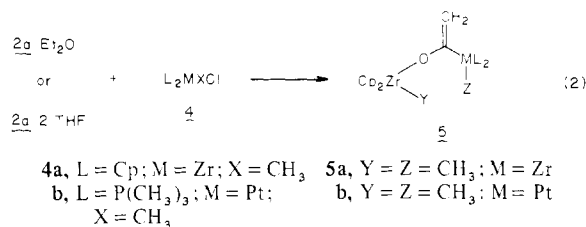


Metallaenolate (ketene) anion **2a** has been found to be of general utility in the preparation of new homo- and heteronuclear bimetallic ketene complexes of interest as models of intermediates implicated in carbon-carbon bond formation in surface-catalyzed carbon monoxide reductions.¹⁵ Reaction of **2a**·Et₂O with zirconocene halide **4a** (eq 2) proceeds rapidly upon dissolution in

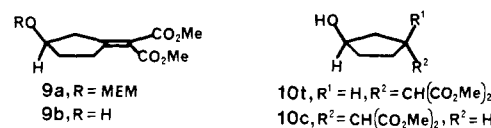


The metallaenolate anions **2** are versatile reagents in organometallic synthesis, particularly for formation of binuclear complexes of relevance to carbon monoxide reduction systems. The reactivity of these new ketene species, including use of complexes such as **2** and **5** in stereospecific organic transformations, is presently being explored.

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Revised Manuscript Received February 3, 1984

Considering the obvious synthetic utility of stereodirected Michael additions, there are remarkably few examples of such processes. To explore the efficacy of various homoallylic substituents as stereodirecting groups, MEM ether **9a** was reduced

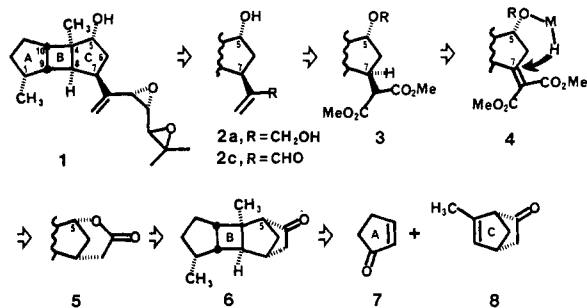
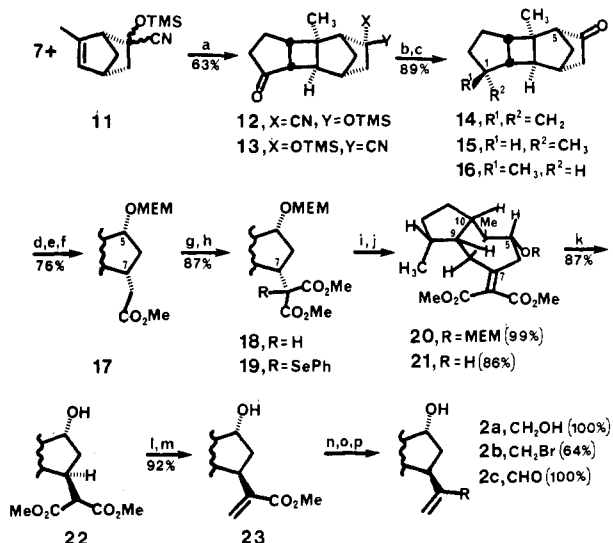


with NaBH₄ in ethanol followed by removal of the MEM protecting group.⁵ A 1:5 mixture of the trans and cis hydroxy malonic esters **10t** and **10c** was obtained. Thus the MEM ether group functions as a bulky steric hindrance to syn approach of the hydride. This contrasts with the syn stereodirecting effect of an allylic MEM ether group which served as a chelating ligand during “heteroconjugate addition” of MeLi.⁶ Most significantly, the stereochemical outcome was reversed by first removing the MEM protecting group. Treatment of **9b** with NaBH₄ in ethanol produced a 1.0:0.67 mixture of **10t** and **10c**. Further improvement in the product ratio to 1.0:0.37 was achieved by using THF as solvent. The solvent effect can be understood in terms of activation

