

Metal Complexes of Alkyne-Bridged α -Amino Acids[☆]Bernd Kayser, Janina Altman, Heinrich Nöth^[2], Jörg Knizek^[2], and Wolfgang Beck*Institut für Anorganische Chemie der Ludwig-Maximilians-Universität,
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The palladium-mediated coupling of *p*-ethynylphenylalanine (*p*-epa) with different halogenated benzenes yielded alkyne-bridged α -amino acids. A series of cationic mono- to hexanuclear $(\text{Ph}_3\text{P})_2\text{Pt}$ complexes with the anions of *p*-ethynylphenylalanine and alkynyl- or benzene-bridged di-, tri-, tetra- and hexa-ethynyl phenylalanines as N,O-chelate ligands was prepared. *N*-*t*-Boc-*p*-ethynylphenylalanine methyl ester was metal-substituted to give

complexes of the types $\text{Ph}_3\text{PAu-C}\equiv\text{C-R}$ and $(\text{Et}_3\text{P})_2\text{Pt-(C}\equiv\text{CR)}_2$. The benzene-bridged di-, tri-, tetra- and hexa-*p*-ethynylphenylalanine methyl esters form Schiff bases with ferrocene aldehyde and a tripodal ligand was obtained from $\text{Ph}_2\text{PCH}_2\text{CH}_2\text{CH}_2\text{NH}_2$ and the benzene-bridged tri-ethynylphenylalanine. The structure of $(\text{Ph}_3\text{P})_2\text{Pt}[\text{NH}_2\text{C(H)-(CH}_2\text{C}_6\text{H}_4\text{C}\equiv\text{CH)CO}_2]^+ \text{BF}_4^-$ was determined by X-ray diffraction.

Non-proteinogenic amino acids influence the biological properties of peptides and are useful for the synthesis of peptidomimetics^[3]. The synthesis of unnatural amino acids is based either on electrophilic^[4] or nucleophilic^[5] equivalents. A valuable building block for the derivatization of phenylalanines has been developed by Schwabacher et al.^[6]. *p*-Iodophenylalanine was used as starting material to build, under palladium catalysis, amino acids which are bridged by a heteroatom^[6] or the iodo substituent was replaced by sulfur-containing fragments or trimethyltin^[7]. The palladium-mediated coupling of terminal acetylides with *p*-iodophenylalanine afforded fluorescent marker molecules^[8a], which find growing attention in organometallic chemistry^[8b-e]. Undheim et al. and Savi et al. reported the Pd-catalyzed synthesis of a series of α,α' -hydrocarbon bis(α -amino acids), including α,α' -alkynyl-bridged bis(glycines) and of alkyne-bridged oxazolidines^[9].

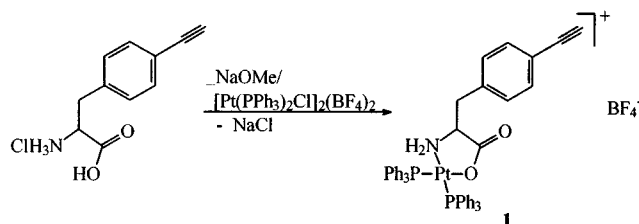
By means of the Heck^[10], especially the Sonogashira reaction^[11], we were able to synthesize various acetylide-bridged phenylalanines^[12a]. *p*-Ethyynylphenylalanine was proven to be a potent inhibitor of the enzyme tryptophan-hydroxylase^[13]; therefore structural details of this compound appear of interest. The bis-, tris-, tetra- and hexapodal^[12b] amino acids should be useful starting compounds in peptide chemistry, for the synthesis of organic as well as metallo-dendrimers^[14], and for linearly-bridged metal complexes with unusual optical or electrical properties^[15]. The scope of organometallic complexes of α -amino acids and peptides has been recently reviewed^[16].

In this paper we wish to present the coordination chemistry of the new amino acid derivatives.

Results and Discussion

Chloro-bridged metal complexes have proven to be ideal starting materials for the synthesis of N,O- α -aminocarboxylate metal compounds^[16]. We have chosen the chloro-bridged complexes $[(\text{Ph}_3\text{P})_2\text{Pt}(\mu\text{-Cl})_2\text{Pt}(\text{PPh}_3)_2]^{2+}$, $(\text{Et}_3\text{P})(\text{Cl})\text{Pd}(\mu\text{-Cl})\text{Pd}(\text{Cl})(\text{PEt}_3)$ and $\text{Cp}^*(\text{Cl})\text{Rh}(\mu\text{-Cl})_2\text{Rh}(\text{Cl})\text{Cp}^*$ from which N,O-chelates with natural α -amino acids have been obtained in our group.^{[16][17][18][19]}

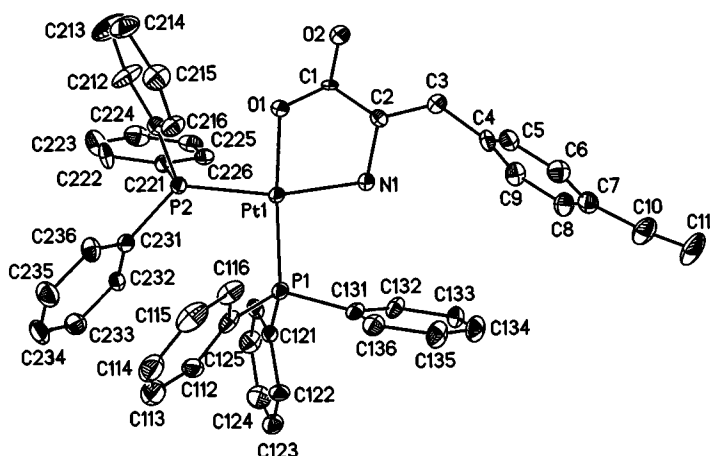
The reaction of the chloro-bridged cationic complex $\{[(\text{Ph}_3\text{P})_2\text{Pt}(\mu\text{-Cl})_2]\}\{\text{BF}_4\}_2$ with the anion of *p*-ethynylphenylalanine yielded the cationic N,O-chelate **1**.



Suitable crystals for structure determination by X-ray diffraction of **1** were obtained by liquid-liquid diffusion of diethyl ether into a methanolic solution of **1** at room temperature. By this means the structure of the new non-proteinogenic amino acid *p*-ethynylphenylalanine (*p*-epa)^[12a] has been established. The Pt-ligand atom distances are close to those of similar complexes, e.g. $(n\text{Bu}_3\text{P})(\text{Cl})\text{Pt}(\text{O}_2\text{CCH}_2\text{N}=\text{C(H)NMe}_2)^{[20a]}$, $(\text{dppe})\text{Pt}[\text{N}(\text{COMe})\text{CH}_2\text{CO}_2]^{[20b]}$ and $(\text{Ph}_3\text{P}(\text{Cl})\text{Pt}(\text{NH}_2\text{CMe}_2\text{CO}_2)^{[20c]}$. The bond angle C11–C10–C7 is not exactly linear.

The analogous reactions of bis-, tris-, tetra- and hexa-ethynylphenylalanine derivatives gave the cationic complexes **2–7**.

[◇] For part CVII see ref.^[1].

