

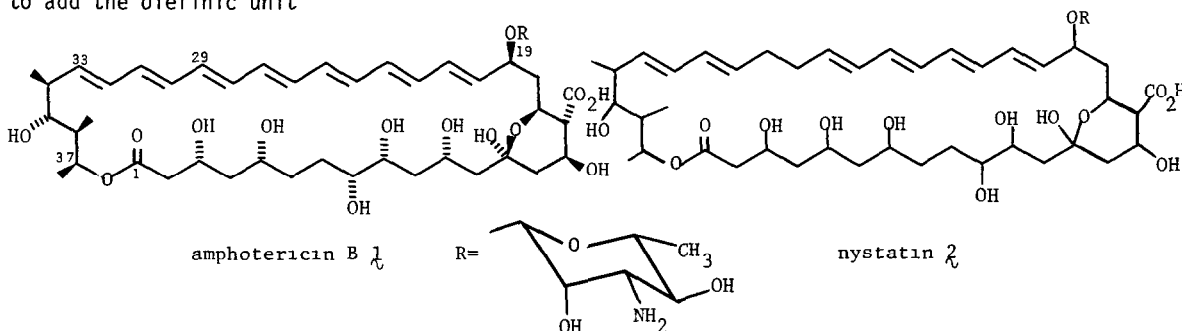
SYNTHETIC STUDIES OF POLYENE MACROLIDES, SYNTHESIS OF A C29-37
 FRAGMENT FOR AMPHOTERICIN B AND NYSTATIN

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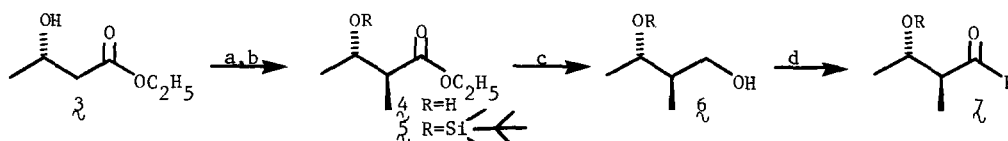
Summary An enantioselective synthesis of a C29-37 fragment common to both amphotericin B **1** and nystatin **2** is described from ethyl 3(S)-hydroxybutanoate

The polyene macrolide antibiotic group of microbial derived natural products, have a rich chemical¹ and medicinal² history. The absolute structure of amphotericin B **1** has been established by X-ray analysis.³ The structures of other members, such as nystatin **2**, have been proposed based on numerous chemical studies together with MS and NMR spectral analysis.⁴ At the present time there is a noticeable lack of synthetic studies of the polyene macrolides.⁵

We wish to report an enantiospecific synthesis of a C29-C37 fragment common to both amphotericin B **1** and nystatin **2**, as the first part of our efforts to develop a convergent total synthesis. Our synthetic scheme utilizes, 1. an efficient microbial reduction of a β -ketoester to generate the required chirality at C37, 2. a stereoselective alkylation of a β -hydroxyester, 3. a stereoselective aldol condensation, and 4. a Wittig-Horner reaction to add the olefinic unit

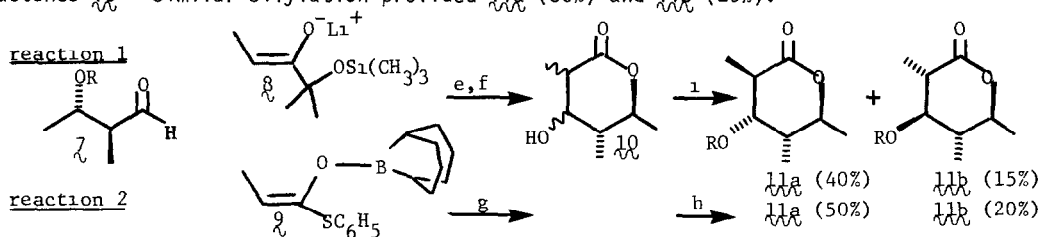


Microbial reduction of ethylacetoacetate with actively fermenting Bakers' yeast provided ethyl 3(S)-hydroxybutanoate **3** as a readily available chiral starting material.⁶ Alkylation of the dianion of **3** with methyl iodide gave (2S,3S)-**4** in 70% yield with greater than 95% stereoselectivity.⁷ The aldehyde **5** was prepared in 60% yield by the following steps, protection of the hydroxy group as a tert-butyldimethylsilyl ether, reduction of the ester **4** with lithium borohydride, and oxidation of the alcohol **6** with pyridinium chlorochromate.



