

Tetrahedron, Vol. 53, No. 22, pp. 7501-7508, 1997 © 1997 Elsevier Science Ltd All rights reserved. Printed in Great Britain 0040-4020/97 \$17.00 + 0.00

PII: S0040-4020(97)00445-6

Novel and Versatile Synthesis of Pyrrolidine Type Azasugars, DAB-1 and LAB-1, Potent Glucosidase Inhibitors[†]

Yong Jip Kim,^a Masaru Kido,^b Masahiko Bando,^b and Takeshi Kitahara*.^a

^aDepartment of Applied Biological Chemistry, Graduate School of Agricultural and Life Sciences, The University of Tokyo, 1-1-1 Yayoi, Bunkyo-ku, Tokyo 113, Japan

^bAnalytical Chemistry, 2nd Tokushima Institute of New Drug Research, Otsuka Pharmaceutical Co. Ltd., Kawauchi, Tokushima 770-01, Japan

Abstract: Synthesis of glucosidase inhibitors, DAB-1 (1) and LAB-1 (2) from dicthyl tartrate is described. The procedure afforded an epimerizable mixture of diastereomeric intermediates 8 and *ent.* 8, and opened the door not only to the selective synthesis of DAB-1 and LAB-1 but for giving various related analogs. © 1997 Elsevier Science Ltd.

INTRODUCTION

Much attention has been paid to naturally occurring polyhydroxylated piperidines and pyrrolidines in view of their remarkable inhibitory activities against glucosidases and/or mannosidases and for their important role in a number of important biological processes.¹⁻² At present, there are numerous known natural azasugars as a glucosidase inhibitor.³

Naturally occurring product DAB-1 (1), isolated from Angylocalyx boutiqueanus and Arachniodes standishii⁴ and its enantiomer LAB-1 (2) are powerful inhibitors of a range of α -glucosidases.⁵ Nectrisine 3, a fungal metabolite isolated from Nectria lucida, is also a potent α -glucosidase and α -mannosidase inhibitor.⁶ There are a variety of syntheses for 1,⁷ 2⁸ and 3⁹ which mostly start from sugars as chiral precursors. We have recently shown that netrisine and 4-epi-nectrisine could be readily synthesized from D-(-)-diethyl tartrate.¹⁰ Herein we wish to describe a novel and versatile synthesis of enantiomeric DAB-1 (1) and LAB-1 (2) by employing an epimerizable mixture of diastereomeric intermediates, respectively.



The synthesis of DAB-1 (1) and LAB-1 (2) could be planned according to the retrosynthetic paths

[†]Synthetic Studies on Enzyme Inhibitors. Part 5. For Part 4, see: Ishigami, K.; Kitahara, T. *Tetrahedron*, **1995**, *51*, 6431.

Y. J. KIM et al.

outlined in Scheme 1. It can be envisioned that DAB-1 (1) would be derived from the pyrrolidine A by reduction of the ester function, followed by deprotection of the *N*-protecting group. The pyrrolidine A can be generated from amino nitriles C by cyclization and methanolysis of nitrile. The diastereomeric mixture of amino nitrile C can be easily prepared by a modified Strecker reaction¹¹ with the known aldehyde E,¹² readily available from a chiral source, D-(-)-diethyl tartrate. LAB-1 (2) must be synthesized starting from L-(+)-diethyl tartrate in the same manner.



