A Linear Synthesis of Branched High-Mannose Oligosaccharides from the HIV-1 Viral Surface Envelope Glycoprotein gp120

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Described is a linear solution-phase synthesis of the HIV-1 viral surface envelope glycoprotein gp120 high-mannose nonasaccharide pentyl glycoside. Envisioning the automated solid-phase assembly of complex carbohydrates, the synthesis of the nonasaccharide and the related tri- and hexa-mannosides demonstrates the facile assembly of highly branched structures in a stepwise fashion incorporating monosaccharide building blocks. A differentially protected core trisaccharide was prepared and further elongated in two

high-yielding tri-mannosylations to furnish the triantennary structure. The tri-, hexa-, and nonamannoside *n*-pentyl glycosides obtained via the described synthesis are currently being used for detailed study of the carbohydrate protein interactions responsible for binding of the anti-HIV protein cyanovirin-*N* to the glycoprotein gp120.

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Introduction

Carbohydrates play a vital role in HIV retroviral pathogenesis. The function of HIV-1 surface envelope glycoprotein gp120 in helper T lymphocyte (T_h) infection has been understood by biologists for some time.^[1,2] Glycoprotein gp120 is highly glycosylated, containing up to 24 *N*-linked high-mannose carbohydrates, which compose 50% of the molecular mass of the glycoprotein.^[3] Gp120 mediates viral fusion with the CD4 receptor on the surface of T_h cells. HIV fusion and subsequent lymphocyte infection occur upon binding of the glycoprotein and the CD4 receptor.

A novel protein has been recently discovered that exhibits potent anti-HIV activity. Isolated from the blue-green algae *Nostoc elliposporum*, cyanovirin-N (CVN) was found to bind gp120, inactivating HIV, and preventing lymphocyte infection.^[2,4–7] CVN is a novel, monomeric, 11-kDa virucidal protein. Both natural and recombinant forms of the protein have been shown to irreversibly inactivate a wide variety of HIV strains while exhibiting minimal toxicity to host cells. The mode of HIV inactivation by CVN has been studied and the protein's affinity for the gp120 high-mannose structure Man₉ 1 was established (Figure 1).^[8] This affinity appears to be the mechanism through which CVN prevents gp120 from interacting with the CD4 receptor of the host lymphocyte. A detailed understanding of the specific interaction between CVN and Man₉ 1 could lead to

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the development of potential HIV preventatives and possibly even therapeutics. Access to synthetically derived mannosides as synthetic tools has therefore become particularly important in order to probe CVN-gp120 binding.

The total synthesis of high-mannose type cell surface glycans, that are found throughout nature as *N*-linked glycoconjugates, has been explored for the past two decades.^[9–12] A synthesis of the high-mannose core structures of Man₉, isolated from calf thyroglobul, was reported by Ogawa in 1981; a decade before the role of such structures in HIV pathogenesis was discovered.^[9] Two successful and highly convergent syntheses of the HIV-1 nonamannoside structure by Fraser-Reid^[13] and Ley^[14–16] were recently completed. However, the convergent nature of these solutionphase syntheses does not allow for their application to automated solid-phase synthesis.

Herein we describe a linear synthesis of the pentyl nonamannoside 2 and the related hexamannoside 3 and trimannoside 4 structures (Figure 1). The synthetic strategy was planned and executed with automated solid-phase synthesis in mind. Using three monomer building blocks the nonamannoside was assembled in four glycosylation events. Two sequential tri-mannosylation reactions, with a single mannosyl donor, allowed for access to the completed structure in a minimal number of steps. The target structures currently serve as tools in biophysical studies focusing on the elucidation of cyanovirin-*N*-binding to branched highmannose structures.

Results and Discussion

Our retrosynthetic analysis of Man_9 analog 2 (Scheme 1) was guided by our long-term goal of developing a synthetic

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Figure 1. High-mannose oligosaccharide targets

strategy that could be applied to the solid support and eventually automated. We planned to obtain the fully protected nonamannoside 5 via simultaneous tri-mannosylation of a hexasaccharide triol derived from 6. This hexasaccharide would in turn be prepared by the tri-mannosylation of a trisaccharide core triol ensuing from 7. The differentially protected trisaccharide 7 would be constructed from three protected monosaccharide building blocks 8, 9, and 10. The stepwise nature of this strategy would facilitate access to a triply branched nonasaccharide using just four glycosylations. A solution-phase synthesis of the β-pentenvl glycoside analog of the natural structure was selected for our studies, based on our strategy for solid phase oligosaccharide synthesis that furnishes the pentenyl glycoside upon cleavage from the solid-support.^[17] In addition to functioning as glycosyl donors, n-pentenyl glycosides are versatile handles, that can be converted into a range of functional groups by transformation of the terminal olefin to a carboxylic acid, aldehyde, ester, thioether, thioester, or hydroxyl group.^[18–20] In this fashion the *n*-pentenyl moiety



Scheme 1. Retrosynthesis of nonamannoside 5

may allow for attachment of the products of our synthetic studies to a linker, fluorescent tag, or biomarker.

Initially, reliable synthetic access to the monosaccharide building blocks had to be established. The core 3,6-differentially protected β -mannoside **8** was the most synthetically challenging building block to be procured. Since glycals have proven to be useful intermediates in oligosaccharide assembly^[21–22] we utilized glycals for the preparation of β mannoside **8**. The synthesis of **8** commenced with benzylation of known glycal **11**^[23] to yield differentially protected



Scheme 2. Synthesis of the core β -mannoside 8

glucal 12 (Scheme 2). Stereospecific epoxidation of glycal 12 by treatment with dimethyldioxirane, followed by opening of the 1,2-anhydrosugar with 4-penten-1-ol in the presence of zinc chloride yielded β -glucoside 13 containing an unprotected C2 hydroxyl group. Inversion of the C2 stereocenter was achieved via an oxidation-reduction se-Oxidation of the C2 quence. hydroxyl under Pfitzner-Moffatt conditions (Ac₂O/DMSO) was followed by stereoselective reduction with sodium borohydride and benzylation to furnish the fully protected β -mannoside 8. Selective removal of the halobenzyl ether was accomplished by palladium-catalyzed amination with N-methyl aniline followed by treatment with dichloroacetic acid to afford monosaccharide acceptor 14.^[23]

At this stage only two other mannosyl monosaccharide building blocks were required for the completion of the nonamannoside; one with a temporary protecting group on C2 (9), and the other incorporating temporary protecting groups on C3 and C6 (10). The C3 hydroxyl of the reducing end mannose is the point of attachment of a linear strand of α -(1 \rightarrow 2) linked mannoses (branch D1). The C6 hydroxyl of the core *n*-pentenyl- β -mannoside is the origin of a 3,6differentiated α -mannose that serves as the core for two branches (D2 and D3) of the triantennary structure (Figure 1). Differentially protected mannopyranosyl trichloroacetimidate $9^{[24]}$ as well as mannosyl donor $10^{[25]}$ were prepared using known procedures. Synthesized on multi gram scale, 9 was about to serve as the source of seven of the nine mannoses in the final Man₉ structure.