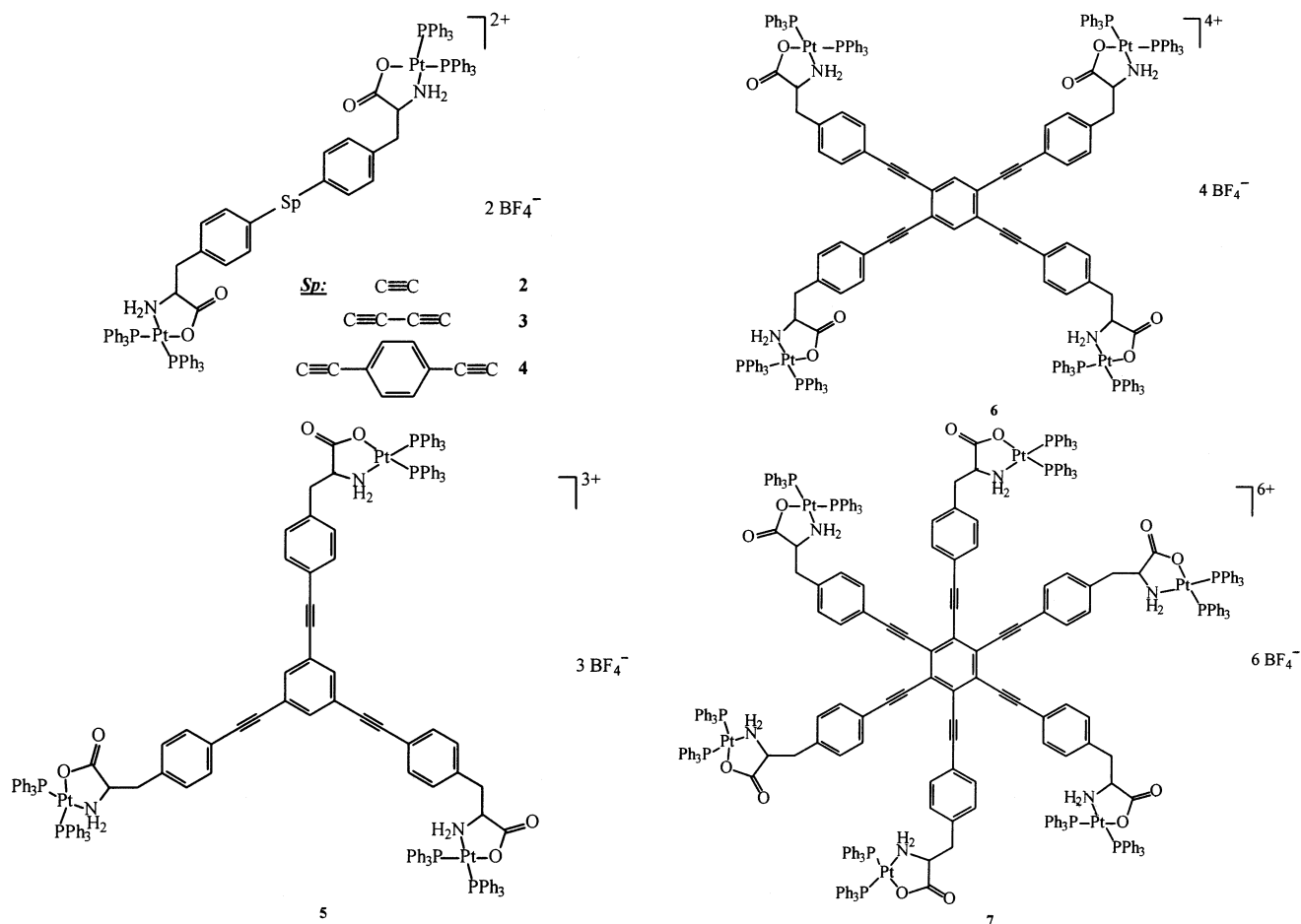
Figure 1. Structure of complex **1**^[a]

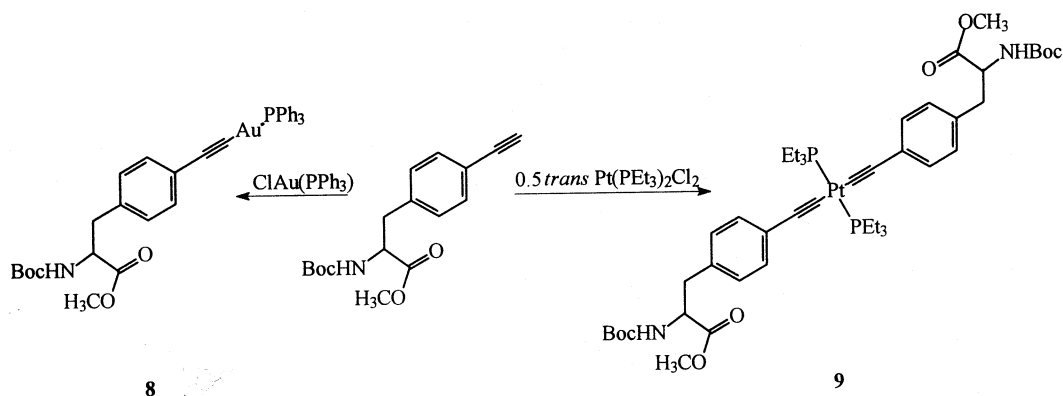
^[a] Selected bond lengths [pm] and angles [°]: Pt–P1 225.3 (2), Pt–P2 225.8 (2), Pt–N1 209.5 (8), Pt–O1 207.1 (7), C10–C11 117.2 (17); N1–Pt–P2 165.9 (2), N1–Pt–P1 95.0 (2), O1–Pt–N1 81.0 (3), O1–Pt–P2 85.18 (19), C7–C10–C11 173.1 (16).

The IR spectra of the compounds **2–7** exhibit characteristic absorptions for η^2 -coordinated α -amino carboxylates at $\nu \approx 1675 \text{ cm}^{-1}$ next to the $\nu(\text{C}\equiv\text{C})$ vibrations at $\nu \approx 2205 \text{ cm}^{-1}$. The high symmetry of **2–7** is manifested in the ^1H - and ^{13}C -NMR spectra. Only one set of signals is observed. The ^{31}P -NMR spectra show a set of double dou-

plets due to the different ligands *trans* to the P atoms with corresponding platinum satellites (^{195}Pt - ^{31}P coupling).

Terminal acetylide functions in organic molecules have successfully been used to build metal acetylide complexes which are also of interest for nonlinear optics. On the other hand the metal atom of bis(acetylide) complexes can func-