2 ===⇒ L-(+)-diethyl tartrate

Scheme 1

RESULTS AND DISCUSSION

Our synthesis is illustrated in Scheme 2. Reaction of the aldehyde 4 with 2.4 eq. of pmethoxybenzylamine and 1.2 eq. of diethyl phosphorocyanidate (DEPC)¹¹ in THF gave aminonitrile 5 (86.7%), as an inseparable diastereometic mixture, which was subsequently deprotected with tetra-*n*butylammonium fluoride (TBAF) in THF to the corresponding amino alcohol 6 (quant.). Esterification of 6 with p-toluenesulfonyl chloride (p-TsCl) in pyridine afforded the tosylate 7 (84%), the precursor of the cyclization reaction. Cyclization was then examined under various basic conditions but proved unsuccessful. Because the 5-membered acetal ring could obstruct this cyclization, we decided to eliminate the acetonide protecting group. Thus, treatment of tosylate 7 with CF₃COOH-H₂O-THF (5:1:1) afforded cyclization products **8a** and **8b** in a ratio of 1:4 (74%). The configuration of the major product **8b** was confirmed by an X-ray structure analysis. Since the desired trans-product 8a was a minor product, the epimerization of 8b to 8a was needed. Faced with this problem, we decided to convert the nitrile group into methyl ester under basic conditions. Thus, the diastereomeric mixture 8a and 8b was treated with 3eq. of sodium methoxide in MeOH at room temperature to give a chromatographically separable mixture of methyl ester 9a (28%), 9b (21%) and the recovered starting material (48.7%). Repeating this reaction with the recovered starting material under the same conditions, we obtained the desired trans-methyl ester 9a (49%), as a slightly major product, and cismethyl ester 9b (38%) and the starting material (6%). In addition, treatment of 9b with NaOMe in MeOH at 65-70°C for 2h afforded a 1:1 mixture of 9a, 9b in 75-80% yield. The configuration of 9a, 9b was assigned by ¹H NMR. The trans configuration of C-3 and C-4 in 9a was evident from the observed coupling constant between trans protons H-3 and H-4, with $J_{3,4}=2.8$ Hz, whereas the coupling constant of 9b was in good agreement with that of the major cis-product 8b (J_{14} =5.7Hz). Reduction of the methyl ester 9a with sodium borohydride (NaBH₄) in ethanol gave the alcohol 10 (89%). Removal of the protecting PMB group in 10 by catalytic hydrogenolysis over Pearlman's catalyst (Pd(OH)₂.C) in ethanol provided DAB-1 (1) which was

conveniently isolated as its crystalline hydrochloride by treatment with conc. HCl (94%). Comparison of the melting point (m.p.= 114-115°C, lit.⁵ m.p.= 113-115°C), specific optical rotation ($[\alpha]_D^{22} = +32.5^\circ$, c=0.5, H₂O, lit.⁵ $[\alpha]_D^{20} = +37.9^\circ$, c=0.53, H₂O), ¹H and ¹³C NMR data of synthetic 1 with those of the literature completely confirmed the identity of 1.



Scheme 2. a) p-(CH₃O)C₆H₄CH₂NH₂, (EtO)₂P(O)CN, THF; 86.7%; b) TBAF, THF; quant.; c) p-TsCl, pyridine; 84%; d) CF₃COOH-H₂O-THF (5:1:1); 70-75%; e) NaOMe, MeOH then 2N HCl; 87%; f) NaOMe, MeOH, 65-70°C, 2h then 2N HCl; 78%; g) NaBH₄, EtOH; 89%; h) H₂, 20% Pd(OH)₂-C, HCOOH, EtOH; i) conc. HCl; 94%

Alternatively, the enantiomeric LAB-1 (2) was then synthesized from L-(+)-diethyl tartrate, following the set of reactions previously described for the DAB-1 (1). LAB-1 (2) was most conveniently isolated as its hydrochloride by treatment with conc. HCl. Comparison of the melting point (m.p.= 109-111°C, lit.⁵ m.p.= 107-111°C), specific optical rotation ($[\alpha]_D^{20} = -36.7^\circ$, c=0.37, H₂O, lit.⁵ $[\alpha]_D^{20} = -34.6^\circ$, c=0.37, H₂O), ¹H and ¹³C NMR of synthetic 2 with those of the literature completely confirmed the identity of LAB-1 (2).

