketone as the key steps. The lithium alkoxide of **18** was coupled with the D-xylofuranose unit, phenyl 2,3-di-*O*-benzyl-5-*O*-trifluoromethanesulfonyl-1-thio- α -D-xylofuranoside which was prepared from 1,2:5,6-di-*O*-isopropylidene- α -D-glucofuranose, via an intermolecular etherification. The resulting coupling product was subjected to de-*O*-tetrahydropyranylation and an intramolecular glycosylation to afford 4,12,13-tribenzyl-6-(benzyloxycarbonyl)AB3217-A (**30**). Final deprotection of **30** furnished AB3217-A.

AB3217-A (**1a**) was isolated in 1989 from the fermentation broth of the strain of *Streptomyces platensis* AB3217.¹⁾ Two new substances, AB3217-B (**1b**) and AB3217-C (**1c**), the C13-ester derivatives of **1a**, were also isolated from the same strain.^{1a,2)} They showed marked activity against the two spotted spider mite, *Tetranychus urticae*.^{1,2)} The structure of **1a** was determined by spectroscopic means and its absolute configuration was determined by X-ray crystallographic analysis.¹⁾ The structures of **1b** and **1c** were determined on the basis of spectroscopic studies in comparison with **1a**.^{1a,2)} The characteristic structure is a novel nine-membered ring built of deacetylanisomycin and β -D-xylofuranose, which are linked through glycosidic and ether bonds. We wish to describe in this full account³⁾ the details of the first total synthesis of AB3217-A (**1a**). It was divided into the deacetylanisomycin unit and the D-xylofuranose unit. On the basis of molecular model studies, we anticipated that the glycosidic bond could be formed in a β -fashion by an intramolecular glycosylation after connecting two units by an intermolecular etherification (Fig. 1). Although glycosylations are among the most widely-used reactions in the synthesis of biologically active substances,⁴⁾ intramolecular glycosylations have been rarely used in the synthesis of natural products.⁵⁾

Results and Discussion

Synthesis of Deacetylanisomycin Unit. Synthesis of the pyrrolidine subunit began with the known diol **2**,⁶⁾ which was prepared from dimethyl L-tartrate in two steps, by the modified Seebach procedures (Scheme 1).⁶⁾ Mono-methoxybenzylation of **2** with 4-methoxybenzyl chloride (MPMCl) and NaH in DMF gave **3** in 76% yield. Tosylation of **3** followed by one-pot transformation of the resulting tosylate with *dl*-10-

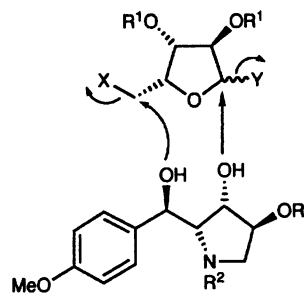
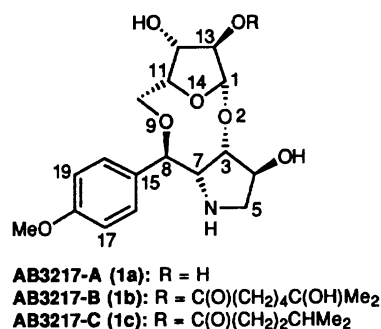
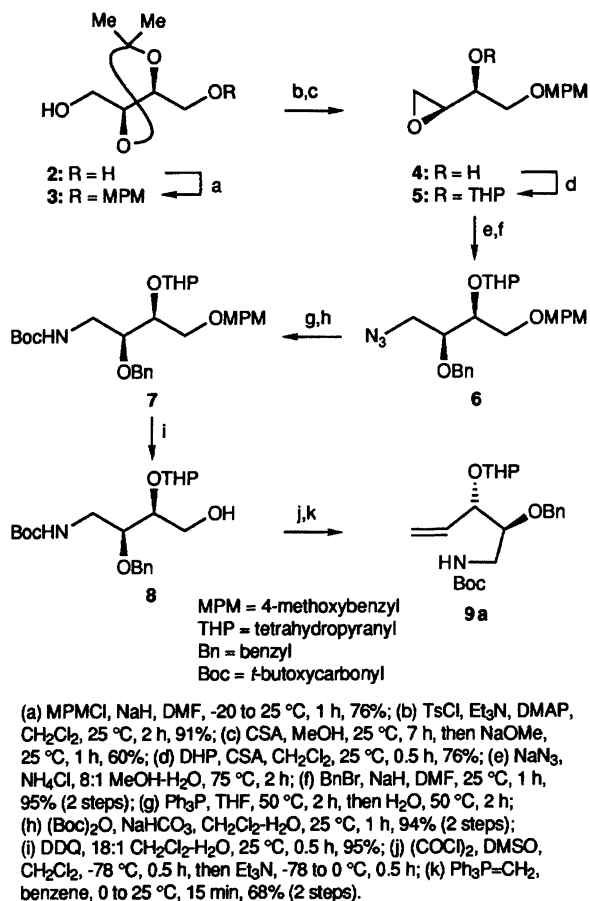


Fig. 1.

camphorsulfonic acid (CSA) and sodium methoxide in MeOH afforded epoxide **4** in 55% yield. Protection of **4** with 3,4-dihydro-2*H*-pyran (DHP) and CSA in CH₂Cl₂ provided **5** in 76% yield. All compounds having tetrahydropyranyl (THP) ether in this account consist of a 3:2—2:1 anomeric mixture, which were not separated. The epoxide-opening of **5** with NaN₃—NH₄Cl⁷⁾ in 8:1 MeOH—H₂O gave azido alcohol as a single isomer, which was benzylated with benzyl bromide and NaH in DMF to afford **6** in 95% yield. The epoxide-opening of other derivatives, having the benzoyl or *t*-butyldimethylsilyl (TBS) protecting group instead of the THP protecting group in **5**, resulted in a concomitant migration of these protecting groups. Reduction of the azido function of **6** with triphenylphosphine in aqueous THF and the resulting amine was protected with the *t*-butoxycar-

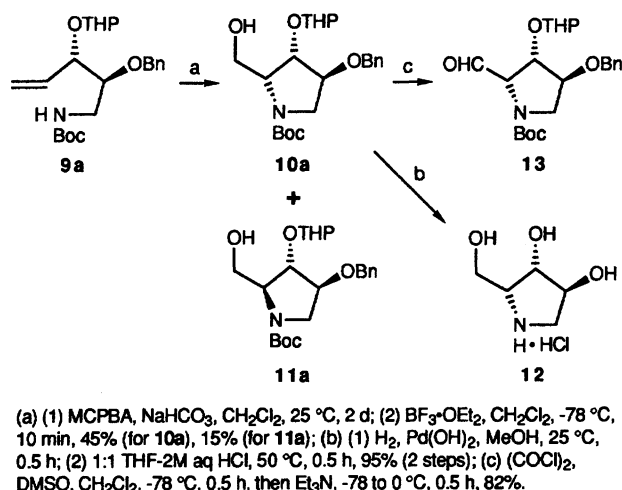
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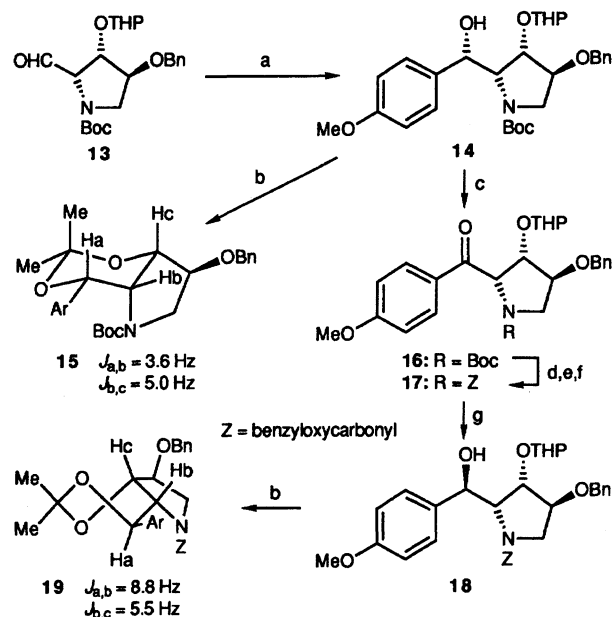
Scheme 1.

bonyl (Boc) group to afford **7** in 94% yield. Deprotection of **7** with 2,3-dichloro-5,6-dicyano-*p*-benzoquinone (DDQ) gave alcohol **8** in 95% yield, which was subjected to Swern oxidation and Wittig olefination with methylenetriphenylphosphorane to give **9a** in 68% yield. The first crucial step for the synthesis of the deacetylanisomycin unit was the pyrrolidine-ring formation, which was realized by a method conceptually similar to those of Joullié⁸) or Takahata-Momose⁹) (Scheme 2). Namely, epoxidation of **9a** with MCPBA followed by BF₃·OEt₂ treatment gave pyrrolidines **10a** and **11a** in 45 and 15% yields, respectively. A nonstereoselective pyrrolidine-ring formation or no reaction was observed when other amino olefins **9b**–**9d** were subjected to the conditions summarized in Table 1. Although the identical isolated yield of **10** was obtained in Entries 1–3, **9a** was the best choice because of total efficiency of the synthesis. The newly generated stereocenter in the major pyrrolidine **10a** was confirmed by its conversion to the known hydrochloride of 1,4-dideoxy-1,4-imino-D-xylitol (**12**)¹⁰) by hydrogenolysis and acidic hydrolysis. Swern oxidation of **10a** gave aldehyde **13** in 82% yield.

The above aldehyde **13** was coupled with 4-anisyllithium prepared from 4-bromoanisole and *n*-BuLi in THF⁹) to afford **14** in 80% yield as a single isomer (Scheme 3). The configuration at the benzylic stereo-



Scheme 2.