- (13) For **2a**: ^1H NMR ($\text{THF}-d_6$) δ -0.68 (s, 3 H), 3.64 (d, $J = 2$ Hz, 1 H), 4.55 (d, $J = 2$ Hz, 1 H), 5.43 (s, 10 H). **2b**: ^1H NMR ($\text{THF}-d_6$) δ -0.68 (s, 3 H), 1.82 (d, $J = 6$ Hz), 5.07 (q, 1 H, $J = 6$ Hz), 5.45 (s, 10 H). **2c**: ^1H NMR ($\text{THF}-d_6$) δ -0.77 (s, 3 H), 1.68 (s, 3 H), 1.83 (s, 3 H), 5.34 (s, 10 H). By modification of reaction conditions it is possible to generate a minor isomer of **2b**.¹⁴
- (14) Ho, S. C. H.; Grubbs, R. H., unpublished results.
- (15) (a) Blyholder, G.; Emmet, P. H. *J. Phys. Chem.* **1960**, *64*, 470 and references therein. (b) Wolczanski, P. T.; Bercaw, J. E. *Acc. Chem. Res.* **1980**, *13*, 121.
- (16) For **5a**: ^1H NMR (C_6D_6) δ -0.17 (s, 3 H), 0.43 (s, 3 H), 4.23 (s, 1 H), 4.61 (s, 1 H), 5.65 (s, 10 H), 5.82 (s, 10 H); ^{13}C NMR (C_6D_6) δ 18.8 (q, $J_{\text{CH}} = 117$ Hz), 33.0 (q, $J_{\text{CH}} = 119$ Hz), 92.7 (dd, $J_{\text{CH}} = 148$, 159 Hz), 107.1 (dm, $J_{\text{CH}} = 172$ Hz), 113.1 (dm, $J_{\text{CH}} = 172$ Hz), 208.9 (pseudotriplet, $J_{\text{CH}} = 9$ Hz); IR (KBr) 1538, 1594 cm^{-1} ($\nu_{\text{C}=\text{C}}$). Anal. $\text{C}_{24}\text{H}_{28}\text{OZr}_2$ (C, H).
- (17) The inequivalence of the two zirconium centers of **5a** in the ^1H NMR at room temperature stands in contrast with the formaldehyde complex ($\text{Cp}_2\text{ZrCl})_2(\mu\text{-OCH}_2)$ (Gambarotta, S.; Floriani, C.; Chiesi-Villa, A.; Guastini, C. *J. Am. Chem. Soc.* **1983**, *105*, 1690) and related complexes (Erker, G.; Kropp, K. *Chem. Ber.* **1982**, *115*, 2437). This might be due to the sp^2 hybridization at the oxygen-bound carbon, which inhibits formal dative Zr-O interaction. Examination at high temperature is not possible because **5a** undergoes bimolecular decomposition to $(\text{Cp}_2\text{Zr}(\text{OCCCH}_2))_n$ (identified by comparison with an authentic sample¹⁴) and $\text{Cp}_2\text{Zr}(\text{CH}_3)_2$ ($k = 1.1 \times 10^{-2} \text{ L mol}^{-1} \text{ s}^{-1}$, 52 $^\circ\text{C}$).
- (18) Binuclear zirconium ketenes with $\text{Y} = \text{OCH}_3$, $\text{Z} = \text{CH}_3$ and $\text{Y} = \text{CH}_3$, $\text{Z} = \text{Cl}$ have also been prepared.
- (19) PtL_2XCl ($\text{L} = \text{P}(\text{CH}_3)_3$, Ph , PCH_3Ph_2 ; $\text{X} = \text{CH}_3$, Cl) react to give similar ketene-bridge complexes.
- (20) For **5b**: ^1H NMR (C_6D_6) δ 6.08 (s, 10 H), 5.06 (ddd, $J_{\text{HH}} = 2$, $J_{\text{HP}} = 13$, 3, $J_{\text{HPi}} = 90$ Hz, 1 H), 4.00 (ddd, $J_{\text{HH}} = 2$, $J_{\text{HP}} = 3$, $J_{\text{HPi}} = 32$ Hz, 1 H), 1.20 (d, $J_{\text{HP}} = 8.5$, $J_{\text{HPi}} = 21.7$ Hz, 9 H), 1.04 (dd, $J_{\text{HP}} = 9.6$, 17.5, $J_{\text{HPi}} = 69.6$ Hz, 3 H), 0.93 (d, $J_{\text{HP}} = 7.8$, $J_{\text{HPi}} = 19.4$ Hz, 9 H), 0.39 (s, 3 H); ^{13}C NMR (C_6D_6) δ 202.1, 152.9, 110.3, 36.4, 17.3, 15.9, 13.9; ^{31}P NMR (C_6D_6) δ -29.3 (d, $J_{\text{PP}} = 12.1$, $J_{\text{PPi}} = 1354$ Hz), -25.0 (d, $J_{\text{PP}} = 12.2$, $J_{\text{PPi}} = 1578$ Hz).