In summary, the synthesis of DAB-1 (1) and LAB-1 (2) achieved in this work has permitted ready utilization of the nonsugar chiral pool, optically active diethyl tartrate, and an efficient and selective routes to

these glucosidase inhibitors has been developed. Because this procedure is simple and moreover, affords the related epimers, we anticipate that it may produce various synthetic intermediates and analogs. Further studies on the syntheses of analogs and thorough biochemical evaluations are in progress.

EXPERIMENTAL

IR: Jasco A-102 spectrometer. – ¹H NMR (in $CDCl_3$, CD_3OD or D_2O): Jeol JNM EX-90 spectrometer (90 MHz), Bruker AC-300 spectrometer (300 MHz) or Jeol JNM– A 500 spectrometer (500MHz). – Optical rotations: Jasco DIP-371 polarmeter. – Column chromatography: Merck Kieselgel 60 (Art. Nr. 7734). – melting points: uncorrected values.

(2R, 3R)-2, 3-Isopropylidenedioxy-4-[(t-butyldimethylsilyl)oxy]-1--cyano-N-(4-methoxybenzyl)butan amine (5). To a mixture of 4 (9.44g, 34.4mmol) and DEPC (6.73g, 41.3mmol) in THF (200ml) was added pmethoxybenzylamine (11.34g, 82.6mmol) in THF (30ml). The mixture was stirred at room temperature for 1h and concentrated *in vacuo*. The residue was chromatographed over SiO₂ (EtOAc/Hexane, 1:6) to give 12.54g (86.7%) of 5 as an oil.; $[\alpha]_D^{20}$ = +44.2 (c=1.0 in CHCl₃). IR (neat): v_{max} (film)/cm⁻¹: 3270, 2920, 2220, 1620, 1520, 1460, 1380, 1250, 1090 and 840. ¹H-NMR (CDCl₃) δ : 0.03 (6H,s), 0.81 (9H, d, J=3.2Hz), 1.39 (6H, d, J=4.9Hz), 2.17 (1H, bs), 3.58-3.68 (2H, m), 3.77 (3H, s), 3.81-4.21 (5H, m), 6.83 (2H, d, J=8.5Hz),7.26 (2H, d, J=8.5Hz). Anal. Cald. for C₂₂H₃₆N₂O₄Si: C, 62.82; H, 8.63; N, 6.66. Found: C, 62.34; H, 8.57; N, 6.56.

(*ent.* 5). In the same manner as described for the preparation of 5, *ent.* 4 (10g) yielded 13.4g (88%) of *ent.* 5 as an oil.; $[\alpha]_D^{22}$ = -40.2 (c=0.6 in CHCl₃). The ¹H and IR spectral data were identical to those of 5. Anal. Cald. for C₂₂H₃₆N₂O₄Si: C, 62.82; H, 8.63; N, 6.66. Found: C, 62.36; H, 8.53; N, 6.62.

(2*R*,3*R*)-2,3-Isopropylidenedioxy-4-hydroxy-1-cyano-N-(4-methoxybenzyl)butanamine (**6**). To a stirred solution of **5** (7.54g, 17.9mmol) in THF (100ml) cooled at 0°C was added a solution of tetrabutylammonium fluoride (5.62g, 21.48mmol) in THF (30ml). The resulting mixture was allowed to warm to room temperature. After being stirred for 2h, water was added and the mixture was extracted with Et₂O. The extract was washed successively with water and brine, dried (MgSO₄) and concentrated under reduced pressure. The residue was purified by SiO₂ column chromatography (EtOAc/Hexane, 1:2) to give 5.5g (quant.) of **6** as an oil.; $[\alpha]_D^{20}$ = +66.1 (c=2.0 in CHCl₃). IR (neat): v_{max} (film)/cm⁻¹: 3440, 3270, 2920, 2220, 1620, 1520, 1460, 1380, 1250, 1170, 1090, 1040 and 840. ¹H-NMR (CDCl₃) δ : 1.41 (6H ,s), 2.15 (1H, bs), 3.63-3.82 (3H, m), 3.78 (3H, s), 3.99-4.25 (5H, m), 6.83 (2H, d, J=8.5Hz), 7.23 (2H, d, J=8.5Hz). Anal. Cald. for C₁₆H₂₂N₂O₄: C, 62.73; H, 7.24; N, 9.14. Found: C, 62.6; H, 7.45; N, 8.72.