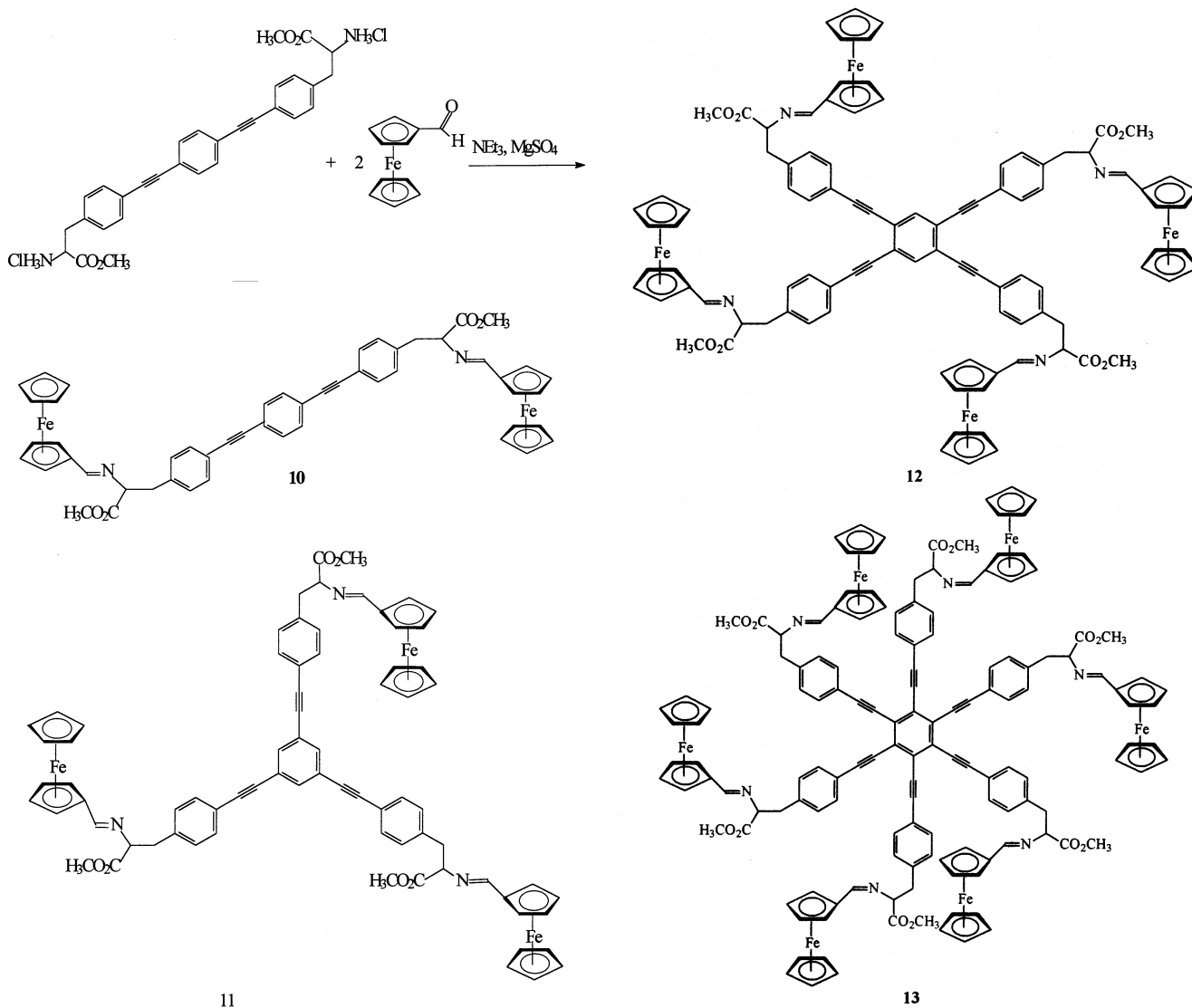


tion as a linker and support the conjugation of biological molecules^[21]. Following known procedures^[22] *p*-epa was treated with Ph_3PAuCl and *trans*- $(\text{Ph}_3\text{P})_2\text{PtCl}_2$ to yield the compounds **8** and **9**.

Complexes **8** and **9** were characterized by IR-, ^1H -, ^{13}C - and ^{31}P -NMR spectroscopy. For **9** the characteristic

$\nu(\text{C}\equiv\text{C})$ absorption is found at 2103 cm^{-1} . The other spectroscopic data could be compared to similar known compounds^[22].

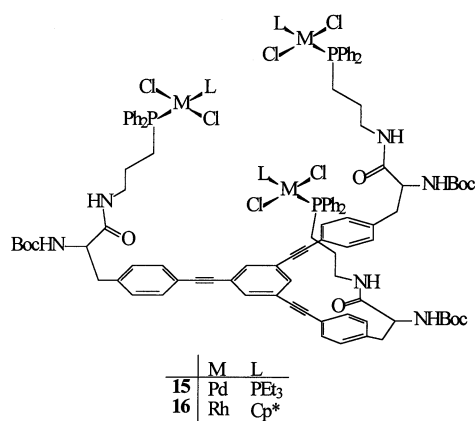
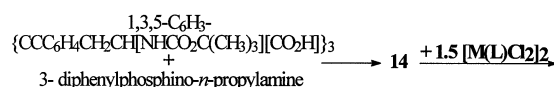
Ferrocenyl Schiff base complexes of amino acids find application as lipophilic and chromophoric groups for masking of peptide bonds^[23a]. Also interactions of the redoxac-



tive ferrocenyl unit and redoxactive enzymes are objects of investigations^[23b]. The condensation between ferrocenecarbaldehyde and the corresponding phenylalanine esters afforded the orange Schiff base complexes **10–13**. For recent examples of star-shaped metal complexes derived from 1,3,5-triethynylbenzene as in **5** and **11** see ref. ^[24].

The successful formation of the Schiff base is proven by the intense $\nu(\text{C}=\text{N})$ IR absorption band at 1636 cm^{-1} and by the imine proton signal ($\delta \approx 7.90$) in the ^1H -NMR spectrum. The ^1H -NMR signal of the H atom at $\alpha\text{-C}$ was not observed. A correlated $^1\text{H}/^1\text{H}$ -NMR experiment showed that the α -proton has the same chemical shift as the OCH_3 group of the methyl ester.

Using common methods of peptide chemistry the tripodal tris amino acid was derivatized and treated with 3-diphenylphosphanyl-*n*-propylamine to give the tripodal phosphane ligand **14** which may function as a tridentate chelate ligand. The reaction of **14** with 0.5 equivalents of $[\text{Rh}(\text{COD})\text{Cl}]_2$ (COD = 1,5-cyclooctadiene) gave no defined compound, probably because the phosphane groups are not pre-organized. On the other hand the reactions of **14** with 1.5 equivalents of $[(\text{Et}_3\text{P})\text{PdCl}(\mu\text{-Cl})_2]$ or $[\text{Cp}^*\text{RhCl}(\mu\text{-Cl})_2]$ (Cp^* = pentamethylcyclopentadienyl) yielded the three core complexes **15** and **16**.



The ^{31}P -NMR spectrum of **15** shows several signal sets due to *cis* and *trans* isomers with the most intensive one having a coupling constant of 538 Hz. This coupling constant is typical for *trans*-situated phosphanes^[25]. In contrast, compound **16** is formed as one isomer with a doublet in the ^{31}P -NMR spectrum due to the $^1J_{\text{Rh-P}}$ coupling of 143 Hz.

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Experimental Section

General: All reactions were carried out in dry solvents under argon. — NMR: Jeol GSX 270 (^1H 270.0; ^{13}C 67.9; ^{31}P 109.3 MHz). Tetramethylsilane as internal standard; H_3PO_4 (85%) as external standard. — IR: 5ZDX FT-IR. — $\text{ClAu}(\text{PPh}_3)$ ^[26], *trans*- $\text{Pt}(\text{PEt}_3)_2\text{Cl}_2$ ^[27a], $\{[(\text{Ph}_3\text{P})_2\text{Pt}(\mu\text{-Cl})_2]\{\text{BF}_4\}_2\}$ ^[28], $[(\text{Et}_3\text{P})\text{PdCl}(\mu\text{-Cl})_2]$ ^[27b], $[\text{Cp}^*\text{RhCl}(\mu\text{-Cl})_2]$ ^[29], 3-diphenylphosphanyl-*n*-propylamine^[30] and the acetylide-substituted phenylalanines were prepared according to literature procedures^[12]. Formylferrocene was purchased from Fluka and stored in the dark under argon.

$\{(\text{Ph}_3\text{P})_2\text{Pt}[(\text{NH}_2)(\text{O}_2\text{C})\text{CHCH}_2\text{C}_6\text{H}_4\text{C}\equiv\text{CH}]/\text{BF}_4\}$ (**1**): To a solution of 34 mg (0.15 mmol) of *p*-ethynylphenylalanine hydrochloride in 10 ml of methanol was added 0.30 mmol of NaOMe and the mixture was stirred for 5 min. at room temperature. After addition of a solution of 120 mg (0.075 mmol) of $\{[(\text{Ph}_3\text{P})_2\text{Pt}(\mu\text{-Cl})_2]\{\text{BF}_4\}_2$ in 10 ml of dichloromethane, the mixture was stirred for additional 3 h. The volatiles were evaporated, the residue was washed two times with 5 ml of water and dried at 50°C in vacuo. The resulting powder was finally recrystallized several times from dichloromethane/diethyl ether. Yield 121 mg (81%), colorless powder, m.p. 250°C (decomp.). — IR (KBr): $\tilde{\nu} = 3295\text{ cm}^{-1}$ (m, NH_2), 1654 s (CO_2), 1084 s (BF_4). — ^1H NMR (CDCl_3): $\delta = 2.85$ (dd, $^2J = 14.6\text{ Hz}$, $^3J = 3.4\text{ Hz}$, 1 H, $\text{CH}'\text{H}$), 3.13 (dd, $^2J = 14.6\text{ Hz}$, $^3J = 7.5\text{ Hz}$, 1 H, CHH'), 3.16 (s, 1 H, CH), $3.19\text{--}3.30$ (m, 1 H, CH), $3.98\text{--}4.12$ (m, 2 H, NH_2), 6.95 (d, $^3J = 8.1\text{ Hz}$, 2 H, C_6H_4), $7.15\text{--}7.50$ (m, 32 H, C_6H_4 , PPh_3). — ^{13}C NMR (CDCl_3): $\delta = 38.90$ (CH_2), 56.58 (CH), 77.76 ($\text{C}\equiv\text{CH}$), 83.41 ($\text{C}\equiv\text{CH}$), 120.85 , 125.33 (d, $^1J = 84.1\text{ Hz}$, *i*- PPh_3), 126.38 (d, $^1J = 83.5\text{ Hz}$, *i*- PPh_3), 128.61 (d, $^3J = 11.4\text{ Hz}$, *m*- PPh_3), 129.17 (d, $^3J = 11.5\text{ Hz}$, *m*- PPh_3), 129.60 , 131.66 (d, $^4J = 2.3\text{ Hz}$, *p*- PPh_3), 132.21 (d, $^4J = 2.2\text{ Hz}$, *p*- PPh_3), 132.66 , 133.91 (d, $^2J = 10.8\text{ Hz}$, *o*- PPh_3), 134.59 (d, $^2J = 10.6\text{ Hz}$, *o*- PPh_3), 136.91 , 182.94 (CO_2). — ^{31}P NMR (CDCl_3): $\delta = 7.70$ (d, $^2J = 24.3\text{ Hz}$, $^1J = 3586\text{ Hz}$), 8.54 (d, $^2J = 24.3\text{ Hz}$, $^1J = 3484\text{ Hz}$). — $\text{C}_{47}\text{H}_{40}\text{BF}_4\text{NO}_2\text{P}_2\text{Pt}$ (994.7): calcd. C 56.75, H 4.05, N 1.41; found C 56.80, H 4.05, N 1.15.