- (1) Gerwick, W. H.; Fenical, W.; Sultanbawa, M. U. S. *J. Org. Chem.* **1981**, *46*, 2233.
- (2) (a) Spatol inhibits cell division in human T242 Melanoma and 224C astrocytoma neoplastic cell lines: Gerwick, W. H.; Fenical, W.; Van Engen, D.; Clardy, J. *J. Am. Chem. Soc.* **1980**, *102*, 7991. (b) Spatol's antimitotic activity apparently results from inhibition of microtubule assembly: Jacobs, R. S.; White, S.; Wilson, L. *Fed. Proc., Fed. Am. Soc. Exp. Biol.* **1981**, *40*, 26.
- (3) Hara, M.; Odaira, Y.; Tsutsumi, S. *Tetrahedron* **1966**, *33*, 95.
- (4) White, J. D.; Gupta, D. N. *J. Am. Chem. Soc.* **1968**, *90*, 6171.
- (5) Corey, E. J.; Gras, J.-L.; Ulrich, P. *Tetrahedron Lett.* **1976**, 809.
- (6) Isohe, M.; Kitamura, M.; Goto, T. *Tetrahedron Lett.* **1979**, 3465.

Scheme I

Scheme II^a

^a Reagents and conditions: (a) $h\nu$ /uranium glass filter/hexane; (b) Ph_3PCH_2 (2.3 equiv)/THF then add H_2O /20 °C, 3 h; (c) H_2 /Pt₂O; (d) $\text{CH}_3\text{CO}_3\text{H}/\text{CH}_3\text{COOH}$; (e) $\text{KOH}/\text{H}_2\text{O}/\text{MeOH}$ then HCl then CH_2N_2 ; (f) $\text{MEMCl}/i\text{-Pr}_2\text{NEt}/\text{CH}_2\text{Cl}_2$; (g) LDA then CO_2 then HCl then CH_2N_2 ; (h) NaH/THF then add PhSeBr ; (i) $\text{H}_2\text{O}_2/\text{CH}_2\text{Cl}_2$; (j) $\text{TiCl}_4/\text{CH}_2\text{Cl}_2$; (k) NaBH_4/THF ; (l) KOH (1.0 equiv)/ $\text{EtOH}/\text{H}_2\text{O}$ then HCl ; (m) $\text{CH}_2\text{O}/\text{H}_2\text{O}/\text{Et}_3\text{NH}/\text{NaOAc}/\text{HOAc}$; (n) $i\text{-Bu}_2\text{AlH}/\text{toluene}$; (o) $\text{Ph}_3\text{P}/\text{CBR}_4/\text{CH}_3\text{CN}$; (p) $\text{MnO}_2/\text{CH}_2\text{Cl}_2$.