(*ent.* 6). In the same manner as described for the preparation of 6, *ent.* 5 (12g) yielded 8.74g (quant.) of *ent.* 6 as an oil.; $[\alpha]_D^{20}$ = -67.6 (c=1.7 in CHCl₃). The ¹H and IR spectral data were identical to those of 6. Anal. Cald. for C₁₆H₂₂N₂O₄: C, 62.73; H, 7.24; N, 9.14. Found: C, 62.40; H, 7.18; N, 8.89.

(2*R*, 3*R*)-2, 3- Isopropy lidenedioxy-4-(*p*-toluene sulfonyl)oxy-1-cyan o-N-(4-m ethoxybenzyl)butanamine (7). *p*-TsCl (4.1g, 21.5mmol) was added to a solution of **6** (5.5g, 17.9mmol) in pyridine (40ml) at room temperature. The resulting reaction mixture was stirred for 4h. After concentration *in vacuo*, the residue was dissolved in Et₂O (50ml) and washed with 1N aqueous HCl, saturated aqueous NaHCO₃ and brine. The organic layer was dried (MgSO₄) and concentrated *in vacuo*. The residue was chromatographed over SiO₂ (EtOAc/ Hexane, 1:4) to give (6.95g, 84%) 7 as an oil.; $[\alpha]_D^{20}$ = +43.4 (c=0.8 in CHCl₃). IR (CHCl₃): *v*_{max} (film)/cm⁻¹: 3270, 2940, 2220, 1620, 1520, 1360, 1250, 1180, 1100, 980, 820 and 760. ¹H-NMR (CDCl₃) δ : 1.36 (6H, d, *J*=4.9Hz), 1.88 (1H, bs), 2.45 (3H, s), 3.54-3.65 (1H, m), 3.81 (3H, s), 4.0-4.39 (6H, m), 6.88 (2H, d, *J*=8.5Hz), 7.28 (4H, t, *J*=8.0Hz), 7.72 (2H, d, *J*=8.5Hz). Anal. Cald. for C₂₃H₂₈N₂O₆S: C, 59.98; H, 6.13; N, 6.08. Found: C, 59.82; H, 6.13; N, 5.86.

(*ent.* 7). In the same manner as described for the preparation of 7, *ent.* 6 (7g) yielded 9.05g (86%) of *ent.* 7 as an oil.; $[\alpha]_D^{20} = -58.3$ (c=1.0 in CHCl₃). The ¹H and IR spectral data were identical to those of 7. Anal. Cald. for $C_{23}H_{28}N_2O_6S$: C, 59.98; H, 6.13; N, 6.08. Found: C, 59.60; H, 5.97; N, 5.63.