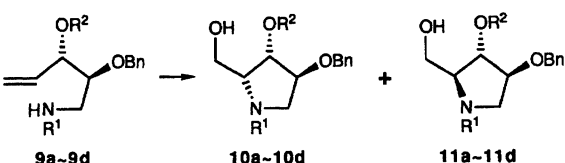


Scheme 3.

center was determined by the coupling constants of the ¹H NMR spectrum of acetone **15** obtained from **14** by acidic treatment and acetone. This remarkable, but undesirable, facial selectivity may arise from NBoc-assisted addition of anisyllithium to the aldehyde.^{8,9,11})

To invert the benzylic configuration, **14** underwent Swern oxidation to give **16** in 88% yield. Acidic treatment of **16** followed by successive *N*-benzyloxycarbonylation and *O*-protection provided **17** in 85% yield. It was necessary to alter the *N*-protecting group from Boc to benzyloxycarbonyl (Z) group because final deprotection of **30** (R²=Boc, vide infra) under acidic conditions resulted in failure. Finally, diisobutylaluminum hydride

Table 1. Pyrrolidine-Ring Formation

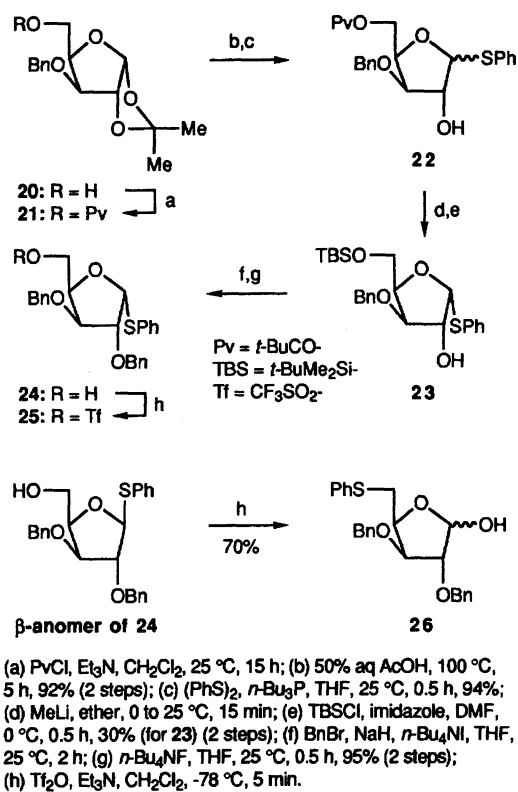


| Entry | 9 | Conditions ^{a)} | Isolated yield | |
|-------|--|--------------------------|--------------------|-----|
| | | | 10 | 11 |
| 1 | 9a : ^{b)} R ¹ =Boc, R ² =THP | A | 45% | 15% |
| 2 | 9b : ^{b)} R ¹ =Boc, R ² =TBS | A | 45% | 47% |
| 3 | 9c : ^{b)} R ¹ =Boc, R ² =H | A | 45% | 47% |
| 4 | 9c | B | Almost no reaction | |
| 5 | 9d R ¹ =Z, R ² =H | B | Almost no reaction | |

a) (A) (1) MCPBA (3 equiv), NaHCO₃ (1.5 equiv), CH₂Cl₂, 25 °C, 2 d; (2) BF₃·OEt₂ (1.2 equiv), CH₂Cl₂, -78 °C, 10 min (see Ref. 8). (B) D-(-)-Diisopropyl tartrate, *t*-butyl hydroperoxide, Ti(O-*i*-Pr)₄, MS3AP, CH₂Cl₂, -20 °C, 15 d (see Ref. 9). b) The corresponding Z-derivatives (R¹=Z) gave low yields of **10** and **11**.

(DIBAL) reduction of **17** afforded **18** in 91% yield as a single isomer. This desirable facial selectivity may arise from NZ-assisted addition of hydride to the ketone.¹²⁾ The newly created stereocenter in **18** was confirmed by the coupling constants of the ¹H NMR spectrum of acetone **19** derived from **18**.¹³⁾

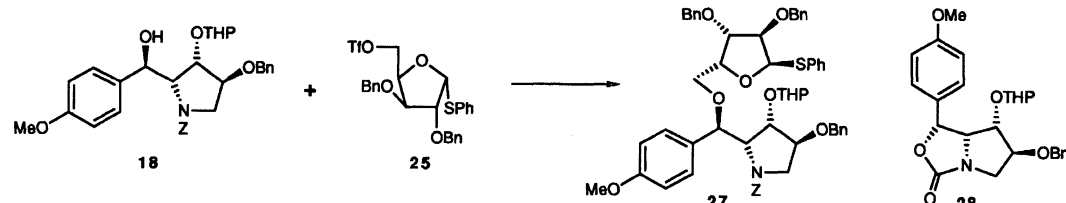
Synthesis of D-Xylofuranose Unit. Synthesis of the D-xylofuranose unit was initiated from **20**, which was derived from 1,2:5,6-di-*O*-isopropylidene- α -D-glucofuranose in four steps (Scheme 4).¹⁴⁾ Pivaloylation of **20** with pivaloyl chloride (PvCl) and triethylamine in CH₂Cl₂ afforded **21**. Although the acetyl derivative of **20** (R=Ac) was used in the initial studies,³⁾ the pivaloate **21** was selected for the large scale preparation because the pivaloyl group was more stable than the acetyl group on the next deacetoization. After deacetoization of the crude **21** in 50% aqueous acetic acid at 100 °C for 5 h, the resulting free sugar (92% yield from **20**) was treated with diphenyl disulfide and tributylphosphine in THF^{15,16)} to afford a 1:2 mixture of α : β anomers **22** in 94% combined yield. Since this mixture could not be separated, it was subjected to de-*O*-pivaloylation with methyllithium in ether and the subsequent silylation with TBSCl to afford the separable phenyl α -thioglycoside **23** and its β -anomer in 30 and 59% yields, respectively. Benzoylation and subsequent de-*O*-silylation of **23** gave **24** in 95% yield. The anomeric configurations of **24** and its β -anomer (and hence **23** and its β -anomer) were determined by their ¹H NMR NOE experiments: for **24**, H-1 \rightarrow H-2, 9.7%. For β -anomer, H-1 \rightarrow H-2, 4.0%; H-1 \rightarrow H-3, 2.1%; H-1 \rightarrow H-4, 2.3%. Finally, introduction of a leaving group to **24** was realized by treatment of **24** with trifluoromethanesulfonic anhydride (Tf₂O) and triethylamine in CH₂Cl₂ at -78 °C for 5 min to afford the extremely labile triflate **25**, which was immediately subjected to the next reaction. Etherification of other derivatives



Scheme 4.

of **24** (R=Ms or Ts, or OR=I) with the model alkoxide, which was derived from 4-methoxybenzyl alcohol or 1-(4-methoxyphenyl)-2-methyl-1-propanol, under a variety of conditions was unsuccessful, giving either the recovered starting materials or decomposition depending on the conditions employed. Triflation of the corresponding β -anomer of **24** was also unsuccessful, giving **26** via 1,5-SPh group migration.¹⁷⁾ The structure of **26** was confirmed by the ¹H NMR and mass spectra. Al-

Table 2. Intermolecular Etherification of the Deacetylanisomycin Unit and the D-Xylofuranose Unit



| Entry | Base ^{a)} | Solvent | Equiv of 25 ^{b)} | Conditions ^{c)} | Isolated yield/% | | |
|-------|--------------------------------------|---------|----------------------------------|--------------------------|------------------|-----------|-----------|
| | | | | | 27 | 18 | 28 |
| 1 | KN(SiMe ₃) ₂ | DMF | 1.5 | A | Trace | 82 | Trace |
| 2 | NaN(SiMe ₃) ₂ | DMF | 1.5 | A | 20 | 0 | 50 |
| 3 | LiN(<i>i</i> -Pr) ₂ | DMF | 1.5 | A | 33 | 57 | 0 |
| 4 | <i>n</i> -BuLi | DMF | 1.5 | A | 35 | 51 | 0 |
| 5 | LiN(<i>i</i> -Pr) ₂ | HMPA | 1.1 | A | 30 | 0 | 56 |
| 6 | LiN(<i>i</i> -Pr) ₂ | HMPA | 1.5 | A | 31 | 42 | 0 |
| 7 | LiN(<i>i</i> -Pr) ₂ | HMPA | 2.1 | A | 30 | 60 | 0 |
| 8 | <i>n</i> -BuLi | DMF | 1.5 | B | 55 | Trace | 33 |
| 9 | <i>n</i> -BuLi | DMF | 3.0 | B | 37 | 57 | Trace |
| 10 | <i>n</i> -BuLi | DMF | 1.5 | C | 64 | 20 | Trace |
| 11 | <i>n</i> -BuLi | DMF | 2.0 | C | 75 | Trace | Trace |

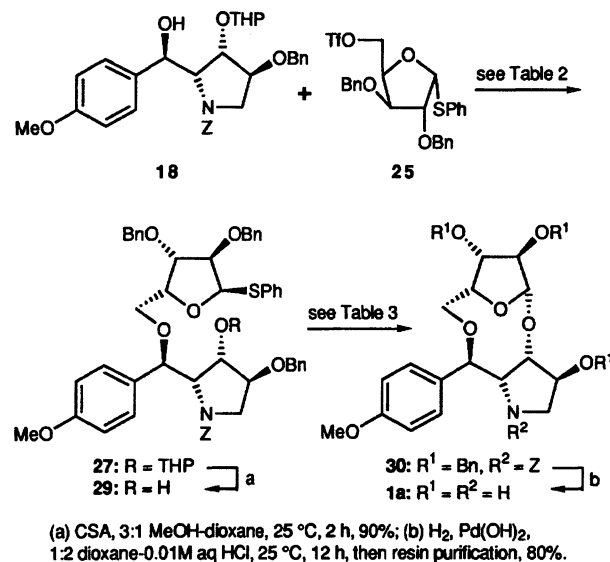
a) 1.5 equiv (for **18**) of base was used. b) Equiv of **25** for **18**. c) (A) Base was added at 0 °C to a mixture of **18**, **25**, MS4AP, and solvent. The reaction mixture was stirred at 25 °C for 0.5 h. (B) Base was added at 0 °C to a mixture of **18**, MS4AP, and DMF. After 5 min at 0 °C, **25** in DMF was added and the mixture was stirred at 25 °C for 0.5 h. (C) After washing a hexane solution of the crude **25** with water, the residual **25** was subjected to the conditions B.

Table 3. Intramolecular Glycosylation of **29** with NBS

| Entry | Solvent | Temp/°C | Time/h | Yield of 30 |
|-------|---------------------------------|---------|--------|--------------------|
| 1 | CH ₂ Cl ₂ | 25 | 1 | Decomp |
| 2 | CH ₃ CN | 25 | 24 | 30% |
| 3 | CH ₃ NO ₂ | 25 | 30 | 29 +Decomp |
| 4 | THF | 25 | 20 | Decomp |
| 5 | Toluene | 60 | 6d | 40% |
| 6 | Toluene | 90 | 24 | 64% |
| 7 | Toluene | 110 | 2 | 46% |

though it was disappointing not to be able to utilize the β -anomer of **24**, it was decided to investigate the intermolecular etherification of the deacetylanisomycin and D-xylofuranose units.

