

Divergent Synthesis of Methylisatoid and Tryptanthrin Derivatives by $\text{Ph}_3\text{P}-\text{I}_2$ -Mediated Reaction of Isatins with and without Alcohols

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Cite This: <https://dx.doi.org/10.1021/acs.joc.0c02403>

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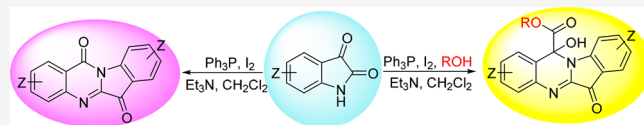


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ABSTRACT: A novel phosphonium-mediated reaction of isatins is described. In the presence of alcohol, the reaction proceeds to furnish C-12 modified tryptanthrin derivatives. Without alcohol, self-dimerization of isatins gives rise to tryptanthrin and its analogs. This divergent and step-economic approach provides a facile access to diverse indoloquinazoline structures including the natural alkaloids, methylisatoid and cephalanthrin B, in high yields from simple precursors under mild and metal-free reaction conditions.



alkaloids, methylisatoid and cephalanthrin B, in high yields from

Tryptanthrin and the related derivatives such as phaitanthrins A–E,^{1a} cephalanthrin A,^{1b} methylisatoid,^{1a–c} and cephalanthrin B^{1b} are naturally occurring indolo[2,1-*b*]-quinazoline alkaloids which have been isolated from various plant sources as well as different cell cultures (Figure 1).^{1d}

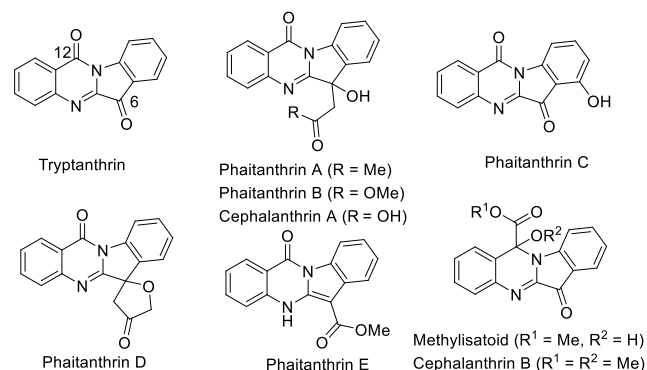


Figure 1. Indoloquinazolines isolated from natural sources.

These compounds have attracted considerable interest as potential therapeutic agents mainly due to their broad spectrum of biological and pharmaceutical activities.² Some of them include antitumor,³ antiparasitic,⁴ antimicrobial,⁵ anti-inflammatory,⁶ antimalarial,⁷ antiviral,⁸ and antitubercular⁹ properties.

Owing to their promising benefits, a number of synthetic approaches have been developed to gain access to these core structures.¹⁰ One of the most straightforward methods toward tryptanthrins is through self-dimerization of isatins. Different reaction conditions have been applied including visible-light mediated photoredox catalysis,¹¹ cathodic reduction,¹² POCl_3 -mediated reaction,¹³ and oxidative cyclization using I_2 ,^{14a,b} KMnO_4 ,^{14c} or $t\text{BuO}_2\text{H}$ ^{14d} as an oxidant. Some recent examples of alternative protocols to install the tryptanthrin framework

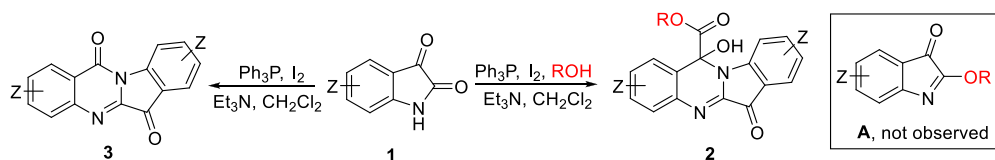
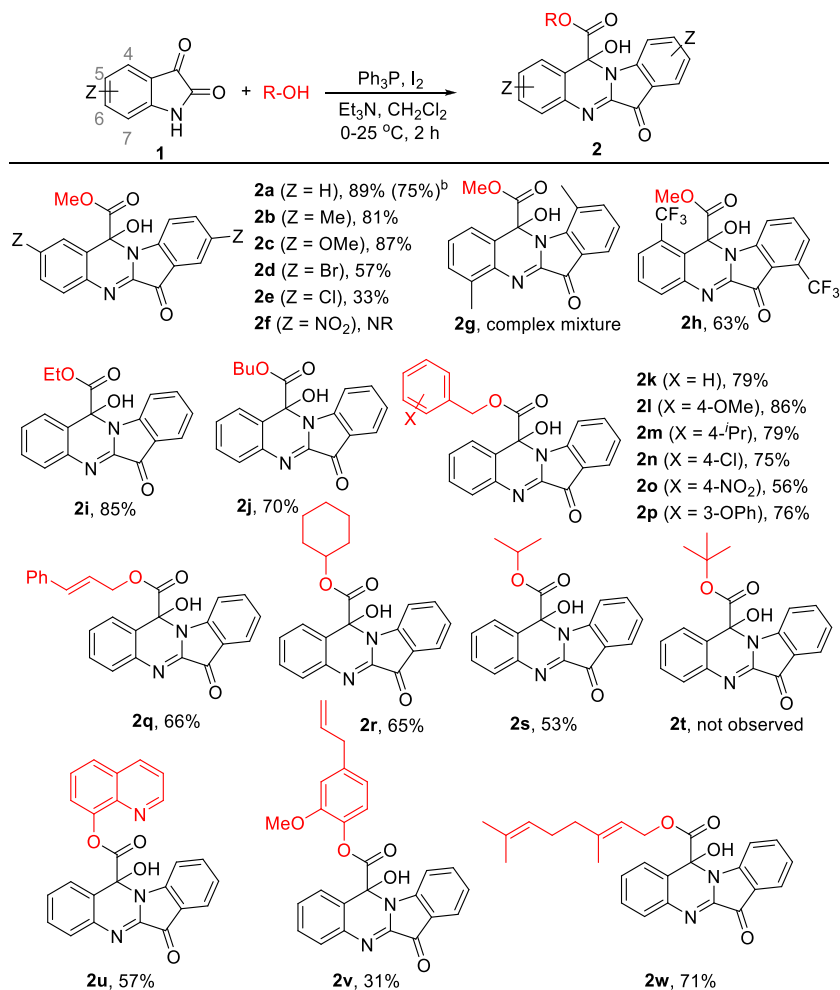
with different aryl substituents include condensation of isatins with isatoic anhydrides,¹⁵ oxidative cyclization of isatoic anhydrides and 2-aminoacetophenones,¹⁶ photochemical reaction of indoles with anthranilic acids,¹⁷ and copper-catalyzed reaction of isatins with indoles.¹⁸

Synthesis of other structurally more diverse indoloquinazolines is considered a challenging task which often requires multistep synthesis or the use of complicated substrates. While a number of methods have been introduced for C-6 functionalization of tryptanthrins,¹⁹ modification at the C-12 position remains largely unexplored.²⁰ Considering these facts and in continuation of our interest in the $\text{Ph}_3\text{P}-\text{I}_2$ -mediated reactions,²¹ herein, we disclose a novel methodology for the divergent synthesis of methylisatoid and tryptanthrin derivatives (**2** and **3**, respectively) from isatins **1** in the presence and in the absence of alcohols as depicted in Scheme 1.

Our initial investigation involved the reaction between isatin (**1a**) and methanol using the combination of $\text{Ph}_3\text{P}-\text{I}_2$ as an activator in the presence of the triethylamine base. To our surprise, when using a 1:1 mol ratio of **1a** and methanol, instead of the expected *O*-methylisatin A ($Z = \text{H}$, $R = \text{Me}$), compound **2a** was isolated in 83% yield. Based on spectroscopic analysis, its structure was identified as 12-hydroxy-12-methoxycarbonyltryptanthrin which is known as methylisatoid.¹ Since *O*-alkylated isatins are highly unstable,²² *O*-methylated product, if formed, may undergo rapid condensation with another isatin molecule to yield **2**. Due to nucleophilicity of the isatin nitrogen, formation of **2** through

Received: October 10, 2020

Scheme 1. Divergent Synthesis of Methylisatoid and Tryptanthrin Derivatives

Scheme 2. Synthesis of Methylisatoid Derivatives^a

^aReaction conditions: **1** (0.68 mmol), PPh₃ (0.51 mmol), I₂ (0.51 mmol), alcohol (0.51 mmol), Et₃N (1.70 mmol), CH₂Cl₂ (5 mL). ^bReaction performed in gram scale using **1a** (10 mmol).

initial condensation of two molecules of **1a** before reacting with the alcohol is also a possible alternative.

Based on this rationale, we decided to further investigate the reaction toward C-12 modified tryptanthrin derivatives **2**. By adjusting the amounts of the substrates (see Table S1), it was found that using a 2:1.5 mol ratio of **1a** and methanol gave the highest yield of the product **2a** (89%). The reaction in the presence of other organic bases including pyridine, imidazole, and diisopropylethylamine, however, failed to give **2a** as substantial amounts of starting material still remained. Several attempts to increase the yield of **2a** via extending reaction times, increasing the temperature, or changing the order of reagent addition also led to unsatisfactory results, while replacing I₂ with other halogenated additives including *N*-

chlorosuccinimide, *N*-bromosuccinimide, and *N*-iodosuccinimide led to recovery of unreacted isatin.