With the three monosaccharide building blocks **8**, **9**, and **10** in hand the assembly of larger structures began. We first focused on the preparation of core trimannoside **7**. The α -selective mannosylation of the C3 hydroxyl of **14** with



Scheme 3. Assembly of core trimannoside 7

donor 9 afforded disaccharide 15 (Scheme 3). Removal of the C6 silyl ether was accomplished in high yield using aqueous trifluoroacetic acid in THF to furnish disaccharide acceptor 16. Installation of the second mannose building block on C6 that will serve as root for branches D2 and D3 required the use of a mannose donor that did not utilize a C2 participating ester group but rather a permanently blocked benzyl ether. Nevertheless, complete α -selectivity was achieved in the TBDMSOTf catalyzed glycosylation of disaccharide acceptor 16 with mannosyl donor 10 to yield the fully-protected core trisaccharide 7. Precedence for α selectivity has been established previously in the case of similar mannosyl donors containing non-participating protecting groups on C2.^[26-27] Treatment of trisaccharide 7 with sodium methoxide accomplished the simultaneous cleavage of one C2 acetate and two benzoates to provide trisaccharide triol 17 in good yield (89%).

Simultaneous extension of the D1, D2, and D3 branches from the core trisaccharide was accomplished via two sequential tri-mannosylations with donor 9 (Scheme 4). Mannosylation of 17 with donor 9 (4.5 equivalents) yielded hexasaccharide 6 in a single step in 94% yield. Deprotection with sodium methoxide produced hexasaccharide triol 18. Trimannosylation of 18 using 9 furnished the fully protected high-mannose nonasaccharide 5 in 80% yield. In just three steps the initial trisaccharide 7 tripled in size to nonasaccharide 5 with 75% overall yield. Liberation of 5 from all protecting groups in two steps by treatment with sodium methoxide to afford triol 19, followed by Pd-catalyzed hydrogenation yielded the desired high-mannose pentyl glycoside 2 (Man₉). Fully deprotected hexasaccharide 3 and core trisaccharide 4 pentyl glycosides were prepared in similar fashion from 17 and 18 (Scheme 4). The three highmannose oligosaccharides 2, 3, and 4 are currently used in biophysical studies to better understand the carbohydrate protein interactions between cyanovirin-N and gp120. Results of these studies are expected to elucidate the formation of very tight carbohydrate-protein interactions that form

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Scheme 4. Assembly of nonasaccharide 6

the basis for novel HIV preventatives and possibly constitute a general principle for protein-carbohydrate interactions.

Conclusion

In summary, we have completed a linear synthesis of the high-mannose nonasaccharide pentyl glycoside 2, and the trimannoside 4 and hexamannoside 3 structures, using just three monomeric glycosyl building blocks. Construction of triantennary nonasaccharide 2 was achieved in four high-yielding glycosidation events; construction of a differentiated core trisaccharide followed by two sequential trimannosylations. Access to these synthetic structures has permitted further study of the anti-HIV micobicide activity exhibited by cyanovirin-N. The strategy described here for the solution phase assembly of three large oligosaccharides constitutes the basis for the solid phase and eventually the automated synthesis of branched high-mannose motifs.

Experimental Section

All commercial materials were used without further purification, unless otherwise noted. Dichloromethane (CH_2Cl_2) and diethyl ether (Et_2O) were purchased from J. T. Baker $(Cycletainer^{TM})$ and passed through neutral alumina columns prior to use. Toluene was purchased from J. T. Baker $(Cycletainer^{TM})$ and passed through neutral alumina and copper(II) oxide columns prior to use. Analytical thin-layer chromatography was performed on E. Merck silica gel 60 F₂₅₄ plates (0.25 mm). Compounds were visualized by dipping the plates in a cerium sulfate-ammonium molybdate solution, followed by heating. Liquid chromatography was performed using forced flow of the indicated solvent on Silicycle Inc. silica gel (230-400 mesh). ¹H NMR spectra were obtained on either a Bruker Avance 400 (400 MHz) or Varian VXR-500 (500 MHz) and are reported in parts per million (δ) relative to chloroform (δ = 7.26). Coupling constants (J) are reported in Hz. ¹³C NMR spectra were recorded on either a Bruker Avance 400 (100 MHz) or a Varian VXR-500 (125 MHz) and are reported in δ relative to $CDCl_3$ ($\delta = 77.23$) as an internal reference. IR spectra were obtained on a Perkin-Elmer 1600 series FTIR spectrometer. ESI Mass spectrometry was performed on a Bruker Daltonics Apex 3 Tesla Fourier Transform Mass Spectrometer. MALDI-TOF mass spectra were obtained on a PerSpective Biosystems Voyager Elite DE Spectrometer using 2,5-dihydroxybenzoic acid or α-cyano-4hydroxycinnamic acid as the matrix.

1,5-Anhydro-4-*O*-benzyl-3-*O*-(4-bromobenzyl)-2-deoxy-6-*O*-triisopropylsilyl-D-*arabino*-hex-1-enitol (12): 1,5-anhydro-3-*O*-(4-bromobenzyl)-2-deoxy-6-*O*-triisopropylsilyl-D-*arabino*-hex-1-enitol 11 (7.80 g, 16.5 mmol) was dissolved in DMF (125 mL) and cooled on an ice-bath to 0 °C. Sodium hydride (0.80 g, 60% in mineral oil, 20 mmol) was carefully added to the solution, and stirred for 20 min at 0 °C. Benzyl bromide (3.39 g, 19.9 mmol) was added to the reaction mixture and slowly warmed to room temperature for 2 h. Methanol (3 mL) was slowly added to quench the reaction, which was further diluted in 150 mL water. The solution was extracted with diethyl ether (3 \times 300 mL). After concentration in vacuo the resulting residue was purified by flash column chromatography on silica gel (2 \rightarrow 5% EtOAc/hexanes) to afford 8.60 g (93%) of **12** as a clear oil. [*a*]_D²⁴ = -19.6 (c = 1.1, CH₂Cl₂). IR (thin film): \tilde{v} = 942, 2865, 1647, 1240, 1101, 682 cm⁻¹. ¹H NMR (CDCl₃): δ = 1.07–1.10 (m, 21 H), 3.93–4.07 (m, 4 H), 4.18–4.20 (m, 1 H), 4.52 (d, J = 12.2 Hz, 1 H), 4.59 (d, J = 11.9 Hz, 1 H), 4.78–4.85 (m, 3 H), 6.41 (dd, J = 1.5, 6.1 Hz, 1 H), 7.20–7.22 (d, J = 8.2 Hz, 2 H), 7.30–7.36 (m, 5 H), 7.44–7.46 (d, J = 8.2 Hz, 2 H). ¹³C NMR (CDCl₃): δ = 12.2, 18.2, 18.2, 62.0, 70.0, 74.1, 74.2, 76.0, 78.3, 99.5, 121.6, 128.0, 128.1, 128.6, 129.5, 137.7, 138.6, 145.1. ESI MS *m/z* (M + Na⁺) calcd. 583.1849, found 583.1838.