Intermolecular Etherification. We examined a number of conditions to achieve this connection; the relevant data are summarized in Table 2. Since the cyclic urethane **28** was formed in 89% yield when **18** was treated with lithium diisopropylamide (LDA) in DMF at 0 to 25 °C for 0.5 h, the base was added after mixing **18**, **25**, molecular sieves 4A powder (MS 4AP), and solvent. It was necessary to add MS 4AP to the reaction mixture because triflate **25** was very unstable in DMF without MS 4AP. Among the bases employed, lithium bases were better than potassium and sodium bases (Entries 1–4). Hexamethylphosphoric triamide (HMPA) could be also used as a solvent (Entries 5–7). It was found that the work-up procedure in the preparation of triflate **25** was important to obtain a good



Scheme 5.

yield of the coupling product (Entries 8–11; see footnote c of Table 2 and Experimental), giving 75% yield of **27** reproducibly (Entry 11). Acidic treatment of **27** afforded **29** in 90% yield (Scheme 5).

Intramolecular Glycosylation, Final Stage. The ultimate intramolecular glycosylation of **29** was accomplished by *N*-bromosuccinimide (NBS)¹⁶ in several solvents (Table 3). The stability of the oxonium ion derived from **29** and NBS depended on the solvent employed. Toluene proved to be the best solvent, providing

30 in 64% yield. A final deprotection of **30** by hydrogenolysis in a mixture of 1:2 dioxane–0.01 M (1 M = 1 mol dm⁻³) aqueous HCl and resin purification furnished AB3217-A (**1a**) in 80% yield. The synthetic **1a** was identical in all respects (¹H NMR, IR, UV, mp, [α]_D, and TLC mobilities) with the natural AB3217-A.¹⁾

Experimental

The melting points were determined on a micro hot-stage Yanaco MP-S3 and were uncorrected. Optical rotations were measured on a JASCO DIP-360 photoelectric polarimeter in chloroform unless otherwise noted. IR spectra were recorded on a BIO RAD DIGILAB FTS-65 spectrometer and ¹H NMR spectra were on a JEOL GSX270 spectrometer in CDCl₃ at 25 °C using TMS as internal standard or in DMSO-*d*₆ at 80 °C (DMSO = 2.50) unless otherwise noted. Mass spectra (EI) were recorded on a JEOL JMS-DX302 mass spectrometer. Silica-gel TLC and column chromatography were performed on a Merck TLC 60F-254 and a Fuji-Davison BW-820MH, respectively. Air- and/or moisture-sensitive reactions were carried out under an atmosphere of argon with oven-dried glassware. In general, the organic solvents were purified and dried by appropriate procedures, and evaporation and concentration were carried out under reduced pressure below 30 °C, unless otherwise noted.

(2S,3S)-2,3-Isopropylidenedioxy-4-(4-methoxybenzyloxy)-1-butanol (3). To a stirred suspension of NaH (2.41 g, 100 mmol) in dry DMF (750 ml) was added at -20 °C a solution of **2** (15.1 g, 93.1 mmol) in dry DMF (75 ml). After 0.5 h at -20 °C, MPMCl (13.2 ml, 97.4 mmol) was added and the mixture was warmed to 25 °C during 1 h. The reaction mixture was concentrated at 45 °C and to the residue were added ether and saturated aqueous NH₄Cl. The aqueous layer was extracted with ether and the combined organic layers were washed with saturated aqueous NaCl, dried, and concentrated. The residue was chromatographed on silica gel (1 kg) with 2:1 hexane–ethyl acetate to afford **3** (20.0 g, 76%) as a colorless syrup: *R*_f = 0.50 (3:2 hexane–ethyl acetate); [α]_D²⁰ +10.6° (*c* 1.06); IR (CHCl₃) 3599, 3459, 3019, 2877, 1613, 1514, 1373, 1249, 1172, 1081, and 1038 cm⁻¹; ¹H NMR (CDCl₃) δ = 1.41 (6H, s, CMe₂), 2.25 (1H, dd, *J* = 8.0 and 4.5 Hz, OH), 3.52 (1H, dd, *J*_{3,4} = 5.9 Hz, *J*_{gem} = 9.9 Hz, H-4), 3.62–3.80 (3H, m, H-1, H-1', and H-4'), 3.81 (3H, s, OMe), 3.92 (1H, dt, *J*_{1,2} = *J*_{1',2} = 4.3 Hz, *J*_{2,3} = 8.2 Hz, H-2), 4.03 (1H, ddd, *J*_{2,3} = 8.2 Hz, *J*_{3,4} = 5.9 Hz, *J*_{3,4'} = 4.9 Hz, H-3), 4.52 (2H, s, OCH₂Ar), 6.88 and 7.24 (each 2H, each d, *J* = 8.5 Hz, aromatic protons). Found: *m/z* 283.1559 (M+H)⁺. Calcd for C₁₅H₂₃O₅: M+1, 283.1546.

(2S,3S)-3,4-Epoxy-1-(4-methoxybenzyloxy)-2-butanol (4). To a stirred solution of **3** (33.1 g, 117 mmol) in dry CH₂Cl₂ (500 ml) were added at 0 °C *p*-toluenesulfonyl chloride (24.6 g, 129 mmol), triethylamine (24.5 ml, 176 mmol), and 4-dimethylaminopyridine (DMAP) (2.86 g, 23.4 mmol). After 2 h at 25 °C, the reaction mixture was poured into cold saturated aqueous NH₄Cl and the new mixture was extracted with ethyl acetate. The extracts were washed with saturated aqueous NaCl, dried, and concentrated. The residue was chromatographed on silica gel (1.2 kg) with 3:1 hexane–ethyl acetate to afford a colorless syrup (46.7 g, 91%). This was dissolved in dry MeOH (935 ml) and to this

was added CSA (4.97 g, 21.4 mmol). After 7 h at 25 °C, the reaction mixture was cooled to 0 °C and to this was added NaOMe (6.94 g, 128 mmol); the new mixture was stirred at 25 °C for 1 h. The reaction mixture was treated with CG-50 in MeOH and the insoluble materials were filtered and washed with MeOH. The combined filtrate and washings were concentrated. The residue was dissolved in ethyl acetate and this was washed with water and saturated aqueous NaCl, dried, and concentrated. The residue was chromatographed on silica gel (1.2 kg) with 3:1 and then 1:1 hexane–ethyl acetate to afford **4** (14.5 g, 60%) as a colorless syrup: *R*_f = 0.35 (1:1 hexane–ethyl acetate); [α]_D²³ +13.6° (*c* 1.03); IR (CHCl₃) 3564, 3019, 1613, 1514, 1250, 1175, 1101, and 1035 cm⁻¹; ¹H NMR (CDCl₃) δ = 2.25 (1H, d, *J* = 6.3 Hz, OH), 2.74–2.81 (2H, m, 2×H-4), 3.10 (1H, dt, *J* = 2.9, 7.8, and 7.8 Hz, H-3), 3.54 (1H, dd, *J*_{gem} = 10.0 Hz, *J*_{1,2} = 6.0 Hz, H-1), 3.59 (1H, dd, *J*_{gem} = 10.0 Hz, *J*_{1',2} = 5.0 Hz, H-1'), 3.76 (1H, m, H-2), 3.81 (3H, s, OMe), 4.50 (2H, s, OCH₂Ar), 6.89 and 7.26 (each 2H, each d, *J* = 8.5 Hz, aromatic protons). Found: *m/z* 224.1061 (M⁺). Calcd for C₁₂H₁₆O₄: M, 224.1048.

(2S,3S)-1,2-Epoxy-4-(4-methoxybenzyloxy)-3-(2-tetrahydropyranyloxy)butane (5). To a stirred solution of **4** (21.4 g, 95.4 mmol) in dry CH₂Cl₂ (430 ml) were added at 25 °C DHP (16.6 ml, 182 mmol) and CSA (4.43 g, 19.1 mmol). After 0.5 h at 25 °C, triethylamine (2.66 ml, 19.1 mmol) was added and the mixture was concentrated. The residue was chromatographed on silica gel (800 g) with 4:1 hexane–ethyl acetate to afford **5** (22.3 g, 76%) as a colorless syrup: *R*_f = 0.60 (2:1 hexane–ethyl acetate); IR (CHCl₃) 3012, 2948, 1613, 1513, 1249, 1175, 1120, 1076, 1034, and 984 cm⁻¹; ¹H NMR (CDCl₃, 3:2 mixture) δ = 1.40–1.95 (6H, m), 2.58 (3/5H, dd, *J* = 5.0 and 3.0 Hz, H-1), 2.73–2.82 (7/5H, m, H-1 and H-1'), 3.11 (2/5H, ddd, *J* = 7.0, 5.0, and 3.0 Hz, H-2), 3.16 (3/5H, ddd, *J* = 5.0, 4.0, and 3.0 Hz, H-2), 3.43–3.74 (4H, m), 3.81 (3H, s, OMe), 3.84–4.01 (1H, m), 4.47 and 4.50 (each 2/5H, ABq, *J* = 12.0 Hz, OCH₂Ar), 4.49 and 4.53 (each 3/5H, ABq, *J* = 12.0 Hz, OCH₂Ar), 4.81 (2/5H, br t, *J* = 4.0 Hz, OCHO), 4.91 (3/5H, dd, *J* = 4.0 and 2.6 Hz, OCHO), 6.88 (2H, d, *J* = 9.0 Hz, aromatic protons), 7.23 and 7.25 (4/5H and 6/5H, each d, *J* = 9.0 Hz, aromatic protons). Found: *m/z* 308.1624 (M⁺). Calcd for C₁₇H₂₄O₅: M, 308.1623.

