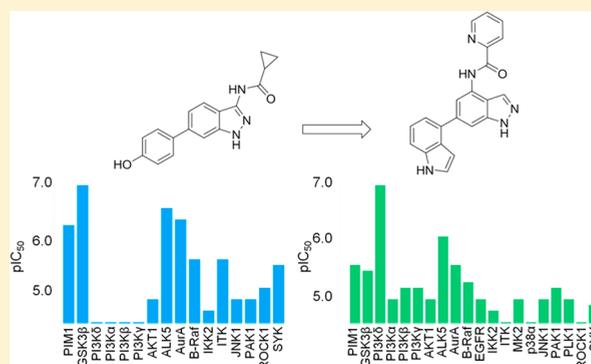


From PIM1 to PI3K δ via GSK3 β : Target Hopping through the KinomeZoë A. Henley,^{*,†} Benjamin D. Bax,^{§,Δ} Laura M. Inglesby,[‡] Aurélie Champigny,[‡] Simon Gaines,^{‡,○} Paul Faulder,^{‡,◇} Joelle Le,[§] Daniel A. Thomas,[#] Yoshiaki Washio,[‡] and Ian R. Baldwin^{*,‡}[†]Refractory Respiratory Inflammation DPU, GlaxoSmithKline R&D, Gunnels Wood Road, Stevenage, SG1 2NY, U.K.[‡]Molecular Discovery Research, [#]Biological Sciences, and [§]Computational Chemistry, Platform Technology & Science, GlaxoSmithKline R&D, Gunnels Wood Road, Stevenage, SG1 2NY, U.K.

Supporting Information

ABSTRACT: Selective inhibitors of phosphoinositide 3-kinase delta are of interest for the treatment of inflammatory diseases. Initial optimization of a 3-substituted indazole hit compound targeting the kinase PIM1 focused on improving selectivity over GSK3 β through consideration of differences in the ATP binding pockets. Continued kinase cross-screening showed PI3K δ activity in a series of 4,6-disubstituted indazole compounds, and subsequent structure–activity relationship exploration led to the discovery of an indole-containing lead compound as a potent PI3K δ inhibitor with selectivity over the other PI3K isoforms.



KEYWORDS: Phosphoinositide-3-kinase delta inhibitor, PI3K δ inhibitor, kinase cross-screening

Kinases play a pivotal role in nearly every aspect of cellular function and represent an important class of biological targets in the development of novel drugs.¹ Kinases modify substrates by chemically adding a phosphate group from adenosine triphosphate (ATP), and this phosphorylation event controls many cellular processes, making them attractive therapeutic targets.²

Identification of starting hits for biological targets can be based on approaches such as high throughput screening (HTS) for diversity or structure-based drug design (SBDD) as a focused design approach,³ but in a target class such as kinases an efficient way of finding hits is from focused or knowledge-based screening. Screening a focused set containing compounds that have previously been shown to have activity against kinases has proven to be a highly successful strategy in discovering hits against new kinases.⁴ This approach predominantly targets small molecules that bind to the ATP-binding pocket, which is well-conserved across the whole kinome.⁵ Therefore, a resultant issue in translating hits into candidates and medicines has been selectivity over other undesired kinase targets, thus avoiding unwanted side effects.

Extensive in-house kinase cross-screening is routine within GSK and ensures any unexpected compound activities against off-targets are identified. This extensive cross-profiling has the benefit of providing unexpected opportunities for other targets. This letter discusses how, by considering a hit compound for PIM1 and issues of selectivity over GSK3 β , kinase cross-screening and subsequent structure–activity relationship (SAR) exploration led to the discovery of a selective lead series for PI3K δ .

The PIM (provirus insertion site of Moloney murine leukemia virus) family of serine/threonine protein kinases is represented by PIM1, PIM2, and PIM3, which are all highly conserved in vertebrates.⁶ PIM proteins have major functions in cell survival, proliferation, and differentiation in response to growth factors and cytokines,⁷ and PIM1 was the first PIM kinase discovered to have a key role as a proto-oncogene.⁸ The overexpression of PIM1 has been found in a wide range of human tumors, including B-cell lymphomas and various types of human leukemia as well as solid tumor cancers.⁹ Therefore, the development of PIM1 inhibitors as new anticancer drugs is of considerable interest.¹⁰

Initial screening of a focused kinase compound set against PIM1 identified indazole compound **1** as a moderately potent inhibitor with a pIC₅₀ of 6.3 (Figure 1).¹¹