$\{(\text{Ph}_3\text{P})_2\text{Pt}[(\text{NH}_2)(\text{CO}_2)\text{CHCH}_2\text{C}_6\text{H}_4\text{C}\equiv\text{CC}_6\text{H}_4\text{CH}_2\text{CH}(\text{CO}_2)(\text{NH}_2)]\text{Pt}(\text{PPh}_3)_2/\text{BF}_4\}_2$ (**2**): The reaction was carried out as described for **1** with 43 mg (0.10 mmol) of the ligand of **2** as dihydrochloride, 0.40 mmol of NaOMe, 160 mg (0.10 mmol) of $\{[(\text{Ph}_3\text{P})_2\text{Pt}(\mu\text{-Cl})_2]\{\text{BF}_4\}_2$ for 16 h. Yield 178 mg (91%), colorless powder, m.p. 220°C (decomp.). — IR (KBr): $\tilde{\nu} = 3303\text{ cm}^{-1}$ (m, NH_2), 1676 s (CO_2), 1084 s (BF_4). — ^1H NMR ($\text{CDCl}_3 + \text{CD}_3\text{OD}$): $\delta = 2.99$ (dd, $^2J = 14.9\text{ Hz}$, $^3J = 4.9\text{ Hz}$, 2 H, $\text{CH}'\text{H}$), 3.11 (dd, $^2J = 14.9\text{ Hz}$, $^3J = 7.6\text{ Hz}$, 2 H, CHH'), $3.97\text{--}4.08$ (m, 2 H, CH), 6.99 (d, $^3J = 8.3\text{ Hz}$, 4 H, C_6H_4), $7.10\text{--}7.47$ (m, 64 H, C_6H_4 , PPh_3). — ^{13}C NMR ($\text{CDCl}_3 + \text{CD}_3\text{OD}$): $\delta = 38.69$ (CH_2), 56.48 (CH), 89.33 ($\text{C}\equiv\text{C}$), 121.75 , 124.94 (d, $^1J = 77.0\text{ Hz}$, *i*- PPh_3), 125.85 (d, $^1J = 74.9\text{ Hz}$, *i*- PPh_3), 128.40 (d, $^3J = 11.4\text{ Hz}$, *m*- PPh_3), 128.97 (d, $^3J = 11.3\text{ Hz}$, *m*- PPh_3), 129.26 , 131.64 , 132.01 (*p*- PPh_3), 132.17 (*p*- PPh_3), 133.53 (d, $^2J = 10.3\text{ Hz}$, *o*- PPh_3), 134.21 (d, $^2J = 10.5\text{ Hz}$, *o*- PPh_3), 135.94 , 183.71 (CO_2). — ^{31}P NMR ($\text{CDCl}_3 + \text{CD}_3\text{OD}$): $\delta = 7.54$ (d, $^2J = 24.1\text{ Hz}$, $^1J = 3610\text{ Hz}$), 8.29 (d, $^2J = 24.1\text{ Hz}$, $^1J = 3466\text{ Hz}$). — $\text{C}_{92}\text{H}_{78}\text{B}_2\text{F}_8\text{N}_2\text{O}_4\text{P}_4\text{Pt}_2$ (1963.3): calcd. C 56.28, H 4.01, N 1.43; found C 55.72, H 4.08, N 1.34.

$\{(\text{Ph}_3\text{P})_2\text{Pt}[(\text{NH}_2)(\text{CO}_2)\text{CHCH}_2\text{C}_6\text{H}_4\text{C}\equiv\text{CC}_6\text{H}_4\text{CH}_2\text{CH}(\text{CO}_2)(\text{NH}_2)]\text{Pt}(\text{PPh}_3)_2/\text{BF}_4\}_2$ (**3**): The reaction was carried out as described for **1** with 45 mg (0.10 mmol) of the ligand of **3** as dihydrochloride, 0.40 mmol of NaOMe, 160 mg (0.10 mmol) of $\{[(\text{Ph}_3\text{P})_2\text{Pt}(\mu\text{-Cl})_2]\{\text{BF}_4\}_2$ for 16 h. Yield 184 mg (93%), colorless powder, m.p. 235°C (decomp.). — IR (KBr): $\tilde{\nu} = 3303$, 3223 cm^{-1} (m, NH_2), 1679 s (CO_2), 1084 s (BF_4). — ^1H NMR

(CDCl₃+CD₃OD): δ = 2.90–3.04 (m, 4 H, CH₂), 3.95–4.03 (m, 2 H, CH), 6.97 (d, 3J = 7.8 Hz, 4 H, C₆H₄), 7.16–7.46 (m, 64 H, C₆H₄, PPh₃). – ¹³C NMR (CDCl₃+CD₃OD): δ = 38.86 (CH₂), 56.66 (CH), 74.22, 81.34 (C≡C), 120.28, 125.05 (d, 1J = 79.8 Hz, *i*-PPh₃), 126.03 (d, 1J = 78.1 Hz, *i*-PPh₃), 128.53 (d, 3J = 11.4 Hz, *m*-PPh₃), 129.09 (d, 3J = 11.2 Hz, *m*-PPh₃), 129.44, 131.74 (*p*-PPh₃), 132.25 (*p*-PPh₃), 132.96, 133.65 (d, 2J = 11.1 Hz, *o*-PPh₃), 134.38 (d, 2J = 10.3 Hz, *o*-PPh₃), 137.34, 183.43 (d, 3J = 5.2 Hz, CO₂). – ³¹P NMR (CDCl₃+CD₃OD): δ = 7.43 (d, 2J = 24.4 Hz, 1J = 3506 Hz), 8.47 (d, 2J = 24.4 Hz, 1J = 3445 Hz). – C₉₄H₇₈B₂F₈N₂O₄P₄Pt₂ × CH₂Cl₂ (2072.3): calcd. C 55.06, H 3.89, N 1.35; found C 54.60, H 3.92, N 1.30.