(2S,3S)-1-Azido-2-benzyloxy-4-(4-methoxybenzyloxy)-3-(2-tetrahydropyranyloxy)butane (6). A mixture of **5** (20.6 g, 66.8 mmol), NH₄Cl (7.86 g, 147 mmol), NaN₃ (21.7 g, 334 mmol), and 8:1 MeOH–water (372 ml) was heated at 75 °C for 2 h. The reaction mixture was concentrated and to the residue were added ethyl acetate and water. The aqueous layer was extracted with ethyl acetate and the combined organic layers were washed with saturated aqueous NaCl, dried and concentrated. The residue (23.5 g) was dissolved in dry DMF (340 ml) and to this was added benzyl bromide (11.9 ml, 100 mmol). To this was added at 0 °C NaH (2.57 g, 107 mmol) and the mixture was stirred at 25 °C for 1 h. MeOH (8 ml) was added at 0 °C and the mixture was stirred at 25 °C for 0.5 h. Saturated aqueous NH₄Cl was added and the mixture was extracted with ether. The extracts were washed with saturated aqueous NaCl, dried, and concentrated. The residue was chromatographed on silica gel (800 g) with 7:1 hexane–ethyl acetate to afford **6** (28.0 g, 95%) as a colorless syrup: *R*_f = 0.50 (4:1 hexane–

acetone); IR (CHCl₃) 3011, 2946, 2103, 1612, 1514, 1249, 1117, 1075, and 1033 cm⁻¹; ¹H NMR (CDCl₃, 3:2 mixture) δ =1.40–1.90 (6H, m), 3.23–4.07 (8H, m), 3.80 and 3.81 (total 3H, each s, OMe), 4.39–4.52 (2H, m), 4.61–4.75 (3H, m), 6.86 (2H, d, J =8.0 Hz, aromatic protons), and 7.19–7.36 (7H, m, aromatic protons). Found: m/z 441.2272 (M⁺). Calcd for C₂₄H₃₁N₃O₅: M, 441.2264.

(2S,3S)-2-Benzoyloxy-1-(*t*-butoxycarbonylamino)-4-(4-methoxybenzyloxy)-3-(2-tetrahydropyranyloxy)butane (7). To a stirred solution of **6** (11.7 g, 26.5 mmol) in dry THF (117 ml) was added at 25 °C triphenylphosphine (7.65 g, 29.2 mmol). After 2 h at 50 °C, water (2.4 ml) was added and the mixture was stirred at 50 °C for 2 h. The reaction mixture was concentrated and the residue was dissolved in CH₂Cl₂ (234 ml) and water (89 ml). To this were added at 0 °C NaHCO₃ (8.91 g, 106 mmol) and (Boc)₂O (12.2 ml, 53.1 mmol). After 1 h at 25 °C, the reaction mixture was concentrated and the residue was dissolved in ethyl acetate. This was washed with water, saturated aqueous NaCl, dried, and concentrated. The residue was chromatographed on silica gel (500 g) with 4:1 and then 3:1 hexane–ethyl acetate to afford **7** (12.9 g, 94%) as a colorless syrup: R_f =0.40 (4:1 hexane–ethyl acetate); IR (CHCl₃) 3449, 3011, 2945, 1706, 1512, 1367, 1249, 1173, 1074, and 1031 cm⁻¹; ¹H NMR (CDCl₃, 3:2 mixture) δ =1.42 and 1.43 (total 9H, each, s, *t*-Bu), 1.40–1.95 (6H, m), 3.10–4.10 (8H, m), 3.79 and 3.80 (total 3H, each s, OMe), 4.38, 4.40 and 4.45, 4.48 (total 2H, each ABq, J =11.0 Hz, OCH₂Ar), 4.50–4.75 (3H, m, OCH₂Ar and OCHO), 4.84 and 5.51 (total 1H, each br, NH), 6.86 and 6.87 (total 2H, each d, J =8.6 Hz, aromatic protons), and 7.18–7.37 (7H, m, aromatic protons). Found: m/z 515.2875 (M⁺). Calcd for C₂₉H₄₁NO₇: M, 515.2883.

(2S,3S)-3-Benzoyloxy-4-(*t*-butoxycarbonylamino)-2-(2-tetrahydropyranyloxy)-1-butanol (8). To a stirred solution of **7** (12.6 g, 24.4 mmol) in 18:1 CH₂Cl₂–water (240 ml) was added at 0 °C DDQ (6.10 g, 26.9 mmol). After 0.5 h at 25 °C, the reaction mixture was passed through Florisil (480 g) and eluted with CH₂Cl₂. The eluate was concentrated and the residue was chromatographed on silica gel (500 g) with 3:2 hexane–ethyl acetate to afford **8** (9.18 g, 95%) as a colorless syrup: R_f =0.20 (2:1 hexane–ethyl acetate); IR (CHCl₃) 3452, 1707, 1506, 1455, 1393, 1368, 1274, 1248, 1163, 1135, 1074, 1028, and 981 cm⁻¹; ¹H NMR (CDCl₃, 3:2 mixture) δ =1.43 and 1.44 (total 9H, each s, *t*-Bu), 1.45–1.90 (6H, m), 3.14 (1H, dt, J =13.8 and 5.8 Hz), 3.30–4.05 (8H, m), 4.50–4.75 (3H, m), 4.82 and 5.35 (total 1H, each br, NH), and 7.28–7.38 (5H, m, Ph). Found: C, 63.26; H, 8.88; N, 3.43%. Calcd for C₂₁H₃₃NO₆: C, 63.78; H, 8.41; N, 3.54%.

(3S,4S)-4-Benzoyloxy-5-(*t*-butoxycarbonylamino)-3-(2-tetrahydropyranyloxy)-1-pentene (9a). A solution of DMSO (6.90 ml, 97.2 mmol) in dry CH₂Cl₂ (17 ml) was added at –78 °C to a stirred solution of oxalyl dichloride (4.24 ml, 48.6 mmol) in dry CH₂Cl₂ (117 ml). After 20 min at –78 °C, a solution of **8** (16.0 g, 40.5 mmol) in dry CH₂Cl₂ (96 ml) was added dropwise and the resulting suspension was stirred at –78 °C for 0.5 h. After addition of triethylamine (22.6 ml, 162 mmol), the mixture was gradually warmed to 0 °C during 0.5 h. The reaction mixture was quenched with water and extracted with 4:1 benzene–ether. The extracts were washed with saturated aqueous

NaCl, dried and concentrated. The residue (15.9 g) was dissolved in dry benzene (470 ml) and to this was added at 0 °C Ph₃P=CH₂ (33.5 g, 121 mmol). After 15 min at 25 °C, saturated aqueous NH₄Cl was added and the mixture was extracted with ethyl acetate. The extracts were washed with saturated aqueous NaCl, dried, and concentrated. The residue was chromatographed on silica gel (750 g) with 6:1 hexane–ethyl acetate to afford **9a** (10.8 g, 68%) as a colorless syrup: R_f =0.70 (3:1 hexane–ethyl acetate); IR (CHCl₃) 3453, 1708, 1506, 1455, 1393, 1368, 1272, 1248, 1168, 1118, 1076, 1023, and 971 cm⁻¹; ¹H NMR (CDCl₃, 2:1 mixture) δ =1.43 (9H, s, *t*-Bu), 1.45–1.95 (6H, m), 3.11 and 3.24 (total 1H, dt, J =13.4 and 6.2 Hz), 3.30–3.65 (3H, m), 3.82–3.95 (1H, m), 4.24 and 4.29 (total 1H, dd, J =6.2 and 6.2 Hz), 4.61, 4.72 and 4.65, 4.75 (total 2H, each ABq, J =11.6 Hz, OCH₂Ph), 4.60–5.10 (2H, m), 5.20–5.43 (2H, m, 2×H-1), 5.79 and 5.94 (total 1H, each ddd, J =17.0, 10.0, and 6.4 Hz, H-2), and 7.28–7.38 (5H, m, Ph). Found: C, 67.13; H, 9.02; N, 3.52%. Calcd for C₂₂H₃₃NO₅: C, 67.49; H, 8.50; N, 3.58%.

(3S,4S,5R)-3-Benzoyloxy-1-(*t*-butoxycarbonyl)-5-hydroxymethyl-4-(2-tetrahydropyranyloxy)pyrrolidine (10a) and Its 5S-Epimer 11a. To a stirred mixture of **9a** (3.85 g, 9.83 mmol) and NaHCO₃ (1.24 g, 14.8 mmol) in dry CH₂Cl₂ (77 ml) was added at 0 °C MCPBA (5.09 g, 29.5 mmol). After 2 d at 25 °C, a 7:1 mixture of saturated aqueous Na₂S₂O₃ and NaHCO₃ was added and the new mixture was extracted with CH₂Cl₂. The extracts were washed with saturated aqueous NaCl, dried, and concentrated. The residue was dissolved in dry CH₂Cl₂ (80 ml) and to this was added at –78 °C BF₃·OEt₂ (1.45 ml, 11.8 mmol). After 10 min at –78 °C, saturated aqueous NH₄Cl was added and the mixture was extracted with CH₂Cl₂. The extracts were washed with saturated aqueous NaCl, dried, and concentrated. The residue was chromatographed on silica gel (400 g) with 5:1 chloroform–ethyl acetate to afford **10a** (1.80 g, 45%) and **11a** (601 mg, 15%) as colorless syrups.

10a: R_f =0.39 (5:1 chloroform–ethyl acetate); IR (CHCl₃) 3445, 1688, 1455, 1398, 1368, 1255, 1169, 1130, 1076, 1035, and 979 cm⁻¹; ¹H NMR (DMSO-*d*₆) δ =1.42 (9H, s, *t*-Bu), 1.35–1.85 (6H, m), 3.27 (1H, dd, J =11.6 and 4.2 Hz), 3.47 (1H, dd, J =11.6 and 5.8 Hz), 3.41–3.52 (1H, m), 3.60–3.75 (2H, m), 3.75–3.92 (2H, m), 4.09 (1H, br t, J =5.8 Hz), 4.13–4.22 (1H, m), 4.23 (1H, dd, J =6.8 and 5.2 Hz), 4.55 (2H, s, OCH₂Ph), 4.72 (1H, br), and 7.25–7.40 (5H, m, Ph). Found: C, 64.64; H, 8.76; N, 3.21%. Calcd for C₂₂H₃₃NO₆: C, 64.84; H, 8.16; N, 3.44%.

11a: R_f =0.25 (5:1 chloroform–ethyl acetate); ¹H NMR (DMSO-*d*₆) δ =1.30–1.80 (6H, m), 1.42 (9H, s, *t*-Bu), 3.20–3.90 (7H, m), 3.96–4.65 (5H, m), 4.70–4.78 (1H, br), and 7.25–7.40 (5H, m, Ph).

1,4-Dideoxy-1,4-imino-D-xylitol Hydrochloride (12). A mixture of **10a** (48.3 mg, 0.119 mmol), Pd(OH)₂ (10 mg), and MeOH (1 ml) was stirred under an atmosphere of hydrogen (1 atm) at 25 °C for 0.5 h. The insoluble materials were filtered and washed with MeOH. The combined filtrate and washings were concentrated and the residue was dissolved in 1:1 THF–2 M aqueous HCl (1 ml). After 0.5 h at 50 °C, the mixture was concentrated to afford **12** (19.1 mg, 95%) as a colorless syrup. The ¹³C NMR spectrum was identical with the reported one:^{10) ¹³C NMR}

(67 MHz, D₂O, dioxane=69.3 ppm) δ =77.1, 65.7, 60.0, and 53.3 [lit.¹⁰ 77.1, 65.7, 60.0, and 53.3].