Compound **1** was an attractive hit with a potential to rapidly expand SAR. However, selectivity data from cross-screening against a panel of in-house kinases revealed a moderate selectivity profile, with inhibition of GSK3 β being one of the most significant off-target activities (Figure 2). GSK3 β is a multifunctional serine/threonine protein kinase, which regulates a wide range of cellular processes, involving various undesirable signaling pathways.^{12,13} ALK5 and AurA are serine/threonine kinases that were also inhibited by compound **1**, and this would also need to be addressed; however, with X-

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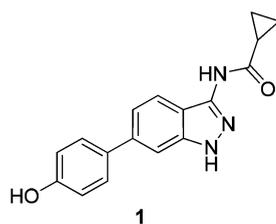


Figure 1. Initial indazole hit compound 1.

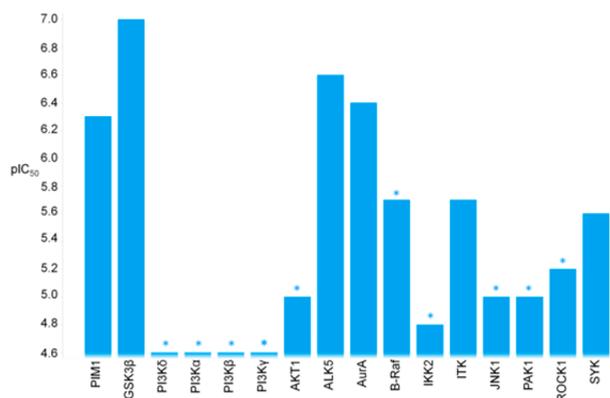


Figure 2. Selectivity profile of compound 1. *Asterisks designate a less than significant value was observed on at least one occasion.

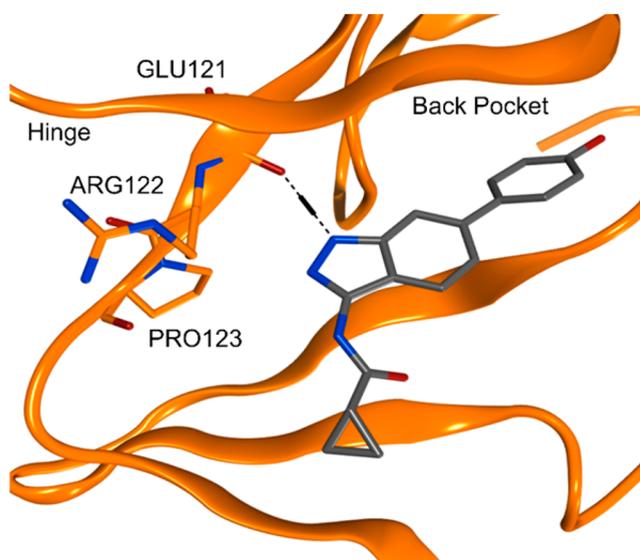


Figure 3. X-ray cocrystal structure of compound 1 in PIM1 with PIM1. Images generated using PyMol.

ray crystallography available in-house for GSK3β, our initial SAR exploration sought to improve selectivity for PIM1 over GSK3β.

Cocrystallization of compound 1 with PIM1 demonstrated that the proton-donor center N1–H of the indazole forms a hydrogen bond with Glu121 (Figure 3).

The greater affinity of compound 1 for GSK3β over PIM1 can be rationalized through consideration of the hinge interactions that are made upon binding to the different kinases. A commonly conserved H-bonding interaction to inhibitors in the hinge region of the ATP site of kinases is that of an NH on the protein backbone interacting with an acceptor motif on the inhibitor. PIM1 is unusual because the presence of