{1,4-C₆H₄[C≡CC₆H₄CH₂CH(CO₂)(NH₂) Pt(PPh₃)₂]₂}/BF₄ (4): The reaction was carried out as described for **1** with 53 mg (0.10 mmol) of the ligand of **4** as dihydrochloride, 0.40 mmol of NaOMe, 160 mg (0.10 mmol) of [(Ph₃P)₂Pt(μ-Cl)]₂{BF₄}₂ for 16 h. Yield 184 mg (89%), colorless powder, m.p. 260 °C (decomp.). – IR (KBr): $\tilde{\nu}$ = 3299, 3223 cm^{−1} (NH₂), 2213 (C≡C), 1666 s (CO₂), 1084 s (BF₄). – ¹H NMR (CDCl₃): δ = 2.89 (dd, 2J = 13.6 Hz, 3J = 4.3 Hz, 2 H, CH'H), 3.13 (dd, 2J = 13.6 Hz, 3J = 7.2 Hz, 2 H, CHH'), 3.32–3.47 (m, 2 H, CH), 3.98–4.19 (m, 4 H, NH₂), 7.01 (d, 3J = 8.3 Hz, 4 H, C₆H₄), 7.12–7.49 (m, 64 H, C₆H₄, PPh₃), 7.59 (s, 4 H, C₆H₄). – ¹³C NMR (CDCl₃): δ = 38.70 (CH₂), 56.50 (CH), 89.50, 90.75 (C≡C), 121.81, 122.88, 125.01 (d, 1J = 78.9 Hz, *i*-PPh₃), 125.90 (d, 1J = 76.4 Hz, *i*-PPh₃), 128.49 (d, 3J = 11.2 Hz, *m*-PPh₃), 129.05 (d, 3J = 11.1 Hz, *m*-PPh₃), 129.28, 131.44, 131.71, 132.08 (*p*-PPh₃), 132.26 (*p*-PPh₃), 133.57 (d, 2J = 10.6 Hz, *o*-PPh₃), 134.27 (d, 2J = 10.8 Hz, *o*-PPh₃), 135.97, 183.75 (d, 3J = 4.0 Hz, CO₂). – ³¹P NMR (CDCl₃): δ = 7.71 (d, 2J = 24.2 Hz, 1J = 3595 Hz), 8.55 (d, 2J = 24.2 Hz, 1J = 3446 Hz). – C₁₀₀H₈₂B₂F₈N₂O₄P₄Pt₂ × 1/2 CH₂Cl₂ (2105.9): calcd. C 57.48, H 3.98, N 1.33; found C 57.29, H 4.23, N 1.25.

{1,3,5-C₆H₃[C≡CC₆H₄CH₂CH(CO₂)(NH₂) Pt(PPh₃)₂]₃}/BF₄ (5): The reaction was carried out as described for **1** with 75 mg (0.10 mmol) of the ligand of **5** as trihydrochloride, 0.60 mmol of NaOMe, 240 mg (0.15 mmol) of [(Ph₃P)₂Pt(μ-Cl)]₂{BF₄}₂ for 16 h. Yield 256 mg (84%), colorless powder, m.p. 270 °C (decomp.). – IR (KBr): $\tilde{\nu}$ = 3300, 3225 cm^{−1} (NH₂), 2208 (C≡C), 1674 s (CO₂), 1085 s (BF₄). – ¹H NMR (CDCl₃+CD₃OD): δ = 2.89–3.15 (m, 6 H, CH₂), 4.03 (pt, 3J = 6.4 Hz, 3 H, CH), 6.98 (d, 3J = 8.0 Hz, 6 H, C₆H₄), 7.10–7.46 (m, 96 H, C₆H₄, PPh₃), 7.69 (s, 3 H, C₆H₃). – ¹³C NMR (CDCl₃+CD₃OD): δ = 38.73 (CH₂), 56.58 (CH), 87.95, 90.17 (C≡C), 121.27, 123.87, 124.99 (d, 1J = 79.0 Hz, *i*-PPh₃), 125.95 (d, 1J = 77.4 Hz, *i*-PPh₃), 128.45 (d, 3J = 11.3 Hz, *m*-PPh₃), 129.02 (d, 3J = 11.4 Hz, *m*-PPh₃), 131.67 (*p*-PPh₃), 132.11 (*p*-PPh₃), 132.18, 133.89 (d, 2J = 10.6 Hz, *o*-PPh₃), 133.89, 134.25 (d, 2J = 10.6 Hz, *o*-PPh₃), 136.41, 183.56 (CO₂). – ³¹P NMR (CDCl₃+CD₃OD): δ = 7.56 (d, 2J = 24.0 Hz, 1J = 3623 Hz), 8.53 (d, 2J = 24.0 Hz, 1J = 3440 Hz). – C₁₄₇H₁₂₀B₃F₁₂N₃O₆P₆Pt₃ × CH₂Cl₂ (3141.1): calcd. C 56.59, H 3.91, N 1.34; found C 56.49, H 4.00, N 1.17.

{1,2,4,5-C₆H₂[C≡CC₆H₄CH₂CH(CO₂)(NH₂) Pt(PPh₃)₂]₄}/BF₄ (6): The reaction was carried out as described for **1** with 73 mg (0.075 mmol) of the ligand of **6** as tetrahydrochloride, 0.60 mmol of NaOMe, 240 mg (0.15 mmol) of [(Ph₃P)₂Pt(μ-Cl)]₂{BF₄}₂ for 16 h. Yield 261 mg (86%), brown powder, m.p. 276 °C (decomp.). – IR (KBr): $\tilde{\nu}$ = 3299, 3225 cm^{−1} (NH₂), 2198 (C≡C), 1674 s (CO₂), 1084 s (BF₄). – ¹H NMR (CDCl₃+CD₃OD): δ = 2.82 (dd, 2J = 14.1 Hz, 3J = 11.2 Hz, 4 H, CH'H), 3.08–3.20 (m, 4 H, CHH'), 4.03–4.17 (m, 4 H, CH), 6.99 (d, 3J = 7.7 Hz, 8 H, C₆H₄), 7.09–7.45 (m, 128 H, C₆H₄, PPh₃), 7.76 (s, 2 H, C₆H₂). – ¹³C NMR (CDCl₃+CD₃OD): δ =

39.04 (CH₂), 56.68 (CH), 87.74, 95.49 (C≡C), 121.05, 125.05 (d, 1J = 89.7 Hz, *i*-PPh₃), 125.25, 126.03 (d, 1J = 88.9 Hz, *i*-PPh₃), 128.38 (d, 3J = 11.6 Hz, *m*-PPh₃), 128.89 (d, 3J = 10.9 Hz, *m*-PPh₃), 129.23, 131.57 (*p*-PPh₃), 131.95, 132.04 (*p*-PPh₃), 133.56 (d, 2J = 10.6 Hz, *o*-PPh₃), 134.02, 134.33 (d, 2J = 10.5 Hz, *o*-PPh₃), 136.89, 183.46 (CO₂). – ³¹P NMR (CDCl₃+CD₃OD): δ = 7.40 (d, 2J = 23.9 Hz, 1J = 3622 Hz), 8.47 (d, 2J = 23.9 Hz, 1J = 3440 Hz). – C₁₉₄H₁₅₈B₄F₁₆N₄O₈P₈Pt₄ × CH₂Cl₂ (4133.7): calcd. C 56.65, H 3.90, N 1.35; found C 56.10, H 3.99, N 1.15.

{1,2,3,4,5,6-C₆[C≡CC₆H₄CH₂CH(CO₂)(NH₂) Pt(PPh₃)₂]₆}/BF₄ (7): The reaction was carried out as described for **1** with 71 mg (0.05 mmol) of the ligand of **7** as hexahydrochloride, 0.60 mmol of NaOMe, 240 mg (0.15 mmol) of [(Ph₃P)₂Pt(μ-Cl)]₂{BF₄}₂ for 16 h. Yield 244 mg (81%), brown powder, m.p. 300 °C (decomp.). – IR (KBr): $\tilde{\nu}$ = 3304, 3231 cm^{−1} (NH₂), 2198 (C≡C), 1675 s (CO₂), 1084 s (BF₄). – ¹H NMR (CDCl₃+CD₃OD): δ = 2.82 (pt, 3J = 11.9 Hz, 6 H, CH'H), 3.19 (pt, 3J = 10.7 Hz, 6 H, CHH'), 4.29 (pt, 3J = 14.7 Hz, 6 H, CH), 7.09–7.51 (m, 204 H, C₆H₄, PPh₃). – ¹³C NMR (CDCl₃+CD₃OD): δ = 39.18 (CH₂), 56.47 (CH), 87.23, 99.70 (C≡C), 120.92, 125.32 (d, 1J = 110.8 Hz, *i*-PPh₃), 126.29 (d, 1J = 108.1 Hz, *i*-PPh₃), 127.82, 128.56 (d, 3J = 10.7 Hz, *m*-PPh₃), 129.02 (d, 3J = 12.8 Hz, *m*-PPh₃), 129.72, 131.59 (*p*-PPh₃), 132.12 (*p*-PPh₃), 132.18, 133.74 (d, 2J = 10.0 Hz, *o*-PPh₃), 134.61 (d, 2J = 8.9 Hz, *o*-PPh₃), 137.63, 183.14 (CO₂). – ³¹P NMR (CDCl₃+CD₃OD): δ = 7.32 (d, 2J = 23.6 Hz, 1J = 3623 Hz), 8.79 (d, 2J = 23.6 Hz, 1J = 3477 Hz). – C₂₈₈H₂₃₄B₆F₂₄N₆O₁₂P₁₂Pt₆ × 2 CH₂Cl₂ (6203.9): calcd. C 56.14, H 3.86, N 1.35; found C 55.53, H 4.09, N 1.18.