(3*S*,4*S*,5*S*)-3-Benzoyloxy-1-(*t*-butoxycarbonyl)-5-formyl-4-(2-tetrahydropyranyloxy)pyrrolidine (13). A solution of DMSO (0.352 ml, 4.96 mmol) in dry CH₂Cl₂ (0.88 ml) was added at -78 °C to a stirred solution of oxalyl dichloride (0.216 ml, 2.48 mmol) in dry CH₂Cl₂ (5.8 ml). After 20 min at -78 °C, a solution of **10a** (504 mg, 1.24 mmol) in dry CH₂Cl₂ (3 ml) was added dropwise and the resulting suspension was stirred at -78 °C for 0.5 h. After addition of triethylamine (1.04 ml, 7.44 mmol), the mixture was gradually warmed to 0 °C during 0.5 h. The reaction mixture was quenched with water and extracted with 4:1 benzene-ether. The extracts were washed with saturated aqueous NaCl, dried, and concentrated. The residue was chromatographed on silica gel (30 g) with 5:1 and then 4:1 hexane-ethyl acetate to afford **13** (412 mg, 82%) as a colorless syrup: R_f =0.57 (2:1 hexane-ethyl acetate); ¹H NMR (DMSO-*d*₆, 2:1 mixture) δ =1.39 (9H, s, *t*-Bu), 1.30–1.65 (6H, m), 3.40–3.70 (4H, m), 4.07–4.16 (1H, m), 4.25 (1H, dd, J =6.0 and 2.0 Hz), 4.50–4.75 (4H, m), 7.27–7.39 (5H, m, Ph), 9.44 and 9.48 (total 1H, each d, J =2.0 Hz, CHO).

(3*S*,4*S*,5*R*)-3-Benzoyloxy-1-(*t*-butoxycarbonyl)-5-[(1*S*)-1-hydroxyl-1-(4-methoxyphenyl)methyl]-4-(2-tetrahydropyranyloxy)pyrrolidine (14). To a stirred solution of 4-bromoanisole (0.507 ml, 4.05 mmol) in dry THF (8.1 ml) was added at -78 °C 1.61 M *n*-BuLi in hexane (2.38 ml, 3.84 mmol). After 0.5 h at -78 °C, a solution of **13** (410 mg, 1.01 mmol) in dry THF (2.1 ml) was added and the mixture was stirred at -78 °C for 0.5 h. Saturated aqueous NH₄Cl was added and the mixture was extracted with ethyl acetate. The extracts were washed with saturated aqueous NaCl, dried, and concentrated. The residue was chromatographed on silica gel (50 g) with 4:1 and then 3:1 hexane-ethyl acetate to afford **14** (420 mg, 80%) as a colorless syrup: R_f =0.20 (3:1 hexane-ethyl acetate); IR (CHCl₃) 3499, 3011, 2950, 1687, 1513, 1397, 1368, 1248, 1175, 1079, and 1035 cm⁻¹; ¹H NMR (DMSO-*d*₆, 2:1 mixture) δ =1.00–1.85 (15H, m), 3.20 (1H, dd, J =11.0 and 5.0 Hz), 3.63 (1H, dd, J =11.0 and 7.2 Hz), 3.73 and 3.74 (total 3H, each s, OMe), 4.05–4.20 (2H, m), 4.55 (2H, s, OCH₂Ph), 4.76 and 5.00 (total 1H, each m), 4.83 (1H, d, J =5.6 Hz), 6.83 and 7.16 (each 2H, ABq, J =8.4 Hz, aromatic protons), and 7.26–7.40 (5H, m, Ph). Found: C, 67.62; H, 7.63; N, 3.09%. Calcd for C₂₉H₃₉NO₇: C, 67.82; H, 7.65; N, 2.73%.

Preparation of 15. A mixture of **14** (15.3 mg, 0.0279 mmol), CSA (1.3 mg, 0.0056 mmol), and MeOH (0.31 ml) was stirred at 25 °C for 0.5 h. Triethylamine (0.002 ml, 0.01 mmol) was added and the mixture was concentrated. The residue was chromatographed on silica gel (2 g) with 2:1 hexane-ethyl acetate to afford a syrup (11.6 mg, 90%). This was dissolved in dry CH₂Cl₂ (0.2 ml) and to this were added 2,2-dimethoxypropane (0.006 ml, 0.05 mmol) and pyridinium *p*-toluenesulfonate (PPTS) (0.3 mg, 0.001 mmol). After 1 d at 25 °C, triethylamine (0.003 ml, 0.02 mmol) was added and the mixture was concentrated. The residue was chromatographed on silica gel (2 g) with 6:1 hexane-ethyl acetate to afford **15** (11.3 mg, 90%) as a colorless syrup: R_f =0.90 (2:1 hexane-ethyl acetate); ¹H NMR (DMSO-*d*₆, 60 °C) δ =1.32 and 1.49 (each 3H, each s, CMe₂), 3.40 (1H, dd, J_{gem} =11.6 Hz, J =2.4 Hz, one of CH₂N),

3.71 (3H, s, OMe), 3.84–3.93 (2H, m, one of CH₂N and CHOBn), 3.91 (1H, dd, J =3.6 and 5.0 Hz, CHN), 4.47 (2H, s, OCH₂Ph), 4.60 (1H, d, J =5.0 and 0 Hz, CHOCMe₂), 5.12 (1H, d, J =3.6 Hz, CHOCMe₂), 6.80 (2H, d, J =9.0 Hz, aromatic protons), and 7.15–7.35 (7H, m, aromatic protons).

(3*S*,4*S*,5*S*)-3-Benzoyloxy-1-(*t*-butoxycarbonyl)-5-(4-methoxybenzoyl)-4-(2-tetrahydropyranyloxy)pyrrolidine (16). A solution of DMSO (0.232 ml, 3.27 mmol) in dry CH₂Cl₂ (0.582 ml) was added at -78 °C to a stirred solution of oxalyl dichloride (0.143 ml, 1.64 mmol) in dry CH₂Cl₂ (3.86 ml). After 20 min at -78 °C, a solution of **14** (420 mg, 0.818 mmol) in dry CH₂Cl₂ (2.5 ml) was added dropwise and the resulting suspension was stirred at -78 °C for 0.5 h. After addition of triethylamine (0.684 ml, 4.91 mmol), the mixture was gradually warmed to 0 °C during 0.5 h. The reaction mixture was quenched with water and extracted with 4:1 benzene-ether. The extracts were washed with saturated aqueous NaCl, dried, and concentrated. The residue was chromatographed on silica gel (35 g) with 3:1 hexane-ethyl acetate to afford **16** (366 mg, 88%) as a colorless syrup: R_f =0.40 (2:1 hexane-ethyl acetate); IR (CHCl₃) 1697, 1602, 1402, 1368, 1257, 1232, 1173, 1126, 1034, and 974 cm⁻¹; ¹H NMR (DMSO-*d*₆, 2:1 mixture) δ =1.00–1.55 (15H, m), 3.10–3.20 (2H, m), 3.50 (1H, dd, J =11.0 and 3.4 Hz), 3.64 and 3.66 (total 1H, each dd, J =11.0 and 5.0 Hz), 3.84 and 3.86 (total 3H, each s, OMe), 4.15 (1H, m), 4.50–4.77 (4H, m), 5.36–5.50 (1H, m, H-5), 7.02 and 7.95 (each 2H, ABq, J =8.6 Hz, aromatic protons), and 7.26–7.40 (5H, m, Ph). Found: C, 67.61; H, 7.44; N, 2.95%. Calcd for C₂₉H₃₇NO₇: C, 68.08; H, 7.29; N, 2.74%.

(3*S*,4*S*,5*S*)-3-Benzoyloxy-1-(benzyloxycarbonyl)-5-(4-methoxybenzoyl)-4-(2-tetrahydropyranyloxy)pyrrolidine (17). To a stirred solution of **16** (366 mg, 0.715 mmol) in CH₂Cl₂ (3.66 ml) was added at 25 °C 1:1 trifluoroacetic acid (TFA)-CH₂Cl₂ (3.66 ml). After 15 min at 25 °C, the mixture was concentrated and the residue was dissolved in 2:1 THF-water (10.5 ml). To this were added at 0 °C K₂CO₃ (148 mg, 1.07 mmol) and benzyl chloroformate (0.122 ml, 0.858 mmol). After 0.5 h at 25 °C, saturated aqueous NH₄Cl was added and the mixture was extracted with ethyl acetate. The extracts were washed with saturated aqueous NaCl, dried, and concentrated. The residue (330 mg) was dissolved in dry CH₂Cl₂ (5 ml) and to this were added at 25 °C DHP (0.130 ml, 1.43 mmol) and CSA (33.2 mg, 0.143 mmol). After 0.5 h at 25 °C, triethylamine (0.020 ml, 0.14 mmol) was added and the mixture was concentrated. The residue was chromatographed on silica gel (20 g) with 2:1 hexane-ethyl acetate to afford **17** (332 mg, 85%) as a colorless syrup: R_f =0.80 (1:1 hexane-ethyl acetate); IR (CHCl₃) 1703, 1602, 1422, 1354, 1261, 1229, 1172, 1126, 1033, and 980 cm⁻¹; ¹H NMR (DMSO-*d*₆, 3:2 mixture) δ =1.10–1.60 (6H, m), 3.00–4.30 (6H, m), 3.85 and 3.87 (total 3H, each s, OMe), 4.55–4.80 (4H, m), 4.90–5.15 (2H, m, COOCH₂Ph), 5.50–5.65 (1H, m, H-5), 7.00 and 7.95 (each 2H, each br, aromatic protons), and 7.05–7.40 (10H, m, 2×Ph). Found: C, 69.86; H, 6.88; N, 2.20%. Calcd for C₃₂H₃₅NO₇: C, 70.44; H, 6.47; N, 2.59%.