4-Pentenyl 4-O-Benzyl-3-O-(4-bromobenzyl)-6-O-triisopropylsilyl-β-D-glucopyranoside (13): Glucal 12 (1.82 g, 3.23 mmol) was dissolved in CH₂Cl₂ (6 mL) and cooled to 0 °C. A 0.08 M solution of dimethyldioxirane in acetone (48.5 mL, 3.88 mmol) was added and the reaction was stirred for 15 min. After the solvent was removed the remaining residue was dried in vacuo for 1.5 h and subsequently dissolved in CH₂Cl₂ (10 mL). The solution was cooled to -78 °C followed by the addition of 4-penten-1-ol (1.61 mL, 16.15 mmol). A 1.0 м solution of ZnCl₂ in diethyl ether (3.55 mL, 3.55 mmol) was added and the reaction was warmed slowly to room temperature and stirred over 16 h. The reaction was diluted with EtOAc (200 mL) and washed with sat. aqueous NaHCO₃ (2 \times 100 mL), water (2 \times 100 mL), and brine (2 \times 100 mL) and dried (Na_2SO_4) . The organic phase was concentrated in vacuo and the resulting residue purified by flash column chromatography on silica gel (15% EtOAc/hexanes) to afford 1.86 g (87%) of 13 as a clear oil. $[\alpha]_{D}^{24} = -20.8 \ (c = 1.3, CH_2Cl_2)$. IR (thin film): $\tilde{\nu} = 456, 2941$, 2865, 1115, 1069, 689 cm⁻¹. ¹H NMR (CDCl₃): $\delta = 0.89 - 1.07$ (m, 21 H), 1.61-1.68 (m, 2 H), 2.02-2.08 (m, 2 H), 2.27-2.28 (m, 1 H), 3.21-3.25 (m, 1 H), 3.51-3.60 (m, 4 H), 3.76-3.92 (m, 3 H), 4.15 (d, J = 7.5, 1 H), 4.63 (d, J = 11.6, 1 H), 4.79 (d, J = 12.5, 11 H), 4.82 (d, J = 11.6, 1 H), 4.81 (d, J = 11.4, 1 H), 4.89–4.98 (m, 2 H), 5.69-5.79 (m, 1 H), 7.16-7.26 (m, 7 H), 7.36 (d, J =6.5 Hz, 2 H). ¹³C NMR (CDCl₃): $\delta = 12.2, 18.2, 29.0, 30.5, 62.6,$ 69.2, 74.4, 75.1, 75.3, 76.4, 77.4, 84.6, 102.6, 115.1, 121.7, 128.0, 128.1, 128.7, 129.8, 131.7, 138.0, 138.3, 138.5. ESI MS m/z (M + Na⁺) calcd. 685.2530, found 685.2532.

4-Pentenyl 2,4-Di-O-benzyl-3-O-(4-bromobenzyl)-6-O-triisopropylsilyl-β-D-mannopyranoside (8): Glucoside 13 (0.56 g, 0.85 mmol) was azeotropically dried with toluene $(3 \times 3 \text{ mL})$ and dissolved in dimethyl sulfoxide (3.5 mL). Acetic anhydride (1.75 mL) was added and the reaction was allowed to stir 24 h at room temperature. After the solvent was removed in vacuo, addition of CH₂Cl₂ (20 mL) was followed by washing with water (2 \times 20 mL) and drying of the organic phase (Na₂SO₄). After concentration in vacuo the residue was dissolved in 1:1 CH2Cl2/MeOH (10 mL) and cooled to 0 °C. NaBH₄ (0.161 g, 4.25 mmol) was slowly added and the reaction was stirred 16 h at room temperature. CH₂Cl₂ (100 mL) was added and the organic phase was washed with water (100 mL), 1% aqueous citric acid (2 \times 100 mL), sat. aqueous NaHCO₃ (100 mL), brine (100 mL), and dried (Na₂SO₄). The organic phase was dried in vacuo to give a clear oil and purified by flash column chromatography on silica gel $(6 \rightarrow 7\%$ EtOAc/hexanes) to afford 0.42 g (74%) of the desired 4-pentenyl 4-O-benzyl-3-O-(4bromobenzyl)-6-O-triisopropylsilyl- β -D-mannopyranoside. $[\alpha]_{D}^{24} =$ -32.0 (c = 1.3, CH₂Cl₂). IR (thin film): $\tilde{v} = 3439$, 2940, 2864, 1638, 1105, 1069, 683 cm⁻¹. ¹H NMR (CDCl₃): $\delta = 0.97 - 1.05$ (m, 2 H), 1.60-1.67 (m, 21 H), 2.04 (m, 2 H), 2.26 (d, J = 2.9 Hz, 1 H), 3.15-3.19 (m, 1 H), 3.40-3.46 (m, 2 H), 3.78-3.92 (m, 4 H), 3.98 (m, 1 H), 4.32 (d, J = 0.9 Hz, 1 H), 4.53 - 4.68 (m, 3 H), 4.80(d, J = 10.9 Hz, 1 H), 4.87 - 4.97 (m, 2 H), 5.68 - 5.78 (m, 1 H),

7.17-7.29 (m, 7 H), 7.36 (d, J = 8.4 Hz, 2 H). ¹³C NMR (CDCl₃): $\delta = 12.4, 18.4, 29.1, 30.7, 63.0, 68.9, 69.1, 71.0, 74.3, 75.6, 76.8,$ 82.0, 99.8, 115.2, 122.0, 128.2, 128.5, 128.9, 129.9, 131.9, 137.5, 138.5, 138.8. ESI MS m/z (M + Na⁺) calcd. 685.2530, found 685.2539. 4-Pentenyl 4-O-benzyl-3-O-(4-bromobenzyl)-6-O-triisopropylsilyl-β-D-mannopyranoside (1.0 g, 1.5 mmol) was azeotropically dried with toluene $(3 \times 3 \text{ mL})$ and dissolved in DMF (15 mL). The solution was cooled to 0 $^{\circ}\mathrm{C}$ and sodium hydride (90 mg, 60% in mineral oil, 1.81 mmol) was carefully added and the mixture was warmed to room temperature. Benzyl bromide (214 μ L, 1.81 mmol) was added to the solution, and stirred for 2 h. The reaction was diluted with diethyl ether (100 mL), washed with water (100 mL), followed by extraction of the combined aqueous phase with diethyl ether (50 mL). The combined organic phase was washed with sat. aqueous NaHCO₃ (100 mL), water (100 mL), brine (100 mL), dried (Na₂SO₄), and concentrated to give a thick oil in vacuo. The residue was purified by flash column chromatography on silica gel (5 \rightarrow 20% EtOAc/hexanes) to afford 1.17 g (98%) of 8. $[\alpha]_{D}^{24} = -54.0^{\circ}$ (c = 1.2, CH₂Cl₂). IR (thin film): $\tilde{\nu} = 2940$, 2865, 1454, 1361, 1107, 1070, 696 cm⁻¹. ¹H NMR (CDCl₃): δ = 1.04-1.16 (m, 21 H), 1.72-1.79 (m, 1 H), 2.18 (m, 2 H), 3.28-3.32 (m, 1 H), 3.43-3.51 (m, 2 H), 3.89-4.05 (m, 5 H), 4.68 (d, J =11.0 Hz, 1 H), 4.85-5.08 (m, 5 H), 5.82-5.92 (m, 1 H), 7.17 (d, J = 8.4 Hz, 2 H), 7.28–7.49 (m, 12 H). ¹³C NMR (CDCl₃): $\delta =$ 12.2, 18.2, 29.2, 30.6, 63.3, 69.1, 70.7, 73.7, 74.0, 75.0, 75.4, 77.4, 82.6, 102.0, 115.1, 121.7, 127.5, 127.9, 128.2, 128.4, 128.6, 129.4, 131.6, 137.6, 138.5, 138.7, 139.1. ESI MS m/z (M + Na⁺) calcd. 775.3000, found 775.3020.