General Procedure for the Preparation of the σ-Acetylide Metal Complexes 8 and 9: To a solution of 99 mg (0.20 mmol) of ClAu(PPh₃) or 100 mg (0.20 mmol) of *trans*-Pt(PtEt₃)₂Cl₂ in 10 ml of diethylamine were added 67 mg (0.22 mmol) or 133 mg (0.44 mmol) of *N*-*t*-Boc-4-ethynyl-L-phenylalanine methyl ester and a catalytic amount of CuI. The solutions were stirred at room temp. for 3 h in the dark and the volatiles were evaporated. The residues were each extracted with a mixture of dichloromethane/diethyl ether (1/3; 5 ml) and the solvent was evaporated. The resulting precipitates were washed with pentane (2 × 10 ml) and dried in vacuo.

(PPh₃)Au{C≡CC₆H₄CH₂CH[NHCO₂C(CH₃)₃](CO₂Me)} (**8**): Yield 131 mg (86%), colorless powder, m.p. 99 °C (decomp.). – IR (KBr): $\tilde{\nu}$ = 3439 cm^{−1} (NH), 1743 m, 1713 s (CO₂, CON). – ¹H NMR (CDCl₃): δ = 1.39 (s, 9 H, CH₃), 2.93–3.09 (m, 2 H, CH₂), 3.65 (s, 3 H, CH₃), 4.53 (pq, 3J = 6.1 Hz, 1 H, CH), 4.92 (d, 3J = 8.4 Hz, 1 H, NH), 6.99 (d, 3J = 8.1 Hz, 2 H, C₆H₄), 7.39–7.56 (m, 17 H, PPh₃, C₆H₄). – ¹³C NMR (CDCl₃): δ = 28.36 [C(CH₃)₃], 38.32 (CH₂), 52.55 (CH₃), 54.39 (CH), 79.94 [C(CH₃)₃], 83.35, 105.09 (C≡C), 123.62, 128.99, 129.21 (d, 3J = 11.4 Hz, *m*-PPh₃), 131.67, 131.80 (d, 1J = 8.9 Hz, *i*-PPh₃), 132.57, 134.35 (d, 2J = 13.7 Hz, *o*-PPh₃), 134.70 (24 C, C₆H₄, PPh₃), 155.14 (CON), 172.40 (CO₂). – ³¹P NMR (CDCl₃): δ = 42.84. – C₃₅H₃₅N₃O₄PAu × 1/2 CH₂Cl₂ (804.1): calcd. C 53.02, H 4.51, N 1.74; found C 52.54, H 4.13, N 1.46.

trans-(PEt₃)₂Pt{C≡CC₆H₄CH₂CH[NHCO₂C(CH₃)₃](CO₂Me)} (**9**): Yield 182 mg (88%), yellow powder, m.p. 135 °C (decomp.). – IR (KBr): $\tilde{\nu}$ = 3443 cm^{−1} (NH), 2102 s (C≡C), 1745 m, 1715 s (CO₂, CON). – ¹H NMR (CDCl₃): δ = 1.46 (dt, 3J = 8.3 Hz, 3J = 7.8 Hz, 18 H, CH₃), 1.35 (s, 18 H, CH₃), 2.10 (tq, 2J = 7.4 Hz, 3J = 7.8 Hz, 12 H, CH₂), 2.88–3.02 (m, 4 H, CH₂), 3.65 (s, 6 H, CH₃), 4.48 (pq, 3J = 7.2 Hz, 2 H, CH), 4.87 (d, 3J = 8.3 Hz, 2 H, NH), 6.89 (d, 3J = 8.0 Hz, 4 H, C₆H₄), 7.13 (d, 3J = 8.0 Hz, 4 H, C₆H₄). – ¹³C NMR (CDCl₃): δ = 8.22 (t, 2J = 12.2

H₂, CH₃), 16.16 (t, ¹J = 17.9 Hz, CH₂), 28.18 [C(CH₃)₃], 37.95 (CH₂), 52.15 (CH₃), 54.23 (CH), 79.87 [C(CH₃)₃], 109.00, 128.99, 131.09, 132.62 (12 C, C₆H₄), 155.23 (CON), 172.55 (CO₂). – ³¹P NMR (CDCl₃): δ = 11.50 (¹J = 2371 Hz). – C₄₆H₇₀N₂O₈P₂Pt (1036.1): calcd. C 53.33, H 6.81, N 2.70; found C 52.71, H 6.67, N 2.69.

1,4-C₆H₄[-C≡CC₆H₄CH₂CH(CO₂CH₃)(N=C-Fc)] (10): To a solution of 112 mg (0.20 mmol) of the ligand of **4** as dihydrochloride dimethyl ester in 20 ml of dichloromethane were added 61.8 μl (0.44 mmol) of triethylamine. The solution was stirred at room temperature for 15 min, 96 mg (0.44 mmol) of ferrocenealdehyde and 150 mg of MgSO₄ were added and stirring was continued for 24 h in the dark. The precipitate was separated by centrifugation, the volatiles were evaporated from the supernatant solution, and the residue was extracted with a mixture of diethyl ether/tetrahydrofuran (3/1; 2 × 15 ml). The solvent was evaporated, the resulting precipitate was washed with pentane (3 × 10 ml) and dried in vacuo. Yield 138 mg (79%), orange powder, m.p. 108°C (decomp.). – IR (KBr): $\tilde{\nu}$ = 2216 cm⁻¹ w (C≡C), 1737 s (CO₂), 1635 s (C=N). – ¹H NMR (CDCl₃): δ = 3.16 (dd, ²J = 14.1 Hz, ³J = 8.8 Hz, 2 H, CH'H), 3.34 (dd, ²J = 14.1 Hz, ³J = 5.1 Hz, 2 H, CHH'), 3.73 (s, 6 H, CH₃), 4.02 (s, 12 H, CH, C₅H₅), 4.36 (s, 4 H, C₅H₅), 4.60 (s, 2 H, C₅H₅), 4.65 (s, 2 H, C₅H₅), 7.19 (d, ³J = 8.1 Hz, 4 H, C₆H₄), 7.42 (d, ³J = 8.1 Hz, 4 H, C₆H₄), 7.47 (s, 4 H, C₆H₄), 7.93 (s, 2 H, CH). – ¹³C NMR (CDCl₃): δ = 39.30 (CH₂), 52.08 (CH₃), 68.70, 68.92, 69.09, 70.79 (C₅H₅), 74.72 (CH), 79.34 (C₅H₅), 88.99, 91.04 (C≡C), 121.21, 123.06, 129.66, 131.45, 137.87, 138.55 (18 C, C₆H₄), 164.71 (C=N), 171.81 (CO₂). – C₅₂H₄₄N₂O₄Fe₂ (872.6): calcd. C 71.57, H 5.08, N 3.21; found C 71.57, H 5.22, N 3.84.