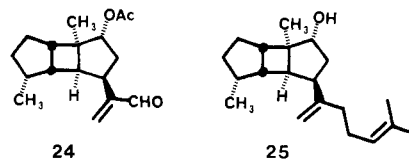
of borohydrides by *B*-alkoxy groups.⁷ In ethanol, ethoxyborohydrides react intermolecularly, whereas in THF, pseudointramolecular reaction is favored for alkoxyborohydride derivatives of the homoallylic hydroxyl substituent.⁸

Our synthesis of the key intermediates **2a** and **2c** is outlined in Scheme II.⁹ Trimethylsilyl cyanohydrin **11** was obtained quantitatively from **8**¹⁰ by reaction with trimethylsilancarbonitrile.¹¹ Although **11** was an epimeric mixture, this choice for masking the carbonyl proved remarkably fortunate (vide infra). Photocycloaddition of **7** with **11** in hexane solution produced an awesome mixture of products. Serendipitously, the required epimers **12** and **13** were readily isolated by crystallization. Thus, **13** crystallized from the photoreaction mixture together with dimers of **7** from which it was readily separated by trituration with boiling hot hexane leaving behind pure dimer. Pure **13** (mp

109–111 °C)¹² was then obtained in 51% yield based on **11** by passage of the partially purified material through a column of silica gel with ethyl acetate–hexane. Column chromatography of the hexane-soluble photoproduct afforded a fraction from which nearly pure **12** crystallized together with a little **13**. This mixture is suitable for Wittig olefination (vide infra) so that the combined isolated yield of **12** plus **13** exceeds 60%. The epimeric relationship between **12** and **13** was proven by production of the same methylenide ketone **14** upon reaction with methylenetriphenylphosphorane followed by hydrolysis. The **13** → **14** conversion was performed as a one-pot procedure, which afforded pure **14** (mp 52–53 °C) in 89% overall yield. Thus, the cyanohydrin silyl ether is sufficiently robust to survive UV irradiation and Wittig olefination, but is readily converted to a carbonyl group by aqueous base.

Catalytic hydrogenation of **14** was expected to deliver hydrogen preferentially from the less congested exo face of the C=C bond to give the *endo*-methyl epimer **15**. With PtO₂ as catalyst precursor, **15** was favored over the *exo*-methyl epimer **16** by about 9:1. The seemingly tedious separation of **15** and **16** is in fact trivial. Thus, pure **15** (mp 53–55 °C) was readily isolated from the mixture by crystallization from pentane at –78 °C. Baeyer–Villiger oxidation of **15** then introduced the C-5 oxygen substituent stereospecifically. The carbon skeleton was completed by carboxylation of ester **17**. Inversion of the configuration at C-7 in diester **18** was then initiated by selenation to give **19**, oxidative deselenation of which afforded the alkylidenemalonate **20**. Removal of the MEM protecting group provided **21**.

The crucial stereocontrolled Michael reduction was then examined. Reduction of the MEM ether **20** with NaBH₄ in ethanol followed by removal of the MEM protecting group afforded a 1:2 mixture of the desired **22** and its C-7 epimer, respectively. As with the model **9**, the MEM ether substituent in **20** sterically hinders the desired syn delivery of hydride. In contrast with **9**, anti delivery of hydride is also hindered for **20** by the hydrogens at positions 9 and 10 resulting in nonstereoselective reduction. Most gratifyingly, the combination of this steric hindrance to anti hydride delivery with the syn stereodirecting influence of a homoallylic hydroxyl substituent results in highly stereoselective reduction of hydroxyalkylidenemalonate **21**. Thus, **21** afforded **22** with no trace of the C-7 epimer upon treatment with NaBH₄ in THF. Selective monosaponification of **22** was readily achieved, and Mannich condensation with subsequent decarboxylative elimination generated α,β -unsaturated ester **23** in a one-pot reaction from the monoacid.¹³ Reduction of ester **23** with diisobutylaluminum hydride gives the target allylic alcohol **2a** (mp 97–98 °C) quantitatively. The diol **2a** was selectively converted to a monobromide **2b** (mp 72–73 °C) upon reaction with triphenylphosphine and carbon tetrabromide.¹⁴ Selective oxidation of **2a** with MnO₂ provided the target aldehyde **2c** (mp 89–91 °C). For comparison with a degradation product from spatol, **2c** was acetylated. The ¹H NMR spectrum of racemic acetate **24** (mp