(3*S*,4*S*,5*R*)-3-Benzoyloxy-1-(benzyloxycarbonyl)-5-[(1*R*)-1-hydroxy-1-(4-methoxyphenyl)methyl]-4-(2-tetrahydropyranyloxy)pyrrolidine (18). To a stirred solution of **17** (225 mg, 0.412 mmol) in dry toluene (4.5 ml) was added at -78 °C 1.02 M DIBAL in toluene (0.810 ml,

0.826 mmol). After 0.5 h at -78°C , MeOH (0.06 ml) and water (0.1 ml) were added and the mixture was warmed to 25°C . To this were added potassium sodium tartrate tetrahydrate (1.16 g, 4.11 mmol) in water (5 ml) and the new mixture was stirred at 25°C for 5 h. The mixture was extracted with ethyl acetate and the extracts were washed with saturated aqueous NaCl, dried, and concentrated. The residue was chromatographed on silica gel (20 g) with 3:1 and then 2:1 hexane–ethyl acetate to afford **18** (206 mg, 91%) as a colorless syrup: $R_f=0.50$ (2:1 hexane–ethyl acetate); IR (CHCl_3) 1675, 1510, 1420, 1240, 1210, 1110, 1035, and 965 cm^{-1} ; $^1\text{H NMR}$ ($\text{DMSO}-d_6$, 2:1 mixture) $\delta=1.20\text{--}1.75$ (6H, m), $3.20\text{--}4.00$ (6H, m), 3.73 and 3.74 (total 3H, each s, OMe), $4.15\text{--}4.60$ (4H, m), $4.70\text{--}5.15$ (4H, m), 6.78 and 6.82 (total 2H, each d, $J=8.4\text{ Hz}$, aromatic protons), and $7.20\text{--}7.40$ (12H, m, aromatic protons). Found: C, 70.04; H, 6.91; N, 2.86%. Calcd for $\text{C}_{32}\text{H}_{37}\text{NO}_7$: C, 70.18; H, 6.81; N, 2.56%.

Preparation of 19. A mixture of **18** (17.6 mg, 0.0343 mmol), CSA (1.6 mg, 0.0069 mmol), and MeOH (0.35 ml) was stirred at 25°C for 0.5 h. Triethylamine (0.003 ml, 0.02 mmol) was added and the mixture was concentrated. The residue was chromatographed on silica gel (2 g) with 2:1 hexane–ethyl acetate to afford a syrup (13.2 mg, 90%). This was dissolved in dry CH_2Cl_2 (0.3 ml) and to this were added 2,2-dimethoxypropane (0.0076 ml, 0.062 mmol) and PPTS (0.4 mg, 0.002 mmol). After 1 d at 25°C , triethylamine (0.003 ml) was added and the mixture was concentrated. The residue was chromatographed on silica gel (2 g) with 6:1 hexane–ethyl acetate to afford **19** (13.0 mg, 90%) as a colorless syrup: $R_f=0.90$ (2:1 hexane–ethyl acetate); $^1\text{H NMR}$ ($\text{DMSO}-d_6$) $\delta=1.36$ and 1.39 (each 3H, each s, CMe_2), 3.48 (1H, dd, $J=12.5$ and 2.5 Hz , one of CH_2N), 3.73 (3H, s, OMe), 3.99 (1H, d, $J=2.5$ and 0 Hz , CHOBN), 4.03 (1H, d, $J=12.5$ and 0 Hz , one of CH_2N), 4.34 (1H, dd, $J=8.8$ and 5.5 Hz , CHN), 4.46 (1H, d, $J=8.8\text{ Hz}$, CHOCMe_2), 4.47 (1H, d, $J_{\text{gem}}=12.5\text{ Hz}$, one of OCH_2Ph), 4.48 (1H, d, $d=5.5$ and 0 Hz , CHOCMe_2), 4.53 (2H, s, OCH_2Ph), 4.86 (1H, d, $J_{\text{gem}}=12.5\text{ Hz}$, one of OCH_2Ph), 6.82 (2H, d, $J=9.0\text{ Hz}$, aromatic protons), and $7.20\text{--}7.40$ (12H, m, aromatic protons).

Phenyl 3-*O*-Benzyl-5-*O*-(*t*-butyldimethylsilyl)-1-thio- α -D-xylofuranoside (23**).** To a stirred solution of **20** (5.53 g, 19.7 mmol) in dry CH_2Cl_2 (110 ml) were added at 25°C triethylamine (5.49 ml, 39.4 mmol) and PvCl (4.37 ml, 35.5 mmol). After 15 h at 25°C , water was added and the mixture was extracted with ethyl acetate. The extracts were washed with saturated aqueous NaCl, dried, and concentrated. The residue (7.19 g) was dissolved in 50% aqueous acetic acid (90 ml) and the mixture was heated at 100°C for 5 h. The reaction mixture was concentrated and the residue was chromatographed on silica gel (300 g) with 3:2 and then 1:1 hexane–ethyl acetate to afford a free sugar (5.86 g, 92%) as a colorless syrup. To a stirred solution of this sample (4.60 g, 14.2 mmol) in dry THF (46 ml) were added at 25°C diphenyl disulfide (4.64 g, 21.3 mmol) and (*n*-Bu) $_3\text{P}$ (5.31 ml, 21.3 mmol). After 0.5 h at 25°C , the mixture was concentrated. The residue was chromatographed on silica gel (300 g) with 8:1 and then 4:1 hexane–ethyl acetate to afford thioglycoside **22** (5.53 g, 94%) as a colorless syrup. To a stirred solution of this sample (5.53 g, 13.3 mmol) in dry ether (55 ml) was added at 0°C 1.15 M MeLi in ether (115

ml, 133 mmol). After 15 min at 25°C , saturated aqueous NH_4Cl was added and the mixture was extracted with ethyl acetate. The extracts were washed with saturated aqueous NaCl, dried, and concentrated. The residue (4.42 g) was dissolved in dry DMF (88 ml) and to this was added at 0°C TBSCl (2.20 g, 14.6 mmol) and imidazole (1.09 g, 16.0 mmol). After 0.5 h at 25°C , water was added and the mixture was extracted with ether. The extracts were washed with saturated aqueous NaCl, dried, and concentrated. The residue was chromatographed on silica gel (600 g) with 40:1 chloroform–ethyl acetate to afford **23** (1.78 g, 30%) as colorless crystals and its β -anomer (3.52 g, 59%) as a colorless syrup.

23: $R_f=0.30$ (40:1 chloroform–ethyl acetate); mp $76\text{--}77^{\circ}\text{C}$ (not recrystallized); $[\alpha]_D^{25}+70.0^{\circ}$ ($c\ 1.02$); IR (CHCl_3) 2955, 2932, 1472, 1257, 1095, 1054, 1007, and 840 cm^{-1} ; $^1\text{H NMR}$ (CDCl_3 , $\text{CHCl}_3=7.26$) $\delta=0.06$ and 0.07 (each 3H, each s, SiMe_2), 0.90 (9H, s, *t*-Bu), 2.43 (1H, d, $J=4.2\text{ Hz}$, OH), 3.84 (1H, dd, $J_{\text{gem}}=10.0\text{ Hz}$, $J_{4,5}=5.8\text{ Hz}$, H-5), 3.90 (1H, dd, $J=10.0\text{ Hz}$, $J_{4,5'}=7.0\text{ Hz}$, H-5'), 4.06 (1H, dd, $J_{3,4}=4.2\text{ Hz}$, $J_{2,3}=2.0\text{ Hz}$, H-3), 4.37 (1H, ddd, $J_{3,4}=4.2\text{ Hz}$, $J_{4,5}=5.8\text{ Hz}$, $J_{4,5'}=7.0\text{ Hz}$, H-4), 4.43 (1H, ddd, $J_{1,2}=4.2\text{ Hz}$, $J_{2,3}=2.0\text{ Hz}$, $J_{2,\text{OH}}=4.2\text{ Hz}$, H-2), 4.64 and 4.66 (each 1H, ABq, $J_{\text{gem}}=11.8\text{ Hz}$, OCH_2Ph), 5.66 (1H, d, $J_{1,2}=4.2\text{ Hz}$, H-1), $7.24\text{--}7.36$ and $7.48\text{--}7.54$ (8H and 2H, each m, $2\times\text{Ph}$). Found: C, 64.19; H, 7.48; S, 6.90%. Calcd for $\text{C}_{24}\text{H}_{34}\text{O}_4\text{SSi}$: C, 64.53; H, 7.67; S, 7.18%.

β -Anomer of 23: $R_f=0.25$ (40:1 chloroform–ethyl acetate); $^1\text{H NMR}$ (CDCl_3 , $\text{CHCl}_3=7.26$) $\delta=0.07$ (6H, s, SiMe_2), 0.91 (9H, s, *t*-Bu), 2.03 (1H, d, $J_{2,\text{OH}}=4.0\text{ Hz}$, OH), 3.85 (1H, dd, $J_{\text{gem}}=10.4\text{ Hz}$, $J_{4,5}=5.4\text{ Hz}$, H-5), 3.94 (1H, dd, $J_{\text{gem}}=10.4\text{ Hz}$, $J_{4,5'}=5.4\text{ Hz}$, H-5'), 4.02 (1H, dd, $J_{2,3}=3.2\text{ Hz}$, $J_{3,4}=5.4\text{ Hz}$, H-3), 4.26 (1H, dt, $J_{3,4}=J_{4,5}=J_{4,5'}=5.4\text{ Hz}$, H-4), 4.36 (1H, ddd, $J_{1,2}=4.0\text{ Hz}$, $J_{2,3}=3.2\text{ Hz}$, $J_{2,\text{OH}}=4.0\text{ Hz}$, H-2), 4.64 and 4.69 (each 1H, ABq, $J_{\text{gem}}=12.0\text{ Hz}$, OCH_2Ph), 5.16 (1H, d, $J_{1,2}=4.0\text{ Hz}$, H-1), $7.20\text{--}7.40$ and $7.48\text{--}7.54$ (8H and 2H, each m, $2\times\text{Ph}$).