Without further optimization of the reaction conditions, the scope and generality of the reaction were then examined with different isatins and alcohols. To determine the scope with regard to the nature of isatin precursors (see Figure S1), methanol was applied as a common nucleophile leading to aryl substituted methylisatoid derivatives. According to Scheme 2, both electronic and steric factors have strong influence on the outcome of the reaction. Isatins with electron-donating group (–Me, –OMe) on the C5, *para* to the nitrogen atom, reacted rapidly to provide the corresponding methyl esters **2b** and **2c** in high yields, whereas those bearing electron-withdrawing groups (–Br and –Cl) at the same position gave lower yields of **2d** and **2e** with some remaining isatin precursors even after

prolonged reaction times (up to 24 h). Owing to the poor nucleophilicity of the aryl amido group, the electron-deficient 5-nitroisatin did not convert to the expected product **2f**. Likewise, the reaction of hindered 7-methylisatin gave rise to a complex mixture in which the expected product **2g** was not isolated. Surprisingly, the presence of CF₃ on the C4 of isatin exerts little effect on the yield of **2h**.

With the scope of the reaction with respect to the isatin structure determined, the effect of the type of alcohols on the formation of **2** was further investigated. As shown in Scheme 2, compounds **2i** and **2j** were afforded in high yields from primary alcohols having different carbon chain lengths. Benzyl alcohol and its derivatives also provided the corresponding hydroxy esters **2k–2p** in satisfactory yields. Adding an electron-donating –OMe group to the benzene ring of the alcohol led to an increased yield of **2l**, whereas the presence of a strong electron-withdrawing nitro group lowered the yield of **2o**. Cinnamyl alcohol having an α,β -unsaturated moiety is also a viable substrate giving the product **2q** in moderate yield. Sterically hindered secondary alcohols such as cyclohexanol and 2-propanol are well tolerated providing the corresponding esters **2r** and **2s** in reasonable yields. However, no reaction toward **2t** was observed when using the bulkier *tert*-butanol. In this case, competitive formation of tryptanthrin **3a** was clearly observed.

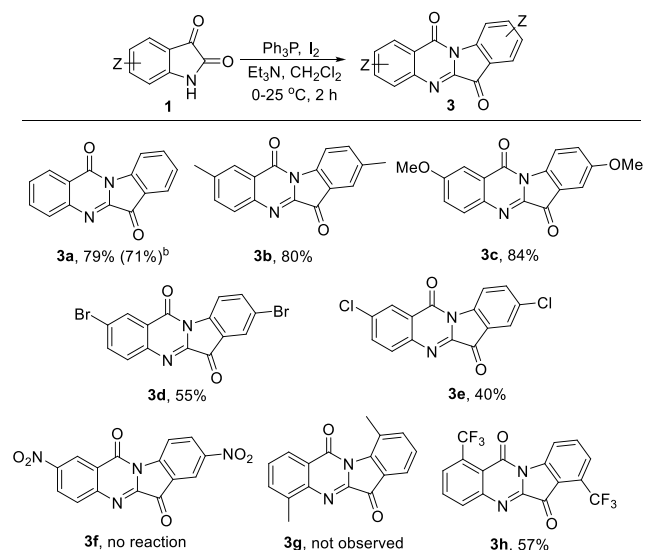
Nevertheless, other more interesting substrates including 8-hydroxyquinoline, a metal chelator with antimicrobial property,²³ eugenol with anticarcinogenic and antitumor actions,²⁴ and bioactive geraniol²⁵ gave rise to the corresponding products **2u**, **2v**, and **2w**, respectively, in moderate to good yields. Since drug combination has been proposed as a promising strategy to overcome monotherapy resistance,²⁶ modification of the tryptanthrin skeleton that would simultaneously provide two bioactive agents would render a great opportunity to achieve new potential drugs having additive and even synergistic properties.

Having successfully prepared methylisatoid derivatives, we then turned our attention to the reaction in the absence of alcohols. Based on the above results, formation of tryptanthrin side product became prominent when poorly nucleophilic alcohols were used. It is thus envisaged that the same reaction conditions without alcohol could be applied to synthesize tryptanthrin derivatives. Indeed, self-dimerization of isatins took place to afford tryptanthrin **3a** and its substituted analogs **3b–3h** in various yields (Scheme 3). In accordance with other studies,^{13b,14a} the reaction is sensitive to both electronic and steric effects. While the self-condensation process toward **3g** is disfavored due to the presence of sterically hindered methyl group, electron-rich isatins react preferably to those electron-deficient substrates and the reaction toward **3f** failed when using the least reactive 5-nitroisatin.

To demonstrate the synthetic practicality of the reaction, gram-scale syntheses of methylisatoid (**2a**) and tryptanthrin (**3a**) were carried out using 10 mmol of isatin. The reaction proceeded smoothly to afford the desired products **2a** and **3a** in 75% and 71% yield, respectively. Moreover, with **2a** in hand, we then take this opportunity to synthesize the natural alkaloid cephalanthrin B (**4**) which was obtained in high yield through methylation of **2a** with methyl iodide (Scheme 4). Compound **4** showed identical NMR data with the reported literature for cephalanthrin B which was isolated in racemic form.^{1b}

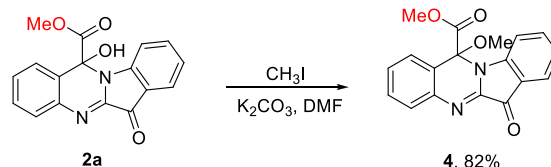
To better understand the reaction mechanism, control experiments were carried out as shown in Scheme 5. Since the

Scheme 3. Synthesis of Tryptanthrin Derivatives^a



^aReaction conditions: **1** (0.68 mmol), PPh₃ (0.51 mmol), I₂ (0.51 mmol), Et₃N (1.70 mmol), CH₂Cl₂ (5 mL). ^bReaction performed in gram scale using **1a** (10 mmol).

Scheme 4. Synthesis of **4** (Cephalanthrin B)

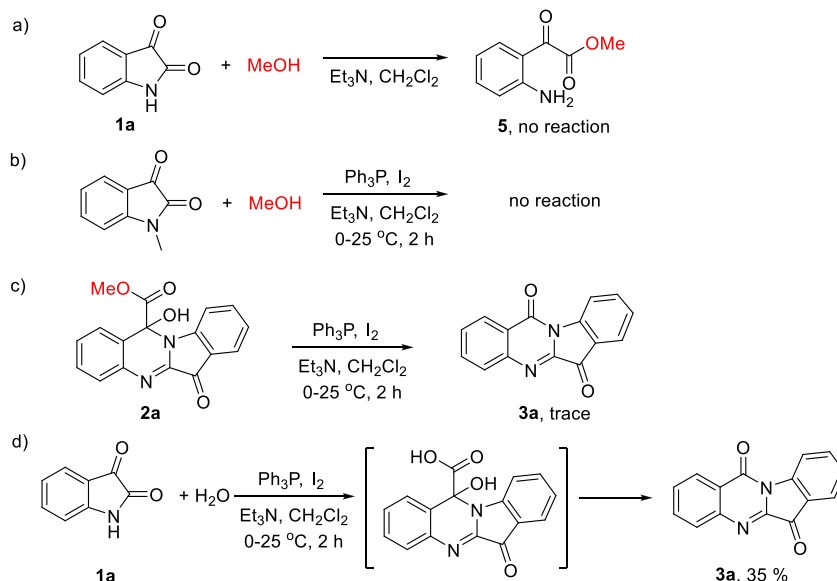


ring-opening of isatins by alcohols could give rise to keto ester **5** which is a known precursor toward **2**,^{20b,c} the reaction of isatin with methanol was examined in the absence of the Ph₃P–I₂ system. Based on Scheme 5a, no reaction was observed suggesting that **5** does not involve in our reaction. It is also well-known that the Ph₃P–I₂ reagent is a C–O bond activator, and the reaction should proceed via an initial activation of the tautomerizable amide of isatin. Indeed, using *N*-methylisatin as the substrate led to no reaction (Scheme 5b). When **2a** was subjected to the standard reaction conditions, only trace amount of **3a** was detected (Scheme 5c) indicating that formation of **3a** through hydrolysis and decarboxylation of **2a** did not occur. In addition, replacing methanol with water would theoretically yield the hydrolysate of methylisatoid. However, possibly due to its high reactivity, only the decarboxylated product **3a** was isolated albeit with low yield (Scheme 5d).

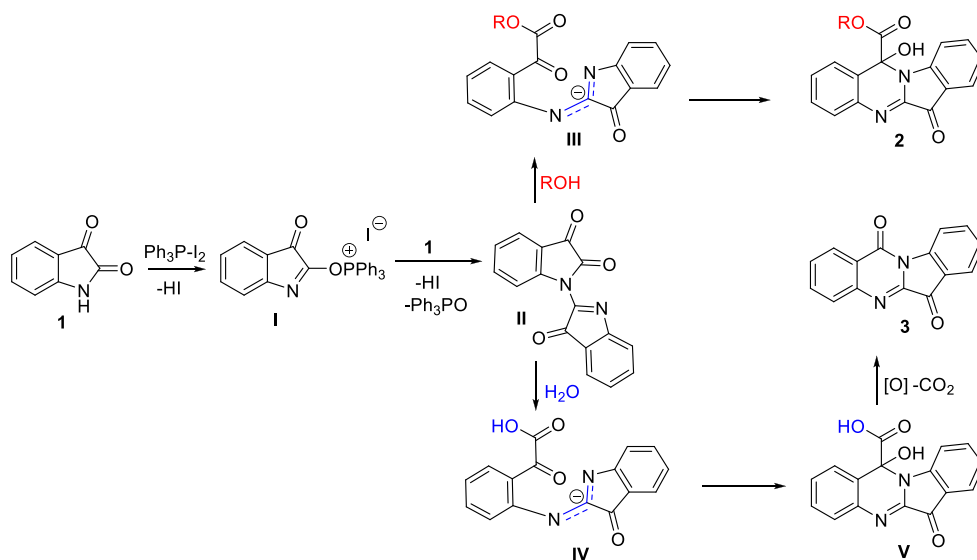
It is important to note that, although *O*-methylisatin (**A**) has been applied as a precursor toward methylisatoids,^{20b,c} under our conditions, no evidence to support its presence could be found. Moreover, based on the ³¹P{¹H} NMR study (Figure S2), triphenylphosphine oxide was the only species observed before adding alcohol, implying a self-dimerized adduct of isatin as the key intermediate. On the basis of the above observation and the previously reported literature,^{13b,20b,c} the mechanism for the formation of **2** and **3** was thus proposed as depicted in Scheme 6.