1,3,5-C₆H₃[-C≡C-C₆H₄-CH₂-CH(CO₂CH₃)(N=C-Fc)] (11): The reaction was carried out as described for **10** with 70 mg (0.10 mmol) of the ligand of **5** as trihydrochloride trimethyl ester, 45.7 μl (0.33 mmol) of triethylamine and 70 mg (0.33 mmol) of ferrocenealdehyde. Yield 107 mg (84%), orange powder, m.p. 123°C (decomp.). – IR (KBr): $\tilde{\nu}$ = 2211 cm⁻¹ w (C≡C), 1740 s (CO₂), 1636 s (C=N). – ¹H NMR (CDCl₃): δ = 3.15 (dd, ²J = 14.0 Hz, ³J = 9.0 Hz, 3 H, CH'H), 3.34 (dd, ²J = 14.0 Hz, ³J = 4.6 Hz, 3 H, CHH'), 3.73 (s, 9 H, CH₃), 4.05 (s, 18 H, CH, C₅H₅), 4.36 (s, 6 H, C₅H₅), 4.58 (s, 3 H, C₅H₅), 4.63 (s, 3 H, C₅H₅), 7.20 (d, ³J = 8.1 Hz, 6 H, C₆H₄), 7.41 (d, ³J = 8.1 Hz, 6 H, C₆H₄), 7.52 (s, 3 H, C₆H₃), 7.93 (s, 3 H, CH). – ¹³C NMR (CDCl₃): δ = 39.30 (CH₂), 52.30 (CH₃), 68.68, 68.92, 69.10, 70.74, 70.79 (C₅H₅), 74.76 (CH), 79.39 (C₅H₅), 87.70, 90.32 (C≡C), 120.96, 123.93, 129.65, 131.66, 133.83, 138.69 (24 C, C₆H₄, C₆H₃), 164.69 (C=N), 171.82 (CO₂). – C₇₅H₆₃N₃O₆Fe₃ × 1/2 CH₂Cl₂ (1312.3): calcd. C 69.09, H 4.92, N 3.20; found C 68.98, H 4.60, N 3.23.

1,2,4,5-C₆H₂[-C≡C-C₆H₄-CH₂-CH(CO₂CH₃)(N=C-Fc)] (12): The reaction was carried out as described for **10** with 51 mg (0.05 mmol) of the ligand of **6** as tetrahydrochloride tetramethyl ester, 30.5 μl (0.22 mmol) of triethylamine and 47 mg (0.22 mmol) of ferrocenealdehyde. Yield 67 mg (80%), orange powder, m.p. 132°C (decomp.). – IR (KBr): $\tilde{\nu}$ = 2204 cm⁻¹ w (C≡C), 1739 s (CO₂), 1636 s (C=N). – ¹H NMR (CDCl₃): δ = 3.06–3.22 (m, 4 H, CH'H), 3.27–3.38 (m, 4 H, CHH'), 3.72 (s, 12 H, CH₃), 4.04 (s, 24 H, CH, C₅H₅), 4.34 (s, 8 H, C₅H₅), 4.57 (s, 4 H, C₅H₅), 4.64 (s, 4 H, C₅H₅), 7.11–7.23 (m, 8 H, C₆H₄), 7.39–7.48 (m, 8 H, C₆H₄), 7.62 (s, 2 H, C₆H₂), 7.92 (s, 4 H, CH). – ¹³C NMR (CDCl₃): δ = 39.41 (CH₂), 52.42 (CH₃), 68.81, 68.91, 69.15, 70.85, 70.88 (C₅H₅), 74.65 (CH), 79.32 (C₅H₅), 87.54, 95.32 (C≡C), 121.20, 125.13, 129.80, 131.77, 134.90, 138.91 (30 C, C₆H₄, C₆H₂), 164.83 (C=N), 171.87 (CO₂). – C₉₈H₈₂N₄O₈Fe₄ × CH₂Cl₂ (1752.1): calcd. C 67.86, H 4.83, N 3.20; found C 67.00, H 4.73, N 3.30.

1,2,3,4,5,6-C₆[-C≡C-C₆H₄-CH₂-CH(CO₂CH₃)(N=C-Fc)] (13): The reaction was carried out as described for **10** with 75 mg (0.05 mmol) of the ligand of **7** as hexahydrochloride hexamethyl ester, 45.7 μl (0.33 mmol) of triethylamine and 70 mg (0.33 mmol) of ferrocenealdehyde. Yield 96 mg (78%), orange powder, m.p. 131°C (decomp.). – IR (KBr): $\tilde{\nu}$ = 2206 cm⁻¹ w (C≡C), 1739 s (CO₂), 1636 s (C=N). – ¹H NMR (CDCl₃): δ = 3.15 (dd, ²J = 13.6 Hz, ³J = 9.2 Hz, 6 H, CH'H), 3.36 (dd, ²J = 13.6 Hz, ³J = 5.2 Hz, 6 H, CHH'), 3.75 (s, 18 H, CH₃), 4.06 (s, 36 H, CH, C₅H₅), 4.37 (s, 12 H, C₅H₅), 4.58 (s, 6 H, C₅H₅), 4.68 (s, 6 H, C₅H₅), 7.16 (d, ³J = 8.2 Hz, 12 H, C₆H₄), 7.44 (d, ³J = 8.2 Hz, 12 H, C₆H₄), 7.95 (s, 6 H, CH). – ¹³C NMR (CDCl₃): δ = 39.34 (CH₂), 52.33 (CH₃), 68.75, 68.96, 69.15, 70.84 (C₅H₅), 74.65 (CH), 79.30 (C₅H₅), 87.23, 99.13 (C≡C), 121.25, 127.22, 129.74, 131.73, 139.05 (42 C, C₆H₄, C₆), 164.73 (C=N), 171.78 (CO₂). – C₁₄₄H₁₂₀N₆O₁₂Fe₆ × 2 CH₂Cl₂ (2631.3): calcd. C 66.63, H 4.75, N 3.19; found C 66.23, H 4.89, N 3.14.

1,3,5-C₆H₃{C≡CC₆H₄CH₂CH[NHCO₂C(CH₃)₃][CO₂-(CH₂)₃PPh₂]}₃ (14): To a solution of 94 mg (0.10 mmol) of the ligand of **5** as *N*-Boc-protected triacid in 5 ml of tetrahydrofuran was added a solution of 98 mg (0.40 mmol) of 3-diphenylphosphanyl-*n*-propylamine in 5 ml of tetrahydrofuran and 38.0 μl (0.30 mmol) of *N*-ethylmorpholine. The solution was cooled to 0°C and 81 mg (0.60 mmol) of 1-hydroxybenzotriazole and 96 mg (0.50 mmol) of 1-[3-(dimethylamino)propyl]-3-ethylcarbodiimide hydrochloride were added. After stirring for 24 h, evaporation of the solvent gave a colorless residue from which the product was isolated by flash chromatography (silica gel, 30 cm, CH₂Cl₂/methanol 30:1). Yield 123 mg (76%), colorless powder, m.p. 185°C. – IR (KBr): $\tilde{\nu}$ = 3322 cm⁻¹ m (NH), 2211 w (C≡C), 1706 s, 1684 s, 1651 s (CO₂, CON). – ¹H NMR (CDCl₃): δ = 1.37 (s, 27 H, CH₃), 1.46–1.60 (m, 6 H, CH₂), 1.93 (pt, ³J = 7.9 Hz, 6 H, CH₂), 3.03 (pd, ³J = 5.3 Hz, 6 H, CH'H, CH₂), 3.24 (dd, ²J = 13.1 Hz, ³J = 7.0 Hz, 6 H, CHH', CH₂), 4.23 (pq, ³J = 7.2 Hz, 3 H, CH), 5.00 (m, 3 H, NH), 5.79 (t, ³J = 6.2 Hz, 3 H, NH), 7.16 (d, ³J = 8.1 Hz, 6 H, C₆H₄), 7.26–7.42 (m, 36 H, C₆H₄, PPh₂), 7.58 (s, 3 H, C₆H₃). – ¹³C NMR (CDCl₃): δ = 24.97 (d, ²J = 10.4 Hz, CH₂), 25.74 (d, ¹J = 18.4 Hz, CH₂), 28.16 [C(CH₃)₃], 38.53 (CH₂), 40.33 (d, ³J = 14.9 Hz, CH₂), 55.73 (CH), 80.28 [C(CH₃)₃], 87.99, 90.27 (C≡C), 121.42, 124.02, 128.50, 128.69 (d, ³J = 10.3 Hz, *m*-PPh₂), 129.52, 131.99, 132.79 (d, ²J = 18.5 Hz, *o*-PPh₂), 134.00, 137.65, 138.30 (d, ¹J = 13.9 Hz, *i*-PPh₂, 60 C, C₆H₄, C₆H₃, PPh₂), 155.42, 171.01 (CON). – ³¹P NMR (CDCl₃): δ = -16.14. – C₉₉H₁₀₅N₆O₉P₃ × 2 H₂O (1654.9): calcd. C 71.98, H 6.65, N 5.08; found C 71.77, H 6.26, N 4.77.