68–71 °C) is identical with that of (+)-**24** derived from spatol.¹ The first total synthesis of (±)-spata-13,17-diene-5-ol (**25**) was completed by copper(I) iodide catalyzed coupling of prenylmagnesium chloride¹⁵ with the allylic bromide **2b**. The ¹H NMR

(7) (a) Brown, H. C.; Mead, E. J.; Shoaf, C. J. *J. Am. Chem. Soc.* **1956**, *78*, 3616. (b) Rickborn, B.; Wuesthoff, M. T. *Ibid.* **1970**, *92*, 6894.

(8) Analogous stereodirecting reactions involving metal hydride derivatives of allylic alcohols were postulated previously: (a) Isobe, M.; Iio, H.; Kawai, T.; Goto, T. *J. Am. Chem. Soc.* **1978**, *100*, 1940. (b) Lansbury, P. T.; Vacca, J. P. *Tetrahedron Lett.* **1982**, *23*, 2623.

(9) All new compounds were thoroughly characterized spectroscopically and by elemental analysis or measurement of the exact mass of the parent ion in the high-resolution mass spectrum.

(10) Brown, H. C.; Peters, E. N.; Ravindranathan, J. *J. Am. Chem. Soc.* **1975**, *97*, 7449.

(11) (a) Evans, D. A.; Truesdale, L. K.; Carroll, G. L. *J. Chem. Soc., Chem. Commun.* **1973**, 55–56; (b) Evans, D. A.; Truesdale, L. K. *Tetrahedron Lett.* **1973**, 4929–4932.

(12) Evidence for the structure assigned to **13** is provided by extensive 200 MHz ¹H and 50 MHz ¹³C–¹H two-dimensional J spectroscopy and ¹³C–¹H two-dimensional shift correlation spectroscopy: Rinaldi, P. L.; Salomon, R. G. *J. Org. Chem.* **1983**, *48*, 3182.

(13) Parker, W. L.; Johnson, F. J. *J. Org. Chem.* **1973**, *38*, 2489.

(14) Nasipuri, D.; Raychaudhuri, S. R. *J. Chem. Soc., Perkin Trans. I* **1975**, 262.

(15) (a) Derguini-Boumechal, F.; Lorne, R.; Linstrumelle, G. *Tetrahedron Lett.* **1977**, 1181. (b) Kwart, H.; Miller, R. K. *J. Am. Chem. Soc.* **1954**, *76*, 5403.

spectrum of racemic **25** is identical with that of (+)-**25** isolated from *Stoechospermum marginatum*.¹

Acknowledgment. This research was assisted financially by Grant CHE8205122 from the National Science Foundation and sabbatical year support of R.G.S. by Case Western Reserve University for which we are grateful. We thank Professor W. Fenical for ¹H NMR spectra of authentic (+)-**24** and (+)-**25**.

Biosynthesis of Cationomycin: Direct and Indirect Incorporation of [¹³C]Acetate and Application of Homoscalar Correlated 2-D ¹³C NMR and Double Quantum Coherence

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Received October 31, 1983

Cationomycin is a polyether ionophore antibiotic produced by a rare actinomycete, *Actinomadura azurea*.^{1,2} It is structurally unique, having an aromatic acyl side chain.³ It binds selectively monovalent cations and is under development as a controlling agent for chicken coccidiosis because of its remarkable activity and relative low toxicity.⁴ As part of the research directed toward chemical and biological modification of this interesting molecule, we report herein the biosynthesis of cationomycin, including the unambiguous assignment of the ¹³C NMR of cationomycin labeled with [1,2-¹³C]acetate by double quantum coherence⁵ and homoscalar correlated 2-D ¹³C NMR (COSY),⁶ and a reasonable explanation for randomization of the [2-¹³C]acetate.