The combination of Ph₃P with I₂ provides reactive phosphonium species which react with isatin **1** to yield

Scheme 5. Control Experiments



Scheme 6. Proposed Mechanism



intermediate I. Substitution with the amide NH of another molecule of isatin provides the dimerized adduct II. An alcohol attack at the lactam carbonyl of II produces the ring-opening intermediate III which upon further cyclization gives rise to the hydroxy ester 2. In the absence of alcohol, II reacts with water, possibly from moist air, to yield IV. Oxidative decarboxylation of the cyclized intermediate V then furnishes tryptanthrin 3.

In summary, this work represents the first concise and efficient synthesis of C12-modified tryptanthrin analogs using cheap and readily available isatins as precursors. This divergent approach not only provides an alternative route toward tryptanthrin framework but also enables a facile one-pot synthesis of methylisatoid and the hydroxy ester derivatives in good yields under mild conditions. Its broad substrate scope, operational simplicity, and easy scale-up make the protocol potentially useful for construction of novel indoloquinazoline derivatives inaccessible by the currently available methods.

EXPERIMENTAL SECTION

General Information. All reagents including isatins 1a–1f were purchased from Sigma-Aldrich or TCI and used without further purification. Compounds 1g^{27a} and 1h^{27b} were synthesized according to the reported procedures. All reactions were run in flame- or oven-dried glassware under N₂ gas. The reaction was monitored by thin-layer chromatography carried out on silica gel plates (60F₂₅₄, MERCK, Germany) and visualized under UV light (254 nm). Column chromatography was performed over silica gel 60 (70–230 mesh, MERCK, Germany). Melting points were determined using Mettler Toledo DSC equipment at a heating rate of 6 °C/min and are uncorrected. NMR spectra were recorded using a Bruker AVANCE (400 and 500 MHz for ¹H). Chemical shifts were reported in parts per million (ppm, δ) downfield from tetramethylsilane (TMS). Splitting patterns are described as singlet (s), doublet (d), triplet (t), quartet (q), quintet (qui), sextet (sex), septet (sep), multiplet (m), broad (br), doublet of doublets (dd), triplet of doublets (td), and doublet of doublet of doublets (ddd). High-resolution mass spectra

(HRMS) were recorded using the Agilent 6546 LC/Q-TOF via the electrospray ionization (ESI).

General Procedure for the Synthesis of Methylisatoid Derivatives 2. To a solution of iodine (129 mg, 0.51 mmol) and triphenylphosphine (134 mg, 0.51 mmol) in freshly distilled dichloromethane (5 mL) were added isatins (0.68 mmol) at 0 °C under N₂. After addition of triethylamine (0.24 mL, 1.70 mmol) and stirring for 10 min, alcohol (0.51 mmol) was added, followed by warming up to room temperature with continuous stirring. After completion of the reaction, the crude mixture was concentrated under reduced pressure before purification by column chromatography (CC) using ethyl acetate/hexanes as the eluent.

Gram scale synthesis of **2a** was carried out as described above using **1a** (1.47 g, 10 mmol), iodine (1.9 g, 7.5 mmol), triphenylphosphine (1.97 g, 7.5 mmol), methanol (0.30 mL, 7.5 mmol), and triethylamine (3.5 mL, 25 mmol) in freshly distilled dichloromethane (50 mL) to provide **2a** in 1.16 g (75%).

Methyl 12-Hydroxy-6-oxo-6,12-dihydroindolo[2,1-b]quinazoline-12-carboxylate (Scheme 2, 2a). Orange solid (0.0932 g, 89% yield); mp 240–242 °C; *R*_f 0.34 (40% EtOAc/hexanes); ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.15 (s, 1H), 7.77 (d, *J* = 7.6 Hz, 1H), 7.74 (t, *J* = 7.6 Hz, 1H), 7.62 (d, *J* = 7.6 Hz, 1H), 7.57–7.45 (m, 3H), 7.24 (t, *J* = 7.6 Hz, 1H), 7.21 (d, *J* = 7.6 Hz, 1H), 3.68 (s, 3H); ¹³C{¹H} NMR (100 MHz, DMSO-*d*₆) δ 184.4, 169.5, 149.5, 144.0, 140.0, 138.7, 131.3, 129.5, 128.9, 126.9, 125.9, 125.3, 124.0, 120.1, 113.1, 82.5, 54.1; HRMS (ESI/QTOF) *m/z* [M + H]⁺ calcd for C₁₇H₁₃N₂O₄ 309.0870, found 309.0870.

Methyl 12-Hydroxy-2,8-dimethyl-6-oxo-6,12-dihydroindolo[2,1-b]quinazoline-12-carboxylate (Scheme 2, 2b). Yellow solid (0.093 g, 81% yield); mp 229–230 °C; *R*_f 0.36 (30% EtOAc/hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.51 (s, 1H), 7.37 (d, *J* = 8.5 Hz, 2H), 7.27 (s, 1H), 7.10 (d, *J* = 8.5 Hz, 1H), 7.01 (d, *J* = 8.5 Hz, 1H), 5.44 (s, 1H), 3.72 (s, 3H), 2.37 (s, 6H); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 183.7, 170.6, 146.9, 143.6, 139.6, 138.2, 137.3, 133.5, 131.9, 129.0, 126.3, 125.5, 123.8, 120.6, 111.8, 81.9, 54.4, 21.4, 20.8; HRMS (ESI/QTOF) *m/z* [M + H]⁺ calcd for C₁₉H₁₇N₂O₄ 337.1183, found 337.1184.

Methyl 12-Hydroxy-2,8-dimethoxy-6-oxo-6,12-dihydroindolo[2,1-b]quinazoline-12-carboxylate (Scheme 2, 2c). Red oil (0.1089 g, 87% yield); *R*_f 0.31 (30% EtOAc/hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.41 (d, *J* = 8.5 Hz, 1H), 7.21 (s, 1H), 7.14 (d, *J* = 8.5 Hz, 1H), 7.04 (d, *J* = 8.5 Hz, 1H), 6.98 (s, 1H), 6.84 (d, *J* = 8.5 Hz, 1H), 5.44 (s, 1H), 3.85 (s, 3H), 3.82 (s, 3H), 3.72 (s, 3H); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 183.6, 170.4, 160.3, 156.3, 143.2, 133.3, 130.6, 125.4, 125.0, 121.3, 117.0, 113.2, 110.7, 108.3, 82.1, 55.9, 55.8, 54.4; HRMS (ESI/QTOF) *m/z* [M + H]⁺ calcd for C₁₉H₁₇N₂O₆ 369.1081, found 369.1082.

Methyl 2,8-Dibromo-12-hydroxy-6-oxo-6,12-dihydroindolo[2,1-b]quinazoline-12-carboxylate (Scheme 2, 2d). Orange solid (0.0901 g, 57% yield); mp 259–260 °C; *R*_f 0.32 (40% EtOAc/hexanes); ¹H NMR (500 MHz, DMSO-*d*₆) δ 8.43 (s, 1H), 7.94 (d, *J* = 2.0 Hz, 1H), 7.91 (dd, *J* = 8.5, 2.0 Hz, 1H), 7.74 (dd, *J* = 8.5, 2.5 Hz, 1H), 7.69 (d, *J* = 2.5 Hz, 1H), 7.49 (d, *J* = 8.5 Hz, 1H), 7.19 (d, *J* = 8.5 Hz, 1H), 3.69 (s, 3H); ¹³C{¹H} NMR (125 MHz, DMSO-*d*₆) δ 182.9, 168.8, 148.1, 144.0, 140.7, 139.3, 134.4, 130.9, 129.5, 127.79, 127.71, 121.93, 121.90, 116.2, 115.4, 82.3, 54.4; HRMS (ESI/QTOF) *m/z* [M + H]⁺ calcd for C₁₇H₁₁⁷⁹Br₂N₂O₄ 464.9080, found 464.9076, calcd for C₁₇H₁₁⁸¹Br₂N₂O₄ 468.9040, found 468.9039.

Methyl 2,8-Dichloro-12-hydroxy-6-oxo-6,12-dihydroindolo[2,1-b]quinazoline-12-carboxylate (Scheme 2, 2e). Orange solid (0.0425 g, 33% yield); mp 257–258 °C; *R*_f 0.35 (30% EtOAc/hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.74 (d, *J* = 2.0 Hz, 1H), 7.59 (d, *J* = 2.5 Hz, 1H), 7.57–7.53 (m, 2H), 7.42 (dd, *J* = 8.5, 2.5 Hz, 1H), 7.30 (s, 1H), 7.21 (d, *J* = 8.5 Hz, 1H), 3.75 (s, 3H); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 169.1, 147.4, 138.2, 137.3, 134.9, 131.3, 130.3, 129.7, 126.3, 125.2, 121.4, 113.9, 54.3; HRMS (ESI/QTOF) *m/z* [M + H]⁺ calcd for C₁₇H₁₁³⁵Cl₂N₂O₄ 377.0090, found 377.0091, calcd for C₁₇H₁₁³⁷Cl₂N₂O₄ 381.0030, found 381.0034.