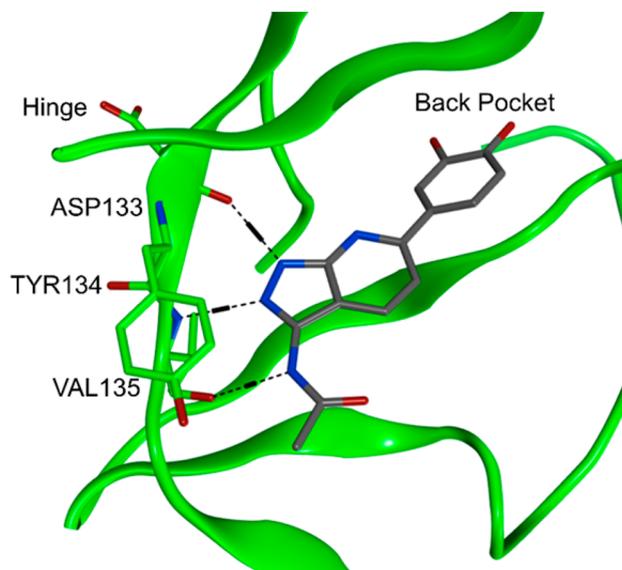
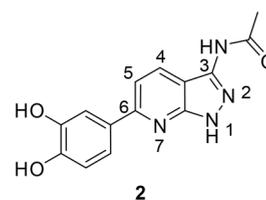
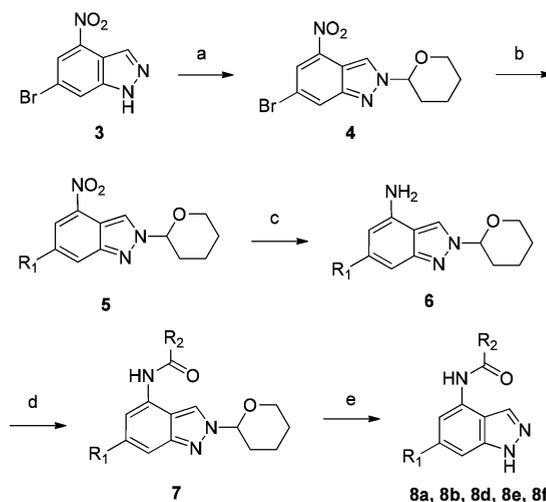


Figure 4. X-ray cocrystal structure of compound 2 with GSK3β. Images generated using PyMol.

Scheme 1. Synthetic Route for Compounds 8a, 8b, 8d, 8e, and 8f²²

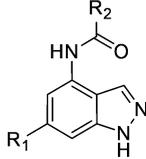


^aReagents and conditions: (a) 3,4-dihydropyran, TFA; (b) boronic acid/ester, Pd(dppf)Cl₂, aq. NaHCO₃, IPA, 150 °C, microwave; (c) H₂, Pd/C, EtOAc; (d) acid chloride, DIPEA, DCM or acid, HATU, DIPEA, DMF; (e) 4 M HCl in dioxane, MeOH. See Table 1 for range of substituents.

a proline residue (Pro123, Figure 3) in the hinge region prevents the formation of this hydrogen bond.

A cocrystal structure of a close analogue of compound 1 with GSK3β confirmed that the two commonly conserved hydrogen bonding interactions are made with Asp133 and Val135 (Figure 4).¹⁴ The accessible carbonyl group of Val135 in fact enables a

Table 1. SAR of 4,6-Disubstituted Indazoles



Cpd	R ¹	R ²	PIM1	GSK3β	pIC ₅₀ ^a			
					δ	α	β	γ
1	See Fig. 1		6.3 (6.4) ^a	7.0	4.6 ^b	<4.6	<4.6	<4.6
8a			6.5 (6.5) ^c	6.2	5.2	4.6 ^d	4.6 ^c	<4.6
8b			6.9	5.8 ^f	6.3	<4.6	<4.6	5.3 ^b
8c			4.8	4.5	4.7 ^b	4.6 ^g	<4.6	4.6 ^g
8d			5.6	5.9	5.4	4.7	4.7 ^b	5.1
8e			5.7 ^c	5.8 ^f	5.8	4.6	4.5	5.1
8f			4.8	5.1	6.0	<4.6	4.6 ^d	<4.6
8g			5.6	5.5 ^h (4.7) ^f	7.0	5.0 ^d	5.2	5.2 ^b
8h			-	<4.6 ^h	5.5	4.6 ⁱ	4.9 ^b	4.8 ^d

^aData is mean of at least three separate test occasions unless stated.

^bData from in-house PIM1 assay, *n* = 1. ^cTested <4.6 on one occasion.

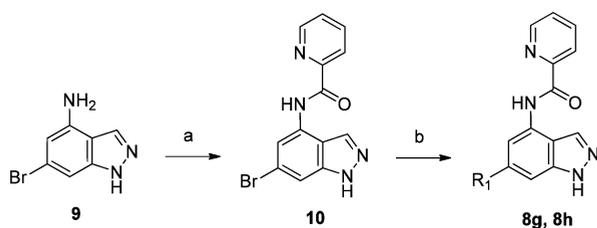
^dData from in-house PIM1 assay. ^eTested <4.6 on two occasions.