Phenyl 2,3-Di-*O*-benzyl-1-thio- α -D-xylofuranoside (24**).** To a solution of **23** (1.50 g, 3.36 mmol), benzyl bromide (0.599 ml, 5.04 mmol), and tetrabutylammonium iodide (62.1 mg, 0.168 mmol) in dry THF (18 ml) was added at 0°C NaH (129 mg, 5.38 mmol). After 2 h at 25°C , ethanol (0.7 ml) was added and the mixture was stirred at 25°C for 0.5 h. Saturated aqueous NH_4Cl was added and the mixture was extracted with ethyl acetate. The extracts were washed with saturated aqueous NaCl, dried, and concentrated. The residue was dissolved in dry THF (30 ml) and to this was added at 25°C 1.00 M (*n*-Bu) $_4\text{NF}$ in THF (3.70 ml, 3.70 mmol). After 0.5 h at 25°C , the reaction mixture was concentrated and the residue was chromatographed on silica gel (50 g) with 4:1 and then 3:1 hexane–ethyl acetate to afford **24** (1.35 g, 95%) as a colorless syrup: $R_f=0.25$ (3:1 hexane–ethyl acetate); IR (CHCl_3) 3552, 3014, 1455, 1358, 1107, 1056, and 1028 cm^{-1} ; $^1\text{H NMR}$ (CDCl_3) $\delta=2.28$ (1H, dd, $J=9.0$ and 4.0 Hz , OH), 3.81 (1H, ddd, $J_{\text{gem}}=12.0\text{ Hz}$, $J_{4,5}=4.0\text{ Hz}$, $J_{5,\text{OH}}=9.0\text{ Hz}$, H-5), 3.88 (1H, ddd, $J_{\text{gem}}=12.0\text{ Hz}$, $J_{4,5'}=4.0\text{ Hz}$, $J_{5',\text{OH}}=4.0\text{ Hz}$, H-5'), 4.29 (1H, dd, $J_{2,3}=4.0\text{ Hz}$, $J_{3,4}=6.0\text{ Hz}$, H-3), 4.33 (1H, dd, $J_{1,2}=5.8\text{ Hz}$, $J_{2,3}=4.0\text{ Hz}$, H-2), 4.45 (1H, ddd, $J_{3,4}=6.0\text{ Hz}$, $J_{4,5}=J_{4,5'}=4.0\text{ Hz}$, H-4), 4.48 , 4.54 , 4.63 , and 4.78 (each 1H, $2\times\text{ABq}$, $J=12.0\text{ Hz}$, $2\times\text{OCH}_2\text{Ph}$), 5.88 (1H, d, $J_{1,2}=5.8\text{ Hz}$, H-1), and $7.20\text{--}7.55$ (15H, m, $3\times\text{Ph}$). Found: C, 71.12;

H, 6.07; S, 7.66%. Calcd for $C_{25}H_{26}O_4S$: C, 71.07; H, 6.20; S, 7.59%

Triflation of the β -Anomer of 24. The β -anomer of **24** [R_f =0.28 (3:1 hexane-ethyl acetate)]; 1H NMR ($CDCl_3$) δ =2.20 (1H, br, OH), 3.80–3.93 (2H, br m, $2\times H-5$), 4.17 (1H, dd, $J_{2,3}$ =2.8 Hz, $J_{3,4}$ =5.8 Hz, H-3), 4.20 (1H, dd, $J_{1,2}$ =4.0 Hz, $J_{2,3}$ =2.8 Hz, H-2), 4.27 (1H, ddd, $J_{3,4}=J_{4,5}=J_{4,5'}=5.8$ Hz, H-4), 4.44, 4.58, 4.60, and 4.69 (each 1H, $2\times ABq$, J =12.0 Hz, $2\times OCH_2Ph$), 5.38 (1H, d, $J_{1,2}$ =4.0 Hz, H-1), and 7.22–7.53 (15H, m, $3\times Ph$) was obtained from the β -anomer of **23** as described above for the preparation of **24** from **23**. To a stirred solution of the β -anomer of **24** (10.7 mg, 0.0253 mmol) and triethylamine (0.0106 mL, 0.0759 mmol) in dry CH_2Cl_2 (0.13 mL) was added at $-78^\circ C$ Tf_2O (0.0040 mL, 0.024 mmol). After 5 min at $-78^\circ C$, saturated aqueous $NaHCO_3$ was added and the mixture was extracted with CH_2Cl_2 . The extracts were washed with saturated aqueous NaCl, dried, and concentrated. The residue was chromatographed on silica gel (2 g) with 4:1 hexane-ethyl acetate to afford **26** (7.5 mg, 70%, $\alpha:\beta$ =1:1.3) as a colorless syrup: R_f =0.56 (3:1 hexane-ethyl acetate);

1H NMR ($CDCl_3$) of α -isomer: δ =3.21 (2H, d, $J_{4,5}$ =7.2 Hz, $2\times H-5$), 3.89 (1H, dd, $J_{1,2}$ =4.0 Hz, H-2), 3.91 (1H, d, $J_{1,OH}$ =10.0 Hz, OH), 4.00 (1H, dd, $J_{2,3}$ =1.4 Hz, $J_{3,4}$ =4.0 Hz, H-3), 4.34 (1H, ddd, $J_{4,5}$ =7.2 Hz, $J_{3,4}$ =4.0 Hz, H-4), 4.45–4.61 (4H, m, $2\times OCH_2Ph$), 5.48 (1H, dd, $J_{1,2}$ =4.0 Hz, $J_{1,OH}$ =10.0 Hz, H-1), and 7.15–7.40 (15H, m, $3\times Ph$);

1H NMR ($CDCl_3$) of β -isomer: δ =3.29 (1H, dd, J_{gem} =13.8 Hz, $J_{4,5}$ =8.2 Hz, H-5), 3.34 (1H, dd, J_{gem} =13.8 Hz, $J_{4,5'}$ =6.2 Hz, H-5'), 3.38 (1H, d, $J_{1,OH}$ =11.2 Hz, OH), 3.94 (1H, br s, H-2), 4.03 (1H, br d, $J_{2,3}$ =0 Hz, $J_{3,4}$ =4.0 Hz, H-3), 4.40 (1H, ddd, $J_{4,5}$ =8.2 Hz, $J_{3,4}$ =4.0 Hz, $J_{4,5'}$ =6.2 Hz, H-4), 4.45–4.61 (4H, m, $2\times OCH_2Ph$), 5.25 (1H, d, $J_{1,2}$ =0 Hz, $J_{1,OH}$ =11.2 Hz, H-1), and 7.15–7.40 (15H, m, $3\times Ph$); M^+ ($\alpha\beta$ mixture), 422.

Intermolecular Etherification. To a stirred solution of **24** (383 mg, 0.906 mmol) and triethylamine (0.379 mL, 2.72 mmol) in dry CH_2Cl_2 (4.53 mL) was added at $-78^\circ C$ Tf_2O (0.152 mL, 0.906 mmol). After 5 min at $-78^\circ C$, saturated aqueous $NaHCO_3$ was added and the mixture was extracted with CH_2Cl_2 . The extracts were concentrated and the residue was dissolved in hexane. The organic layer was washed with water and saturated aqueous NaCl, dried, and concentrated. The residue was dried under vacuum for 0.5 h. To a stirred suspension of **18** (248 mg, 0.453 mmol) and MS 4AP (906 mg) in dry DMF (4.53 mL) was added at $0^\circ C$ 1.63 M n -BuLi in hexane (0.417 mL, 0.680 mmol). After 5 min at $0^\circ C$, a solution of the above triflate in dry DMF (2.27 mL) was added and the mixture was stirred at $25^\circ C$ for 0.5 h. Saturated aqueous NH_4Cl was added and the mixture was extracted with ether. The extracts were washed with saturated aqueous NaCl, dried, and concentrated. The residue was chromatographed on silica gel (40 g) with 12:1 toluene-ethyl acetate to afford **27** (323 mg, 75%) as a colorless syrup: R_f =0.60 (8:1 chloroform-ethyl acetate); IR ($CHCl_3$) 1690, 1510, 1450, 1350, 1240, 1210, 1085, 1050, and 1035 cm^{-1} ; 1H NMR ($DMSO-d_6$, 2:1 mixture) δ =1.20–1.70 (6H, m), 3.72 (3H, s, OMe), 4.08 (1H, dd, J =4.0 and 2.0 Hz), 4.98 and 5.05 (each 1H, ABq, J =12.0 Hz, $COOCH_2Ph$), 5.84 and 5.85 (total 1H, each d, J =5.6 Hz, $CHSPh$), 6.74 (2H, d, J =8.8 Hz, aromatic protons), and 7.10–7.50 (27H, m, aromatic protons). Found: C, 71.45; H, 6.59; N, 1.29%. Calcd

for $C_{57}H_{61}NO_{10}S$: C, 71.90; H, 6.46; N, 1.47%.

Preparation of 28. To a stirred solution of **18** (20.0 mg, 0.0365 mmol) in dry DMF (0.2 mL) was added at $0^\circ C$ 1 M LDA in THF (0.0365 mL, 0.0365 mmol). After 0.5 h at $25^\circ C$, saturated aqueous NH_4Cl was added and the mixture was extracted with ethyl acetate. The extracts were washed with saturated aqueous NaCl, dried, and concentrated. The residue was chromatographed on silica gel (2 g) with 10:1 chloroform-ethyl acetate to afford **28** (14.2 mg, 89%) as a colorless syrup: R_f =0.29 (10:1 chloroform-ethyl acetate); 1H NMR ($DMSO-d_6$) δ =0.80–1.60 (6H, m), 3.15 and 3.18 (total 1H, each d, J =0 and 12.6 Hz), 3.18–3.30 (1H, m), 3.51 (0.5H, dd, J =2.8 and 5.2 Hz) 3.53–3.65 (1H, m), 3.75 and 3.77 (total 3H, each s, OMe), 3.78 (0.5H, dd, J =5.0 and 12.6 Hz), 3.87 and 4.02 (total 1H, each d, J =0 and 2.0 Hz), 4.23 and 4.31 (total 1H, each d, J =0, 0, and 5.0 Hz), 4.47 and 4.51 (total 1H, each dd, J =2.0 and 7.8 Hz), 4.55 and 4.56 (total 2H each s, OCH_2Ph), 5.92 (1H, d, J =7.8 Hz), 6.94 and 6.95 (total 2H, each d, J =9.0 Hz), and 7.26–7.39 (7H, m); M^+ , 439.

Acid-Hydrolysis of 27. To a stirred solution of **27** (132 mg, 0.139 mmol) in dioxane (0.66 mL) and MeOH (1.98 mL) was added at $0^\circ C$ CSA (6.5 mg, 0.028 mmol). After 2 h at $25^\circ C$, triethylamine (0.005 mL) was added and the mixture was concentrated. The residue was chromatographed on silica gel (12 g) with 4:1 hexane-ethyl acetate to afford **29** (109 mg, 90%) as a colorless syrup: R_f =0.38 (3:1 hexane-ethyl acetate); $[\alpha]_D^{25}+50.6^\circ$ (c 1.25); IR ($CHCl_3$) 3444, 3012, 1694, 1513, 1455, 1414, 1354, 1249, 1110, 1060, and 1030 cm^{-1} ; 1H NMR ($DMSO-d_6$) δ =3.32–3.37 (2H, m), 3.41 (1H, dd, J =10.6 and 6.0 Hz), 3.63 (1H, dd, J =10.6 and 5.0 Hz), 3.73 (3H, s, OMe), 3.89 (1H, m), 4.07–4.23 (3H, m), 4.35–5.05 (13H, m), 5.83 (1H, d, $J_{1,13}$ =5.0 Hz, H-1), 6.76 (2H, d, J =8.0 Hz, H-17 and H-19), and 7.17–7.51 (27H, m, aromatic protons). Found: C, 71.76; H, 6.33; N, 1.77; S, 3.93%. Calcd for $C_{52}H_{53}NO_9S$: C, 71.95; H, 6.15; N, 1.61; S, 3.69%.