4-Pentenyl 2,4-Di-O-benzyl-6-O-triisopropylsilyl-B-D-mannopyranoside (14): Mannoside 8 (0.51 g, 0.67 mmol) was azeotropically dried with toluene $(3 \times 3 \text{ mL})$ then for 1 h in vacuo. The residue was dissolved in toluene (2 mL) followed by the addition of N-methyl aniline (86 mg, 0.805 mmol). An oven-dried Schlenk flask was evacuated and backfilled with argon (5 \times). The flask was charged with Pd₂(dba)₃ (12.3 mg, 2 mol %), (o-biphenyl)P(tBu)₂ (4 mol %), and NaOtBu (0.090 g, 0.94 mmol), evacuated and backfilled with argon (5 \times). A rubber septum was installed and the aryl bromide/ amine solution was added via cannula. The flask was sealed using a Teflon screwcap and the reaction mixture was heated to 80 °C with stirring. After 18 h, the reaction was cooled to room temperature, diluted with diethyl ether (20 mL), filtered through a pad of celite, and concentrated in vacuo. The crude product was purified by flash column chromatography on silica gel $(2 \rightarrow 5\% \text{ EtOAc}/$ hexanes) to yield 0.483 g (92%) of 4-pentenyl 4-O-benzyl-3-O-[4-(N-methyl-N-phenylamino)benzyl]-6-O-triisopropylsilyl-β-Dmannopyranoside. $[\alpha]_{D}^{24} = -52.6$ (c = 1.5, CH₂Cl₂). IR (thin film): $\tilde{v} = 2940, 2865, 1595, 1497, 1105, 1067, 696 \text{ cm}^{-1}$. ¹H NMR $(CDCl_3): \delta = 1.10 - 1.16 \text{ (m, 21 H)}, 1.71 - 1.79 \text{ (m, 2 H)}, 2.15 - 2.21$ (m, 2 H), 3.29-3.35 (m, 4 H), 3.43-3.49 (m, 1 H), 3.55 (dd, J =3.0, 9.4 Hz, 1 H), 3.90-4.06 (m, 5 H), 4.40 (s, 1 H), 4.47 (d, J =11.5 Hz, 1 H), 4.54 (d, J = 11.5 Hz, 1 H), 4.67 (d, J = 10.9 Hz, 1 H), 4.89-5.09 (m, 5 H), 5.82-5.93 (m, 1 H), 6.97-7.08 (m, 4 H), 7.23-7.37 (m, 13 H), 7.51 (d, J = 6.9 Hz, 2 H). ¹³C NMR (CDCl₃): $\delta = 12.2, 18.2, 29.2, 30.6, 40.5, 63.4, 69.0, 71.5, 73.7, 74.1, 75.0,$ 75.4, 82.6, 102.0, 115.1, 120.6, 120.9, 121.6, 127.4, 127.8, 128.2, 128.4, 128.5, 129.2, 129.4, 131.0, 138.5, 138.9, 139.3, 148.9, 149.3. ESI MS m/z (M + Na⁺) calcd. 802.4479, found 802.4473. The aminated product (68 mg, 0.088 mmol) was dissolved in CH₂Cl₂ (3 mL) followed by the addition of dichloroacetic acid (72 μ L, 0.88 mmol), resulting in a transparent blue color. The reaction was stirred for 30 min at room temperature then diluted with CH₂Cl₂ (20 mL). Washing with sat. aqueous NaHCO₃ (2×30 mL), brine (30 mL), was followed by drying (Na₂SO₄) and concentration in

vacuo. The crude product was purified by flash column chromatography on silica gel (2 \rightarrow 5% EtOAc/ toluene) to afford 46 mg (90%) of **14**. [α]_D²⁴ = -31.4° (c = 1.3, CH₂Cl₂). IR (thin film): \tilde{v} = 446, 2940, 2865, 1734, 1455, 1249, 1085, 695 cm⁻¹. ¹H NMR (CDCl₃): δ = 0.94–1.07 (m, 21 H), 1.62–1.67 (m, 2 H), 2.03–2.08 (m, 2 H), 2.48 (d, J = 9.7 Hz, 1 H), 3.17–3.20 (m, 1 H), 3.35–3.41 (m, 1 H), 3.53–3.64 (m, 2 H), 3.73 (d, J = 3.1 Hz, 1 H), 3.83–3.96 (m, 3 H), 4.39 (d, J = 0.4 Hz, 1 H), 4.53–4.56 (m, 2 H), 4.80 (d, J = 11.1 Hz, 1 H), 4.87–5.00 (m, 3 H), 5.69–5.80 (m, 1 H), 7.17–7.32 (m, 10 H). ¹³C NMR (CDCl₃): δ = 12.5, 18.2, 29.2, 30.6, 63.2, 69.1, 74.1, 74.6, 75.0, 76.7, 76.9, 77.9, 101.7, 115.0, 127.9, 128.2, 128.5, 138.4, 138.7, 138.8. ESI MS m/z (M + Na⁺) calcd. 607.3425, found 607.3440.

2-O-Acetyl-3,4,6-tri-O-benzyl-α-D-mannopyranosyl-**4-Pentenyl** $(1\rightarrow 3)$ -2,4-di-O-benzyl-6-O-triisopropylsilyl- β -D-mannopyranoside (15): Mannosyl trichloroacetimidate 9 (65 mg, 0.10 mmol) and glycosyl acceptor 14 (30 mg, 0.051 mmol) were azeotropically dried with toluene $(3 \times 3 \text{ mL})$ and dissolved in CH₂Cl₂ (1 mL). The solution was cooled to -20 °C for 15 min, followed by the addition TBDMSOTf (2.4 μ L, 0.010 mmol), and stirred for 30 min at -20°C. The reaction was quenched by addition of Et_3N (50 µL), and dried in vacuo. The crude product was purified by flash column chromatography on silica gel (2 \rightarrow 5% EtOAc/toluene) to afford 54 mg (99%) disaccharide 15. $[\alpha]_D^{24} = -15.6$ (c = 1.4, CH₂Cl₂). IR (thin film): $\tilde{v} = 940, 2865, 1745, 1235, 1078, 697 \text{ cm}^{-1}$. ¹H NMR $(CDCl_3)$: $\delta = 0.93 - 1.04$ (m, 21 H), 1.57 - 1.64 (m, 2 H), 1.99 - 2.06(m, 5 H), 3.13-3.17 (m, 1 H), 3.27-3.33 (m, 1 H), 3.51-3.52 (m, 2 H), 3.54-3.93 (m, 9 H), 4.29 (s, 1 H), 4.35-4.70 (m, 8 H), 4.79 (d, J = 11.1 Hz, 1 H), 4.87-4.95 (m, 3 H), 5.12 (d, J = 1.2 Hz, 1 Hz, 1 Hz)H), 5.43-5.44 (m, 1 H), 5.69-5.79 (m, 1 H), 7.04-7.32 (m, 25 H). ¹³C NMR (CDCl₃): $\delta = 12.5, 18.2, 21.2, 29.2, 30.6, 63.0, 69.0,$ 69.1, 69.2, 72.1, 72.2, 73.6, 73.8, 73.9, 74.5, 75.0, 75.3, 77.2, 77.7, 78.3, 80.6, 99.9, 101.7, 114.9, 127.2, 127.6, 127.7, 127.8, 127.9, 127.9, 128.2, 128.3, 128.4, 128.4 128.5, 128.6, 138.1, 138.3, 138.4, 138.4, 138.8, 139.2, 170.5. ESI MS m/z (M + Na⁺) calcd. 1081.5468, found 1081.5428.