^fTested <4.6 on three occasions. ^gGSK3β data was generated at

Reaction Biology Corporation. ^hTested <4.6 on four occasions. ⁱ*n* = 1.

^j*n* = 2.

Scheme 2. Synthesis of Compounds 8g and 8h^a



^aReagents and conditions: (a) 2-pyridinecarbonyl chloride hydrochloride, DIPEA, DCM; (b) boronic acid, Pd(dppf)Cl₂, Na₂CO₃, 1,4-dioxane, H₂O. See Table 1 for the range of substituents.

donor–acceptor–donor interaction between the amido-indazole ring and the protein (Figure 4), which is not possible with PIM1.

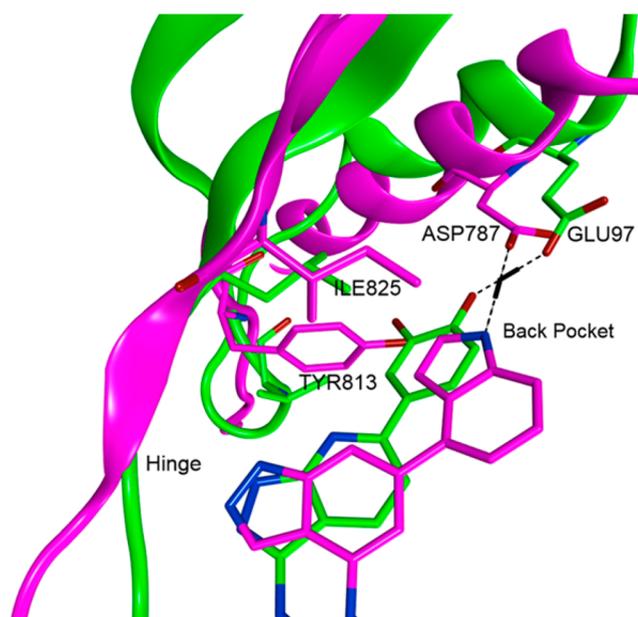


Figure 5. Close-up of the hinge and back pocket region of compound 8f docked in PI3Kδ (magenta) and overlay with compound 2 cocrystallized in GSK3β (green).

We looked to alter the PIM1/GSK3β selectivity profile in favor of PIM1 by disrupting the additional H-bonding interaction between Val135 and the amido-NH observed in GSK3β (Figure 4) by moving the amido-substituent to the indazole 4-position.

To access the desired 4,6-disubstituted indazoles, a routine synthesis was employed that began with conversion of the commercially available 4-nitro-6-bromoindazole (3) to the corresponding THP-protected indazole intermediate 4 (general synthesis shown in Scheme 1). Installation of the 6-aryl groups under standard palladium-catalyzed cross-coupling conditions with the required boronic acid or boronate ester yielded compounds of structure 5 (Scheme 1). The THP-protected phenolic boronic acids were employed when introducing phenolic boronic substituted aromatic groups (see Supporting Information). Reduction of the nitro group afforded aromatic amine intermediates of structure 6, which allowed the introduction of different amide substituents by coupling with the corresponding acid chlorides. Subsequent THP-deprotection gave the target compounds 8a, 8b, 8d, 8e, and 8f (see Table 1 for the range of substituents). Compound 8c was synthesized using an analogous route with SEM-protection of the indazole (see Supporting Information).

A shortened synthesis from commercially available 4-amino-6-bromoindazole (9), where amide formation was carried out as the first step before introduction of the required aryl groups via palladium-catalyzed cross-coupling, was followed for the synthesis of compounds 8g and 8h (Scheme 2, see Table 1 for range of substituents).

Consideration of the residue differences in the hinge region of PIM1 and GSK3β quickly led to the design of compound 8a. The initial data for compound 8a was encouraging, with GSK3β potency reduced but PIM1 activity maintained (Table 1). The introduction of a picolinamide substituent at the indazole 4-position in compound 8b showed a slight increase in PIM1 activity, while a corresponding increase in potency for GSK3β was not observed (Table 1).