Methyl 12-Hydroxy-6-oxo-3,9-bis(trifluoromethyl)-6,12-dihydroindolo[2,1-b]quinazoline-12-carboxylate (Scheme 2, 2h). Yellow solid (0.0951 g, 63% yield); mp 230–231 °C; *R*_f 0.32 (30% EtOAc/hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.87 (d, *J* = 8.0 Hz, 1H), 7.82 (d, *J* = 8.0 Hz, 1H), 7.76 (d, *J* = 8.0 Hz, 1H), 7.71 (t, *J* = 8.0 Hz, 1H), 7.59 (t, *J* = 8.0 Hz, 1H), 7.47 (d, *J* = 8.0 Hz, 1H), 5.39 (s, 1H), 3.73 (s, 3H); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 179.6, 167.8, 149.8, 142.1, 142.0, 137.2, 134.1, 130.9, 128.7 (q, ³*J*_{CF} = 6.25 Hz), 128.0 ((q, ²*J*_{CF} = 36.25 Hz), 123.9 (q, ¹*J*_{CF} = 235.0 Hz), 122.3, 121.1 (q, ³*J*_{CF} = 6.25 Hz), 118.3, 116.8, 83.0, 54.7; HRMS (ESI/QTOF) *m/z* [M + H]⁺ calcd for C₁₉H₁₁F₆N₂O₄ 445.0618, found 445.0618.

Ethyl 12-Hydroxy-6-oxo-6,12-dihydroindolo[2,1-b]quinazoline-12-carboxylate (Scheme 2, 2i). Orange solid (0.0933 g, 85% yield); mp 240–242 °C; *R*_f 0.34 (40% EtOAc/hexanes); ¹H NMR (500 MHz, DMSO-*d*₆) δ 8.12 (s, 1H), 7.77 (d, *J* = 7.5 Hz, 1H), 7.74 (t, *J* = 7.5 Hz, 1H), 7.63 (d, *J* = 7.5 Hz, 1H), 7.55–7.51 (m, 2H), 7.48–7.44 (m, 1H), 7.24 (d, *J* = 7.5 Hz, 1H), 7.23 (t, *J* = 7.5 Hz, 1H), 4.21–4.08 (m, 2H), 1.02 (t, *J* = 7.0 Hz, 3H); ¹³C{¹H} NMR (125 MHz, DMSO-*d*₆) δ 184.5, 168.9, 149.7, 144.1, 140.1, 138.6, 131.2, 129.5, 128.9, 126.9, 126.1, 125.3, 124.0, 120.1, 113.2, 82.6, 62.9, 14.2; HRMS (ESI/QTOF) *m/z* [M + H]⁺ calcd for C₁₈H₁₅N₂O₄ 323.1026, found 323.1028.

Butyl 12-Hydroxy-6-oxo-6,12-dihydroindolo[2,1-b]quinazoline-12-carboxylate (Scheme 2, 2j). Red solid (0.0836 g, 70% yield); mp 171–173 °C; *R*_f 0.42 (30% EtOAc/hexanes); ¹H NMR (400 MHz, CDCl₃) δ 7.69 (d, *J* = 7.6 Hz, 1H), 7.58 (t, *J* = 7.6 Hz, 1H), 7.51 (dd, *J* = 7.6, 1.6 Hz, 1H), 7.43–7.40 (m, 1H), 7.33–7.21 (m, 2H), 7.18–7.14 (m, 2H), 5.84 (s, 1H), 4.20–4.06 (m, 2H), 1.41–1.34 (m, 2H), 0.99–0.89 (m, 2H), 0.65 (t, *J* = 7.2 Hz, 4H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 183.5, 169.6, 149.0, 143.9, 139.4, 137.6, 130.9, 129.1, 128.9, 126.0, 125.4, 124.2, 123.7, 120.3, 112.4, 82.0, 67.5, 30.0, 18.5, 13.3; HRMS (ESI/QTOF) *m/z* [M + H]⁺ calcd for C₂₀H₁₉N₂O₄ 351.1339, found 351.1340.

Benzyl 12-Hydroxy-6-oxo-6,12-dihydroindolo[2,1-b]quinazoline-12-carboxylate (Scheme 2, 2k). Orange solid (0.1031 g, 79% yield); mp 175–177 °C; *R*_f 0.32 (30% EtOAc/hexanes); ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.22 (s, 1H), 7.76 (d, *J* = 7.6 Hz, 1H), 7.65–7.49 (m, 5H), 7.45–7.42 (m, 1H), 7.26–7.17 (m, 5H), 7.05 (d, *J* = 7.6 Hz, 2H), 5.19 (d, *J* = 12.4 Hz, 1H), 5.13 (d, *J* = 12.4 Hz, 1H); ¹³C{¹H} NMR (100 MHz, DMSO-*d*₆) δ 189.2, 173.4, 154.3, 148.9, 144.9, 143.3, 140.3, 136.1, 134.2, 133.7, 133.5, 133.4, 132.9, 131.7, 130.7, 130.0, 128.8, 128.0, 124.8, 118.1, 117.4, 87.6, 72.7; HRMS (ESI/QTOF) *m/z* [M + H]⁺ calcd for C₂₃H₁₇N₂O₄ 385.1183, found 385.1185.

4-Methoxybenzyl 12-Hydroxy-6-oxo-6,12-dihydroindolo[2,1-b]quinazoline-12-carboxylate (Scheme 2, 2l). Orange solid (0.1211 g, 86% yield); mp 178–181 °C; *R*_f 0.37 (30% EtOAc/hexanes); ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.16 (s, 1H), 7.75 (d, *J* = 7.2 Hz, 1H), 7.64–7.53 (m, 4H), 7.45–7.41 (m, 1H), 7.21 (t, *J* = 7.2 Hz, 1H), 7.12 (d, *J* = 8.0 Hz, 1H), 7.03 (d, *J* = 8.4 Hz, 2H), 6.77 (d, *J* = 8.4 Hz, 2H), 5.14 (d, *J* = 12.0 Hz, 1H), 5.05 (d, *J* = 12.0 Hz, 1H), 3.72 (s, 3H); ¹³C{¹H} NMR (100 MHz, DMSO-*d*₆) δ 184.4, 168.7, 159.7, 149.5, 144.1, 140.1, 138.5, 131.2, 130.3, 129.4, 128.9, 127.4, 126.9, 126.0, 125.2, 123.9, 120.0, 114.1, 113.2, 82.7, 67.9, 55.5; HRMS (ESI/QTOF) *m/z* [M + H]⁺ calcd for C₂₄H₁₉N₂O₅ 415.1288, found 415.1289.

4-Isopropylbenzyl 12-Hydroxy-6-oxo-6,12-dihydroindolo[2,1-b]quinazoline-12-carboxylate (Scheme 2, 2m). Orange solid (0.1145 g, 79% yield); mp 190–192 °C; *R*_f 0.36 (30% EtOAc/hexanes); ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.19 (s, 1H), 7.75 (d, *J* = 7.2 Hz, 1H), 7.61 (d, *J* = 7.6 Hz, 2H), 7.54 (d, *J* = 2.8 Hz, 2H), 7.46–7.42 (m, 1H), 7.21 (t, *J* = 7.2 Hz, 1H), 7.15 (d, *J* = 8.0 Hz, 1H), 7.09 (d, *J* = 8.0 Hz, 2H), 6.99 (d, *J* = 8.0 Hz, 2H), 5.17 (d, *J* = 12.4 Hz, 1H), 5.08 (d, *J* = 12.4 Hz, 1H), 2.83 (sep, *J* = 7.2 Hz, 1H), 1.16 (d, *J* = 7.2 Hz, 6H); ¹³C{¹H} NMR (100 MHz, DMSO-*d*₆) δ 184.4, 168.7, 149.6, 149.0, 144.1, 140.1, 138.5, 132.9, 131.3, 129.5, 128.9, 128.4, 127.0, 126.7, 126.0, 125.2, 124.0, 120.1, 113.3, 82.8, 68.0, 33.6, 24.3, 24.2; HRMS (ESI/QTOF) *m/z* [M + H]⁺ calcd for C₂₆H₂₃N₂O₄ 427.1652, found 427.1654.

4-Chlorobenzyl 12-Hydroxy-6-oxo-6,12-dihydroindolo[2,1-*b*]quinazoline-12-carboxylate (Scheme 2, 2n). Orange solid (0.0821 g, 75% yield); mp 212–215 °C; R_f 0.34 (30% EtOAc/hexanes); ^1H NMR (400 MHz, DMSO- d_6) δ 8.23 (s, 1H), 7.75 (dd, J = 8.0, 1.2 Hz, 1H), 7.62 (t, J = 8.0 Hz, 1H), 7.61 (d, J = 7.6 Hz, 1H), 7.54 (d, J = 7.6 Hz, 2H), 7.47–7.42 (m, 1H), 7.27 (d, J = 8.4 Hz, 2H), 7.22 (t, J = 7.6 Hz, 1H), 7.15 (d, J = 8.0 Hz, 1H), 7.09 (d, J = 8.4 Hz, 2H), 5.21 (d, J = 12.4 Hz, 1H), 5.12 (d, J = 12.4 Hz, 1H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, DMSO- d_6) δ 184.4, 168.6, 149.5, 144.1, 140.1, 138.5, 134.5, 133.4, 131.3, 130.2, 129.5, 128.9, 128.8, 126.9, 125.9, 125.3, 124.0, 120.1, 113.2, 82.8, 67.2; HRMS (ESI/QTOF) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{23}\text{H}_{16}^{35}\text{ClN}_2\text{O}_4$ 419.0793, found 419.0793, calcd for $\text{C}_{23}\text{H}_{16}^{37}\text{ClN}_2\text{O}_4$ 421.0763, found 421.0766.