2-O-Acetyl-3,4,6-tri-O-benzyl-α-D-mannopyranosyl-**4-Pentenyl** $(1\rightarrow 3)$ -2,4-di-*O*-benzyl- β -D-mannopyranoside (16): To a solution of disaccharide 15 (0.108 g, 0.103 mmol) in THF (2 mL), water (2 mL) and trifluoroacetic acid (0.8 mL) were added. The turbid white solution was stirred for 2 h at room temperature. The reaction was diluted with diethyl ether (30 mL) and washed with sat. aqueous NaHCO₃ (2 \times 20 mL), brine (20 mL), dried (Na₂SO₄), and concentrated. The residue was purified by flash column chromatography on silica gel (10 \rightarrow 30% EtOAc/hexanes) to afford 84 mg (91%) of disaccharide 16. $[\alpha]_{D}^{24} = -24.0$ (c = 0.5, CH₂Cl₂). IR (thin film): $\tilde{v} = 443$, 2089, 1639, 1234, 698 cm⁻¹. ¹H NMR (CDCl₃): $\delta = 1.70 - 1.78$ (m, 2 H), 2.12-2.22 (m, 6 H), 3.31-3.39 (m, 1 H), 3.41-3.44 (m, 1 H), 3.63-3.69 (m, 2 H), 4.75-3.43 (m, 9 H), 4.43-4.73 (m, 8 H), 4.78-4.84 (m, 2 H), 4.91 (d, J =11.0 Hz, 1 H), 4.97-5.09 (m, 3 H), 5.26 (d, J = 1.5 Hz, 1 H), 5.54-5.55 (m, 1 H), 5.80-5.90 (m, 1 H), 7.17-7.42 (m, 25 H). ¹³C NMR (CDCl₃): $\delta = 21.2, 29.0, 30.4, 62.3, 68.9, 69.2, 69.7, 72.1,$ 72.3, 74.4, 74.4, 75.0, 75.4, 76.0, 77.5, 78.1, 80.0, 99.8, 101.8, 115.4, 127.6, 127.7, 127.8, 127.8, 127.8, 128.0, 128.0, 128.1, 128.3, 128.4, 128.2, 128.6, 128.6, 137.9, 137.9, 138.2, 138.3, 138.6, 138.7, 170.5. ESI MS m/z (M + Na⁺) calcd. 925.4133, found 925.4141.

4-Pentenyl 2-*O*-Acetyl-3,4,6-tri-*O*-benzyl-α-D-mannopyranosyl- $(1\rightarrow 3)$ -[2,4-di-*O*-benzyl-3,6-di-*O*-benzoyl-α-D-mannopyranosyl- $(1\rightarrow 6)$]-2,4-di-*O*-benzyl-β-D-mannopyranoside (7): Mannosyl trichloroacetimidate 10 (94 mg, 0.15 mmol) and disaccharide 15 (80 mg, 0.088 mmol) were combined, azeotropically dried with toluene (3

 \times 3 mL) and dissolved in diethyl ether (2 mL). The solution was cooled to -20 °C for 15 min, followed by the addition of TBDMSOTf (4 μ L, 0.018 mmol), and stirred for 30 min at -20 °C. The reaction was quenched by the addition of Et_3N (50 µL), and dried in vacuo. The crude product was purified by flash column chromatography on silica gel (5 \rightarrow 10% EtOAc/toluene) to afford 120 mg (93%) of trisaccharide 7. $[\alpha]_{D}^{24} = +9.1$ (c = 1.0, CH₂Cl₂). IR (thin film): $\tilde{v} = 917, 1721, 1452, 1270, 1097, 1070, 698 \text{ cm}^{-1}$. ¹H NMR (CDCl₃): $\delta = 1.42 - 1.51$ (m, 2 H), 1.88 - 1.94 (m, 2 H), 1.97 (s, 3 H), 3.21-3.27 (m, 2 H), 3.46-3.53 (m, 2 H), 3.65-4.03 (m, 12 H), 4.16-4.35 (m, 5 H), 4.40-4.62 (m, 10 H), 4.73-4.85 (m, 5 H), 5.11 (s, 1 H), 5.40-5.42 (m, 1 H), 5.55-5.64 (m, 2 H), 6.83-6.86 (m, 1 H), 6.95-7.25 (m, 36 H), 7.31-7.49 (m, 4 H), 7.92-7.98 (m, 4 H). ¹³C NMR (CDCl₃): $\delta = 21.5, 29.3, 30.6, 64.0,$ 66.5, 69.3, 69.4, 69.6, 70.4, 72.5, 72.5, 72.5, 73.6, 73.8, 74.4, 74.6, 74.8, 75.2, 75.4, 75.5, 75.7, 76.1, 77.1, 77.8, 78.4, 80.7, 98.6, 100.1, 101.9, 115.2, 127.6, 127.7, 127.8, 128.0, 128.1, 128.2, 128.2, 128.4, 128.5, 128.6, 128.7, 128.7, 128.8, 128.8, 128.9, 128.9, 130.2, 130.5, 130.5, 133.3, 133.5, 137.9, 138.3, 138.3, 138.6, 138.8, 139.0, 139.0, 166.0, 166.8, 170.4. ESI MS m/z (M + Na⁺) calcd. 1475.6130, found 1475.6141.