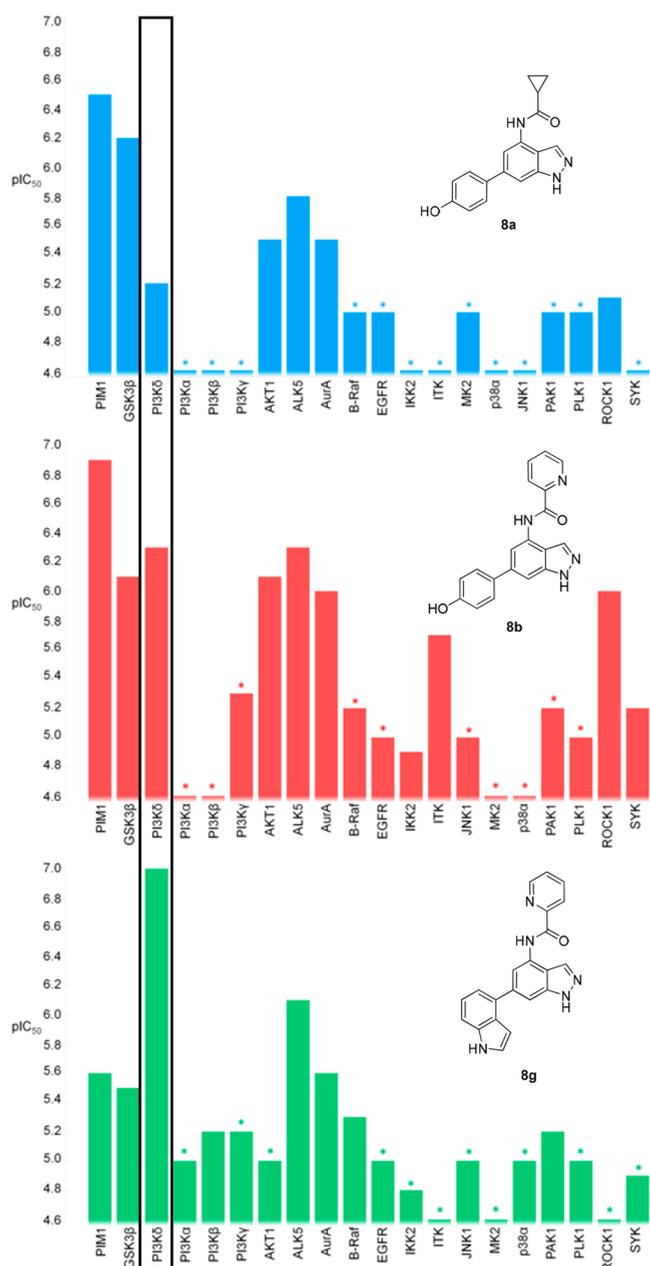


Figure 6. Kinase selectivity profiles of compounds **8a**, **8b**, and **8g**. *Asterisks designate a less than significant value was observed on at least one occasion.

Continued kinase profiling also highlighted some interesting SARs against the PI3Ks. The phosphoinositide 3-kinases (PI3Ks) are a family of enzymes that are central to signaling involving intracellular lipids.¹⁵ The Class 1 PI3K family includes phosphoinositide 3-kinase delta (PI3Kδ) and the closely related isoforms α , β , and γ , which convert phosphatidylinositol-4,5-bisphosphate to phosphatidylinositol-3,4,5-trisphosphate *in vivo*.¹⁶ This leads to downstream signaling cascades that control cell growth, cell cycle entry, and cell survival. PI3Kδ inhibition is of interest for oncology indications, with the approval of GS-1101 (idelalisib) recently reported for the treatment of patients with relapsed chronic lymphoid leukemia and non-Hodgkin lymphoma.¹⁷ Since PI3Kδ is primarily expressed in leukocytes and plays a key role in immune signaling and inflammatory processes, it is also an attractive

target for respiratory diseases such as asthma¹⁸ and chronic obstructive pulmonary disease (COPD).¹⁹ PI3Kγ is also enriched in leukocytes and has been targeted for the treatment of inflammatory disease,²⁰ whereas the α and β isoforms are ubiquitously expressed and the α and β murine knockout phenotypes are embryonically lethal.²¹ This demonstrates the potential requirement for isoform selectivity when trying to treat chronic disease.

Compound **8a** was the first in the series to show significant PI3Kδ activity (pIC₅₀ = 5.2) compared with the original hit compound **1**, which was inactive. The increased potency for PI3Kδ over the other class 1 PI3K isoforms was a key observation, as this was the first compound to show this selectivity profile from many compound series screened in-house against the PI3Ks. These initial results from kinase cross-screening and recognition of the therapeutic potential of selective PI3Kδ inhibitors led to a shift in target emphasis from PIM1 to PI3Kδ for this series of compounds.