(2R,3R)-N-(4-Methoxybenzyl)-2,3-dihydroxy-4-cyanopyrrolidine (8a, b). A solution of 7 (2g, 4.3mmol) in 20ml of CF₃COOH-H₂O-THF (8:1:1) was stirred for 20h at room temperature. The reaction mixture was concentrated in vacuo and 50ml of EtOAc was added to the residue and the mixture was poured into saturated aqueous NaHCO₃ (100ml) and extracted with EtOAc (three times). The extract was washed with saturated aqueous NaHCO₃ and brine, dried (MgSO₄) and concentrated in vacuo. The residue was purified by SiO₂ column chromatography (EtOAc/Hexane, 2:1) to give 0.16g (15%) of 8a as an oil and 0.61g (57%) of 8b as a white crystalline solid; 8a; $[\alpha]_{D^{2}} = -33.4$ (c=0.5 in MeOH). IR (neat): ν_{max} (film)/cm⁻¹: 3400, 2940, 2220, 1680, 1520, 1250, 1180 and 760. H-NMR (CDCl₃) δ: 2.65 (1H, dd, J=2.8Hz, 8.0Hz), 3.15 (1H, dd, J=6.5Hz), 3.47 (1H, d, J= 2.4Hz), 3.62 (1H, d, J=12.8Hz), 3.80 (3H, s), 3.90 (1H, d, J=12.8Hz), 4.15 (1H, m), 4.35 (1H, bs), 6.87 (2H, d, J=8.5Hz), 7.24 (2H, d, J=8.5Hz). Anal. Cald. for $C_{13}H_{16}N_{2}O_{3}$: C, 62.89; H, 6.50; N, 11.28. Found: C, 62.57; H, 6.40; N, 11.25.; **8b**; m.p.= 110-113°C; [α]₀²¹= +34.2 (c=0.5 in MeOH). IR (CHCl₃): v_{max} (film)/cm⁻¹: 3400, 2940, 2220, 1680, 1520, 1250, 1180 and 760. ¹H-NMR (CDCl₃) δ: 1.61 (1H, br), 2.73 (1H, dd, J=2.8Hz, 8.0Hz), 3.09 (1H, dd, J=6.5Hz), 3.62 (1H, d, J=12.8Hz), 3.80 (3H, s), 3.84 (1H, d, J=12.8Hz), 3.93 (1H, d, J=5.6Hz), 4.22-4.27 (2H, m), 6.87 (2H, d, J=8.5Hz), 7.24 (2H, d, J=8.5Hz). Anal. Cald. for C₁₃H₁₆N₂O₃: C, 62.89; H, 6.5; N, 11.28. Found: C, 62.47; H, 6.50; N, 11.28.

X-ray analysis of **8b**: Crystal size $0.5 \times 1.0 \times 1.0$ mm. All data were obtained Rigaku AFC-5S automated four-circle diffractometer with graphite-monochromated Mo K α radiation. Final lattice parameters were obtained from a least-squares refinement using 25 reflections. Crystal data: C₁₃H₁₆N₂O₃, *M*r=248.28, monoclinic, space group P2₁, a=7.383(5)Å, b=6.534(4)Å, c=13.723(3)Å, β=104.26(3)°, V=641.6(5) Å³, Z=2, *D*x=1.285 g/cm³, F(000)=264, and μ (MoK α)=0.86cm⁻¹. The intensities were measured using ω /2 θ scan up to 50°. Three standard reflections were monitored every 150 measurements. The data were corrected for Lorentz and polarization factors. Decay and absorption corrections were applied. Of the 1348 independent reflections which were collected, 1248 reflections with $I > 3.0\sigma$ (*I*) were used for structure determination and refinement. The structure was solved by direct method using TEXSAN crystallographic software package.¹³ All non-H atoms were found in Fourier map. All H atoms were calculated at geometrical positions and refined isotropically. The refinement of atomic parameters was carried out by the full matrix least-squares refinement, using anisotropically temperature factors for all non-H atoms. The final refinement converged with R=0.036 and Rw=0.041 for 226 parameters. The minimum and maximum peaks in the final difference Fourier map were -0.14 and 0.23 eÅ⁻³. Atomic scattering factors were taken from "International Tables for X-ray Crystallography."¹⁴

(*ent.* **8a**, **b**). In the same manner as described for the preparation of a mixture of **8a**, **b**, *ent.* **7** (6g) yielded 2.4g (74.7%) of a mixture of *ent.* **8a**, **b** as an oil. Further purification was not occurred.