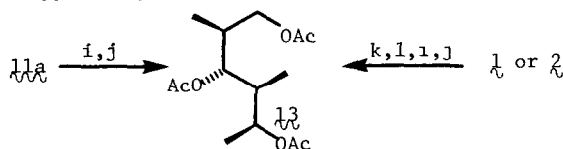
a) (1) lithium diisopropylamide (LDA) (2.2 equiv.), tetrahydrofuran (THF), -50°C , 1h. (2) add iodomethane (2.0 equiv.) and hexamethylphosphoramide (5.0 equiv.), -50°C , 1h, 70% b) t-butyl-dimethylsilylchloride (TBDMSCl) (1.1 equiv.), imidazole (3.0 equiv.), dimethylaminopyridine (DMAP) (0.1 equiv.), dimethylformamide (DMF), 60°C , 16h. c) LiBH_4 (2 equiv.), THF, reflux, 16h d) pyridinium chlorochromate (PCC) (1.5 equiv.), CH_2Cl_2 , 25°C , 2h, 60% overall from **4**.

The aldol condensation⁸ of chiral aldehyde **7** was studied with two different nucleophiles:
 1 the lithium enolate of 2-methyl-2-trimethylsiloxy-3-pentanone **8** (reaction 1),⁹ and
 2 the E-vinyloxyborane of S-phenyl propanethioate **9** (reaction 2).¹⁰ The adducts from reaction 1 were directly treated with periodic acid to provide a mixture of isomeric lactones **10**. Silylation provided derivatives which could be readily separated by gas and liquid chromatography.¹¹ Two major products **11a** (40%) and **11b** (15%) were obtained.¹² The structures were initially assigned on the basis of their 470MHz ^1H NMR spectra.¹³ The adducts from reaction 2 were directly treated with aqueous methanolic hydrogen peroxide to provide the lactones **10**. Similar silylation provided **11a** (50%) and **11b** (20%).¹²



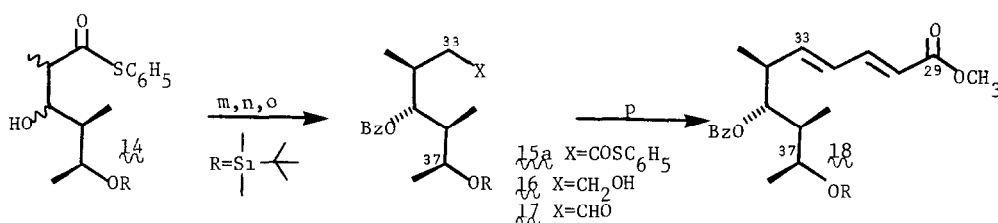
e) **8** generated by LDA (1.0 equiv.), THF, -78° , add **7**, stir 10 min at -78°C , quench with 1N HCl (3.0 equiv H^+). f) HIO_6 (5 equiv.), aqueous methanol (1 l), stir at 25°C , 16h g) **9** prepared as in ref 10, ether, 0°C , add **7**, stir 30 min at 25°C , quench with methanolic H_2O_2 (10 equiv), stir 1h at 50°C h) TBDMSCl (1.2 equiv.), imidazole (3.0 equiv), DMAP (0.1 equiv.), DMF, 60°C , 24h

The chiral lactone **11a** represents a potential precursor for the C33-37 fragment of both **1** and **2**. In order to confirm the absolute structure of **11a**, it was converted to the triacetate **13** and compared to the corresponding triacetate derived from degradation of both **1** and **2**.¹⁴ All three triacetates showed identical ^1H and ^{13}C NMR spectra¹⁵ and specific rotations as follows, synthetic **13**, $[\alpha]_D^{23} = +4.2^{\circ} \pm 0.2$, **13** from **1**, $+4.4^{\circ} \pm 0.2$ (lit ref 14, $+4.6^{\circ} \pm 0.5$ (c = 0.51, ether), **13** from **2**, $+4.3^{\circ} \pm 0.2$ (all c = 1.0, ether)



i) LiAlH_4 (4 equiv), THF, 25°C , 1h. j) acetic anhydride (10 equiv), pyridine (20 equiv), 25°C , 16h k) O_3 , methanol, -78°C , 15 min l) H_2 , PtO_2 , methanol, 25°C , 2h.