An assignment of ¹³C NMR of cationomycin⁷ was based on that of structurally related laidlomycin,⁸ INEPT ¹³C NMR analysis,⁹ and calculation with substituent parameters.¹⁰ [1-¹³C]Acetate, [1-¹³C]propionate, [3-¹³C]propionate, and [methyl-¹³C]-L-methionine were incorporated as expected.¹¹ However,

(1) Nakamura, G.; Kobayashi, K.; Sakurai, T.; Isono, K. *J. Antibiot.* **1981**, *34*, 1513.

(2) Nakamura, G.; Isono, K. *J. Antibiot.* **1983**, *36*, 1468.

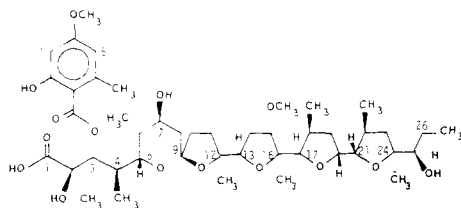
(3) Sakurai, T.; Kobayashi, K.; Nakamura, G.; Isono, K. *Acta Crystallogr., Sect. B* **1982**, *B38*, 2471.

(4) Nakamura, G.; Kobayashi, K.; Sakurai, T.; Isono, K. *Antimicrob. Agents Chemother.* **1982**, *22*, no. 170.

(5) (a) Bax, A.; Freeman, R.; Kempell, S. P. *J. Am. Chem. Soc.* **1980**, *102*, 4849. (b) Mackenzie, N. E.; Baxter, R. L.; Scott, A. I.; Fagners, P. E. *J. Chem. Soc., Chem. Commun.* **1982**, 145. (c) Bacher, A.; LeVan, Q.; Bühler, M. *J. Am. Chem. Soc.* **1982**, *104*, 3754.

(6) Though 2-D INADEQUATE experiments have generally been applied to assignment of double-labeled compounds,⁵ satisfactory data were obtained by homoscalar correlated 2-D ¹³C NMR (COSY) experiment in this case.

(7) The numbering was conventionally adopted as follows:



(8) Seto, H.; Otake, N. *Heterocycles* **1982**, *17*, 555.

(9) (a) Morris, G. A.; Freeman, R. *J. Am. Chem. Soc.* **1979**, *101*, 760. (b) Doddrell, D. M.; Pegg, D. T. *J. Am. Chem. Soc.* **1980**, *102*, 6388. INEPT ¹³C NMR spectra were obtained at 25 MHz using Jeol FX 100.

(10) Wehrli, F. W.; Wirthlin, T. "Interpretation of Carbon-13 NMR Spectra"; Heyden: London, 1976.

(11) The experiment was done by feeding ¹³C-labeled compounds (100–400 mg, 90% enriched) in two portions at 36 and 48 h after inoculation to a shaking culture of *A. azurea* in an organic medium (350 mL). After a total 144-h fermentation, cationomycin was isolated as described before,¹ average yield ca. 20 mg.

(12) Bax, A.; Freeman, R. *J. Magn. Reson.* **1981**, *42*, 164.