(2R,3R)-N-(4-Methoxybenzyl)-2,3-dihydroxy-4-methoxycarbonylpyrrolidine (9a,b). Sodium (0.19g, 8.3 mmol) was dissolved in 15ml of MeOH. To this solution was added a mixture of 8a and 8b (0.7g, 2.8mmol) in 10ml of MeOH. After 5h at room temperature, the ice-cooled reaction mixture was acidified to pH5 with 2M HCl. Then 30ml of saturated aqueous NH₄Cl and 30ml of EtOAc were added. The aqueous layer was extracted twice with EtOAc. The combined organic layers were washed with saturated aqueous NaHCO3 and brine, dried (MgSO₄) and concentrated in vacuo. The residue was chromatographed over SiO₂ (EtOAc/Hexane, 2:1) to give 0.34g (48.7%) of recovered 8, and 0.22g (27.8%) of 9a as an oil and 0.17g (21%) of **9b** as a solid.; **9a**; $[\alpha]_{b^{21}} = -27.4$ (c=0.9 in MeOH). IR (neat): v_{max} (film)/cm⁻¹: 3360, 2920, 1740, 1520, 1460, 1380, 1250, 1040 and 910. ¹H-NMR (CDCl₃) δ: 2.89 (1H, dd, J=4.5Hz, 10.5Hz), 2.98 (1H, d. J=10.5Hz), 3.22 (1H, d, J= 2.8Hz), 3.66 (1H, d, J=12.9Hz), 3.67 (3H, s), 3.80 (3H, s), 3.78-3.87 (3H, m), 3.97 (1H, d, J=4.4 Hz), 4.22 (1H, br, s), 6.84 (2H, d, J=8.5Hz), 7.22 (2H, d, J=8.5 Hz). Anal. Cald. for C-₁₄H₁₉NO₅: C, 59.78; H, 6.81; N, 4.98. Found: C, 59.40; H, 6.83; N, 5.09.; **9b**; m.p.=134-137°C; [α]_D²¹= +15.1 (c=0.42 in MeOH). IR (CHCl₃): v_{max} (film)/cm⁻¹: 3400, 2220, 1518, 1250, 1180 and 760. ¹H-NMR (CDCl₃) δ: 2.35 (1H, dd, J=4.5Hz), 3.40 (1H, d, J=3.5Hz, 6.0Hz), 3.52 (2H, d, J= 12.8Hz), 3.62 (1H, d, J=5.7Hz), 3.73 (3H, s), 3.80 (3H, s), 3.87 (1H, d, J=12.8Hz), 4.20 (2H, m), 6.85 (2H, d, J=8.5Hz), 7.22 (2H, d, J=8.5 Hz). Anal. Cald. for C14H19NO5: C, 59.78; H, 6.81; N, 4.98. Found: C, 60.05; H, 6.75; N, 5.06.

Epimerization of **9b** *to* **9a**. Sodium (0.06g, 2.6mmol) was dissolved in 10ml of MeOH. To this solution was added a solution of **9b** (0.25g, 0.89mmol) in 5ml of MeOH. After the mixture was heated under 65-70°C for 2h, the ice-cooled reaction mixture was acidified to pH5 with 2M HCl. Then 30ml of saturated aqueous NH₄Cl and 30ml of EtOAc were added. The aqueous layer was extracted twice with EtOAc. The combined organic layers were washed with saturated aqueous NaHCO₃ and brine, dried (MgSO₄) and concentrated *in vacuo*. The residue was chromatographed over SiO₂ (EtOAc/Hexane, 2:1) to give 0.10g (40%) of **9a**, 0.09g (38%) of **9b**

(ent. 9a, 9b). In the same manner as described for the preparation of 9a and 9b, ent. 8 (2.2g) yielded 1.03g (46.8%) of recovered ent. 8 and 0.72g (29%) of ent. 9a as an oil, 0.57g (23%) of ent. 9b as a solid.;

ent. **9a**; $[\alpha]_D^{21} = +25.4$ (c=0.9 in MeOH). The ¹H and IR spectral data were identical to those of **9a**. Anal. Cald. for C₁₄H₁₉NO₅: C, 59.78; H, 6.81; N, 4.98. Found: C, 59.47; H, 6.82; N, 5.10. *ent.* **9b**; m.p.=142-143°C; $[\alpha]_D^{21} = -17.3$ (c=0.42 in MeOH). The ¹H and IR spectral data were identical to those of **9b**. Anal. Cald. for C₁₄H₁₉NO₅: C, 59.78; H, 6.81; N, 4.98. Found: C, 59.50; H, 6.78; N, 5.02.