4-Nitrobenzyl 12-Hydroxy-6-oxo-6,12-dihydroindolo[2,1-*b*]quinazoline-12-carboxylate (Scheme 2, 2o). Orange solid (0.0817 g, 56% yield); mp 205–207 °C; R_f 0.30 (40% EtOAc/hexanes); ^1H NMR (400 MHz, DMSO- d_6) δ 8.30 (s, 1H), 8.06 (d, J = 8.0 Hz, 2H), 7.77 (d, J = 7.5 Hz, 1H), 7.64 (t, J = 7.5 Hz, 1H), 7.63 (d, J = 7.5 Hz, 1H), 7.56 (s, 2H), 7.49–7.46 (m, 1H), 7.32 (d, J = 8.0 Hz, 2H), 7.22 (t, J = 7.5 Hz, 1H), 7.18 (d, J = 7.5 Hz, 1H), 5.35 (d, J = 13.5 Hz, 1H), 5.27 (d, J = 13.5 Hz, 1H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, DMSO- d_6) δ 184.3, 168.5, 149.5, 147.6, 144.2, 143.1, 140.1, 138.5, 131.4, 129.6, 129.1, 129.0, 127.0, 125.8, 125.3, 124.0, 123.8, 120.1, 113.3, 82.8, 66.7; HRMS (ESI/QTOF) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{23}\text{H}_{16}\text{N}_3\text{O}_6$ 430.1034, found 430.1034.

3-Phenoxybenzyl 12-Hydroxy-6-oxo-6,12-dihydroindolo[2,1-*b*]quinazoline-12-carboxylate (Scheme 2, 2p). Orange oil (0.1228 g, 76% yield); R_f 0.36 (30% EtOAc/hexanes); ^1H NMR (400 MHz, DMSO- d_6) δ 8.25 (s, 1H), 7.73 (d, J = 7.2 Hz, 1H), 7.64–7.56 (m, 2H), 7.55–7.50 (m, 2H), 7.41–7.35 (m, 3H), 7.24 (t, J = 8.0 Hz, 1H), 7.21–7.13 (m, 3H), 6.93–6.84 (m, 3H), 6.73 (s, 1H), 5.19 (d, J = 12.8 Hz, 1H), 5.12 (d, J = 12.8 Hz, 1H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, DMSO- d_6) δ 184.4, 168.6, 156.9, 156.8, 149.5, 144.1, 140.1, 138.9, 138.5, 137.7, 131.3, 130, 129.4, 128.9, 126.9, 125.8, 125.3, 124.0, 123.2, 120.1, 119.0, 118.9, 118.4, 113.2, 82.8, 67.5; HRMS (ESI/QTOF) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{29}\text{H}_{21}\text{N}_2\text{O}_5$ 477.1445, found 477.1446.

Cinnamyl 12-Hydroxy-6-oxo-6,12-dihydroindolo[2,1-*b*]quinazoline-12-carboxylate (Scheme 2, 2q). Yellow solid (0.0921 g, 66% yield); mp 197–198 °C; R_f 0.30 (30% EtOAc/hexanes); ^1H NMR (400 MHz, DMSO- d_6) δ 8.23 (s, 1H), 7.77 (d, J = 7.5 Hz, 1H), 7.68 (t, J = 7.0 Hz, 2H), 7.58–7.53 (m, 2H), 7.49–7.45 (m, 1H), 7.34–7.26 (m, 6H), 7.21 (t, J = 7.5 Hz, 1H), 6.37 (d, J = 16.0 Hz, 1H), 6.18 (dt, J = 16.0, 5.8 Hz, 1H), 4.80 (m, 2H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, DMSO- d_6) δ 184.5, 168.6, 149.6, 144.1, 140.1, 138.5, 136.0, 133.5, 131.3, 129.5, 129.1, 129.0, 128.6, 127.0, 126.9, 126.0, 125.3, 124.0, 123.0, 120.1, 113.3, 82.7, 66.6; HRMS (ESI/QTOF) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{25}\text{H}_{19}\text{N}_2\text{O}_4$ 411.1339, found 411.1335.

Cyclohexyl 12-Hydroxy-6-oxo-6,12-dihydroindolo[2,1-*b*]quinazoline-12-carboxylate (Scheme 2, 2r). Orange oil (0.0832 g, 65% yield); R_f 0.36 (30% EtOAc/hexanes); ^1H NMR (500 MHz, CDCl_3) δ 7.76 (d, J = 7.5 Hz, 1H), 7.59 (dd, J = 7.5, 1.5 Hz, 1H), 7.57 (td, J = 7.5, 1.5 Hz, 1H), 7.46 (dd, J = 7.5, 1.5 Hz, 1H), 7.39 (td, J = 7.5, 1.5 Hz, 1H), 7.34 (td, J = 7.5, 1.5 Hz, 1H), 7.17 (t, J = 7.5 Hz, 1H), 7.15 (d, J = 7.5 Hz, 1H), 4.87–4.82 (m, 1H), 1.60–1.52 (m, 2H), 1.38–1.33 (m, 3H), 1.25–1.09 (m, 5H); ^{13}C NMR (125 MHz, CDCl_3) δ 183.9, 169.5, 149.1, 144.0, 139.7, 137.5, 130.9, 129.3, 129.1, 125.52, 125.49, 124.4, 123.8, 120.5, 112.2, 82.0, 76.9, 30.8, 30.6, 24.8, 22.9; HRMS (ESI/QTOF) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{22}\text{H}_{21}\text{N}_2\text{O}_4$ 377.1496, found 377.1498.

Isopropyl 12-Hydroxy-6-oxo-6,12-dihydroindolo[2,1-*b*]quinazoline-12-carboxylate (Scheme 2, 2s). Orange solid (0.0607 g, 53% yield); mp 205–207 °C; R_f 0.34 (30% EtOAc/hexanes); ^1H NMR (500 MHz, CDCl_3) δ 7.74 (d, J = 7.5 Hz, 1H), 7.59–7.54 (m, 2H), 7.49–7.45 (m, 1H), 7.37–7.31 (m, 2H), 7.16 (t, J = 7.5 Hz, 1H), 7.14 (d, J = 7.5 Hz, 1H), 5.06 (sep, J = 6.5 Hz, 1H), 1.06 (d, J = 6.5 Hz, 2H), 1.01 (d, J = 6.5 Hz, 2H); $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3) δ 183.8, 169.5, 149.0, 143.9, 139.6, 137.5, 130.9, 129.2, 129.1, 125.6, 125.5, 124.3, 123.7, 120.4, 112.2, 81.9, 72.4, 21.3, 21.1; HRMS

(ESI/QTOF) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{19}\text{H}_{17}\text{N}_2\text{O}_4$ 337.1183, found 337.1184.

Quinolín-8-yl 12-Hydroxy-6-oxo-6,12-dihydroindolo[2,1-*b*]quinazoline-12-carboxylate (Scheme 2, 2u). Orange solid (0.0818 g, 57% yield); mp 236–237 °C; R_f 0.35 (30% EtOAc/hexanes); ^1H NMR (500 MHz, CDCl_3) δ 9.16 (dd, J = 4.2, 1.5 Hz, 1H), 8.30 (dd, J = 8.5, 1.5 Hz, 1H), 7.80 (ddd, J = 7.5, 1.2, 1.0 Hz, 1H), 7.75 (d, J = 7.5, 1H), 7.74 (d, J = 8.5 Hz, 1H), 7.66 (dd, J = 8.5, 4.5 Hz, 1H), 7.44–7.40 (m, 2H), 7.29–7.25 (m, 2H), 7.18 (td, J = 7.5, 1.2 Hz, 1H), 7.08 (td, J = 7.5, 1.0 Hz, 1H), 6.74 (dd, J = 8.0, 1.5 Hz, 1H), 6.27 (d, J = 8.0 Hz, 1H); $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3) δ 183.2, 172.0, 152.3, 148.7, 148.4, 144.3, 139.0, 137.9, 136.7, 134.9, 131.0, 130.8, 130.6, 129.8, 126.6, 126.4, 125.9, 125.7, 123.8, 123.4, 121.3, 120.7, 110.3, 65.4; HRMS (ESI/QTOF) m/z $[\text{M} - \text{OH}]^+$ calcd for $\text{C}_{25}\text{H}_{14}\text{N}_3\text{O}_3$ 404.1030, found 404.1030.

4-Allyl-2-methoxyphenyl 12-Hydroxy-6-oxo-6,12-dihydroindolo[2,1-*b*]quinazoline-12-carboxylate (Scheme 2, 2v). Orange solid (0.0466 g, 31% yield); mp 236–237 °C; R_f 0.42 (30% EtOAc/hexanes); ^1H NMR (500 MHz, DMSO- d_6) δ 8.32 (s, 1H), 7.84 (td, J = 8.0, 2.0 Hz, 1H), 7.81–7.78 (m, 2H), 7.61–7.54 (m, 3H), 7.44 (d, J = 8.0 Hz, 1H), 7.28 (t, J = 8.0 Hz, 1H), 6.90 (d, J = 2.0 Hz, 1H), 6.79 (d, J = 8.0 Hz, 1H), 6.72 (dd, J = 8.0, 2.0 Hz, 1H), 5.97–5.89 (m, 1H), 5.09–5.03 (m, 2H), 3.52 (s, 3H), 3.33 (d, J = 7.0 Hz, 2H); $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, DMSO- d_6) δ 184.4, 167.1, 150.7, 149.5, 144.0, 140.1, 138.4, 137.7, 137.4, 131.4, 129.5, 129.0, 127.3, 125.7, 125.2, 124.1, 122.1, 120.8, 120.2, 116.6, 113.8, 113.6, 82.7, 55.9, 40.3; HRMS (ESI/QTOF) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{26}\text{H}_{21}\text{N}_2\text{O}_5$ 441.1445, found 441.1446.