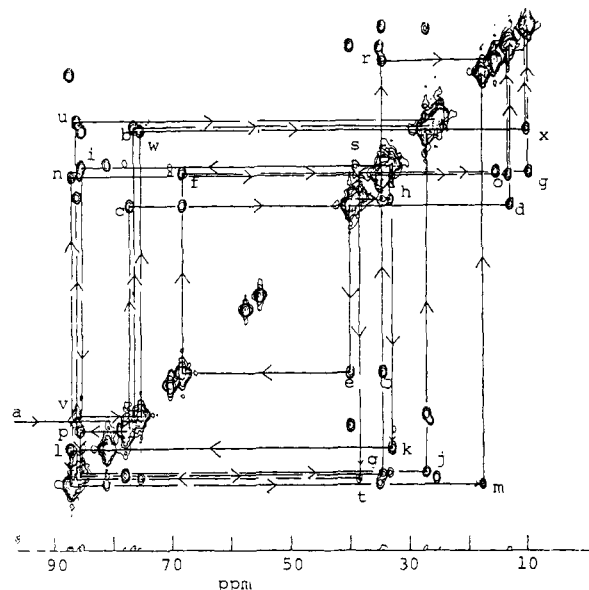
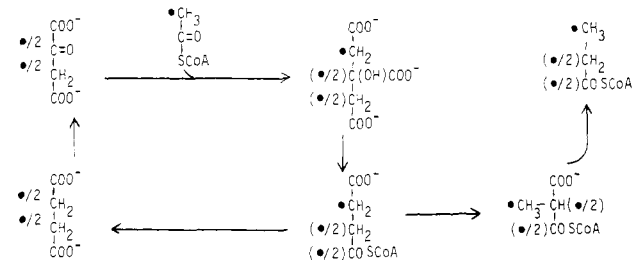


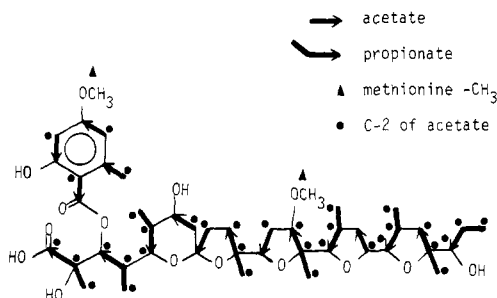
Figure 1. Homoscalar correlated 2-D ¹³C NMR of cationomycin labeled with [2-¹³C]acetate. The spectrum was obtained by COSY sequence¹² on ¹³C nucleus with ¹H decoupling through the experiment at 100 MHz using Jeol GX 400 (acquisition time ca. 40 h, dimension of matrix 256 × 1024, dimension of transformation 512 × 1024, amount of the compound used ca. 20 mg). (a) Correlation of C-1 with C-2, (b) C-2 with 2-Me, (c) C-3 with C-4, (d) C-4 with 4-Me, (e) C-4 with C-5, (f) C-5 with C-6, (g) C-6 with 6-Me, (h) C-10 with C-11, (i) C-11 with C-12, (j) C-12 with 12-Me, (k) C-14 with C-15, (l) C-15 with C-16, (m) C-16 with 16-Me, (n) C-17 with C-18, (o) C-18 with 18-Me, (p) C-20 with C-21, (q) C-21 with C-22, (r) C-22 with 22-Me, (s) C-22 with C-23, (t) C-23 with C-24, (u) C-24 with 24-Me, (v) C-24 with C-25, (w) C-25 with C-26, (x) C-26 with C-27.

Scheme I. Pathway for Propionate from [2-¹³C]Acetate through the Krebs Cycle^a



^a Parentheses show the labeling pattern for the second cycle.

Scheme II. Biogenesis of Cationomycin



feeding of [2-¹³C]acetate resulted in considerable randomization. In the ¹³C NMR spectrum of cationomycin labeled with [1,2-¹³C]acetate, the application of double quantum coherence and homoscalar 2-D ¹³C NMR revealed eight pairs of ¹³C-¹³C coupling, $J_{1',CO,1'}$ (= 76 Hz), $J_{2',3'}$ (= 70.8 Hz), $J_{4',5'}$ (= 65.8 Hz), $J_{6',6'-Me}$ (= 42.7 Hz), $J_{7,8}$ (= 37.8 Hz), $J_{9,10}$ (= 41.5 Hz), $J_{13,14}$ (= 36.6 Hz), and $J_{19,20}$ (= 36.6 Hz).

In the case of [2-¹³C]acetate, the carbons that should be derived from C-1, C-2, and C-3 of propionate were also enriched. Homoscalar correlated 2-D ¹³C NMR and double quantum coherence