We next investigated the application of the Wittig-Horner reaction to add an olefinic unit representing C29-32 to a chiral aldehyde representing C33-37 of **1** and **2**. The thiol ester adduct **14** obtained by treatment of the initial adduct from reaction 2 with cold pH 7 buffered aqueous methanolic hydrogen peroxide served as an efficient precursor to the desired aldehyde **17** as follows. Protection of the hydroxy group of **14** as a benzoate ester allowed simple purification of the major desired isomer **15a** by column chromatography.¹⁶ Selective reduction of the thiol ester group with lithium borohydride gave the alcohol **16** which was oxidized with pyridinium chlorochromate to provide the aldehyde **17** in 40% overall yield from **14**. Condensation of the sodium anion of methyl 4-(diisopropylphosphiny)2-butenate with **17** gave the desired (E,E)-dieneester **18** in 65% yield¹⁷ which represents a C29-37 synthetic fragment of **1** and **2**.



m) benzoyl chloride (1.2 equiv), pyridine (1.2 equiv), DMAP (0.1 equiv), 25°C, 24h n) LiBH_4 (2 equiv), THF, 0°C, 2h. o) PCC (1.5 equiv), dichloromethane, 25°C, 1h, 40% overall from **14**.
 p) $(\text{tC}_3\text{H}_7\text{O})_3\text{POCH}_2\text{CH}=\text{CHCOOCH}_3$ (1.2 equiv), NaH (1.2 equiv), toluene, 25°C, 30 min

Further work toward the total synthesis of polyene macrolides is in progress

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References and Notes

- 1 a) For a historical account of the discovery of the first polyene macrolide antibiotic, nystatin see: Baldwin, R. S. "The Fungus Fighters", Cornell University Press, 1981, Ithaca, New York. b) For a review of chemistry and biology of polyene macrolides see: Hamilton-Miller, J. M. T. Bact. Rev. 1973, **37**, 166. c) For recent structural studies see: 1) Borowski, E., Golik, J., Zielinski, J., Falkowski, L., Kolodziejczyk, P., Pawlak, J. J. of Antibiotics 1978, **37**, 117. 2) Padics, L., Incze, M., Dornberger, K., Thrum, H. Tetrahedron 1982, **38**, 183.
- 2 Several polyene macrolides are used clinically in the treatment of systemic mycotic infections. For accounts of medicinal studies see: a) reference 1b, b) Hermans, P. E. Mayo Clin. Proc. 1977, **52**, 587, c) Medoff, G., Kobayashi, G. A. J. Am. Med. Assoc. 1975, **232**, 619.
- 3 Gannis, T., Artibabile, G., Mechliniski, W., Schaffner, C. P. J. Am. Chem. Soc. 1971, **93**, 4560.
- 4 Borowski, E., Zielinski, Z., Falkowski, J., Ziminski, T., Golik, J., Kolodziejczyk, P., Jerceczek, E., Gdulewicz, M., Shennin, Yu., Kotienko, T. Tetrahedron Letter, 1971, 685.
- 5 Recent report describing synthetic studies to prepare polyene macrolide mimics, Floyd, D. M., Fritz, A. W. Tetrahedron Letters, 1981, 2847, prompts us to describe our initial synthetic studies.