A significant increase in PI3Kδ potency was observed for the picolinamide compound **8b**, and subsequent SAR exploration has shown that the internal hydrogen bond was key to maintaining the planarity of the amide leading to stabilization of the preferred binding conformation to PI3Kδ, which was confirmed through crystallography as described by Down et al.²²

Removal of the phenolic hydroxyl group in compound **8c** caused a reduction in potency against PIM1, GSK3β, and PI3Kδ, but moving the phenol from the *para*- to the *meta*-position in compounds **8d** and **8e** was more detrimental to PIM1 activity than to that of GSK3β and PI3Kδ.

Replacement of the phenol with a bioisosteric indole moiety in compound **8f** resulted in reduced activities for PIM1 and GSK3β but increased activity for PI3Kδ. Combining the previously recognized 4-position picolinamide with 6-position indole moieties led to compounds **8g** and **8h**, where the 4-substituted indole displayed much more potent PI3Kδ inhibition than the 5-substituted analogue. Significantly, there was a continued lack of activity of the 4,6-disubstituted indazole compounds against the other PI3K isoforms.

Due to the lack of a suitable crystal structure of PI3Kδ at this time, a homology model was built to aid the rationalization of the SAR.²⁴ The model suggested that the indazole nitrogen engaged in the hinge-binding interaction but that the core was shifted significantly when compared with the indazole binding in GSK3β and PIM1 (overlay of PI3Kδ homology model with GSK3β cocrystal structure shown in Figure 5). This movement is hypothesized to be due to changes in the gatekeeper residue and nearby back pocket residues between the three kinases. The gatekeeper residue is a single residue in the ATP pocket that has been shown to control sensitivity to small molecule kinase inhibitors since it influences the size and shape of the back cavity that is not occupied by ATP.⁵ The PI3Kδ Ile825 gatekeeper residue (PIM1 = Leu120, GSK3β = Leu132) and Tyr813 residue (PIM1 = Ile104, GSK3β = Val110) extend further into the back pocket than the corresponding residues of PIM1 and GSK3β. The movement of the indazole core, to avoid clashing with the aryl substituent of Tyr813, orients the back pocket substituent such that a key H-bond interaction with Asp787 can be made, and this is achieved when the indole substituent is substituted at the 4-position (Figure 5). This differs from the *para*-phenol substituents that are preferred for PIM1 and GSK3β, which give the optimal orientation for an H-

bond interaction with a glutamic acid residue in both kinases (Glu97 of GSK3 β shown in Figure 5).

Since the compounds were not expected to extend into areas of the protein where there are significant residue differences between the PI3K isoforms, the PI3K selectivity profile was unexpected. Subsequent SAR investigation showed that the PI3K activities were greatly affected by the back pocket substituent.²¹ Since the residues are conserved in this region, it suggested a more complex mechanism of achieving selectivity than could be explained by modeling. Potentially, a subtle change in the extended H-bonding network could be responsible for the isoform selectivity observed.²⁵

In addition to the highly encouraging selective PI3K δ inhibitory characteristics of compound **8g**, in-house kinase cross-screening indicated a good overall kinase selectivity profile compared with the initial 4-substituted indazole compounds **8a** and **8b** (Figure 6). This identified the indole-substituted indazole as a potentially novel PI3K δ selective template. ALK5 remained the only kinase where moderate selectivity (<10-fold) was observed and would need to be addressed by further optimization.²²

In summary, continued kinase cross-screening of indazole compounds focused on PIM1 antagonism identified the 4,6-disubstituted indazole series to be a novel, potent, and selective class of PI3K δ inhibitors. The promising selectivity profile of compound **8g** for PI3K δ , specifically against the other PI3K isoforms, led to its further development and enabled the discovery of potent PI3K δ inhibitors with therapeutic potential.²²

■ ASSOCIATED CONTENT

📄 Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acsmchemlett.7b00296.

Biological assays and experimental procedures (PDF)

Accession Codes

PDB codes to be deposited on acceptance.

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Author Contributions

I.B. oversaw the medicinal chemistry. S.G. and Y.W. designed compounds, and L.I., A.C., and S.G. performed chemical synthesis. B.B. carried out protein crystallography. P.F. and J.L. carried out computational modeling and molecular design. D.T. oversaw and analyzed *in vitro* assay data. Z.H. and I.B. wrote the

manuscript. All authors have given approval to the final version of the manuscript.

Notes

The authors declare the following competing financial interest(s): All authors were employees of GlaxoSmithKline at the time the work was carried out.