4-Pentenyl 3,4,6-Tri-O-benzyl- α -D-mannopyranosyl- $(1 \rightarrow 3)$ -[2,4-di-*O*-benzyl-α-D-mannopyranosyl-(1→6)]-2,4-di-*O*-benzyl-β-D-mannopyranoside (17): Trisaccharide 7 (100 mg, 0.0687 mmol) was dissolved in CH₂Cl₂/MeOH (4 mL, 1:1). A solution of sodium methoxide in MeOH (450 µL, 25% w/v, 2 mmol) was added and the reaction was heated on an oil-bath to 45 °C for 1.5 h. The reaction was quenched with DOWEX-50 W-hydrogen strongly acidic ionexchange resin, filtered, and dried in vacuo. The resulting crude product was purified by flash column chromatography on silica gel $(10 \rightarrow 40\% \text{ EtOAC/toluene})$ to afford 73 mg (89%) of trisaccharide **17.** $[\alpha]_{D}^{24} = 11.0$ (c = 1.3, CH₂Cl₂). IR (thin film): $\tilde{v} = 470$, 2922, 1453, 1364, 1072, 697 cm⁻¹. ¹H NMR (CDCl₃): $\delta = 1.50 - 1.56$ (m, 2 H), 1.84 (br. s, 1 H), 1.94-2.00 (m, 2 H), 2.23 (br. s, 2 H), 3.20-3.29 (m, 2 H), 3.49-3.54 (m, 2 H), 3.55-3.83 (m, 13 H), 3.86-3.92 (m, 3 H), 4.25-4.58 (m, 10 H), 4.64-4.76 (m, 3 H), 4.83–4.91 (m, 4 H), 5.11 (d, J = 0.97 Hz, 1 H), 5.18 (d, J = 1.2 Hz, 1 H), 5.61-5.72 (m, 1 H), 7.05-7.30 (m, 35 H). ¹³C NMR $(CDCl_3): \delta = 29.3, 30.6, 62.7, 66.5, 69.1, 69.4, 69.7, 71.7, 72.2,$ 72.2, 72.6, 73.0, 73.7, 75.2, 75.4, 75.7, 76.1, 76.9, 77.6, 77.9, 79.1, 80.3, 80.6, 98.0, 101.7, 102.1, 115.3, 127.8, 127.9, 127.9, 128.0, 128.1, 128.1, 128.1, 128.1, 128.3, 128.3, 128.4, 128.6, 128.7, 128.7, 128.8, 128.9, 129.0, 129.0, 138.2, 138.2, 138.4, 138.4, 138.4, 138.6, 138.9, 138.9, 139.0. ESI MS m/z (M + Na⁺) calcd. 1225.5500, found 1225.5497.

4-Pentenyl 2-O-Acetyl-3,4,6-tri-O-benzyl-α-D-mannopyranosyl- $(1\rightarrow 2)$ -3,4,6-tri-*O*-benzyl- α -D-mannopyranosyl- $(1\rightarrow 3)$ -[2-*O*-acetyl-3,4,6-tri-O-benzyl-α-D-mannopyranosyl-(1→3)-[2-O-acetyl-3,4,6-tri-*O*-benzyl- α -D-mannopyranosyl- $(1\rightarrow 6)$]-2,4-di-*O*-benzyl- α -D-mannopyranosyl- $(1\rightarrow 6)$]-2,4-di-O-benzyl- β -D-mannopyranoside (6): Trisaccharide acceptor 17 (25 mg, 0.021 mmol) and mannosyl trichloroacetimidate 9 (60 mg, 0.093 mmol, 4.5equiv.) were combined, azeotropically dried with toluene $(3 \times 3 \text{ mL})$ and dissolved in CH₂Cl₂ (2 mL). The solution was cooled to -20 °C for 15 min, followed by the addition of TMSOTf (2.2 µL, 0.012 mmol). The reaction mixture was stirred and warmed to room temperature over 40 min. The reaction was quenched by the addition of Et_3N (100 μ L), and dried in vacuo. The crude product was purified by flash column chromatography on silica gel $(2 \rightarrow 20\%$ EtOAc/toluene) to afford 51 mg (94%) of hexasaccharide 6. $[\alpha]_{D}^{24} = +34.9 (c = 1.3, CH_2Cl_2).$ IR (thin film): $\tilde{v} = 029, 2916, 1744, 1235, 1076, 736, 697 \text{ cm}^{-1}$. ¹H NMR (CDCl₃): $\delta = 1.56 - 1.60$ (m, 2 H), 2.01 - 2.04 (m, 2 H), 2.09 (s, 3 H), 2.13 (s, 3 H), 2.16 (s, 3 H), 3.19-3.22 (m, 1 H), 3.30-3.36 (m, 2 H), 3.54-3.62 (m, 7 H), 3.67-3.72 (m, 6 H), 3.78-3.87 (m, 6 H), 3.89-3.99 (m, 10 H), 4.00-4.07 (m, 2 H), 4.09-4.13 (m, 1 H), 4.23-4.28 (m, 2 H), 4.34-4.37 (m, 1 H), 4.39-4.50 (m, 10 H), 4.53-4.59 (m, 6 H), 4.61-4.68 (m, 6 H), 4.73 (app. s, 1 H), 4.76 (d, J = 2.3 Hz, 1 H), 4.81-4.89 (m, 5 H), 4.90-4.94 (m, 3 H),4.98-5.03 (m, 3 H), 5.20-5.24 (m, 2 H), 5.52-5.53 (m, 3 H), 5.70-5.77 (m, 1 H), 7.10-7.40 (m, 80 H). ¹³C NMR (CDCl₃): $\delta =$ 21.8, 21.9, 29.6, 31.0, 66.1, 66.3, 68.4, 68.7, 68.8, 69.0 69.4, 69.5, 71.0, 71.4, 71.7, 72.0, 72.0, 72.0, 72.1, 72.3, 72.4, 72.6, 73.4, 73.4, 73.5, 73.6, 74.1, 74.2, 74.7, 74.9, 75.0, 75.2, 75.2, 75.3, 75.4, 77.5, 77.6, 77.8, 78.2, 78.4, 79.8, 81.7, 97.0, 98.4, 99.6, 100.0, 101.2, 101.7, 114.9, 127.4, 127.4, 127.5, 127.5, 127.6, 127.6, 127.6, 127.6, 127.7, 127.7, 127.7, 127.7, 127.8, 127.8, 127.9, 127.9, 127.9, 128.0, 128.0, 128.1 128.2, 128.3, 128.3, 128.4, 128.4, 128.4, 128.4, 128.5, 128.5, 128.5, 128.6, 128.6, 128.7, 128.7, 129.2, 138.0, 138.0, 138.2, 138.3, 138.4, 138.4, 138.5, 138.5, 138.5, 138.7, 138.8, 138.9, 139.0, 170.3, 170.3, 170.4; HSQC anomeric cross-peaks (CDCl₃) δ (4.23) \times 101.8), (4.93 \times 97.0), (5.02 \times 98.4), (5.06 \times 99.5), (5.21 \times 99.9), (5.22×101.1) . ESI MS m/z (M + Na⁺) calcd. 2648.1622, found 2648.1530.