(*E*)-3,7-Dimethylocta-2,6-dien-1-yl 12-Hydroxy-6-oxo-6,12-dihydroindolo[2,1-*b*]quinazoline-12-carboxylate (Scheme 2, 2w). Yellow solid (0.1037 g, 71% yield); mp 104–105 °C; R_f 0.32 (30% EtOAc/hexanes); ^1H NMR (500 MHz, CDCl_3) δ 7.74 (dd, J = 7.5, 1.2 Hz, 1H), 7.57–7.54 (m, 2H), 7.46–7.44 (m, 1H), 7.37–7.31 (m, 2H), 7.16 (td, J = 7.5, 1.2 Hz, 1H), 7.11 (d, J = 8.0 Hz, 1H), 5.39 (s, 1H), 5.09–5.06 (m, 1H), 4.99–4.95 (m, 1H), 4.69 (dd, J = 12.0, 7.2 Hz, 1H), 4.58 (dd, J = 12.0, 7.2 Hz, 1H), 1.94–1.89 (m, 2H), 1.87–1.83 (m, 2H), 1.65 (d, J = 1.0 Hz, 3H), 1.55 (s, 3H), 1.45 (d, J = 1.0 Hz, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3) δ 183.7, 170.0, 148.9, 145.3, 143.8, 139.6, 137.4, 132.0, 130.9, 129.2, 129.0, 125.8, 125.4, 124.2, 123.7, 123.4, 120.4, 116.2, 112.3, 81.9, 64.5, 39.3, 26.1, 25.7, 17.7, 16.4; HRMS (ESI/QTOF) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{26}\text{H}_{27}\text{N}_2\text{O}_4$ 431.1965, found 431.1966.

General Procedure for the Synthesis of Tryptanthrins 3. To a solution of iodine (129 mg, 0.51 mmol) and triphenylphosphine (134 mg, 0.51 mmol) in freshly distilled dichloromethane (5 mL) were added isatins (0.68 mmol), followed by triethylamine (0.24 mL, 1.70 mmol) at 0 °C under N_2 . The reaction mixture was allowed to warm up to room temperature with continuous stirring. After completion of the reaction, the crude mixture was concentrated under reduced pressure before purification by column chromatography (CC) using ethyl acetate/hexanes as the eluent. Gram scale synthesis of **3a** was carried out as described above using **1a** (1.47 g, 10 mmol), iodine (1.9 g, 7.5 mmol), triphenylphosphine (1.97 g, 7.5 mmol), and triethylamine (3.5 mL, 25 mmol) in freshly distilled dichloromethane (50 mL) to yield **3a** in 0.8815 g (71%).

Indolo[2,1-*b*]quinazoline-6,12-dione (Tryptanthrin) (Scheme 3, 3a). Yellow solid (0.0667 g, 79% yield); mp 259–260 °C (lit.^{14a} mp 260–262 °C); R_f 0.36 (20% EtOAc/hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.62 (d, J = 8.5 Hz, 1H), 8.44 (d, J = 8.0 Hz, 1H), 8.03 (d, J = 8.0 Hz, 1H), 7.92 (d, J = 7.5 Hz, 1H), 7.85 (t, J = 7.5 Hz, 1H), 7.79 (t, J = 8.0 Hz, 1H), 7.67 (t, J = 7.5 Hz, 1H), 7.43 (t, J = 7.5 Hz, 1H); $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3) δ 182.6, 158.1, 146.6, 146.4, 144.3, 138.3, 135.2, 130.7, 130.3, 127.6, 127.2, 125.4, 123.8, 121.9, 118.0.

2,8-Dimethylindolo[2,1-*b*]quinazoline-6,12-dione (Scheme 3, 3b). Yellow solid (0.0752 g, 80% yield); mp 248–250 °C (lit.^{11b} mp 249.8–251.6 °C); R_f 0.40 (20% EtOAc/hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.48 (d, J = 8.0 Hz, 1H), 8.21 (d, J = 2.0 Hz, 1H), 7.90 (d, J = 8.0 Hz, 1H), 7.69 (d, J = 2.0 Hz, 1H), 7.64 (dd, J = 8.0,

2.0 Hz, 1H), 7.57 (dd, $J = 8.0$, 2.0 Hz, 1H), 2.55 (s, 3H), 2.45 (s, 3H).

2,8-Dimethoxyindolo[2,1-*b*]quinazoline-6,12-dione (Scheme 3, 3c). Yellow solid (0.0883 g, 84% yield); mp 284–285 °C (lit.¹⁸ mp 285–288 °C); R_f 0.39 (30% EtOAc/hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.51 (d, $J = 9.0$ Hz, 1H), 7.93 (d, $J = 9.0$ Hz, 1H), 7.81 (d, $J = 3.0$ Hz, 1H), 7.38 (dd, $J = 9.0$, 3.0 Hz, 1H), 7.37 (d, $J = 3.0$ Hz, 1H), 7.30 (dd, $J = 9.0$, 3.0 Hz, 1H), 3.99 (s, 3H), 3.89 (s, 3H).

2,8-Dibromoindolo[2,1-*b*]quinazoline-6,12-dione (Scheme 3, 3d). Yellow solid (0.0762 g, 55% yield); mp 317–319 °C (lit.^{13b} mp 319–320 °C); R_f 0.39 (20% EtOAc/hexanes); ^1H NMR (500 MHz, $\text{CDCl}_3 + \text{CD}_3\text{OD}$ 4 drops) δ 8.50 (d, $J = 2.5$ Hz, 1H), 8.45 (d, $J = 8.5$ Hz, 1H), 7.98 (d, $J = 2.0$ Hz, 1H), 7.91 (dd, $J = 8.5$, 2.5 Hz, 1H), 7.87 (dd, $J = 8.5$, 2.0 Hz, 1H), 7.83 (d, $J = 8.5$ Hz, 1H).

2,8-Dichloroindolo[2,1-*b*]quinazoline-6,12-dione (Scheme 3, 3e). Yellow solid (0.0432 g, 40% yield); mp 288–290 °C (lit.¹⁸ mp 289–291 °C); R_f 0.39 (20% EtOAc/hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.53 (d, $J = 8.5$ Hz, 1H), 8.35 (d, $J = 2.5$ Hz, 1H), 7.93 (d, $J = 8.5$ Hz, 1H), 7.84 (d, $J = 2.0$ Hz, 1H), 7.78 (dd, $J = 8.5$, 2.5 Hz, 1H), 7.73 (dd, $J = 8.5$, 2.0 Hz, 1H).

3,9-Bis(trifluoromethyl)indolo[2,1-*b*]quinazoline-6,12-dione (Scheme 3, 3h). Orange solid (0.0744 g, 57% yield); mp 240–242 °C; R_f 0.34 (40% EtOAc/hexanes); ^1H NMR (500 MHz, CDCl_3) δ 9.03 (d, $J = 8.0$ Hz, 1H), 8.27 (d, $J = 8.0$ Hz, 1H), 8.10 (d, $J = 8.0$ Hz, 1H), 7.96 (t, $J = 8.0$ Hz, 1H), 7.92 (t, $J = 8.0$ Hz, 1H), 7.74 (d, $J = 8.0$ Hz, 1H); $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3) δ 179.6, 167.8, 149.8, 142.08, 142.02, 137.2, 134.1, 130.9, 128.7 (q, $^3J_{\text{CF}} = 6.25$ Hz), 128.0 (q, $^2J_{\text{CF}} = 33.75$ Hz), 122.9 (q, $^1J_{\text{CF}} = 233.75$ Hz), 122.3, 121.1 (q, $^3J_{\text{CF}} = 6.25$ Hz), 118.3, 116.8, 83.0, 54.7; HRMS (ESI/QTOF) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{17}\text{H}_7\text{F}_6\text{N}_2\text{O}_2$ 385.0406, found 385.0407.

Synthesis of Methyl 12-Methoxy-6-oxo-6,12-dihydroindolo[2,1-*b*]quinazoline-12-carboxylate (Cephalanthrine B, Scheme 4, 4). To a solution of **2a** (105 mg, 0.34 mmol) in anhydrous DMF (3 mL) were added iodomethane (42 μL , 0.68 mmol) and potassium carbonate (94 mg, 0.68 mmol) at 0 °C. After warming up to room temperature, the reaction mixture was stirred for 2 h. The crude mixture was filtered washed with ethyl acetate. The organic layer was concentrated under reduced pressure before purification by column chromatography (CC) using ethyl acetate/hexanes as the eluent to provide compound **4** as a yellow solid (0.0898 g, 82% yield); mp 214–216 °C (lit.^{1b} mp 215–217 °C); R_f 0.41 (20% EtOAc/hexanes); ^1H NMR (500 MHz, CDCl_3) δ 7.83 (d, $J = 7.5$ Hz, 1H), 7.72 (d, $J = 8.0$ Hz, 1H), 7.63–7.59 (m, 2H), 7.52 (t, $J = 7.5$ Hz, 1H), 7.42 (t, $J = 7.5$ Hz, 1H), 7.24 (d, $J = 8.5$ Hz, 1H), 7.21 (t, $J = 7.5$ Hz, 1H), 3.71 (s, 3H), 3.07 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3) δ 184.0, 167.8, 148.9, 144.7, 141.9, 138.0, 131.5, 129.7, 129.4, 126.1, 125.5, 124.0, 120.5, 120.3, 112.3, 87.4, 53.8, 50.6.

■ ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.joc.0c02403>.

Copies of ^1H NMR, $^{13}\text{C}\{^1\text{H}\}$ NMR spectra of all new products, optimization data, and $^{31}\text{P}\{^1\text{H}\}$ NMR reaction monitoring study (PDF)

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Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

This work is partially supported by Chiang Mai University, Thailand and The Thailand Research Fund through the Royal Golden Jubilee Ph.D. Program (Grant No. PHD/0072/2559 to N.W.).

■ REFERENCES

- (1) (a) Jao, C.-W.; Lin, W.-C.; Wu, Y.-T.; Wu, P.-L. Isolation, Structure Elucidation, and Synthesis of Cytotoxic Tryptanthrin Analogues from *Phaius mishmensis*. *J. Nat. Prod.* **2008**, *71*, 1275. (b) Chang, C.-F.; Hsu, Y.-L.; Lee, C.-Y.; Wu, C.-H.; Wu, Y.-C.; Chuang, T.-H. Isolation and cytotoxicity evaluation of the chemical constituents from *Cephalanthropsis gracilis*. *Int. J. Mol. Sci.* **2015**, *16*, 3980. (c) Jao, C.-W.; Hung, T.-H.; Chang, C.-F.; Chuang, T.-H. Chemical constituents of *Phaius mishmensis*. *Molecules* **2016**, *21*, 1605. (d) Kaur, R.; Manjal, S. K.; Rawal, R. K.; Kumar, K. Recent synthetic and medicinal perspectives of tryptanthrin. *Bioorg. Med. Chem.* **2017**, *25*, 4533.
- (2) (a) Michael, J. P. Quinoline, quinazoline, and acridone alkaloids. *Nat. Prod. Rep.* **2007**, *24*, 223. (b) Jahng, Y. Progress in the studies on tryptanthrin, an alkaloid of history. *Arch. Pharmacol. Res.* **2013**, *36*, 517.
- (3) (a) Yang, S.; Li, X.; Hu, F.; Li, Y.; Yang, Y.; Yan, J.; Kuang, C.; Yang, Q. Discovery of Tryptanthrin Derivatives as Potent Inhibitors of Indoleamine 2,3-Dioxygenase with Therapeutic Activity in Lewis Lung Cancer (LLC) Tumor-Bearing Mice. *J. Med. Chem.* **2013**, *56*, 8321. (b) Yang, D.; Zhang, S.; Fang, X.; Guo, L.; Hu, N.; Guo, Z.; Li, X.; Yang, S.; He, J. C.; Kuang, C.; Yang, Q. *N*-Benzyl/Aryl Substituted Tryptanthrin as Dual Inhibitors of Indoleamine 2,3-Dioxygenase and Tryptophan 2,3-Dioxygenase. *J. Med. Chem.* **2019**, *62*, 9161. (c) Jun, K.-Y.; Park, S.-E.; Liang, J. L.; Jahng, Y.; Kwon, Y. Benzo[*b*]tryptanthrin Inhibits MDRI, Topoisomerase Activity, and Reverses Adriamycin Resistance in Breast Cancer Cells. *ChemMedChem* **2015**, *10*, 827.
- (4) (a) Krivogorsky, B.; Grundt, P.; Yolken, R.; Jones-Brando, L. Inhibition of *Toxoplasma gondii* by indirubin and tryptanthrin analogs. *Antimicrob. Agents Chemother.* **2008**, *52*, 4466. (b) Scovill, J.; Blank, E.; Konnick, M.; Nenortas, E.; Shapiro, T. Antitrypanosomal activities of tryptanthrins. *Antimicrob. Agents Chemother.* **2002**, *46*, 882.
- (5) (a) Bandekar, P. P.; Roopnarine, K. A.; Parekh, V. J.; Mitchell, T. R.; Novak, M. J.; Sinden, R. R. Antimicrobial Activity of Tryptanthrins in *Escherichia coli*. *J. Med. Chem.* **2010**, *53*, 3558. (b) Kamal, A.; Reddy, B. V. S.; Sridevi, B.; Ravikumar, A.; Venkateswarlu, A.; Sravanthi, G.; Sridevi, J. P.; Yogeewari, P.; Sriram, D. Synthesis and biological evaluation of phaitanthrin congeners as anti-mycobacterial agents. *Bioorg. Med. Chem. Lett.* **2015**, *25*, 3867.
- (6) (a) Schepetkin, I. A.; Khlebnikov, A. I.; Potapov, A. S.; Kovrizhina, A. R.; Matveevskaya, V. V.; Belyanin, M. L.; Atochin, D. N.; Zanoza, S. O.; Gaidarzhy, N. M.; Lyakhov, S. A.; Kirpotina, L. N.; Quinn, M. T. Synthesis, biological evaluation, and molecular modeling of 11H-indeno[1,2-*b*]quinoxalin-11-one derivatives and