Epimerization of ent. **9b** to *ent.* **9a** In the same manner as described for the epimerization of **9b** to **9a**, *ent.* **9b** (0.3g) yielded 0.11g (38%) of *ent.* **9a** and 0.12g (41%) of *ent.* **9b**.

(2R,3R,4R)-N-(4-Methoxybenzyl)-2,3-dihydroxy-4-hydroxymethylpyrrolidine (10). To a stirred solution of **9a** (0.25g, 0.89mmol) in 10ml of EtOH was added NaBH₄ (0.067g, 1.78mmol). After the addition, the mixture was heated under reflux and vigorously stirred for 1h under Ar. After cooling, the reaction mixture was evaporated, diluted with water, and extracted with EtOAc. The extract was washed with water and brine, dried (MgSO₄) and concentrated *in vacuo*. The residue was chromatographed over SiO₂(CHCl₃/MeOH, 5:2) to give 0.2g (89%) of **10** as an oil.; $[\alpha]_D^{21}$ = -52.3 (c=0.5 in MeOH). IR (CHCl₃): ν_{max} (film)/cm⁻¹: 3360, 2960, 1610, 1520, 1460, 1250, 1040 and 820. ¹H-NMR (CD₃OD) δ : 2.50 (1H, dd, *J*=4.5Hz), 2.64 (1H, dd, *J*=5.0Hz, 10.5Hz), 2.78 (1H, d, *J*=10.5Hz), 3.29 (1H, m), 3.38 (1H, d, *J*=11Hz), 3.64 (2H, m), 3.76 (3H, s), 3.86 (1H, m), 3.91 (1H, m), 3.96 (1H, d, *J*=12.8 Hz), 6.85 (2H, d, *J*= 8.5Hz), 7.25 (2H, d, *J*= 8.5Hz). Anal. Cald. for C₁₃H₁₉NO₄: C, 61.64; H, 7.56; N, 5.53. Found: C, 61.85; H, 7.78; N, 5.50.

(*ent.* 10). In the same manner as described for the preparation of 10, *ent.* 9a (0.3g) yielded 0.24g (89%) of *ent.* 10 as an oil.; $[\alpha]_D^{22}$ = +52.1 (c=0.5 in MeOH). The ¹H and IR spectral data were identical to those of 10. Anal. Cald. for C₁₃H₁₉NO₄: C, 61.64; H, 7.56; N, 5.53. Found: C, 61.30; H, 7.73; N, 5.39.

Synthesis of DAB-1 (1). 10 (0.1g, 0.39mmol) dissolved in EtOH (5ml) was stirred under a hydrogen atmosphere in the presence of 20% Pd(OH)₂-C (0.01g) and HCOOH (catalytic amount) for 1h. The catalyst was removed by filtration and the reaction mixture was concentrated, removing residual water by azeotroping with toluene. The residue was purified by flash chromatography using CHCl₃-MeOH-28% aq. NH₄OH (5:3:1) as the eluent. The appropriate fractions were combined and concentrated *in vacuo* to provide the free amine as an oil. This oil was dissolved in MeOH (5ml) and aqueous hydrochloric acid (12N, 0.2ml) was slowly added. After cooling to 5°C, the solid was collected by filtration and dried in a vacuum oven to give hydrogen chloride of DAB-1 (1), (0.063g, 94%) as a white solid.; m. p.=114-115°C; $[\alpha]_D^{22}$ = +32.5 (c=0.5 in H₂O). IR (CHCl₃): ν_{max} (film)/cm⁻¹: 3360, 2960, 1580, 1400, 1260, 1080, 1010 and 960; ¹H-NMR (D₂O) δ : 3.21 (1H, dd, *J*=12.5Hz, 2.6Hz), 3.46 (2H, m), 3.69 (1H, dd, *J*=12.2Hz, 8.1Hz), 3.81 (1H, dd, *J*=4.7Hz), 3.95 (1H, m), 4.19 (1H, m). ¹³C NMR (D₂O) d 50.90 (t, CH₂N), 59.83 (t, CH₂OH), 67.53 (d, CHN), 75.18 (d, CHOH), 76.56 (d, CHOH).