4-Pentenyl 3,4,6-Tri-O-benzyl-α-D-mannopyranosyl-(1→2)-3,4,6-tri-*O*-benzyl-α-D-mannopyranosyl-(1→3)-[3,4,6-tri-*O*-benzyl-α-Dmannopyranosyl- $(1\rightarrow 3)$ -[3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1→6)]-2,4-di-O-benzyl-α-D-mannopyranosyl-(1→6)]-2,4-di-Obenzyl-β-D-mannopyranoside (18): Hexasaccharide 6 (46 mg, 0.018 mmol) was azeotropically dried with toluene $(3 \times 3 \text{ mL})$ and dissolved in CH₂Cl₂ (1.5 mL). MeOH (5 mL) was added followed by a solution of sodium methoxide in MeOH (120 µL, 25% w/v, 0.525 mmol). The reaction was stirred for 1 h, quenched with DOWEX-50 W-hydrogen strongly acidic ion-exchange resin, filtered, and dried in vacuo. The resulting residue was purified by flash column chromatography on silica gel (5 \rightarrow 20% EtOAc/toluene) to afford 46 mg (quant.) of hexasaccharide triol 18. $[\alpha]_{D}^{24} =$ +34.3 (c = 1.0, CH₂Cl₂). IR (thin film): $\tilde{v} = 448$, 3029, 2916, 1453, 1053, 697 cm⁻¹. ¹H NMR (CDCl₃): $\delta = 1.43 - 1.53$ (m, 2 H), 1.89-1.95 (m, 2 H), 2.07-2.28 (br. s, 3 H), 3.10-3.13 (m, 1 H), 3.19-3.24 (m, 1 H), 3.33-3.36 (m, 1 H), 3.43-3.62 (m, 13 H), 3.64-3.81 (m, 14 H), 3.82-3.92 (m, 3 H), 3.99-4.02 (m, 4 H), 4.13 (app. s, 1 H), 4.22-4.30 (m, 3 H), 4.35-4.40 (m, 8 H), 4.45-4.51 (m, 12 H), 4.53-4.65 (m, 4 H), 4.69-4.75 (m, 5 H), 4.78-4.83 (m, 3 H), 4.86-4.97 (m, 2 H), 5.02 (app. s, 1 H), 5.11 (app. s, 1 H), 5.16 (app. s, 1 H), 5.57-5.67 (m, 1 H), 6.81-7.48 (m, 80 H). ¹³C NMR (CDCl₃): $\delta = 14.9, 21.8, 22.2, 69.4, 69.5, 69.7,$ 69.9, 70.1, 71.9, 72.0, 72.0, 72.3, 72.5, 72.6, 72.7, 72.8, 73.0, 73.2, 73.9, 74.0, 74.1, 74.2, 74.6, 74.9, 74.9, 75.2, 75.4, 75.5, 75.7, 75.8, 75.9, 76.2, 77.9, 78.4, 78.5, 80.3, 80.5, 80.8, 82.5, 97.5, 100.5, 101.8, 101.8, 102.1, 102.1, 115.4, 127.9, 128.0, 128.1, 128.1, 128.2, 128.3, 128.3, 128.4, 128.4, 128.4, 128.5, 128.5, 128.6, 128.6, 128.8, 128.9, 128.9, 129.0, 129.0, 129.0, 129.1, 129.1, 129.1, 129.2, 129.3, 138.6, 138.7, 138.7, 138.8, 138.8, 138.9, 139.0, 139.0, 139.1, 139.3, 139.4, 139.5. ESI MS m/z (M + Na⁺) calcd. 2522.1305, found 2522.1307.

4-Pentenyl 2-O-Acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)-[2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)-[2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 6)]-2,4-di-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 6)]-2,4-di-O-benzyl- β -D-mannopyranoside 5. Hexasaccharide 18 (19 mg, 0.0075 mmol) and mannosyl trichloroacetimidate 9 (36 mg, 0.056 mmol, 7.5equiv.) were azeotropically dried with toluene (3 \times 3 mL), dried an addi-

tional 1.5 h in vacuo and dissolved in diethyl ether (2 mL). The solution was cooled to -20 °C for 15 min, followed by the addition of TMSOTf (1 µL, 0.005 mmol), and stirred for 30 min. The reaction was quenched by the addition of Et₃N (50 µL), and dried in vacuo. The crude product was purified by flash column chromatography on silica gel (2 \rightarrow 18% EtOAc/toluene) affording 24 mg (80%) of nonasaccharide 5. $[\alpha]_{D}^{24} = +24.7$ (c = 0.9, CH₂Cl₂). IR (thin film): $\tilde{v} = 029, 2864, 1744, 1453, 1137, 1056, 736, 697 \text{ cm}^{-1}$. ¹H NMR (CDCl₃): $\delta = 1.48 - 1.55$ (m, 2 H) 1.94 - 1.98 (m, 2 H), 2.08 (s, 3 H), 2.12 (s, 3 H), 2.13 (s, 3 H), 3.13-3.18 (m, 2 H), 3.26-3.30 (m, 1 H), 3.35-3.46 (m, 5 H), 3.50-3.52 (m, 3 H), 3.55-3.67 (m, 10 H), 3.73-3.78 (m, 4 H), 3.82-3.94 (m, 21 H), 3.97-4.06 (m, 5 H), 4.08-4.13 (m, 4 H), 4.16-4.22 (m, 3 H), 4.30-4.43 (m, 12 H), 4.45-4.51 (m, 9 H), 4.54-4.55 (m, 6 H), 4.56-4.58 (m, 4 H), 4.60-4.61 (m, 2 H), 4.63-4.68 (m, 4 H), 4.71-4.73 (m, 1 H), 4.74-4.77 (m, 2 H), 4.80-4.89 (m, 7 H), 4.95-5.00 (m, 2 H), 5.08-5.11 (m, 3 H), 5.14 (m, 1 H), 5.17 (m, 1 H), 5.23-5.24 (m, 1 H), 5.52-5.55 (m, 4 H), 5.60-5.69 (m, 1 H), 7.00-7.02 (m, 1 H), 7.07-7.34 (m, 124 H). ¹³C NMR (CDCl₃): $\delta = 20.5, 21.0, 21.3, 22.0, 29.1, 30.3, 66.6, 66.6, 66.6, 66.7, 68.2,$ 68.2, 68.8, 68.8, 68.8, m68.9, 69.0, 69.2, 69.2, 69.2, 69.4, 70.5, 70.8, 70.9, 71.4, 71.4, 71.9, 72.0, 72.0, 72.1, 72.2, 72.3, 72.7, 72.8, 73.3, 73.3, 73.4, 73.5, 73.5, 74.2, 74.9, 74.9, 75.2, 75.2, 75.2, 75.4, 75.4, 75.5, 77.4, 78.3, 78.4, 79.1, 79.2, 79.5, 79.9, 80.1, 82.2, 97.0, 99.3, 99.5, 99.5, 99.7, 100.7, 101.1, 101.4, 101.4, 115.0, 125.7, 126.2, 126.7, 126.6, 127.3, 128.0, 128.0, 128.3, 128.7, 129.3, 129.8, 130.3, 131.3, 132.5, 171.0, 171.0, 171.1; HSQC anomeric cross-peaks $(CDCl_3) \delta (4.16 \times 102.0), (4.86 \times 97.0), (5.00 \times 99.6), three (5.10)$ \times 99.6), (5.12 \times 100.7), (5.18 \times 101.2), (5.21 \times 101.3). ESI MS m/z (M + Na⁺) calcd. 3944.7437, found 3944.7440.