- tryptanthrin-6-oxime as c-Jun N-terminal kinase inhibitors. *Eur. J. Med. Chem.* **2019**, *161*, 179. (b) Lee, S.; Kim, D.-C.; Baek, H. Y.; Lee, K.-D.; Kim, Y.-C.; Oh, H. Anti-neuroinflammatory effects of tryptanthrin from *Polygonum tinctorium* Lour. in lipopolysaccharide-stimulated BV2 microglial cells. *Arch. Pharmacol. Res.* **2018**, *41*, 419. (c) Hamburger, M. Isatis tinctoria - from the rediscovery of an ancient medicinal plant towards a novel anti-inflammatory phytopharmaceutical. *Phytochem. Rev.* **2002**, *1*, 333.
- (7) Bhattacharjee, A. K.; Hartell, M. G.; Nichols, D. A.; Hicks, R. P.; Stanton, B.; Van Hamont, J. E.; Milhous, W. K. Structure-activity relationship study of antimalarial indolo[2,1-*b*]quinazoline-6,12-diones (tryptanthrins). Three dimensional pharmacophore modeling and identification of new antimalarial candidates. *Eur. J. Med. Chem.* **2004**, *39*, 59.
- (8) Mani, J. S.; Johnson, J. B.; Steel, J. C.; Broszczak, D. A.; Neilsen, P. M.; Walsh, K. B.; Naiker, M. Natural product-derived phytochemicals as potential agents against coronaviruses: A review. *Virus Res.* **2020**, *284*, 197989.
- (9) (a) Hwang, J.-M.; Oh, T.; Kaneko, T.; Upton, A. M.; Franzblau, S. G.; Ma, Z.; Cho, S.-N.; Kim, P. Design, Synthesis, and Structure-Activity Relationship Studies of Tryptanthrins As Antitubercular Agents. *J. Nat. Prod.* **2013**, *76*, 354. (b) Mitscher, L. A.; Baker, W. Tuberculosis: a search for novel therapy starting with natural products. *Med. Res. Rev.* **1998**, *18*, 363. (c) Mitscher, L. A.; Baker, W. R. A search for novel chemotherapy against tuberculosis amongst natural products. *Pure Appl. Chem.* **1998**, *70*, 365.
- (10) Tucker, A. M.; Grundt, P. The chemistry of tryptanthrin and its derivatives. *ARKIVOC* **2012**, *2012*, 546.
- (11) (a) Sultan, S.; Gupta, V.; Shah, B. A. Photoredox-Catalyzed Isatin Reactions: Access to Dibenzo-1,7-Naphthyridine Carboxylate and Tryptanthrin. *ChemPhotoChem.* **2017**, *1*, 120. (b) Hou, H.; Li, H.; Han, Y.; Yan, C. Synthesis of visible-light mediated tryptanthrin derivatives from isatin and isatoic anhydride under transition metal-free conditions. *Org. Chem. Front.* **2018**, *5*, 51.
- (12) Batanero, B.; Barba, F. Electrosynthesis of tryptanthrin. *Tetrahedron Lett.* **2006**, *47*, 8201.
- (13) (a) Moskovkina, T. V. New synthesis of 6,12-dihydro-6,12-dioxindolo[2,1-*b*]quinazoline (tryptanthrine, couropitine A). *Russ. J. Org. Chem.* **1997**, *33*, 125. (b) Moskovkina, T. V.; Kalinovskii, A. I.; Makhankov, V. V. Synthesis of tryptanthrin (couropitine) derivatives by reaction of substituted isatins with phosphoryl chloride. *Russ. J. Org. Chem.* **2012**, *48*, 123.
- (14) (a) Amara, R.; Awad, H.; Chaker, D.; Bentabed-Ababsa, G.; Lassagne, F.; Erb, W.; Chevallier, F.; Roisnel, T.; Dorcet, V.; Fajloun, Z.; Vidal, J.; Mongin, F. Conversion of Isatins to Tryptanthrins, Heterocycles Endowed with a Myriad of Bioactivities. *Eur. J. Org. Chem.* **2019**, *2019*, 5302. (b) Brandao, P.; Pinheiro, D.; Sergio Seixas de Melo, J.; Pineiro, M. I₂/NaH/DMF as oxidant trio for the synthesis of tryptanthrin from indigo or isatin. *Dyes Pigm.* **2020**, *173*, 107935. (c) Moskovkina, T. V.; Denisenko, M. V.; Kalinovskii, A. I.; Stonik, V. A. Synthesis of substituted tryptanthrins via oxidation of isatin and its derivatives. *Russ. J. Org. Chem.* **2013**, *49*, 1740. (d) Jia, F.-C.; Zhou, Z.-W.; Xu, C.; Wu, Y.-D.; Wu, A.-X. Divergent Synthesis of Quinazolin-4(3*H*)-ones and Tryptanthrins Enabled by a *tert*-Butyl Hydroperoxide/K₃PO₄-Promoted Oxidative Cyclization of Isatins at Room Temperature. *Org. Lett.* **2016**, *18*, 2942.
- (15) (a) Mane, A. H.; Patil, A. D.; Kamat, S. R.; Salunkhe, R. S. Biocatalyst Mediated Synthesis of Tryptanthrins Performed Under Ultrasonication. *ChemistrySelect* **2018**, *3*, 6454. (b) Kumar, A.; Tripathi, V. D.; Kumar, P. β -Cyclodextrin catalyzed synthesis of tryptanthrin in water. *Green Chem.* **2011**, *13*, 51. (c) Rai, B.; Shukla, R. D.; Kumar, A. Zinc oxide-NP catalyzed direct indolation of *in situ* generated bioactive tryptanthrin. *Green Chem.* **2018**, *20*, 822.
- (16) Reddy, B. V. S.; Reddy, D. M.; Reddy, G. N.; Reddy, M. R.; Reddy, V. K. Domino Oxidative Cyclization of 2-Amino-acetophenones for the One-Pot Synthesis of Tryptanthrin Derivatives. *Eur. J. Org. Chem.* **2015**, *2015*, 8018.
- (17) Li, X.; Huang, H.; Yu, C.; Zhang, Y.; Li, H.; Wang, W. Synthesis of Tryptanthrins by Organocatalytic and Substrate Co-catalyzed Photochemical Condensation of Indoles and Anthranilic Acids with Visible Light and O₂. *Org. Lett.* **2016**, *18*, 5744.
- (18) Wang, C.; Zhang, L.; Ren, A.; Lu, P.; Wang, Y. Cu-Catalyzed Synthesis of Tryptanthrin Derivatives from Substituted Indoles. *Org. Lett.* **2013**, *15*, 2982.
- (19) (a) Gahtory, D.; Chouhan, M.; Sharma, R.; Nair, V. A. Total Synthesis of a Pyrroloindoloquinazoline Alkaloid. *Org. Lett.* **2013**, *15*, 3942. (b) Gao, H.; Luo, Z.; Ge, P.; He, J.; Zhou, F.; Zheng, P.; Jiang, J. Direct Catalytic Asymmetric Synthesis of β -Hydroxy Acids from Malonic Acid. *Org. Lett.* **2015**, *17*, 5962. (c) Kang, G.; Luo, Z.; Liu, C.; Gao, H.; Wu, Q.; Wu, H.; Jiang, J. Amino Acid Salts Catalyzed Asymmetric Aldol Reaction of Tryptanthrin: A Straightforward Synthesis of Phaitanthrin A and Its Derivatives. *Org. Lett.* **2013**, *15*, 4738. (d) Liao, H.; Peng, X.; Hu, D.; Xu, X.; Huang, P.; Liu, Q.; Liu, L. CoCl₂-promoted TEMPO oxidative homocoupling of indoles: access to tryptanthrin derivatives. *Org. Biomol. Chem.* **2018**, *16*, 5699. (e) Vaidya, S. D.; Argade, N. P. Aryne Insertion Reactions Leading to Bioactive Fused Quinazolinones: Diastereoselective Total Synthesis of Cruciferane. *Org. Lett.* **2013**, *15*, 4006. (f) Yavari, I.; Solgi, R.; Khajeh-Khezri, A.; Askarian-Amiri, M.; Halvagar, M. R. Synthesis of spiroindolo[2,1-*b*]quinazolines from Huisgen zwitterions and tryptanthrin-malononitrile adducts. *J. Heterocycl. Chem.* **2019**, *56*, 3396.
- (20) (a) Bird, C. W. Structure of methylisatoid. *Tetrahedron* **1963**, *19*, 901. (b) Cornforth, J. W. Structures of isamic acid and methylisatoid. *J. Chem. Soc., Perkin Trans. 1* **1976**, 2004. (c) Cornforth, S. J.; Hitchcock, P. B.; Rozos, P. Isatin chloride: a phantom. Reactions of 2-(2,2-dichloro-2,3-dihydro-3-oxindol-1-yl)-3*H*-indol-3-one. *J. Chem. Soc., Perkin Trans. 1* **1996**, 2787. (d) Kimura, N.; Fujita, M. Novel Indoloquinazoline compound and method for producing same, WO2018123467A1, 2018.
- (21) (a) Pattarawarapan, M.; Wiriya, N.; Yimklan, S.; Wangngae, S.; Phakhodee, W. Zwitterionic Ring-Opened Oxyphosphonium Species from the Ph₃P-I₂ Mediated Reactions of Benzo[d]oxazol-2(3*H*)-ones with Secondary Amines. *J. Org. Chem.* **2020**, *85*, 6151. (b) Pattarawarapan, M.; Yamano, D.; Wiriya, N.; Phakhodee, W. Metal-Free Synthesis of 2-*N,N*-Dialkylaminobenzoxazoles Using Tertiary Amines as the Nitrogen Source. *J. Org. Chem.* **2019**, *84*, 6516. (c) Phakhodee, W.; Wangngae, S.; Pattarawarapan, M. Approach to the Synthesis of 2,3-Disubstituted-3*H*-quinazolin-4-ones Mediated by Ph₃P-I₂. *J. Org. Chem.* **2017**, *82*, 8058. (d) Wangngae, S.; Pattarawarapan, M.; Phakhodee, W. Ph₃P/I₂-Mediated Synthesis of *N,N,N'*-Substituted Guanidines and 2-Iminoimidazolin-4-ones from Aryl Isothiocyanates. *J. Org. Chem.* **2017**, *82*, 10331.
- (22) (a) Ault, R. G.; Hirst, E. L.; Morton, R. A. Absorption spectra in relation to the constitution of derivatives of isatin and carbostyryl. *J. Chem. Soc.* **1935**, 1653. (b) Kennewell, P. D.; Miller, D. J.; Scowston, R. M.; Westwood, R. Reactions of diazo compounds with isatin and its derivatives. *J. Chem. Res., Synop.* **1995**, 396. (c) Moriconi, E. J.; Murray, J. J. Pyrolysis and photolysis of 1-methyl-3-diazooxindole. Base decomposition of isatin 2-tosylhydrazone. *J. Org. Chem.* **1964**, *29*, 3577. (d) Shmidt, M. S.; Perillo, I. A.; Gonzalez, M.; Blanco, M. M. Reaction of isatin with alkylating agents with acidic methylenes. *Tetrahedron Lett.* **2012**, *53*, 2514.
- (23) (a) Netopilova, M.; Houdkova, M.; Urbanova, K.; Rondevoldova, J.; van Damme, P.; Kokoska, L. In vitro antimicrobial combinatory effect of Cinnamomum cassia essential oil with 8-hydroxyquinoline against *Staphylococcus aureus* in liquid and vapour phase. *J. Appl. Microbiol.* **2020**, *129*, 906. (b) Phopin, K.; Ruankham, W.; Prachayasittikul, S.; Prachayasittikul, V.; Tantimongkolwat, T. Insight into the molecular interaction of cloxyquin (5-chloro-8-hydroxyquinoline) with bovine serum albumin: biophysical analysis and computational simulation. *Int. J. Mol. Sci.* **2020**, *21*, 249. (c) Yin, X.; Ma, K.; Wang, Y.; Sun, Y.; Shang, X.-F.; Zhao, Z.; Wang, R.; Chen, Y.; Zhu, J.; Liu, Y.-Q. Design, Synthesis and Antifungal Evaluation of 8-hydroxyquinoline Metal Complexes against Phytopathogenic Fungi. *J. Agric. Food Chem.* **2020**, *68*, 11096.
- (24) Bezerra, D. P.; Militao, G. C. G.; de Moraes, M. C.; de Sousa, D. P. The dual antioxidant/prooxidant effect of eugenol and its action in cancer development and treatment. *Nutrients* **2017**, *9*, 1367.

(25) (a) Lei, Y.; Fu, P.; Jun, X.; Cheng, P. Pharmacological Properties of Geraniol - A Review. *Planta Med.* **2019**, *85*, 48. (b) Lira, M. H. P. d.; Andrade Junior, F. P. d.; Moraes, G. F. Q.; Macena, G. d. S.; Pereira, F. d. O.; Lima, I. O. Antimicrobial activity of geraniol: an integrative review. *J. Essent. Oil Res.* **2020**, *32*, 187. (c) Maczka, W.; Winska, K.; Grabarczyk, M. One hundred faces of geraniol. *Molecules* **2020**, *25*, 3303.

(26) (a) Vlot, A. H. C.; Aniceto, N.; Menden, M. P.; Ulrich-Merzenich, G.; Bender, A. Applying synergy metrics to combination screening data: agreements, disagreements and pitfalls. *Drug Discovery Today* **2019**, *24*, 2286. (b) Bhatia, K.; Bhumika; Das, A. Combinatorial drug therapy in cancer - New insights. *Life Sci.* **2020**, *258*, 118134.

(27) (a) Reddy Panyam, P. K.; Ugale, B.; Gandhi, T. Palladium(II)/N-Heterocyclic Carbene Catalyzed One-Pot Sequential α -Arylation/Alkylation: Access to 3,3-Disubstituted Oxindoles. *J. Org. Chem.* **2018**, *83*, 7622. (b) Zhang, A.; Yu, M.; Lan, T.; Liu, Z.; Mao, Z. Novel synthesis of 4- or 6-substituted indirubin derivatives. *Synth. Commun.* **2010**, *40*, 3125.