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■ ABBREVIATIONS

AKT1, v-Akt murine thymoma viral oncogene homologue 1; ALK5, activin receptor-like kinase 5; AurA, Aurora kinase A; B-Raf, v-Raf murine sarcoma viral oncogene homologue B; COPD, chronic obstructive pulmonary disease; dppf, 1,1'-bis(diphenylphosphino)ferrocene; EGFR, epidermal growth factor receptor; GSK3 β , glycogen synthase kinase 3 beta; HATU, 1-[bis(dimethylamino)methylene]-1H-1,2,3-triazolo-[4,5-*b*]pyridinium 3-oxid hexafluorophosphate; IKK2, inhibitor of nuclear factor kappa-B kinase subunit beta; ITK, interleukin-2-inducible T-cell kinase; IPA, isopropyl alcohol; JNK1, c-Jun N-terminal kinase 1; MK2, mitogen-activated protein kinase-activated protein kinase-2; p38 α , p38-mitogen-activated protein kinase alpha; PAK1, p21-activated kinase 1; PI3K, phosphoinositide 3-kinase; PIM, provirus insertion site of Moloney murine leukemia virus; PLK1, polo-like kinase 1; ROCK1, Rho-associated protein kinase 1; SEM, 2-(trimethylsilyl)-ethoxymethyl; SYK, spleen tyrosine kinase

■ REFERENCES

- (1) Cohen, P.; Alessi, D. R. Kinase Drug Discovery – What's Next in the Field. *ACS Chem. Biol.* **2013**, *8*, 96–104.
- (2) Wu, P.; Nielsen, T. E.; Clausen, M. H. Small-molecule kinase inhibitors: an analysis of FDA-approved drugs. *Drug Discovery Today* **2016**, *21*, 5–10.
- (3) Jhoti, H.; Rees, S.; Solari, R. High-throughput screening and structure-based approaches to hit discovery: is there a clear winner? *Expert Opin. Drug Discovery* **2013**, *8*, 1449–1453.
- (4) Bamborough, P.; Christopher, J. A.; Cutler, G. J.; Dickson, M. C.; Mellor, G. W.; Morey, J. V.; Patel, C. B.; Shewchuk, L. M. 5-(1H-Benzimidazol-1-yl)-3-alkoxy-2-thiophenecarbonitriles as potent, selective, inhibitors of IKK- ϵ kinase. *Bioorg. Med. Chem. Lett.* **2006**, *16*, 6236–6240.
- (5) Zuccotto, F.; Ardini, E.; Casale, E.; Angiolini, M. Through the "Gatekeeper Door": Exploiting the Active Kinase Conformation. *J. Med. Chem.* **2010**, *53*, 2681–2694.
- (6) Mikkers, H.; Nawijn, M.; Allen, J.; Brouwers, C.; Verhoeven, E.; Jonkers, J.; Berns, A. Mice deficient for all PIM kinases display reduced body size and impaired responses to haematopoietic growth factors. *Mol. Cell. Biol.* **2004**, *24*, 6104–6115.
- (7) Brault, L.; Gasser, C.; Bracher, F.; Huber, K.; Knapp, S.; Schwaller, J. PIM serine/threonine kinases in the pathogenesis and therapy of hematologic malignancies and solid cancers. *Haematologica* **2010**, *95*, 1004–1015.
- (8) Cuypers, H. T.; Selden, G.; Quint, W.; Zijlstra, M.; Mandaag, E. R.; Boelens, W. Murine leukemia virus-induced T-cell lymphomagenesis: integration of proviruses in a distinct chromosomal region. *Cell* **1984**, *37*, 141–150.
- (9) Nawijn, M. C.; Alendar, A.; Berns, A. For better or for worse: the role of Pim oncogenes in tumorigenesis. *Nat. Rev. Cancer* **2011**, *11*, 23–34.