4-Pentenyl 3,4,6-Tri-O-benzyl-α-D-mannopyranosyl-(1→2)-3,4,6-tri-O-benzyl-α-D-mannopyranosyl-(1→2)-3,4,6-tri-O-benzyl-α-Dmannopyranosyl-(1→3)-[3,4,6-tri-O-benzyl-α-D-mannopyranosyl- $(1\rightarrow 2)$ -3,4,6-tri-*O*-benzyl- α -D-mannopyranosyl- $(1\rightarrow 3)$ -[3,4,6-tri-*O*benzyl-α-D-mannopyranosyl-(1->2)-3,4,6-tri-O-benzyl-α-D-mannopyranosyl- $(1\rightarrow 6)$]-2,4-di-O-benzyl- α -D-mannopyranosyl- $(1\rightarrow 6)$]-2,4di-O-benzyl-B-D-mannopyranoside (19): Nonasaccharide 5 (14 mg, 0.0035 mmol) was dissolved in a mixture of CH₂Cl₂/MeOH (3 mL, 1:2) and cooled to 0 °C. A solution of sodium methoxide in MeOH (35 µL, 25% w/v) was added and the reaction was slowly warmed to room temperature over 1 h, quenched with DOWEX-50 W-hydrogen strongly acidic ion-exchange resin, filtered, and dried in vacuo. The resulting residue was purified by flash column chromatography on silica gel (5 \rightarrow 30% EtOAc/toluene) affording 12 mg (90%) of nonasaccharide triol 19. ¹H NMR (CDCl₃): δ = 1.37-1.43 (m, 2 H), 1.85-1.88 (m, 2 H), 2.23 (br. s, 1 H), 2.29 (br. s, 2 H), 3.07-3.27 (m, 3 H), 3.30-4.27 (m, 56 H), 4.28-4.66 (m, 41 H), 4.70-4.77 (m, 9 H), 4.79 (d, J = 10.5 Hz, 1 H), 4.94 (app. s, 1 H), 5.02 (app. s, 1 H), 5.08-5.09 (m, 3 H), 5.13 (m, 2 H), 5.51-5.60 (m, 1 H), 6.92-7.44 (m, 125 H).

n-Pentyl *a*-D-Mannopyranosyl-(1 \rightarrow 3)-[*a*-D-mannopyranosyl-(1 \rightarrow 6)]*β*-D-mannopyranoside (4): Activated palladium on carbon (100 mg, 10%) was suspended in ethanol (10 mL) and exposed to an atmosphere of hydrogen gas (balloon). After 30 min, trisaccharide triol 17 (70 mg, 0.058 mmol) in EtOAc (5 mL) was added by cannula and stirred for 48 h under an atmosphere of hydrogen. The product was filtered through celite, dried in vacuo, to afford 6 mg (79%) trisaccharide 4. [α]_D²⁴ = +35.0 (c = 0.4, H₂O/EtOH 1:1). ¹H NMR (D₂O): δ = 0.70–0.73 (m, 3 H), 1.15–1.17 (m, 4 H), 1.43–1.46 (m, 2 H), 3.37–3.40 (m, 1 H), 3.47–3.83 (m, 19 H), 3.90–3.91 (m, 1 H), 3.98 (d, J = 1.6 Hz, 1 H), 4.52 (app. s, 1 H), 4.75 (app. s, 1 H), 4.94 (app. s, 1 H). ¹³C NMR (D₂O): δ = 13.6, 22.1, 27.2, 28.7, 61.2, 65.8, 66.1, 67.0, 67.1, 70.2, 70.3, 70.4, 70.6, 70.7, 72.9, 73.6, 74.4, 81.1, 99.7, 100.0, 102.7. MALDI-TOF *m*/*z* (M + Na⁺) calcd. 595.22, found 596.77.

n-Pentyl α-D-Mannopyranosyl-(1 \rightarrow 2)-α-D-mannopyranosyl-(1 \rightarrow 3)-[α-D-mannopyranosyl-(1 \rightarrow 3)-[α-D-mannopyranosyl-(1 \rightarrow 6)]-α-Dmannopyranosyl-(1 \rightarrow 6)]-β-D-mannopyranoside (3): Activated palladium on carbon (50 mg, 10%) was suspended in ethanol (5 mL) and exposed to an atmosphere of hydrogen gas (balloon). After 30 min, hexasaccharide triol **18** (35 mg, 0.014 mmol) in EtOAcc (2 mL) was added by cannula and stirred for 48 h under an atmosphere of hydrogen. The product was filtered through celite, dried in vacuo, to afford 12 mg (81%) hexasaccharide **3**. [α]_D²⁴ = +54.5 (c = 0.5, H₂O/EtOH 1:1). ¹H NMR (D₂O): $\delta = 0.70-0.74$ (m, 3 H), 1.15-1.17 (m, 4 H), 1.43-1.46 (m, 2 H), 3.38-3.43 (m, 1 H), 3.46-3.89 (m, 33 H), 3.89-3.92 (m, 2 H), 3.96-3.97 (m, 2 H), 4.00 (app. s, 1 H), 4.51 (app. s, 1 H), 4.72 (app. s, 1 H), 4.75 (app. s, 1 H), 4.89 (app. s, 1 H), 4.98 (app. S, 1 H), 5.19 (app. s, 1 H), MALDI-TOF m/z (M + Na⁺) calcd. 1081.38, found 1082.54.

n-Pentyl α -D-Mannopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl- $(1\rightarrow 3)$ - $[\alpha$ -D-mannopyranosyl- $(1\rightarrow 2)$ - α -D-mannopyranosyl- $(1 \rightarrow 3)$ - $[\alpha$ -D-mannopyranosyl- $(1 \rightarrow 2)$ - α -D-mannopyranosyl- $(1\rightarrow 6)$]- α -D-mannopyranosyl- $(1\rightarrow 6)$]- β -D-mannopyranoside (2): Activated palladium on carbon (50 mg, 10%) was suspended in ethanol (5 mL) and exposed to an atmosphere of hydrogen gas (balloon). After 30 min, nonasaccharide triol 19 (12 g, 0.0031 mmol) in EtOAc (2 mL) was added by cannula and stirred for 48 h under an atmosphere of hydrogen. The product was separated by filtration through celite and dried in vacuo to yield 4 mg (88%) of fully deprotected nonasaccharide 2. $[\alpha]_{D}^{24} = +38.0$ (c = 0.05, H₂O/EtOH 1:1). ¹H NMR (D₂O): $\delta = 0.71 - 0.74$ (m, 3 H), 1.16-1.18 (m, 5 H), 1.44-1.46 (m, 2 H), 3.38-3.40 (m, 1 H), 3.46-3.52 (m, 5 H), 3.52-3.57 (m, 5 H), 3.58-3.66 (m, 12 H), 3.68-3.72 (m, 6 H), 3.73-3.76 (m, 6 H), 3.79-3.82 (m, 3 H), 3.84-3.87 (m, 4 H), 3.91-3.99 (m, 7 H), 4.00 (app. s, 1 H), 4.51 (app. s, 1 H), 4.72 (app. s, 1 H), 4.89 (app. s, 3 H), 5.00 (app. s, 1 H), 5.16 (app. s, 1 H), 5.19 (app. s, 1 H), 5.26 (app. s, 1 H). ¹³C NMR (CDCl₃): $\delta = 13.4, 21.9, 23.3, 27.5, 28.4, 61.1, 61.1, 61.2,$ 61.2, 65.6, 65.6, 65.7, 65.7, 65.7, 65.9, 66.9, 67.1, 69.7, 70.1, 70.3, 70.3, 70.3, 70.3, 70.3, 70.5, 71.2, 72.8, 73.3, 73.3, 74.2, 78.6, 78.8, 78.8, 79.1, 98.2, 99.7, 99.9, 100.8, 100.8, 100.9, 102.4, 102.4, 102.4; HSQC anomeric cross-peaks (CDCl₃) δ (4.51 \times 99.9), (4.72 \times 99.8), three (4.89 \times 102.4), (5.00 \times 98.1), (5.16 \times 100.8), (5.19 \times 100.8), (5.26 \times 100.9). MALDI-TOF m/z (M + Na⁺) calcd. 1567.54, found 1569.09.

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