- 6 Deol, B S , Ridley, D D , Simpson, G W Aust J Chem. 1976, 29, 2459
- 7 Frater, G Helv Chim Acta , 1979, 62, 2825
- 8 For a review of stereoselective aldol condensations see Evans, D A , Nelson, J.V , Taber, T R. "Topics in Stereochemistry", Vol 13, Allinger, N L , Eliel, E L , Wilen, S H Ed., John Wiley and Sons, New York, 1982
- 9 Heathcock, C.H ; Buse, C T , Kleschick, W A , Pirrung, M C , Sohn, J E , Lampe, J , J Org Chem 1980, 45, 1066.
- 10 a. VanHorn, D.E , Masamune, S Tetrahedron Letters, 1979, 2229-2232
b. Hiram, M , Garvey, D S , Lu, L D -L , Masamune, S Tetrahedron Letters, 1979, 3937-3940
- 11 G C conditions used were 6 ft stainless steel 1/8 in o d column containing 10% SE -30 on 80-100 mesh Chromasorb W, 150° for 2 min, then a 16° increase per min to 230° Retention times were 6 65 min for 11a and 6 96 min for 11b Liquid chromatography was performed with a Merck "Lobar" B column with a flow rate of 10 mL/min with a gradient of 10-30% ether in hexane The undesired isomer 11b eluted first
- 12 The percent yield indicated is the average overall isolated yield from aldehyde 7
- 13 11a: $[\alpha]_D^{23} = -0.8^\circ$ (c=1.0, CHCl₃) ¹H NMR (470MHz, CDCl₃) 0.06 (s, 3H), 0.07 (s, 3H), 0.89 (s, 9H), 1.0 (d, J=6.77 Hz, 3H), 1.27 (d, J=7.63 Hz, 3H), 1.35 (d, J=6.45 Hz, 3H), 1.77-1.87 (m, 1H), 2.64 (qd, J=6.0, 2.3 Hz, 1H), 3.64 (t, J=2.3 Hz, 1H), 4.45 (dq, J=10.0, 6.0 Hz, 1H), ¹³C NMR (50.3 MHz, CDCl₃) -4.8 (q), -4.5 (q), 13.9 (q), 16.5 (q), 18.0 (s), 19.9 (q), 25.7 (q), 36.2 (d), 44.2 (d), 74.3 (d), 77.3 (d), 174.2 (s), Anal. Calcd for C₁₄H₂₈O₃Si: C, 61.72, H, 10.36, Si, 10.31
- 14 Cope, A C , Axen, U , Burrows, E P , Weinlich, J J Am Chem Soc 1966, 88, 4228
- 15 13: ¹H NMR (470 MHz, CDCl₃) δ 0.92 (d, J=5.1 Hz, 3H), 0.93 (d, J=5.0 Hz, 3H), 1.15 (d, J=6.5 Hz, 3H), 2.03 (s, 3H), 2.06 (s, 3H), 2.09 (s, 3H), 2.08-2.16 (m, 2H), 3.84 (dd, J=11.0, 6.2 Hz, 1H), 3.94 (dd, J=11.0, 7.8 Hz, 1H), 4.90-4.95 (m, 2H), ¹³C NMR (50.3 MHz, CDCl₃) 10.23 (q), 10.6 (q), 14.4 (q), 20.8 (q), 20.9 (q), 21.4 (q), 33.9 (d), 38.3 (d), 66.2 (t), 70.5 (d), 73.5 (d), 170.2 (s), 170.6 (s), 171.0 (s), Anal. Calcd for C₁₄H₂₄O₆: C, 58.32, H, 8.39. Found C, 58.15, H, 8.64
- 16 15a: $[\alpha]_D^{23} = +23.3^\circ$ (c=0.63, CHCl₃) ¹H NMR (470 MHz, CDCl₃) δ -0.002 (s, 3H), 0.008 (s, 3H), 0.87 (s, 9H), 0.99 (d, J=7.05 Hz, 3H), 1.11 (d, J=6.23 Hz, 3H), 1.35 (d, J=7.04 Hz, 3H), 2.0-2.15 (m, 1H), 3.19 (qd, J=7.04, 3.9 Hz, 1H), 3.97 (qd, J=6.3, 3.7 Hz, 1H), 5.52 (dd, J=5.62, 3.65 Hz, 1H), 7.37 (m, 5H), 7.44 (t, J=7.6 Hz, 2H), 7.56 (t, J=7.6 Hz, 1H), 8.04 (d, J=7.6 Hz, 2H) ¹³C NMR (50.3 MHz, CDCl₃) -4.8 (q), -4.5 (q), 10.0 (q), 11.0 (q), 18.0 (q), 18.1 (s), 25.9 (q), 42.1 (d), 49.8 (d), 67.7 (d), 75.4 (d), 127.5 (s), 128.4 (d), 129.1 (d), 129.3 (d), 129.7 (d), 129.8 (s), 133.0 (d), 134.6 (d), 165.6 (s), 198.6 (s).
- 17 18: $[\alpha]_D^{23} = +42.8^\circ$ (c=2.62, CHCl₃) ¹H NMR (470 MHz, CDCl₃) δ -0.028 (s, 3H), -0.020 (s, 3H), 0.847 (s, 9H), 0.94 (d, J=7.0 Hz, 3H), 1.07 (d, J=6.3 Hz, 3H), 1.14 (d, J=6.8 Hz, 3H), 2.00-2.15 (m, 1H), 2.70-2.80 (m, 1H), 3.71 (s, 3H), 3.92 (m, 1H), 5.11 (dd, J=8.9, 4.1 Hz, 1H), 5.75 (d, J=15.4 Hz, 1H), 6.08 (dd, J=15.3, 7.2 Hz, 1H), 6.19 (dd, J=15.3, 10.7 Hz, 1H), 7.19 (dd, J=15.4, 10.7 Hz, 1H), 7.45 (dd, J=7.5, 7.4 Hz, 2H), 7.57 (t, J=7.5 Hz, 1H), 8.02 (d, J=7.4 Hz, 2H), ¹³C NMR (50.3 MHz, CDCl₃) -4.8 (q), -4.5 (q), 9.7 (q), 13.4 (q), 18.0 (q), 18.0 (s), 25.8 (q), 38.8 (d), 41.9 (d), 51.5 (d), 67.6 (d), 77.5 (q), 119.9 (d), 128.3 (d), 128.4 (d), 129.5 (d), 130.0 (s), 133.0 (d), 144.8 (d), 145.6 (d), 165.9 (s), 167.4 (d)

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