- (10) Anizon, F.; Shtil, A. A.; Danilenko, V. N.; Moreau, P. Fighting tumor cell survival: advances in the design and evaluation of pim inhibitors. *Curr. Med. Chem.* **2010**, *17*, 4114–4133.
- (11) $pIC_{50} = -\log_{10}IC_{50}$; where the IC_{50} is the concentration of compound required to inhibit the kinase activity by 50%.
- (12) Leloir, L. F.; Olavaria, S. H.; Goldenberg, S. H.; Carminatti, H. Biosynthesis of Glycogen from Uridine Diphosphate Glucose. *Arch. Biochem. Biophys.* **1959**, *81*, 508.
- (13) Mandarino, L.; Wright, K.; Verity, L.; Nichols, J.; Bell, J.; Kolterman, O.; Beck-Nielsen, H. Effects of insulin infusion on human skeletal muscle pyruvate dehydrogenase, phosphofructokinase, and glycogen synthase. Evidence for their role in oxidative and non-oxidative glucose metabolism. *J. Clin. Invest.* **1987**, *80*, 655.
- (14) Witherington, J.; Bordas, V.; Gaiba, A.; Garton, N. S.; Naylor, A.; Rawlings, A. D.; Slingsby, B. P.; Smith, D. G.; Takle, A. K.; Ward, R. W. *Bioorg. Med. Chem. Lett.* **2003**, *13*, 3055–3057.
- (15) Cantley, L. C. The phosphoinositide 3-kinase pathway. *Science* **2002**, *296*, 1655–1657.
- (16) Vanhaesebroeck, B.; Guillermet-Guibert, J.; Graupera, M.; Bilanges, B. The emerging mechanisms of isoform-specific PI3K signalling. *Nat. Rev. Mol. Cell Biol.* **2010**, *11*, 329–341.
- (17) Dienstmann, R.; Rodon, J.; Serra, V.; Tabernero, J. Picking the point of inhibition: A comparative review of PI3K/AKT/mTOR pathway inhibitors. *Mol. Cancer Ther.* **2014**, *13*, 1021–1031.
- (18) Rowan, W. C.; Smith, J. L.; Affleck, K.; Amour, A. Targeting phosphoinositide 3-kinase δ for allergic asthma. *Biochem. Soc. Trans.* **2012**, *40*, 240–245.
- (19) Sriskantharajah, S.; Hamblin, N.; Worsley, S.; Calver, A. R.; Hessel, E. M.; Amour, A. Targeting phosphoinositide 3-kinase δ for the treatment of respiratory diseases. *Ann. N. Y. Acad. Sci.* **2013**, *1280*, 35–39.
- (20) Rückle, T.; Schwarz, M. K.; Rommel, C. PI3K γ inhibition: towards an ‘aspirin of the 21st century’? *Nat. Rev. Drug Discovery* **2006**, *5*, 903–918.
- (21) Vanhaesebroeck, B.; Ali, K.; Bilancio, A.; Geering, B.; Foukas, L. C. Signalling by PI3K isoforms: insights from gene-targeted mice. *Trends Biochem. Sci.* **2005**, *30*, 194–204.
- (22) Down, K. D.; Amour, A.; Baldwin, I. R.; Cooper, A. W. J.; Deakin, A. M.; Felton, L. M.; Guntrip, S. B.; Hardy, C.; Harrison, Z. A.; Jones, K. A.; Jones, P.; Keeling, S. E.; Le, J.; Livia, S.; Lucas, F.; Luniss, C. J.; Parr, N. J.; Robinson, E.; Rowland, P.; Smith, S.; Thomas, D. A.; Vitulli, G.; Washio, Y.; Hamblin, J. N. Optimization of Novel Indazoles as Highly Potent and Selective Inhibitors of Phosphoinositide 3-Kinase δ for the Treatment of Respiratory Disease. *J. Med. Chem.* **2015**, *58*, 7381–7399.
- (23) <http://www.reactionbiology.com>.
- (24) A homology model of PI3K δ was developed using an X-ray structure of PI3K γ (1vgt) as a template and using the Biopolymer Composer module in Sybyl v7.2.
- (25) Sutherlin, D. P.; Baker, S.; Bisconte, A.; Blaney, P. M.; Brown, A.; Chan, B. K.; Chantry, D.; Castanedo, G.; DePledge, P.; Goldsmith, P.; Goldstein, D. M.; Hancox, T.; Kaur, J.; Knowles, D.; Kondru, R.; Lesnick, J.; Lucas, M. C.; Lewis, C.; Murray, J.; Nadin, A. J.; Nonomiya, J.; Pang, J.; Pegg, N.; Price, S.; Reif, K.; Safina, B. S.; Salphati, L.; Staben, S.; Seward, E. M.; Shuttleworth, S.; Sohal, S.; Sweeney, Z. K.; Ultsch, M.; Waszkowycz, B.; Wei, B. Potent and selective inhibitors of PI3K δ : Obtaining isoform selectivity from the affinity pocket and tryptophan shelf. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 4296–4302.