Synthesis of LAB-1 (2). In the same manner as described for the preparation of DAB-1 (1), ent. 10 (0.1g) yielded 0.61g (91%) of hydrogen chloride of LAB-1 (2) as a white solid; m. p.=109-111°C; $[\alpha]_D^{20}$ = -36.5 (c=0.37 in H₂O). The ¹H and IR spectral data were identical to those of hydrogen chloride of DAB-1 (1).

ACKNOWLEDGMENTS

We thank Dr. A. Takatsuki, a chief scientist of RIKEN, The Institute of the Physical and Chemical Research, for biological study of the synthetic samples. The financial support by the Special Coordination Funds of the Science and Technology Agency of the Japanese Government is acknowledged. We are indebted to Mrs. Y. Naito for the elemental analysis.

REFERENCES

- a) Fleet, G.W.J.; Smith, P.W. Tetrahedron, 1986, 42, 5685. b) Fleet, G.W.J.; Witty, D.R. Tetrahedron; Asymmetry, 1990, 1, 119.
- a) Hung, R.R.; Straub, J.A.; Whitesides, G.M. J. Org. Chem., 1991, 56, 3849. b) Hecquet, H.; Lemaire, M.; Bolte, J.; Demuynck, C. Tetrahedron Lett., 1994, 35, 8791 and references cited therein.
- 3) Overkleeft, H.S.; Wiltenburg, J.V.; Pandit, U.K. Tetrahedron, 1994, 50, 4215.
- a) Furukawa, J.; Okuda, S.; Saito, K.; Hatanaka, S.I.; *Phytochemistry*, **1985**, 24, 593. b) Nash,
 R.J.; Bell, E.A.; Williams, J.M. *Phytochemistry*, **1985**, 24, 1620.
- 5) Fleet, G.W.J.; Nicholas, S.I.; Smith, P.W.; Evans, S.V.; Fellows, L.E.; Nash, R.J. Tetrahedron Lett., 1985, 26, 3127.
- Shibata, T.; Nakayama, O.; Tsurumi, Y.; Okuhara, M.; Terano, H. and Kohsaka, M. J. Antibiot., 1988, 41, 296.
- 7) Brunet, D.G.; Langlois, N. Tetrahedron Lett., 1994, 35, 2889 and references cited therein.
- Behling, J.R.; Campbell, A.L.; Babiak, K.A.; Ng, J.S.; Medich, J.; Farid, P.; Fleet, G.W.J. *Tetrahedron*, 1993, 49, 3359 and references cited therein.
- 9) a) Chen, S.H.; Danishefsky, S.J. Tetrahedron Lett., 1990, 31, 2229. b) Kayakiri, H.; Nakamura, K.; Takase, S.; Setoi, H.; Uchida, I.; Terano, H.; Hashimoto, M.; Tada, T.; Koda, S. Chem. Pharm. Bull., 1991, 39, 2807.
- 10) Kim, Y.J.; Kitahara, T. Tetrahedron Lett., in press.
- 11) Harusawa, S.; Hamada, Y.; Shioiri, T. Tetrahedron Lett., 1979, 48, 4663.
- 12) Iida, H.; Yamazaki, N.; Kibayashi, C. J. Org. Chem., 1987, 52, 3337.
- "TEXAN, TEXRAY Structure Analysis PacKage," Molecular Structure Corporation, 3200 Research Forest Drive, The Woodland, TX 77381, USA., 1985.
- "International Tables for X-ray Crystallography," Kynoch Press, Birmingham, England, 1964, Vol. N, Table 2.2A.

(Received in Japan 19 March 1997; accepted 17 April 1997)