Intramolecular Glycosylation. To a stirred suspension of **29** (109 mg, 0.126 mmol) and MS 4AP (1.26 g) in dry toluene (12.6 mL) was added at $25^\circ C$ NBS (33.5 mg, 0.188 mmol). After 1 h at $25^\circ C$, the mixture was heated at $90^\circ C$ for 24 h. The reaction mixture was cooled to ambient temperature and the insoluble materials were filtered out. The filtrate was washed with toluene and the combined filtrate and washings were concentrated. The residue was chromatographed on silica gel (15 g) with 35:2 and then 10:1 chloroform-ethyl acetate to afford **30** (61.1 mg, 64%) as a colorless foam: R_f =0.42 (10:1 chloroform-ethyl acetate); $[\alpha]_D^{25}+30.4^\circ$ (c 1.02); IR ($CHCl_3$) 1696, 1513, 1454, 1419, 1356, 1248, 1096, 1058, 1030, and 979 cm^{-1} ; 1H NMR ($DMSO-d_6$) δ =3.16 (1H, dd, J =11.8 and 6.0 Hz), 3.31 (1H, dd, J =11.8 and 7.8 Hz), 3.71 (3H, s, OMe), 3.87 (1H, dd, J =13.0 and 11.4 Hz), 3.99 (1H, dd, J =13.0 and 6.0 Hz), 4.15–4.85 (15H, m), 5.31 (1H, s, H-1), 6.72 (2H, d, J =9.0 Hz, H-17 and H-19), 7.13 (2H, d, J =9.0 Hz, H-16 and H-20), and 7.20–7.40 (20H, m, $4\times Ph$). Found: C, 72.71; H, 6.24; N, 2.17%. Calcd for $C_{46}H_{47}NO_9$: C, 72.90; H, 6.25; N, 1.85%.

AB3217-A (1a). A mixture of **30** (53 mg, 0.0699 mmol), $Pd(OH)_2$ (159 mg), dioxane (3.71 mL), and 0.01 M aqueous HCl (7.42 mL) was stirred under an atmosphere of hydrogen (1 atm) at $25^\circ C$ for 12 h. The insoluble materi-

als were filtered and thoroughly washed with MeOH. The combined filtrate and washings were concentrated and the residue was dissolved in 0.1 M aqueous NaOH (0.74 ml) and this was stirred at 25 °C for 0.5 h. This was passed through CM-Sephadex (C-25) with 50% aqueous MeOH and then triethylamine-CO₂ buffer in 50% aqueous MeOH. The latter eluate was concentrated to afford **1a** (19.8 mg, 80%) as colorless crystals. The analytical sample of **1a** was obtained by recrystallization from MeOH: R_f = 0.32 (1:1 ethyl acetate-MeOH); mp 238–239 °C [mp of natural AB3217-A: 241 °C, lit.¹⁾ 241 °C]; mixed mp 238–239 °C; $[\alpha]_D^{26}$ –61.3° (c 0.46, H₂O) [$[\alpha]_D^{26}$ of natural AB3217-A: –61.0° (c 0.41, H₂O), lit.¹⁾ $[\alpha]_D^{24}$ –52.5 °C (c 1.0, H₂O)]; UV (H₂O) λ_{max} nm (ϵ) 226 (9200) and 272 (1160) [lit.¹⁾ λ_{max} nm (ϵ) 226 (11400), 272 (1100), and 278 (900)]; IR (KBr) 3511, 3416, 3339, 2940, 2920, 2890, 1610, 1510, 1455, 1390, 1370, 1300, 1250, 1240, 1175, 1120, 1100, 1080, 1065, 1020, 1010, 1000, and 890 cm^{–1}. IR (KBr) of natural AB3217-A: 3510, 3416, 3339, 2945, 2920, 2890, 1610, 1510, 1455, 1390, 1370, 1305, 1250, 1240, 1170, 1120, 1105, 1080, 1065, 1025, 1015, 1000, and 890 cm^{–1}; ¹H NMR (DMSO-*d*₆, 25 °C) δ = 1.50 (1H, br, NH), 2.42 (1H, dd, J = 11.0 and 4.0 Hz, H-5), 2.85 (1H, dd, J = 11.0 and 5.7 Hz, H-5'), 3.41 (1H, m, H-7), 3.65–3.75 (2H, m, 2×H-10), 3.73 (3H, s, OMe), 3.94 (1H, m, H-4), 4.00 (2H, m, H-12 and H-13), 4.09 (1H, dd, J = 6.4 and 3.6 Hz, H-3), 4.18 (1H, m, H-11), 4.40 (1H, d, J = 9.6 Hz, H-8), 4.87 (1H, br, OH), 4.90 (1H, s, H-1), 5.40 (2H, br, 2×OH), 6.85 (2H, d, J = 8.6 Hz, H-17 and H-19), and 7.21 (2H, d, J = 8.6 Hz, H-16 and H-20).

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References

- 1) a) K. Kanbe, A. Takahashi, K. Sato, T. Tamamura, Y. Sato, H. Naganawa, and T. Takeuchi, *Jpn. Kokai Tokkyo Koho JP 232889*; b) K. Kanbe, Y. Mimura, T. Tamamura, S. Yatagai, Y. Sato, A. Takahashi, K. Sato, H. Naganawa, H. Nakamura, T. Takeuchi, and I. Iitaka, *J. Antibio.*, **45**, 458 (1992).
- 2) K. Kanbe, A. Takahashi, T. Tamamura, K. Sato, H. Naganawa, and T. Takeuchi, *J. Antibio.*, **45**, 568 (1992).
- 3) M. Nakata, T. Tamai, T. Kamio, M. Kinoshita, and K. Tatsuta, *Tetrahedron Lett.*, **35**, 3099 (1994).
- 4) K. Toshima and K. Tatsuta, *Chem. Rev.*, **93**, 1503 (1993).
- 5) a) For cyclo-glycosylations to yield cyclo-(1→4)-glycohexaoses, see: H. Kuyama, T. Nukada, Y. Nakahara, and T. Ogawa, *Tetrahedron Lett.*, **34**, 2171 (1993), and references cited therein; b) For a spacer-mediated intramolecular glycosylation to yield oligosaccharides, see: T. Takahashi, K. Imamura, T. Harada, and H. Yamada, "34th Symposium on the Chemistry of Natural Product," Tokyo, October 1992, Symposium papers p. 15; c) For a nonselective intramolecular glycosylation via Mitsunobu reaction to yield an (–)-ovatolide model compound, see: A. Delgado and J. Clardy, *J. Org. Chem.*, **58**, 2862 (1993).
- 6) E. Hungerbühler and D. Seebach, *Helv. Chim. Acta*, **64**, 687 (1981).
- 7) M. Caron and K. B. Sharpless, *J. Org. Chem.*, **50**, 1557 (1985).
- 8) W. R. Ewing, B. D. Harris, K. L. Bhat, and M. M. Joullie, *Tetrahedron*, **42**, 2421 (1986).
- 9) H. Takahata, Y. Banba, M. Tajima, and T. Momose, *J. Org. Chem.*, **56**, 240 (1991).
- 10) D. W. C. Jones, R. J. Nash, E. A. Bell, and J. M. Williams, *Tetrahedron Lett.*, **26**, 3125 (1985).
- 11) J. Jurczak and A. Golebiowski, *Chem. Rev.*, **89**, 149 (1989).
- 12) R. Polt, M. A. Peterson, and L. D. Young, *J. Org. Chem.*, **57**, 5469 (1992).
- 13) In the preliminary communication,³⁾ we pictured acetone **19** adopting a chair conformation. But the twist-boat conformation depicted in Scheme 3 is favored for **19** because of 1,3-diaxial interaction in the chair conformation. We thank a referee for this suggestion. See: S. D. Rychnovsky, G. Yang, and J. P. Powers, *J. Org. Chem.*, **58**, 5251 (1993), and references cited therein.
- 14) G. W. J. Fleet and D. R. Witty, *Tetrahedron: Asymmetry*, **1**, 119 (1990).
- 15) R. B. Woodward, E. Logusch, K. P. Nambiar, K. Sakan, D. E. Ward, B.-W. Au-Yeung, P. Balaram, L. J. Browne, P. J. Card, C. H. Chen, R. B. Chênevert, A. Fliri, K. Frobé, H.-J. Gais, D. G. Garratt, K. Hayakawa, W. Heggie, D. P. Hesson, D. Hoppe, I. Hoppe, J. A. Hyatt, D. Ikeda, P. A. Jacobi, K. S. Kim, Y. Kobuke, K. Kojima, K. Krowicki, V. J. Lee, T. Leutert, S. Malchenko, J. Martens, R. S. Matthews, B. S. Ong, J. B. Press, T. V. Rajan Babu, G. Rousseau, H. M. Sauter, M. Suzuki, K. Tatsuta, L. M. Tolbert, E. A. Truesdale, I. Uchida, Y. Ueda, T. Uyehara, A. T. Vasella, W. C. Vladuchick, P. A. Wade, R. M. Williams, and H. N.-C. Wong, *J. Am. Chem. Soc.*, **103**, 3215 (1981).
- 16) K. C. Nicolaou, S. P. Seitz, and D. P. Papahatjis, *J. Am. Chem. Soc.*, **105**, 2430 (1983).
- 17) For 1,2-SPh group migration, see: K. C. Nicolaou, T. Ladduwahetty, J. L. Randall, and A. Chucholowski, *J. Am. Chem. Soc.*, **108**, 2466 (1986); H. M. Zuurmond, P. A. M. van der Klein, G. A. van der Marel, and J. H. van Boom, *Tetrahedron Lett.*, **33**, 2063 (1992); H. M. Zuurmond, P. A. M. van der Klein, G. A. van der Marel, and J. H. van Boom, *Tetrahedron*, **49**, 6501 (1993).