General Procedure for the Preparation of the Complexes 15 and 16: To a solution of 81 mg (0.05 mmol) of ligand **14** in 10 ml of dichloromethane were added 45 mg of [(Et₃P)PdCl(μ-Cl)]₂ or 46 mg of [Cp*RhCl(μ-Cl)]₂ (0.075 mmol each). The solutions were stirred at room temp. for 16 h, concentrated in vacuo and the residues were washed with 10 ml of diethyl ether and 30 ml of pentane.

1,3,5-C₆H₃{C≡CC₆H₄CH₂CH[NHCO₂C(CH₃)₃][CO₂-(CH₂)₃PPh₂]}Pd(PEt₃)(Cl)₂ (15): Yield 123 mg (98%), yellow powder, m.p. 183°C (decomp.). – IR (KBr): $\tilde{\nu}$ = 3428 cm⁻¹ m (NH), 2213 (C≡C), 1713, 1677 s (CON), 354, 329 w (Pd-Cl). – ¹H NMR (CDCl₃): δ = 0.93 (dt, ³J_{PH} = 17.8 Hz, ³J_{HH} = 7.7 Hz, 27 H, CH₃, B), 1.16 (dt, ³J_{PH} = 16.5 Hz, ³J_{HH} = 7.5 Hz, 27 H, CH₃, A), 1.37 (s, 18 H, CH₃, A+B), 1.51–1.65 (m, 6 H, CH₂, A+B), 1.81–1.95 (m, 18 H, CH₂, A+B), 2.14–2.32 (m, 6 H, CH₂, A+B), 2.89–3.41 (m, 12 H, CH₂, A+B), 4.25–4.40 (m, 3 H, CH, A+B), 5.09 (d, ³J = 6.8 Hz, 3 H, NH, A+B), 6.52 (t, ³J = 5.0 Hz, 3 H, NH, A+B), 7.19 (d, ³J = 8.2 Hz, 4 H, C₆H₄), 7.29–7.84 (m,

39 H, C₆H₄, C₆H₃, PPh₂). – ¹³C NMR (CDCl₃): δ = 8.23, 8.60 (d, ²J = 1.5 Hz; 3.2 Hz, CH₃, A+B), 14.33 (dd, ¹J = 25.7 Hz, ³J = 3.1 Hz, CH₂, A), 16.70 (d, ¹J = 30.4 Hz, CH₂, B), 21.99 (d, ¹J = 27.8 Hz, CH₂, A+B), 24.00 (CH₂, A+B), 28.26 [C(CH₃)₃, A+B], 39.20 (CH₂, A+B), 39.74 (d, ²J = 14.4 Hz, CH₂, A+B), 55.64 (CH, A+B), 79.83 [C(CH₃)₃, A+B], 87.77, 90.55 (C≡C, A+B), 121.07, 124.02, 128.39–133.69 (m, 6 C, C₆H₄, C₆H₃, PPh₂, A+B), 133.95, 137.93, 155.12 (CON, A+B), 170.93 (CON, A+B). – ³¹P NMR (CDCl₃): δ = 11.37 (d, ²J_{trans} = 538 Hz), 14.85 (d, ²J_{cis} = 234 Hz), 25.09 (d, ²J_{trans} = 538 Hz), 30.15 (d, ²J_{cis} = 234 Hz). – C₁₁₇H₁₅₀Cl₆N₆O₉P₆Pd₃ (2502.3): calcd. C 56.16, H 6.04, N 3.36; found C 56.51, H 5.95, N 3.34.

1,3,5-C₆H₃{C≡CC₆H₄CH₂CH[NHCO₂C(CH₃)₃][CO₂-(CH₂)₃PPh₂]}Rh(Cp*)(Cl)₂/3 (**16**): Yield 123 mg (97%), orange powder, m.p. 209°C (decomp.). – IR (KBr): ν̄ = 3341 cm⁻¹ m (NH), 2213 (C≡C), 1713, 1674 s (CON). – ¹H NMR (CDCl₃): δ = 1.29 (s, 18 H, CH₃), 1.31 (s, 15 H, CH₃), 1.42–1.58 (m, 6 H, CH₂), 2.47–3.19 (m, 18 H, CH₂), 4.29 (pq, ³J = 6.7 Hz, CH), 5.13–5.37 (m, 3 H, NH), 6.24–6.46 (m, 3 H, NH), 7.18 (d, ³J = 7.6 Hz, 6 H, C₆H₄), 7.32 (d, ³J = 7.6 Hz, 6 H, C₆H₄), 7.38–7.52 (m, 18 H, PPh₂), 7.58 (s, 3 H, C₆H₃), 7.69–7.86 (m, 12 H, PPh₂). – ¹³C NMR (CDCl₃): δ = 8.62 (Cp*), 23.48 (CH₂), 26.74 (d, ¹J = 28.7 Hz, CH₂), 28.20 [C(CH₃)₃], 38.68 (CH₂), 39.86 (d, ²J = 13.4 Hz, CH₂), 55.34 (CH), 79.57 [C(CH₃)₃], 87.51, 90.69 (C≡C), 98.69 (dd, J_{PC} = 6.7 Hz, J_{RhC} = 2.4 Hz, cp*), 120.72, 124.11, 128.35 (dd, J_{PC} = 9.8 Hz, J_{RhC} = 3.9 Hz, PPh₂), 129.62, 130.89 (d, J_{PC} = 12.3 Hz, PPh₂), 131.53, 133.28 (d, J_{PC} = 9.6 Hz, PPh₂), 133.67, 133.92 (d, J_{PC} = 9.0 Hz, PPh₂), 138.38, 155.19 (CON), 170.01 (CON). – ³¹P NMR (CDCl₃): δ = 27.34 (d, J_{RhP} = 143 Hz). – C₁₂₉H₁₅₀Cl₆N₆O₉P₃Rh₃ × CH₂Cl₂ (2627.9): calcd. C 59.42, H 5.83, N 3.19; found C 59.59, H 6.08, N 3.10.

X-ray-Structural Determination of 1: C₄₇H₄₀BF₄NO₂P₂Pt, MW = 994.64, crystal size 0.15 × 0.15 × 0.1 mm³. The slightly yellow prism was covered with perfluoropolyether oil, mounted on a glass fiber and centered at 183(2) K. All reflections in the range 2 θ = 13.7–49.4° (index ranges –33 < h < 33, –33 < k < 33, –10 < l < 17) were measured by using a Siemens P4 diffractometer equipped with a CCD area detector. Hexagonal, space group R3̄, a = 28.708(5), b = 28.708(5), c = 14.458(3) Å, V = 10320(4) Å³, Z = 9, d(calcd.) = 1.440 Mg/m³, = 3.182 mm⁻¹, F(000) = 4446, independent reflections 6091 [R(int) = 0.0534], 5890 observed with I > 4 σ(I); at semiempirical absorption correction, max./min. transmission 0.972/0.698. The structure was solved by the heavy-atom method, and the non-hydrogen atoms were refined anisotropically. H atoms in calculated positions were included – the final refinement against F² using a riding model (SHELX93 programs). 523 variables, R1 = 0.0408, wR2 = 0.1015, largest residual peak: 2.624 e/Å³ close to Pt.

Crystallographic data (excluding structure factors) for the structures reported in this paper have been deposited with the Cambridge Crystallographic Data Center as supplementary publication no. CCDC-102301. Copies of the data can be obtained free of charge on application to CCDC, 12 Union Road, Cambridge CB2 1EZ, UK [fax: (+44)1223-336-033; e-mail: deposit@ccdc.cam.ac.uk].

* Dedicated to Professor Meinhart Zenk on the occasion of his 65th birthday.

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[2] X-ray structure analysis of **1**.

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